Successful Treatment of Carbapenem-Resistant *Klebsiella pneumoniae* Meningitis with Combination Therapy of Meropenem and Amikacin

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Abstract

**Introduction:** The widespread use of carbapenems increased the prevalence of Carbapenem-resistant *Enterobacteriaceae* with subsequent increases in mortality due to extremely limited treatment options. Following neurosurgical procedures, carbapenem-resistant *Klebsiella pneumoniae* (CRKP) is the main cause of central nervous system (CNS) infections. The optimal antimicrobial treatment of such infections has not yet been defined.

**Case Presentation:** We present a 4-month-old boy with an extra-ventricular drain-related central nervous system (CNS) infection with CRKP. Although meropenem minimum inhibitory concentration (MIC) for the bacterial isolate was \( \geq 16 \) mg/L and it harbored *blaNDM*, *blaVIM*, and *blaOXA-48*-like Carbapenemase genes, this infection was effectively treated with a combination therapy of intravenous (IV) double dose extended infusion of meropenem in addition to amikacin.

**Conclusions:** This successful treatment regimen for CRKP-causing meningitis may pave the way to manage severe CNS infections with extensive-drug-resistant bacteria in infants and children without inserting an external drain or intra-thecal antibiotic administration.

**Keywords:** Carbapenem-resistant *Klebsiella*, Intravenous, Extended Infusion, Meropenem, Amikacin

1. Introduction

Healthcare-associated ventriculitis and meningitis (VM) are some of the most serious complications that drastically impact the prognosis following neurosurgical procedures. Recently, multidrug-resistant *Enterobacteriaceae*, especially carbapenem-resistant *Klebsiella pneumonia*, became an important cause of central nervous system infection, representing a major public health threat (1, 2). Such organisms represent a therapeutic contest as carbapenemase can hydrolyze all carbapenems, cephalosporins, and Beta-lactams (3). The most common *K. pneumoniae* carbapenemase (KPC) is New Delhi metallo-\( \beta \)-lactamase (NDM), oxacillinase (OXA-48-like), and imipenemase (IMP) (4).

A few drugs, such as colistin, tigecycline, and aminoglycosides, and in some strains, ceftazidime/avibactam, can effectively treat CPKP. Due to the poor penetration of the blood-brain barrier, it is difficult for most medications to reach the minimal inhibitory concentration (MIC) in the cerebrospinal fluid (CSF). To overcome these disadvantages, many cases and clinical studies have attempted to treat CNSIs caused by multidrug-resistant bacteria using intraventricular (IVT) injections (4).

The best treatment for CNS infection caused by carbapenem-resistant *K. pneumoniae* is still controversial. The current case elaborated that the combination regimen of parental, extended infusion high-dose meropenem, and amikacin could be a useful treatment option.

1.1. Ethical Consideration

Written informed consent was obtained from the patient’s parent for publishing this case report.
2. Case Presentation

A 4-month-old boy with hydrocephalus was admitted to our institute for the treatment of a suspected shunt infection that had previously been treated for three weeks with various antibiotic courses with no clinical improvement. On admission, the child was hypoactive, with a temperature of 37°C, heart rate (HR) of 130 beats/min, respiratory rate (RR) of 30 cycles/min, and oxygen saturation of 99% in room air. His complete blood count (CBC) revealed an increase in C-reactive protein (CRP) level of 6.61 × 10³/cm³, absolute neutrophil count of 1.75 × 10³/cm³, hemoglobin 10.5 g/dL, platelet count of 376 × 10³/cm³, and C-reactive protein (CRP) 9.4 mg/dL. Cerebrospinal fluid (CSF) revealed no growth, and a new antibiotic-impregnated VP shunt was inserted after ensuring the successful treatment of the shunt infection. New labs revealed an increase in C-reactive protein (CRP) level of 140 mg/dL, and his complete blood count (CBC) showed an increase in C-reactive protein (CRP) level of 6.61 × 10³/cm³, absolute neutrophil count of 1.75 × 10³/cm³, hemoglobin 10.5 g/dL, platelet count of 376 × 10³/cm³, and C-reactive protein (CRP) 9.4 mg/dL.

Cerebrospinal fluid (CSF) analysis was done (Table 1) and culture were withdrawn. The infected shunt was removed, an extra-ventricular drain was inserted, and empiric treatment with vancomycin 15 mg/kg/dose Q8h and Meropenem 40 mg/kg/dose Q8h extended infusion over 4 hours accompanied with amikacin at dose 7.5 mg/kg/dose Q8hrs, and treatment was continued for 21 days. On the seventh day of treatment, a cerebrospinal fluid culture revealed no growth, and treatment was continued for another 14 days after the negative culture.

On the ninth day of admission, the child became hypoactive and febrile (his temperature was 38°C). The child had a white blood cell count (WBC) of 6.61 × 10³/cm³, absolute neutrophil count of 1.75 × 10³/cm³, hemoglobin 10.5 g/dL, platelet count of 376 × 10³/cm³, and C-reactive protein (CRP) 9.4 mg/dL. Cerebrospinal fluid (CSF) analysis was done (Table 1) and culture were withdrawn. The infected shunt was removed, an extra-ventricular drain was inserted, and empiric treatment with vancomycin 15 mg/kg/dose Q8h and Meropenem 40 mg/kg/dose Q8h extended infusion over 4 hours accompanied with amikacin at dose 7.5 mg/kg/dose Q8hrs, and treatment was continued for 21 days. On the seventh day of treatment, a cerebrospinal fluid culture revealed no growth, and treatment was continued for another 14 days after the negative culture.

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In the laboratory, 2 mL of the Cerebrospinal Fluid was injected into a Bact-Alert Blood culture bottle (bioMérieux, France). Another portion of the CSF was cultured on Blood agar, MacConkey’s agar, Chocolate agar, and Sabouraud Dextrose agar (Condalab, Madrid, Spain), and a direct Gram-stained smear was examined microscopically.

*Klebsiella pneumoniae* subsp. Pneumoniae was primarily identified by colony morphology and then microscopically by Gram-staining. Further identification and antibiotic susceptibility testing were done with the Vitek 2 Compact system (bioMérieux, France) according to the manufacturer’s protocol. The culture results showed extensive drug-resistant (XDR) carbapenem-resistant *K. pneumoniae* (Table 2). Ceftazidime-Avibactam (CAZ-AVI) E-test (Liofilchem®, Italy), containing CAZ (0.016 - 256 µg/mL) – AVI (4 µg/mL), was used to determine CAZ-AVI sensitivity. The isolate was found to be resistant to this drug.

For genomic identification of Carbapenemase genes in this bacterial isolate, bacterial genomic DNA was extracted using Thermo Scientific GeneJET Genomic DNA Purification Kit (K0721, Thermo Fisher, USA) according to manufacturer’s instructions.

SYBR Green real-time PCR was performed to detect Carbapenemases genes (*blaNDM, blaVIM, blaIMP, blaKPC, blaGES*, and *blaOXA-48-like*) using Mx3000P™ real-time PCR thermal cycler instrument (Strategene, USA). A total of 6 µL of template DNA was added to the reaction mix, which was composed of 10 µL Maxima SYBR Green Master Mix (2X) (K0251 et al., USA) and 0.6 µL for each forward and reversed primers (5-7), and 2.8 µL nuclease-free water. The reaction began with an initial denaturation step at 95°C for 10 min. This was followed by 45 cycles of DNA denaturation at 95°C for 15 s, primer annealing at 55°C for 30 seconds, and primer extension at 72°C for the 30 s. This was followed by dissociation curve analysis consisting of 1 cycle at 95°C for 1min, then at 55°C for the 30 s, and finally at 95°C for 30 s. The results of the molecular study showed that the bacterial isolate was positive for *blaNDM, blaVIM, and blaOXA-48-like.*

It was negative for *blaIMP, blaKPC,* and *blaGES* genes.

To start the effective treatment regimen, the extra-ventricular drain (EVD) was removed as biofilm formation from bacteria will hinder antibiotic penetration and lead to treatment failure. Also, inserting a new EVD was discouraged, as it may be a new source of acquiring other infections. Thus, the clinical decision was to start the treatment regimen through the parenteral route.

Following removal of the drain, we started therapy with meropenem 40 mg/kg/dose Q8h extended infusion over 4 hours accompanied with amikacin at dose 7.5 mg/kg/dose Q8hrs, and treatment was continued for 21 days. On the seventh day of treatment, a cerebrospinal fluid culture (CSF) revealed no growth, and treatment was continued for another 14 days after the negative culture.

Another two cultures were withdrawn one week apart, revealing no growth, and a new antibiotic-impregnated VP shunt was inserted after ensuring the successful treatment of bacterial meningitis.

3. Discussion

Healthcare-associated meningitis and ventriculitis are rising concurrently with an increase in neurosurgery procedures. Multidrug-resistant (MDR) *K. pneumoniae* CNS infection has significant morbidity and mortality (8). Meningitis caused by CRKP post-neurosurgery has been reported in the USA, Turkey, and China (9-11). The main cause of carbapenem resistance in *K. pneumoniae* is the production of carbapenemases, which are mainly acquired through horizontal gene transfer (12). The type
Table 1. Cerebrospinal Fluid Analyses

<table>
<thead>
<tr>
<th>Aspect</th>
<th>Day 1</th>
<th>Day 9</th>
<th>Day 16</th>
<th>Day 23</th>
</tr>
</thead>
<tbody>
<tr>
<td>Glucose, mg/dL</td>
<td>18.59</td>
<td>30.57</td>
<td>19.24</td>
<td>40</td>
</tr>
<tr>
<td>Protein, mg/dL (15.00 - 45.00)</td>
<td>214.43</td>
<td>178.50</td>
<td>223.18</td>
<td>213.2</td>
</tr>
<tr>
<td>LDH, U/L (0.00 - 26.00)</td>
<td>86.67</td>
<td>197.30</td>
<td>151.30</td>
<td>113.7</td>
</tr>
<tr>
<td>Chloride, mmol/L (110.0 - 130.0)</td>
<td>115.40</td>
<td>105.20</td>
<td>104.40</td>
<td>104</td>
</tr>
<tr>
<td>Neutrophil, /cmm</td>
<td>120</td>
<td>44</td>
<td>25</td>
<td>0</td>
</tr>
<tr>
<td>Lymphocyte, /cmm</td>
<td>15</td>
<td>16</td>
<td>75</td>
<td>4</td>
</tr>
<tr>
<td>Culture result</td>
<td>Staphylococcus epidermidis</td>
<td>Klebsiella pneumoniae</td>
<td>No growth</td>
<td>No growth</td>
</tr>
</tbody>
</table>

Table 2. The antibiogram of Klebsiella pneumoniae isolated on day 9

<table>
<thead>
<tr>
<th>Antibiotic</th>
<th>MIC (mg/L)</th>
<th>Interpretation</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ampicillin</td>
<td>≥ 32</td>
<td>R</td>
</tr>
<tr>
<td>Ampicillin/sulbactam</td>
<td>≥ 32</td>
<td>R</td>
</tr>
<tr>
<td>Piperacillin/tazobactam</td>
<td>≥ 128</td>
<td>R</td>
</tr>
<tr>
<td>Cefazidime</td>
<td>≥ 64</td>
<td>R</td>
</tr>
<tr>
<td>Ceftriaxone</td>
<td>≥ 64</td>
<td>R</td>
</tr>
<tr>
<td>Cefepime</td>
<td>≥ 64</td>
<td>R</td>
</tr>
<tr>
<td>Meropenem</td>
<td>≥ 16</td>
<td>R</td>
</tr>
<tr>
<td>Amikacin30</td>
<td>16</td>
<td>S</td>
</tr>
<tr>
<td>Gentamicin</td>
<td>≤ 1</td>
<td>S</td>
</tr>
<tr>
<td>Ciprofloxacin</td>
<td>&gt; 2</td>
<td>R</td>
</tr>
<tr>
<td>Levofloxacin</td>
<td>&gt; 2</td>
<td>R</td>
</tr>
<tr>
<td>Trimethoprim +sulfamethoxazole</td>
<td>≥ 320</td>
<td>R</td>
</tr>
<tr>
<td>Colistin</td>
<td>≤ 0.5</td>
<td>S</td>
</tr>
<tr>
<td>Tigecycline</td>
<td>&lt; 2</td>
<td>S</td>
</tr>
</tbody>
</table>

Abbreviations: R, resistant; S, sensitive; MIC, minimum inhibitor concentration.

of carbapenemase varies greatly in different geographical locations. The carbapenemases include K. pneumoniae carbapenemases (KPCs), metallo-β-lactamases (MBLs), and oxacillinas (OXAs) (13, 14).

Unfortunately, which antibiotics should be used to treat CRKP infections is still unclear. Recent evidence suggests that antimicrobial combination therapy may be more effective than monotherapy and that, whenever possible, it is recommended to include meropenem as the primary component of combination regimens (15).

Parental colistin alone in treating meningitis is discouraged due to poor blood-brain barrier penetration, leading to low-dose drug concentrations in the CSF and treatment failure. Moreover, there is an increased risk of nephrotoxicity and, less commonly, neurotoxicity, even when administered in appropriate doses (16, 17). Based on the recommendation of the Infectious Disease Society of America, intraventricular treatment is necessary for healthcare-associated ventriculitis and meningitis where systemic antibiotic therapy is ineffective (18).

Given the data mentioned above, and though meropenem resistance (MIC ≥ 16mg/L) to the isolated CRKP strain, in this case, the selected treatment regimen with high dose extended infusion of meropenem and amikacin was administered, and the patient was effectively cured.

The rationale for using this regimen depends on the fact that carbapenems have time-dependent antibacterial activity; the antibacterial activity is correlated with the length of time that free concentrations persist above the minimum inhibitory concentration (MIC) (time of free concentration (%fT) > MIC). For carbapenems to be effective against bacteria in severe infections, larger levels (%fT > MIC, 70 to 80%), or even higher, are required. As a result, adopting the maximal dose typically necessitates extending the infusion period (3 to 4 h or continuous infusion) for improved pharmacokinetic (PK) and pharmacodynamic (PD) outcomes (19).

In concordance with our case, He et al. also reported successful treatment of carbapenem-resistant Enterobacter cloacae (MIC of imipenem ≥ 16mg/L) with high dose and prolonged-infusion regimen of IV meropenem and Amikacin plus Intraventricular Amikacin (20). Different treatment regimens were used in different other studies, including parental tigecycline and amikacin with intraventricular amikacin, meropenem and intraventricular colistin, and IV ceftazidime/avibactam and intraventricular gentamicin (9-11).

3.1. Conclusions

This case report demonstrates the possibility of using a high dose of extended infusion meropenem and Amikacin in treating CRKP Meningitis (MIC ≥ 16 mg/L) precisely with

limited treatment options. Further studies are urgently needed to determine the most effective treatment of CRKP meningitis. This case report sheds light and paved the way in treating CRKP Meningitis and proves treating CRKP Meningitis and proves that antimicrobial combination therapy will be more effective than monotherapy. It is recommended to include meropenem as the primary component of combination regimens.

Acknowledgments

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Footnotes

Authors’ Contribution: Eman Hamza, study conception and design, data collection and clinical outcome monitoring, final interpretations, and draft manuscript preparation; Hassan Eshra, draft manuscript preparation; Shahinda Rezk, practical microbiology and molecular diagnostics, data collection, draft manuscript preparation, and final revision; Heba M. Selim, draft manuscript preparation and revision; Mohamed Turkey, practical microbiology and molecular diagnostics, and draft manuscript preparation.

Conflict of Interests: The authors declare that they have no competing interests.

Data Reproductibility: The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Ethical Approval: The report applied the World Medical Association Helsinki Declaration to study human participants. Then took the approval of the Ethics Committee of the Medical Research Institute, Alexandria University (IORG#: IORG0008812).

Funding/Support: There is no funding for the case report.

Informed Consent: Written informed consent was obtained from the patient’s parent for participating in this case report.

References


