Published online 2023 January 31.

**Research Article** 

# Modulatory Effect of Neurotrophic Factors on the TRPV1 Expression: Possible Mechanisms Involved in the Antiepileptic Effect of Exercise

Azam Navazesh 💿<sup>1, 2</sup>, Homa Rasoolijazi 💿<sup>2</sup>, Ghazal Rahmani 💿<sup>3</sup>, Saad Bavi 💿<sup>2</sup>, Gelareh Vahabzadeh 🔟 <sup>1, 4</sup>, Mansoureh Soleimani 🔟 <sup>5, 2</sup> and Fariba Karimzadeh 🔟 <sup>5, \*</sup>

<sup>1</sup>Razi Drug Research Center, Iran University of Medical Sciences, Tehran, Iran <sup>2</sup>Department of Anatomy, School of Medicine, Iran University of Medical Sciences, Tehran, Iran

<sup>3</sup>Department of Physiology, Medical School, Iran University of Medical Sciences, Tehran, Iran

<sup>4</sup>Department of Pharmacology, School of Medicine, Iran University of Medical Sciences, Tehran, Iran

<sup>5</sup>Cellular and Molecular Research Center, Iran University of Medical Sciences, Tehran, Iran

Corresponding author: Cellular and Molecular Research Center, Iran University of Medical Sciences, Tehran, Iran. Email: fariba\_karimzade@yahoo.com

Received 2023 January 01; Revised 2023 January 11; Accepted 2023 January 15.

# Abstract

Background: Epilepsy is one of the most important diseases of the central nervous system, for which has no definitive treatment. Neurotrophic factors increase the survival of nerve cells and improve the treatment of neurological diseases. Identifying factors that affect the increase of neurotrophins in the brain is an important goal for brain health and function.

**Objectives:** This study aimed to investigate the effectiveness of exercise on neurotrophic factors by influencing the expression of vanilloid receptor type 1 (TRPV1).

Methods: Convulsions were induced by injecting pentylenetetrazol (PTZ; 35 mg/kg) five hours after exercise. Animals were divided into five groups: sham (Sham), seizure (PTZ), exercise (EX), exercise with seizure induction (EX+PTZ), and exercise before seizure induction (EX-PTZ). The exercise was 30 minutes of forced running on a treadmill, five days a week for four weeks.

Results: The average percentage of NGF cells in the exercise groups (EX), exercise with seizure induction (EX+PTZ), and exercise before seizure induction (EX-PTZ), and GDNF in the exercise group with seizure induction (EX+PTZ) had a significant increase compared to the seizure group (PTZ). Also, TRPV1 activity in exercise groups (EX), exercise with seizure induction (EX+PTZ), and exercise before seizure induction (EX-PTZ) showed a significant increase compared to the seizure group (PTZ).

Conclusions: Our findings suggested the possible antiepileptic and antiepileptogenesis effects of exercise through activation of neurotrophic factors and TRPV1 modulation.

Keywords: Epilepsy, Exercise, Seizure, Hippocampus, NGF, GDNF, TRPV1

#### 1. Background

Epilepsy is one of the brain disorders characterized by frequent and sudden seizures and affects half to one percent of the world population (1). In epilepsy, the excitability of neurons is permanently increased in the central nervous system (CNS) following an imbalance between the excitatory and inhibitory systems (2), thus causing functional alteration of ion channels and neurotransmitter receptors and increasing the permeability of the blood-brain barrier and expression changes of the glutamate receptor subunits (3, 4). In more than 70% of patients with epilepsy, seizures are suppressed by antiepileptic drugs (5). Some major side effects of antiepileptic drugs, such as neurotoxicity and hepatotoxicity, have attracted the attention of researchers to find non-medicinal alternatives (6,7).

New studies show that sports interventions can improve cognitive functions, increase brain activity, and re-

duce psychological symptoms (8, 9). Both high- and moderate-intensity aerobic exercises increase the level of neurotrophins and neurotransmitters and improve psycho-cognitive performance at all ages (10, 11). Also, exercise can reduce the severity of seizures and delay their onset (2). One of the mechanisms through which exercise leads to the improvement of brain function is the potentiation of neurotrophic factors (NTFs). Neurotrophic factors are small proteins that have key roles in the regeneration of nerve cells and neural survival (12). Nerve growth factor (NGF) is essential for the growth and maintenance of the phenotype of neurons in the peripheral nervous system and the functional integrity of cholinergic neurons in the CNS (13). Also, glial cell-derived growth factor (GDNF) is critical for neuronal survival and neurogenesis (14).

Previous studies have reported exercise stimulates the regeneration of nerve cells by increasing the expression of

Copyright © 2023, Author(s). This is an open-access article distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 International License (http://creativecommons.org/licenses/by-nc/4.0/) which permits copy and redistribute the material just in noncommercial usages, provided the original work is properly cited.

NGF (15). Exercises also lead to the production of GDNF factor produced by the glial cells of the substantia nigra, where dopaminergic neurons are located. Therefore, exercise improves plasticity in dopaminergic neurons by increasing the production of GDNF (16). TRPV1 is one of the transient potential receptors (TRP) that belongs to the family of non-specific voltage-gated and ligand-gated ion channels and is more permeable to calcium than other cations. The expression level of TRPV1 in the CNS is significantly lower than in the peripheral system, but this small amount plays an important and significant effect on the central (17, 18). There is evidence that TRPV1 receptors in the brain are involved in many fundamental functions, including synaptic transmission and synaptic plasticity. TRPV1 expression is variable and depends on the presence or absence of growth factors. Considerable studies on TRPV1 receptors indicate that these receptors can be considered specific targets for antiepileptic compounds (19). Evidence suggests that NTFs influence seizure-related processes through the regulation of TRPV1 receptors (20).

Even though the nervous system is one of the parts that has always been of interest in adapting to exercises. There has not been enough research regarding clarifying the influence and possible compatibility of TRPV1 with sports training. But the role of this receptor is prominent in the goals related to the health of sports.

# 2. Objectives

This study aimed to investigate the effectiveness of exercise on neurotrophic factors by influencing the expression of vanilloid receptor type 1 (TRPV1). Considering the role of moderate physical exercise on seizure alleviation and also its role in the activation of neurotrophic factors, this study aimed to investigate whether TRPV1 modulation through neurotrophic factors activation in the hippocampus may be one of the underlying mechanisms for exercise effectiveness in epilepsy or not.

# 3. Methods

#### 3.1. Animals

Thirty adult male Wistar-Bauzen rats weighing 250 to 300 g were placed in a controlled environment with a lightdark cycle at a temperature of  $22 \pm 1^{\circ}$ C for one week before the beginning of the experiment. All experiments were performed according to the protocol approved by the ethical principles of working with laboratory animals of Iran University of Medical Sciences (IR.IUMS.REC.1399.023). Animals were randomly divided into five groups (n = 6). (1) Seizure group: Seizure was induced by intraperitoneal (IP) injection of Pentylenetetrazol (PTZ) every other day for four weeks. (2) Sham group: 0.9% normal saline was injected according to the PTZ protocol. (3) Exercise group (EX): The animals in this group had to run on a treadmill for 30 minutes five days a week in weeks one to four. (4) Exercise and seizure group (EX+PTZ): Seizure and exercise were performed for four weeks. Animals were forced to run on a treadmill five days a week, and seizures were performed five hours after exercise according to the seizure group protocol. (5) Exercise before seizure induction group (EX-PTZ): Animals in this group were forced to do exercise for four weeks like the exercise group (without PTZ injection). In weeks five to eight, seizure induction and exercise were done according to the protocol of the exercise and seizure group (EX + PTZ).

#### 3.2. Treadmill Exercise Protocol

Rats in EX, EX+PTZ, and EX-PTZ groups were forced to run on a treadmill for 30 minutes, five days a week. The speed of the treadmill was gradually increased at a zerodegree incline (5 m/min every five min, up to 25 m/min)(2). The first and last three minutes were considered warm-up and cool-down, respectively.

#### 3.3. Immunohistofluorescence

After the last injection and exercise, the animals were anesthetized with 350 mg/kg chloroform (Sigma-Aldrich). After complete anesthesia, heart perfusion was performed first with 250 mL of saline and then with 400 mL of paraformaldehyde (PFA) solution. After the completion of perfusion, the head of the animal was detached, and the brain was removed from the skull and placed in 4% paraformaldehyde at 4°C for one week (21).

Tissue samples were embedded in paraffin blocks and then coronally cut with a thickness of 8  $\mu$ m by microtome (21). Three parts of each brain were selected and deparaffinized at a distance of 30  $\mu$ m. Then they were dehydrated by xylene and alcohol and washed three times with phosphate-buffered saline (PBS). Then the slices were washed three times in Tris wash buffer (Three minutes each time) and with a secondary antibody (Goat polyclonal Anti Rabbit IgG) conjugated with DyLight 484 (Cat.no: GTX213110-04) for one hour at room temperature. The chamber was incubated and washed three more times in Tris wash buffer (for three minutes each). Samples were photographed for statistical analysis using a digital camera (Nikon, objective lens  $40 \times$ ) connected to a fluorescent microscope (100 Ts Nikon Florescent).

### 3.4. Statistical Analysis

All the values were presented using Mean  $\pm$  standard error of mean (S.E.M). The data were analyzed by SPSS software. One-way ANONA analysis was used to compare the experimental groups and followed by post-hoc Tukey's test for between groups comparison. The significance level of the tests was considered P< 0.05.

# 4. Results

4.1. The Effect of Treadmill Exercise on the Distribution of NGF, GDNF, and TRPV1 Receptor

# 4.1.1. Distribution of NGF in Hippocampal Regions

The mean percentage of NGF cells in the CA1 region in the PTZ group  $(0.4 \pm 0.02)$  was significantly decreased compared to the Sham group  $(0.7 \pm 0.03)$  (P < 0.01). There was no significant difference compared to the Sham group in EX+PTZ ( $0.6 \pm 0.03$ ), EX ( $0.7 \pm 0.02$ ), and EX-PTZ ( $0.7 \pm 0.02$ ) groups, (respectively, P = 0.4 and P = 0.9). Also, in the EX+PTZ group, EX and EX-PTZ showed a significant increase compared to the PTZ group (P < 0.01) (Figure 1B).

The mean percentage of NGF cells in the CA3 region in the PTZ group was  $(0.1 \pm 0.01)$  compared to the Sham group ( $0.3 \pm 0.03$ ), which has decreased significantly (P < 0.05). However, in EX+PTZ ( $0.3 \pm 0.0$ ), EX ( $0.3 \pm 0.06$ ), and EX-PTZ  $(0.3 \pm 0.06)$  groups, there was no significant difference compared to the Sham group (P = 0.6 and P = 1, respectively). Also, in the EX+PTZ, EX, and EX-PTZ groups, there was no significant difference compared to the PTZ group (P = 0.3 and P = 0.06, respectively). The distribution of NGF in the CA1 and CA3 regions in the PTZ group was significantly reduced compared to the Sham group. The distribution of NGF in the CA1 region showed a significant increase in the EX+PTZ, EX, and EX-PTZ groups compared to the PTZ group. In the bar chart, a and aa indicate P< 0.05 and P< 0.01 compared to the Sham group, respectively and bb indicates P < 0.01 compared to the PTZ group (Figure 1).

#### 4.1.2. Distribution of GDNF in Hippocampal Regions

The mean  $\pm$  standard deviation (SD) percentage of GDNF cells in the CA1 region in the PTZ group (0.2  $\pm$  0.03) showed a significant decrease compared to the Sham group (0.5  $\pm$  0.06) (P < 0.01). But in the EX+PTZ (0.5  $\pm$  0.03), EX (0.4  $\pm$  0.0), and EX-PTZ (0.4  $\pm$  0.0) groups, it did not show any significant difference compared to the Sham group (P = 1 and P = 0.1, respectively). It also showed a significant increase in the EX+PTZ group compared to the PTZ group (P < 0.01). However, there was no significant difference in EX and EX-PTZ groups compared to the PTZ group (P = 0.2) (Figure 2B).

The mean  $\pm$  SD percentage of GDNF cells in the CA3 region in the PTZ group (0.8  $\pm$  0.03) compared to the Sham group (0.9  $\pm$  0.0) did not show a significant difference (P = 0.3). Also, in the EX+PTZ (0.7  $\pm$  0.0), EX (0.7  $\pm$  0.03), and EX-PTZ (0.7  $\pm$  0.03) groups, it showed a significant decrease compared to the Sham group (P < 0.01). In addition, it showed a significant decrease in the EX+PTZ group compared to the PTZ group (P < 0.05). However, there was no significant difference in EX and EX-PTZ groups compared to the PTZ group (P = 0.09).

The distribution of GDNF in the CA1 region In the PTZ group and the CA3 region in the EX+PTZ, EX, and EX-PTZ

groups was significantly reduced compared to the Sham group. It showed a significant increase and decrease in the CA1 and CA3 regions in the EX+PTZ group compared to the PTZ group, respectively. In the bar chart, aa indicates P < 0.01 compared to the Sham group, and b and bb indicate P < 0.05 and P < 0.01 compared to the PTZ group, respectively.

#### 4.1.3. Distribution of TRPV1 Receptor in Hippocampal Regions

The mean  $\pm$  SD percentage of TRPV1 receptor distribution in the CA1 region in the PTZ (0.1  $\pm$  0.01), EX (0.3  $\pm$  0.0), and EX-PTZ (0.3  $\pm$  0.0) groups did not show any significant difference compared to the Sham group (0.2  $\pm$  0.03) (P = 0.1 and P = 0.4, respectively). But in the EX+PTZ group (0.4  $\pm$  0.02), it showed a significant increase compared to the Sham group (P < 0.01). Also, the mean percentage of TRPV1 receptor distribution in the EX+PTZ, EX, and EX-PTZ groups showed a significant increase compared to the PTZ group (P < 0.001 and P < 0.05, respectively) (Figure 3B).

The mean  $\pm$  SD percentage of TRPV1 receptor distribution in the CA3 region in PTZ (0.3  $\pm$  0.03), EX+PTZ (0.2  $\pm$  0.0), EX (0.2  $\pm$  0.03), and EX-PTZ (0.2  $\pm$  0.03) groups compared to Sham group (0.2  $\pm$  0.03) showed no significant difference (P = 0.1 and P = 0.8, respectively). Also, in the EX+PTZ, EX, and EX-PTZ groups, there was no significant difference compared to the PTZ group (P = 0.06 and P = 0.4, respectively).

#### 4.2. Correlation Between Expression of NGF/GDNF and TRPV1

The relationship between NGF/GDNF and TRPV1 expression in CA1 and CA3 regions is presented (Figure 4). The correlation between NGF and TRPV1 expression in the CA1 region showed that high NGF expression (Figure 4A; r = 0.36, P < 0.05) was associated with high TRPV1 level, but there was no significant correlation between GDNF and TRPV1 expression (Figure 4B; r = 0.32, P = 0.05). The correlation between the expression of NGF/GDNF and TRPV1 in the CA3 region showed that there was no significant correlation between the expression of NGF and TRPV1 (Figure 4C; r = 0.27, P = 0.08) and also between the expression of GDNF and TRPV1 (Figure 4D; r = 0.26, P = 0.08). (A) and B) scatter plots show the correlation between NGF/GDNF and TRPV1 expression in the CA1 region. A strong correlation between NGF and TRPV1 expression was shown in the CA1 region. (C) and (D) scatter plots show the correlation between NGF/GDNF and TRPV1 expression in the CA3 region. There was no significant correlation between the expression of NGF/GDNF and TRPV1 in the CA3 region.

#### 5. Discussion

5.1. Regulating the Neurotrophic Factors by Doing Exercise in Epilepsy

Our results showed a significant increase in the expression of NGF in the CA1 region in the EX, EX+PTZ, and



Figure 1. The distribution of NGF in hippocampal regions. (A) Immunofluorescent microscopic image of NGF cells, (B) Bar chart of the effect of PTZ and exercise on the distribution of NGF.

EX-PTZ groups, as well as an increase in the expression of GDNF in the EX+PTZ group compared to the PTZ group. Seizures are followed by excessive immune responses and imbalances in synaptic transmitters (22). Immune activation by increasing the production of pro-inflammatory cytokines increases the potential for new convulsions (23). After seizure induction, inflammatory mediators released

from glial cells, including interleukin-1beta (IL-1 $\beta$ ), IL-6, IL-10, and tumor necrosis factor-alpha (TNF- $\alpha$ ), increase in the brain, which directly leads to activation of their pathways involved in apoptosis through their receptors in neurons (24). They stimulate convulsions by affecting ion channels, and neurotransmitter receptors and increasing the permeability of the blood-brain barrier (25). On the contrary,



Figure 2. GDNF distribution in hippocampal regions. (A) Immunofluorescent microscopic image of GDNF cells. (B) Bar chart of the effect of PTZ and exercise on GDNF distribution.

long-term and moderate-intensity exercise training (65% VO2 Max) causes a clear decrease in the level of inflammatory cytokines, including IL-1 $\beta$  and IL-6, in the hippocampus and cerebellum (26, 27), and by improving the inflammatory condition and decreasing oxidative stress (28). It increases the expression of neurotrophic factors in the hippocampus of mice and rats (29).

One of the mechanisms of exercise that leads to the improvement of brain function is neurotrophic factors, which play an important role in the survival of neurons that are affected by the processes of neuronal destruction. Neurotrophic factors increase the growth and function of damaged neurons and increase neuronal survival and neurogenesis by preventing nerve cell death (30). Several previous studies have reported that regular aerobic exercise is effective in enhancing neuroprotection by increasing NGF secretion and leading to NGF upregulation (15). Also, increasing NGF secretion has a direct relationship with memory improvement and an inverse relationship with oxidative stress (31). Four weeks of endurance exercise on a tread-



Figure 3. TRPVI distribution in hippocampal regions. (A) Immunofluorescent microscopic image of TRPVI cells. (B) Bar chart of the effect of PTZ and exercise on TRPVI distribution. aa indicates P < 0.01 compared to the Sham group, b and bbb indicate P < 0.05 and P < 0.001 compared to the PTZ group, respectively.

mill has been shown that is related to the reduction of the effects of age-related progressive neurodegenerative diseases such as Parkinson's by increasing striatal NGF levels (32). Studies have shown that moderate-intensity treadmill exercise increases the level of NGF and prevents the apoptosis of nerve cells in nerve-damaged mice, and reduces the death and demyelination of neurons activating P-PL3K (33). Aerobic exercise can play a significant role in improv-

ing learning, increasing GDNF expression, and reducing inflammation caused by glial cells (34). Also, GDNF can have an anti-inflammatory and anti-neurotoxic role. GDNF leads to a decrease in apoptosis through reducing inflammatory factors such as TNF- $\alpha$  and IL-1 $\beta$  and, finally, causes cell survival of neurons and improves memory and learning (3, 34).

It has been shown that the content of GDNF in the



Figure 4. The correlation between protein expression of NGF/GDNF and TRPV1. Scatter plots are shown in the CA1 region (A and B). There is a strong correlation between NGF and TRPV1 expression, but there was no significant correlation between the expression of NGF/GDNF and TRPV1 in the CA3 region (C and D, respectively).

sciatic nerve (35), substantia nigra (36), and striatum (37) increases significantly following 12 and 18 weeks of moderate-intensity exercise. In contrast, two other studies reported that four weeks of low-to-moderate exercise training did not affect the protein content of the striatum and substantia nigra (38) and the GDNF content of the lumbar spine of rats (39). Therefore, increasing the expression of GDNF and improving learning can be influenced by the duration of exercise (3). Previous evidence and our findings show that regular exercise with moderate intensity causes some kind of adaptation in the antioxidant system, can increase resistance to oxidative stress, and prevent excessive excitability of neurons. Also, as a strong factor, it can play a role in cell proliferation of neurotrophic factors and suppression of accelerating factors in the process of neuronal destruction diseases, including inflammation caused by the activity of microglia and inhibition of NMDA receptors (2).

5.2. TRPV1 Receptor Distribution with Exercise Training in Epilepsy

Our results showed that the increase in TRPV1 expression in the EX, EX+PTZ, and EX-PTZ groups was significantly different compared to the PTZ group. It has been found that TRPV1 receptor protein expression increases as a result of moderate-intensity exercise. Sensory nerves adapt to exercise by increasing the expression of the TRPV1 receptor protein (40). There is increasing evidence that exercise increases NGF and neurogenesis (41). Recent reports have shown that NGF is upstream of the TRPV1 channel and increased NGF levels facilitate TRPV1 expression (42). On the other hand, there are also reports that NGF decreases in the DRG of rats with heart failure due to a decrease in muscle volume (43). At the same time, along with the decrease in NGF level, a decrease in TRPV1 levels has also been observed, and the evidence indicates a relationship between these two neural factors (44).

The relationship between delayed muscle pain and TRPV1 and the essential role of this receptor in delayed pain have been reported that usually occurs hours after intense or long-term activities. The results showed that delayed muscle soreness after the prolonged contraction was not increased in mice without TRPV1: however, the level of mRNA related to NGF increased three hours after muscle activity. These findings indicate that NGF upregulation is not impaired in the absence of TRPV1. Also, TRPV1 expression is NGF downstream in delayed muscle pain conditions (45). Due to the prominent role of this receptor on pain, the TRPV1 channel is one of the therapeutic targets for pain relief methods, which is done either by desensitization of the channel with an agonist or by blocking the channel with the use of an antagonist (46). Dysfunction of the synaptic plasticity of TRPV1 receptors has been reported in diseases such as Alzheimer's, Parkinson's, and epilepsy (47). Since the role of sports activities in increasing the expression of NGF has been confirmed in several reports, it can be concluded that the increase in the expression of NGF in cell signaling pathways and calciumdependent pathways is related to the increase in the expression of the TRPV1 receptor, which is a cation dependent receptor. From a functional point of view and according to the available evidence on the relationship between TRPV1 and some sports biomarkers, it can be concluded that the role of this receptor is significant in regulating the factors involved in energy production. The findings of this research show that it is possible to consider TRPV1 as a neural marker in adaptation to exercise.

#### 5.3. Signaling Correlation Between NGF and TRPV1

The results of our study showed a high correlation between NGF and TRPV1 in the CA1 region. Research results have shown that among the neurotrophic factors, NGF plays a more fundamental role in regulating the expression of TRPV1 and is involved in the expression of TRPV1 in the posterior root ganglion (DRG) (48). Exogenous NGF increases TRPV1 mRNA content, transport, and presence in the plasma membrane, as well as sensitivity to capsaicin in DRG neurons in a culture medium. Also, NGF upregulates TRPV1 during inflammation (49). On the other hand, in cases where nerve cutting occurs, such as axotomy of the sciatic nerve, downregulation of TRPV1 is seen, which is probably due to the lack of NGF. It determines the importance of the presence of NGF on the expression of TRPV1 (50). The mechanisms through which NGF facilitates TRPV1 expression remained unclear. Recently, it has been shown that NGF activates a small protein, GTPase Ras,

whose activation is essential for the regulation of TRPV1 expression. Ras activation is an important component of the downstream pathway of inflammatory TRPV1 induction by NGF. Also, NGF may regulate TRPV1 expression by modulating the membrane potential, which leads to a depolarizing change in the membrane potential (50). In addition, the role of TRPV1 receptors in epileptogenesis has been reported. Hippocampal TRPV1 receptors are highly expressed in people with epilepsy suffering from temporal lobe epilepsy (51). Functional disorders of TRPV1 receptors cause changes in seizure induction in an animal model of tonic-clonic epilepsy (52). In contrast, TRPV1 agonist delays epileptogenesis in the kainic acid model of epilepsy (53). Therefore, our findings suggest that exercise modulates TRPV1 function in sensory neurons by increasing NGF expression.

### 5.4. Conclusions

These findings suggest that increased level of neurotrophins following moderate-intensity exercise may be involved in the antiepileptic effect of exercise through modulation of TRPV1 pathway. This study's findings may be another emphasis on regulating moderate exercise as a supplementary treatment for patients who suffer from epileptic seizures. Also, considering the role of physical exercise on TRPV1 modulation can point out new approaches that may lead to the development of a new class of antiepileptic drugs for clinical applications.

#### 5.5. Limitations

The limitation of this study is that the kind of cells (glia or neurons) involved in the TRPV1 modulation have not been distinguished. It seems that more studies in this field will help understand the mechanisms underlying the beneficial effects of exercise.

# Acknowledgments

This research was supported by Iran University of Medical Sciences; grant number, 98-4-21-16659.

## Footnotes

Authors' Contribution: Conception and study design: Fariba Karimzadeh; administrative, technical, and material support: Azam Navazesh and Homa Rasoolijazi; Performing experiments and acquisition of data: Ghazal Rahmani and Saad Bavi; Analysis and interpretation of data: Azam Navazesh and Gelareh Vahabzadeh; drafting of the manuscript: Mansoureh Soleimani; Critical revision of the manuscript for important intellectual content: Fariba Karimzadeh. **Conflict of Interests:** There is no conflict of interest in the funding, personal financial interests, or other properties.

**Data Reproducibility:** The data set presented in the study is available on request from the corresponding author during submission or after publication.

Ethical Approval:Ethical approvalcode:IR.IUMS.REC.1399.023;Link:ethics.research.ac.ir/EthicsProposalView.php?&code=IR.IUMS.REC.1399.023

**Funding/Support:** This research was supported by Iran University of Medical Sciences; grant number, 98-4-21-16659.

#### References

- Eftekhari S, Mehrabi S, Karimzadeh F, Joghataei MT, Khaksarian M, Hadjighassem MR, et al. Brain Derived Neurotrophic Factor Modification of Epileptiform Burst Discharges in a Temporal Lobe Epilepsy Model. *Basic Clin Neurosci.* 2016;7(2):115– 20. [PubMed ID: 27303606]. [PubMed Central ID: PMC4892316]. https://doi.org/10.15412/J.BCN.03070205.
- Barzroodi Pour M, Bayat M, Golab F, Eftekharzadeh M, Katebi M, Soleimani M, et al. The effect of exercise on GABA signaling pathway in the model of chemically induced seizures. *Life Sci.* 2019;232:116667. [PubMed ID: 31326567]. https://doi.org/10.1016/j.lfs.2019.116667.
- 3. Ghasemi P, Gharakhanlou R, Shamsi MM, Khodadadi D. The Effect of 4 Weeks Aerobic Exercise on Gene Expression of Glial Cell\_Derived Neurotrophic Factor, TNF- $\alpha$  and Cognition in Rat's Hippocampus with Alzheimer's Disease Induced by Amyloid Beta. *Sport Phys.* 2021;**13**(49):169–98.
- Paudel YN, Shaikh MF, Shah S, Kumari Y, Othman I. Role of inflammation in epilepsy and neurobehavioral comorbidities: Implication for therapy. *Eur J Pharmacol.* 2018;837:145–55. [PubMed ID: 30125565]. https://doi.org/10.1016/j.ejphar.2018.08.020.
- Duchatel RJ, Shannon Weickert C, Tooney PA. White matter neuron biology and neuropathology in schizophrenia. NPJ Schizophr. 2019;5(1):10. [PubMed ID: 31285426]. [PubMed Central ID: PMC6614474]. https://doi.org/10.1038/s41537-019-0078-8.
- Zimcikova E, Simko J, Karesova I, Kremlacek J, Malakova J. Behavioral effects of antiepileptic drugs in rats: Are the effects on mood and behavior detectable in open-field test? *Seizure*. 2017;**52**:35–40. [PubMed ID: 28957723]. https://doi.org/10.1016/j.seizure.2017.09.015.
- Chen B, Choi H, Hirsch LJ, Katz A, Legge A, Buchsbaum R, et al. Psychiatric and behavioral side effects of antiepileptic drugs in adults with epilepsy. *Epilepsy Behav.* 2017;**76**:24–31. [PubMed ID: 28931473]. https://doi.org/10.1016/j.yebeh.2017.08.039.
- Carrizosa-Moog J, Ladino LD, Benjumea-Cuartas V, Orozco-Hernandez JP, Castrillon-Velilla DM, Rizvi S, et al. Epilepsy, Physical Activity and Sports: A Narrative Review. *Can J Neurol Sci.* 2018;45(6):624–32. [PubMed ID: 30430971]. https://doi.org/10.1017/cjn.2018.340.
- Hafele CA, Rombaldi AJ, Feter N, Hafele V, Gervini BL, Domingues MR, et al. Effects of an exercise program on health of people with epilepsy: A randomized clinical trial. *Epilepsy Behav.* 2021;**117**:107904. [PubMed ID: 33740496]. https://doi.org/10.1016/j.yebeh.2021.107904.
- Hung CL, Tseng JW, Chao HH, Hung TM, Wang HS. Effect of Acute Exercise Mode on Serum Brain-Derived Neurotrophic Factor (BDNF) and Task Switching Performance. J Clin Med. 2018;7(10). [PubMed ID: 30249981]. [PubMed Central ID: PMC6209934]. https://doi.org/10.3390/jcm7100301.
- Jeon YK, Ha CH. The effect of exercise intensity on brain derived neurotrophic factor and memory in adolescents. *Environ Health Prev Med.* 2017;22(1):27. [PubMed ID: 29165142]. [PubMed Central ID: PMC5664787]. https://doi.org/10.1186/s12199-017-0643-6.

- Mitre M, Mariga A, Chao MV. Neurotrophin signalling: novel insights into mechanisms and pathophysiology. *Clin Sci (Lond)*. 2017;131(1):13– 23. [PubMed ID: 27908981]. [PubMed Central ID: PMC5295469]. https://doi.org/10.1042/CS20160044.
- Faustino C, Rijo P, Reis CP. Nanotechnological strategies for nerve growth factor delivery: Therapeutic implications in Alzheimer's disease. *Pharmacol Res.* 2017;**120**:68–87. [PubMed ID: 28351757]. https://doi.org/10.1016/j.phrs.2017.03.020.
- Paolone G, Falcicchia C, Lovisari F, Kokaia M, Bell WJ, Fradet T, et al. Long-Term, Targeted Delivery of GDNF from Encapsulated Cells Is Neuroprotective and Reduces Seizures in the Pilocarpine Model of Epilepsy. J Neurosci. 2019;39(11):2144-56. [PubMed ID: 30665947]. [PubMed Central ID: PMC6507083]. https://doi.org/10.1523/JNEUROSCI.0435-18.2018.
- Mokhtarzadeh M, Ranjbar R, Majdinasab N, Negaresh R. [The Role of Physical Activity on Modulation of Nerve Growth Factors [Brain-Derived Neurotrophic Factor (BDNF) and Nerve Growth Factor (NGF)] in Patients with Multiple Sclerosis: A Systematic Review]. J Isfahan Med Sch. 2017; 35(438):842–54. Persian.
- Ferreira AF, Real CC, Rodrigues AC, Alves AS, Britto LR. Short-term, moderate exercise is capable of inducing structural, BDNFindependent hippocampal plasticity. *Brain Res.* 2011;1425:111-22. [PubMed ID: 22035567]. https://doi.org/10.1016/j.brainres.2011.10.004.
- Chen HC, Chen MY, Hsieh CL, Wu SY, Hsu HC, Lin YW. TRPV1 is a Responding Channel for Acupuncture Manipulation in Mice Peripheral and Central Nerve System. *Cell Physiol Biochem*. 2018;49(5):1813–24. [PubMed ID: 30231245]. https://doi.org/10.1159/000493627.
- Bonacin YS, Marques ICS, Garcia SB, Silva SBG, Canola PA, Marques JA. The role of vanilloid receptor type 1 (TRPV1) in hyperalgesia related to bovine digital dermatitis. *J Dairy Sci.* 2020;**103**(8):7315–21. [PubMed ID: 32505399]. https://doi.org/10.3168/jds.2019-17035.
- Carletti F, Gambino G, Rizzo V, Ferraro G, Sardo P. Involvement of TRPV1 channels in the activity of the cannabinoid WIN 55,212-2 in an acute rat model of temporal lobe epilepsy. *Epilepsy Res.* 2016;**122**:56–65. [PubMed ID: 26970948]. https://doi.org/10.1016/j.eplepsyres.2016.02.005.
- Shutov LP, Warwick CA, Shi X, Gnanasekaran A, Shepherd AJ, Mohapatra DP, et al. The Complement System Component C5a Produces Thermal Hyperalgesia via Macrophage-to-Nociceptor Signaling That Requires NGF and TRPVI. J Neurosci. 2016;36(18):5055-70. [PubMed ID: 27147658]. [PubMed Central ID: PMC4854968]. https://doi.org/10.1523/[NEUROSCI.3249-15.2016.
- Barzroodi Pour M, Bayat M, Navazesh A, Soleimani M, Karimzadeh F. Exercise Improved the Anti-Epileptic Effect of Carbamazepine through GABA Enhancement in Epileptic Rats. *Neurochem Res.* 2021;46(8):2112–30. [PubMed ID: 34008120]. https://doi.org/10.1007/s11064-021-03349-3.
- Castaneda-Cabral JL, Urena-Guerrero ME, Beas-Zarate C, Colunga-Duran A, Nunez-Lumbreras MLA, Orozco-Suarez S, et al. Increased expression of proinflammatory cytokines and iNOS in the neocortical microvasculature of patients with temporal lobe epilepsy. *Immunol Res.* 2020;68(3):169–76. [PubMed ID: 32542572]. https://doi.org/10.1007/s12026-020-09139-3.
- Bauer J, Becker AJ, Elyaman W, Peltola J, Ruegg S, Titulaer MJ, et al. Innate and adaptive immunity in human epilepsies. *Epilepsia*. 2017;58 Suppl 3(Suppl Suppl 3):57-68. [PubMed ID: 28675562]. [PubMed Central ID: PMC5535008]. https://doi.org/10.1111/epi.13784.
- Rana A, Musto AE. The role of inflammation in the development of epilepsy. J Neuroinflammation. 2018;15(1):144. [PubMed ID: 29764485]. [PubMed Central ID: PMC5952578]. https://doi.org/10.1186/s12974-018-1192-7.
- Clark LF, Kodadek T. The Immune System and Neuroinflammation as Potential Sources of Blood-Based Biomarkers for Alzheimer's Disease, Parkinson's Disease, and Huntington's Disease. ACS Chem Neurosci. 2016;7(5):520-7. [PubMed ID: 27046268]. https://doi.org/10.1021/acschemneuro.6b00042.
- 26. Azizzadeh T, Zolfaghari MR, Fattahi A, Khadem VK. Effect of moderateintensity continuous training and high-intensity interval training with astaxanthin supplementation on inflammatory factors of cardiac tissue in type 2 diabetic rats. *Ebnesina*. 2021;**23**(3):12–22.

- Chennaoui M, Drogou C, Gomez-Merino D. Effects of physical training on IL-1beta, IL-6 and IL-1ra concentrations in various brain areas of the rat. *Eur Cytokine Netw.* 2008;**19**(1):8-14. [PubMed ID: 18299269]. https://doi.org/10.1684/ecn.2008.0115.
- Radak Z, Ihasz F, Koltai E, Goto S, Taylor AW, Boldogh I. The redox-associated adaptive response of brain to physical exercise. Free Radic Res. 2014;48(1):84–92. [PubMed ID: 23870001]. https://doi.org/10.3109/10715762.2013.826352.
- Gibbons TE, Pence BD, Petr G, Ossyra JM, Mach HC, Bhattacharya TK, et al. Voluntary wheel running, but not a diet containing (-)-epigallocatechin-3-gallate and beta-alanine, improves learning, memory and hippocampal neurogenesis in aged mice. *Behav Brain Res.* 2014;**272**:131–40. [PubMed ID: 25004447]. [PubMed Central ID: PMC4428596]. https://doi.org/10.1016/j.bbr.2014.05.049.
- Sullivan AM, O'Keeffe GW. Neurotrophic factor therapy for Parkinson's disease: past, present and future. *Neural Regen Res.* 2016;**11**(2):205-7. [PubMed ID: 27073356]. [PubMed Central ID: PMC4810967]. https://doi.org/10.4103/1673-5374.177710.
- Chen P, Li X. Study on Effect of Striatal mGluR2/3 in Alleviating Motor Dysfunction in Rat PD Model Treated by Exercise Therapy. Front Aging Neurosci. 2019;11:255. [PubMed ID: 31632264]. [PubMed Central ID: PMC6783497]. https://doi.org/10.3389/fnagi.2019.00255.
- Hamidi Perchikolaei O, Fallah Mohammadi Z, Haji Zadeh Moghadam A. The effect of treadmill running with consumption of vitamin D3 on NGF levels in Parkinsonian rat's striatum. Sport Physiology. 2016;8(29):91–102.
- Hong YP, Lee HC, Kim HT. Treadmill exercise after social isolation increases the levels of NGF, BDNF, and synapsin I to induce survival of neurons in the hippocampus, and improves depression-like behavior. J Exerc Nutrition Biochem. 2015;19(1):11-8. [PubMed ID: 25960950]. [PubMed Central ID: PMC4424441]. https://doi.org/10.5717/jenb.2015.19.1.11.
- Wu HM, Tzeng NS, Qian L, Wei SJ, Hu X, Chen SH, et al. Novel neuroprotective mechanisms of memantine: increase in neurotrophic factor release from astroglia and anti-inflammation by preventing microglial activation. *Neuropsychopharmacology*. 2009;**34**(10):2344– 57. [PubMed ID: 19536110]. [PubMed Central ID: PMC3655438]. https://doi.org/10.1038/npp.2009.64.
- Groover AL, Ryals JM, Guilford BL, Wilson NM, Christianson JA, Wright DE. Exercise-mediated improvements in painful neuropathy associated with prediabetes in mice. *Pain*. 2013;**154**(12):2658-67. [PubMed ID: 23932909]. [PubMed Central ID: PMC3844098]. https://doi.org/10.1016/j.pain.2013.07.052.
- Lau YS, Patki G, Das-Panja K, Le WD, Ahmad SO. Neuroprotective effects and mechanisms of exercise in a chronic mouse model of Parkinson's disease with moderate neurodegeneration. *Eur J Neurosci.* 2011;33(7):1264–74. [PubMed ID: 21375602]. [PubMed Central ID: PMC3079264]. https://doi.org/10.1111/j.1460-9568.2011.07626.x.
- Saavedra A, Baltazar G. GDNF and PD: Less Common Points of View. Towards New Therapies for Parkinson's Disease. IntechOpen; 2011. https://doi.org/10.5772/17816.
- Wu SY, Wang TF, Yu L, Jen CJ, Chuang JI, Wu FS, et al. Running exercise protects the substantia nigra dopaminergic neurons against inflammation-induced degeneration via the activation of BDNF signaling pathway. *Brain Behav Immun.* 2011;25(1):135–46. [PubMed ID: 20851176]. https://doi.org/10.1016/j.bbi.2010.09.006.
- McCullough MJ, Gyorkos AM, Spitsbergen JM. Short-term exercise increases GDNF protein levels in the spinal cord of young and old rats. *Neuroscience*. 2013;240:258–68. [PubMed ID: 23500094]. [PubMed Central ID: PMC3637874]. https://doi.org/10.1016/j.neuroscience.2013.02.063.

- Khorshidvand M, Ghara Khanlou R, Hassan Sajedi R. The Effect of Moderate Continuous Training on TRPVI Protein Expression in Slow-Contraction Muscles of Wistar Rats. J Sport Bioscie. 2019;11(1):83–96. https://doi.org/10.22059/jsb.2019.269023.1319.
- Santoso DII, Yolanda S, Redjeki S, Andraini T, Ivanali K. Continuous environmental enrichment and aerobic exercise improves spatial memory: focus on rat hippocampal BDNF and NGF. *Comp Exerc Physiol.* 2020;16(2):121-8. https://doi.org/10.3920/cep190036.
- Schultz MG, La Gerche A, Sharman JE. Blood Pressure Response to Exercise and Cardiovascular Disease. *Curr Hypertens Rep.* 2017;**19**(11):89. [PubMed ID: 29046978]. https://doi.org/10.1007/s11906-017-0787-1.
- Kalfon L, Azran A, Farajun Y, Golan-Hamu O, Toben A, Abramov L, et al. Localized Provoked Vulvodynia: Association With Nerve Growth Factor and Transient Receptor Potential Vanilloid Type 1 Genes Polymorphisms. J Low Genit Tract Dis. 2019;23(1):58–64. [PubMed ID: 30418350]. https://doi.org/10.1097/LGT.000000000000445.
- 44. Mizumura K, Taguchi T. Delayed onset muscle soreness: Involvement of neurotrophic factors. J Physiol Sci. 2016;**66**(1):43–52. [PubMed ID: 26467448]. https://doi.org/10.1007/s12576-015-0397-0.
- Baskaran P, Krishnan V, Ren J, Thyagarajan B. Capsaicin induces browning of white adipose tissue and counters obesity by activating TRPV1 channel-dependent mechanisms. *Br J Pharmacol.* 2016;**173**(15):2369–89. [PubMed ID: 27174467]. [PubMed Central ID: PMC4945767]. https://doi.org/10.1111/bph.13514.
- Nanou E, Catterall WA. Calcium Channels, Synaptic Plasticity, and Neuropsychiatric Disease. *Neuron*. 2018;98(3):466–81. [PubMed ID: 29723500]. https://doi.org/10.1016/j.neuron.2018.03.017.
- 47. Amaya F, Shimosato G, Nagano M, Ueda M, Hashimoto S, Tanaka Y, et al. NGF and GDNF differentially regulate TRPV1 expression that contributes to development of inflammatory thermal hyperalgesia. *Eur J Neurosci.* 2004;20(9):2303-10. [PubMed ID: 15525272]. https://doi.org/10.1111/j.1460-9568.2004.03701.x.
- Shimizu N, Wada N, Shimizu T, Suzuki T, Takaoka EI, Kanai AJ, et al. Effects of nerve growth factor neutralization on TRP channel expression in laser-captured bladder afferent neurons in mice with spinal cord injury. *Neurosci Lett.* 2018;683:100–3. [PubMed ID: 29960052]. [PubMed Central ID: PMC6448406]. https://doi.org/10.1016/j.neulet.2018.06.049.
- Sapio MR, Neubert JK, LaPaglia DM, Maric D, Keller JM, Raithel SJ, et al. Pain control through selective chemo-axotomy of centrally projecting TRPVI+ sensory neurons. *J Clin Invest.* 2018;**128**(4):1657– 70. [PubMed ID: 29408808]. [PubMed Central ID: PMC5873867]. https://doi.org/10.1172/JCI94331.
- Obeidat AM, Donner A, Miller RE. An update on targets for treating osteoarthritis pain: NGF and TRPV1. *Curr Treatm Opt Rheumatol.* 2020;6(3):129–45. [PubMed ID: 34178580]. [PubMed Central ID: PMC8223492]. https://doi.org/10.1007/s40674-020-00146-x.
- Shamsizadeh A, Fatehi F, Khajehasani F, Hassanshahi G, Kazemi Arababadi M. Expression of TRPV1 Receptors Increased in Hippocampus Following Pentylenetetrazole-Induced Kindling in Male Rats. Arch Neurosci. 2016;3(3). https://doi.org/10.5812/archneurosci.35459.
- 52. Vilela LR, Lima IV, Kunsch EB, Pinto HPP, de Miranda AS, Vieira ELM, et al. Anticonvulsant effect of cannabidiol in the pentylenetetrazole model: Pharmacological mechanisms, electroencephalographic profile, and brain cytokine levels. *Epilepsy Behav*. 2017;**75**:29–35. [PubMed ID: 28821005]. https://doi.org/10.1016/ji.yebeh.2017.07.014.
- Lee TH, Lee JG, Yon JM, Oh KW, Baek IJ, Nahm SS, et al. Capsaicin prevents kainic acid-induced epileptogenesis in mice. *Neurochem Int.* 2011;58(6):634–40. [PubMed ID: 21333704]. https://doi.org/10.1016/j.neuint.2011.01.027.