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**Research Article** 

# Prevalence and Antibiotic Susceptibility Patterns of *Salmonella* and *Shigella* Species Isolated from Pediatric Diarrhea in Tehran

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### Abstract

**Background:** Salmonella and Shigella infections are important public health concerns among children. The emergence of antibiotic resistance amongst Salmonella and particularly Shigella isolates has posed serious problems in antimicrobial treatment worldwide. Data on local antibiotic resistance patterns are essential to design suitable antibiotic treatment guidelines.

**Objectives:** The aim of the present study was to determine the prevalence and drug susceptibility patterns of *Shigella* and *Salmonella* species in addition to the detection of extended-spectrum  $\beta$ -lactamase producing isolates among diarrhea samples of pediatric patients.

**Methods:** A total of 5300 diarrheic samples from children were examined for the presence of *Salmonella* and *Shigella* species. Biochemical and microbial tests, as well as specific polyvalent antisera, were used for the identification of the bacterial species. Antibiotic susceptibility tests and extended-spectrum  $\beta$ -lactamase detection were conducted by disc diffusion and combination disc methods, respectively.

**Results:** A total of 371 (7%) and 472 (8.9%) samples were positive for *Salmonella* and *Shigella* species, respectively. The most prevalent *Salmonella* isolate was serogroup D (n: 176, 47.5%). Of the *Shigella* isolates, 60.8% were found as *Shigella* sonnei and 39.2% as *Shigella flexneri*. The highest level of antibiotic resistance was noted among *Shigella flexneri* isolates. Overall, 35.7% of the *Shigella flexneri* isolates and 31% of the *Shigella* sonnei isolates produced extended-spectrum  $\beta$ -lactamases.

**Conclusions:** This study provided information on the prevalence and antimicrobial susceptibility patterns of *Salmonella* and *Shigella* isolates in Iran. It also indicated a high-level resistance among *Shigella* isolates to co-trimoxazole and ampicillin and among *Salmonella* isolates to nalidixic acid.

Keywords: Prevalence, Drug Resistances, Pediatric, Diarrhea, Shigella, Salmonella

### 1. Background

Diarrhea, as a major health problem, is considered a leading cause of morbidity and mortality, especially among children, in developing countries (1). *Salmonella* and *shigella* species are two major enteropathogenic bacteria, which, along with Enterotoxigenic Escherichia coli (ETEC) and Campylobacter, account for the most diarrheal infections in the developing world. Human salmonellosis, being more prevalent among children under 5 years of age, causes more than one billion disease cases annually, 3 million of which lead to death across the world (2, 3).

In addition, about 200 million *Shigella* infections and 3 - 5 million deaths occur annually in developing countries, most of which affect children under 5 years of age

(2). Epidemiological data have shown that *Shigella sonnei* (*S. sonnei*) is the predominating *Shigella* species in Europe and USA while *Shigella flexneri* (*S. flexneri*) is more frequent among Asian and African countries (4). In Tehran, until 2003, the dominant species was *S. flexneri*, but it was later replaced by *S. sonnei* (5).

Salmonella and particularly Shigella species are increasingly acquiring resistance to commonly used antibiotics, most importantly to third-generation cephalosporins (3GC), as the main drug of choice for the treatment of Salmonella and Shigella infections, through the production of extended-spectrum  $\beta$ -lactamases (ESBLs) (6-9). Data on local antibiotic resistance patterns are essential to have an empiric antibiotic treatment guideline adapted to the

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local microbial epidemiology (10).

## 2. Objectives

The aim of the present study was to determine the prevalence and drug susceptibility patterns of *Shigella* and *Salmonella* isolates in addition to the detection of ESBL producing isolates among diarrhea samples of pediatric patients.

## 3. Methods

## 3.1. Sample Collection

A cross-sectional study was conducted from 2012 to 2016 on pediatric patients admitted to the children's medical center (CMC) hospital in Tehran, Iran, to assess the prevalence and drug susceptibility patterns of *Salmonella* and *Shigella* species, in addition to the detection of ESBL producing isolates. Single stool samples from each of the 5,300 diarrheic children were examined for the presence of *Salmonella* and *Shigella* species. All the patients, admitted with the clinical signs of acute diarrhea or enteric fever, had an Iranian nationality with an age range of 1-10 years old.

## 3.2. Bacterial Analysis

Stool specimens were cultured on MacConkey (Merck Co., Germany) and Xylose-Lysin-Deoxycholate (XLD) selective media (Merck Co., Germany) and incubated at 35 - 37° C for 1 to 4 days. Inoculation into Selenite-F enrichment broth (Oxoid Co., UK) and subculture on XLD after 8 - 12 hours were then performed to improve the recovery of *Salmonella* spp.

All the suspected colonies grown on XLD were analyzed by routine biochemical and microbiological tests (11). Specific polyvalent antisera (Baharafshan Co., Iran) were used for the serotype identification of *Salmonella* and *Shigella* species.

## 3.3. Antimicrobial Susceptibility Testing and Detection of ESBLs

Antibiotic susceptibility tests were performed by the Kirby-Bauer disc diffusion method on Mueller Hinton agar (Merck Co., Germany), as recommended by the clinical and laboratory standards institute (CLSI) (12). The antimicrobial agents (MAST Co., UK) used in this study were ampicillin (10  $\mu$ g), nalidixic acid (30  $\mu$ g), co-trimoxazole (25  $\mu$ g), and cefotaxime (30  $\mu$ g). *E. coli* ATCC (American type culture collection) 25922 was used as the control.

The phenotypic detection of ESBL production was carried out for *Salmonella* and *Shigella* spp. that were resistant to cefotaxime and ceftazidime by the combination disc method using the cefotaxime (30  $\mu$ g), cefpodoxime (30  $\mu$ g), and ceftazidime (30  $\mu$ g) discs (MAST Co., UK), with or without 10  $\mu$ g of clavulanic acid. A  $\geq$  5 mm increase in the zone diameter of ceftazidime, cefpodoxime, or cefotaxime tested in combination with clavulanic acid versus its zone tested without clavulanic acid was indicative of ESBL production. The ESBL-producing strain *K. pneumoniae* ATCC 700603 was used as the positive control (12).

## 3.4. Statistical Analysis

The Chi-Square test was used to analyze the correlation between the prevalence of *Salmonella* and *Shigella* species and the age and gender of the patients, as well as the resistance rate of *Salmonella* and *Shigella* species with respect to each antibiotic.

SPSS version 22 was used for the analyses and P value  $\leq$  0.05 was considered significant.

## 4. Results

## 4.1. Bacterial Isolation

Out of the 5300 stool samples collected, 371 (7%) and 472 (8.9%) were positive for *Salmonella* and *Shigella* species, respectively. Of the 371 *Salmonella* isolates, 64 (17.2%) belonged to group B, 131 (35.3%) to group C, and 176 (47.5%) to group D.

Of the 472 Shigella isolates, 287 (60.8%) were identified as *S. sonnei* and the remaining 185 (39.2%) as *S. flexneri*. In this study, the prevalence of *S. flexneri* and *S. sonnei* was significantly higher among patients older than 5 years of age (P < 0.001). All the *Salmonella* serogroups were significantly higher in prevalence among patients in the age group of 1-5 years (P < 0.001).

The distribution of *Salmonella* and *Shigella* species according to the sex and age of the patients and their distribution among different hospital's wards are respectively shown in Tables 1 and 2.

## 4.2. Antibiotic Resistance Patterns

With 96.2% resistance to ampicillin, 85.4% to cotrimoxazole, 39.5% to cefotaxime, and 37.3% to nalidixic acid, *S. flexneri* was the most resistant isolate among all the *Shigella* isolates tested.

Among the *Salmonella* serogroups, group C had the highest level of antibiotic resistance (68.7% resistance to nalidixic acid and 48.8% to co-trimoxazole). Resistance to ampicillin and co-trimoxazole was significantly higher among *Shigella* spp. than among different *Salmonella* serogroups (P < 0.001).

	S. flexneri		S. sonnei		S. Serogroup B		S. Serogroup C		S. Serogroup D	
	Male	Female	Male	Female	Male	Female	Male	Female	Male	Female
Age group, y										
< 1	2 (40)	3 (60)	4 (80)	1(20)	9 (50)	9 (50)	27 (64.3)	15 (35.7)	28 (56)	22(44)
1-5	46 (52.3)	42 (47.7)	69 (53.1)	61(46.9)	19 (50)	19 (50)	34 (53.1)	30 (46.9)	52 (50)	52 (50)
5 - 10	52 (56.5)	40 (43.5)	87 (57.2)	65 (42.8)	5 (62.5)	3 (37.5)	16 (64)	9 (36)	12 (54.5)	10 (45.5)
Total	185		287			64	131		176	

Abbreviations: S. flexneri, Shigella flexneri; S. sonnei, Shigella sonnei; S. serogroup B, Salmonella serogroup B; S. serogroup C, Salmonella serogroup C; S. serogroup D, Salmonella serogroup D.
<sup>a</sup>Values are expressed as No. (%).

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#### Table 2. The Distribution of Salmonella and Shigella Species Among Different Hospital Wards<sup>a</sup>

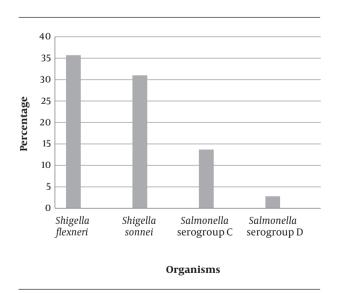
Organisms	Hospital Wards							
	Outpatients	Emergency	Infectious Diseases	Gastroenterology	Rheumatology	ICU	Total	
S. sonnei	177 (61.7)	100 (34.8)	5 (1.7)	2 (0.7)	2 (0.7)	1(0.3)	287	
S. flexneri	99 (53.5)	60 (32.4)	19 (10.3)	6 (3.3)	-	1(0.5)	185	
S. serogroup B	44 (68.7)	12 (18.7)	3 (4.7)	3 (4.7)	1(1.6)	1(1.6)	64	
S. serogroup C	78 (59.5)	25 (19.1)	9 (6.9)	3 (2.3)	14 (10.7)	2 (1.5)	131	
S. serogroup D	106 (60.2)	45 (25.6)	8(4.6)	11 (6.2)	3 (1.7)	3 (1.7)	176	

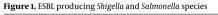
Abbreviations: S. flexneri, Shigella flexneri; S. serogroup B, Salmonella serogroup B; S. serogroup C, Salmonella serogroup C; S. serogroup D, Salmonella serogroup D; S. sonnei, Shigella sonnei; ICU, intensive care unit.

<sup>a</sup> Values are expressed as No. (%).

On the other hand, *Salmonella* serogroups had significantly higher resistance to nalidixic acid compared to *Shigella* species (P < 0.001).

The results of the antibiotic resistance patterns and ESBL detection tests for the *Salmonella* and *Shigella* isolates are presented in Table 3 and Figure 1.





## 5. Discussion

Salmonella and Shigella infections are global public health concerns that are increasingly acquiring resistance to commonly used antibiotics, posing serious problems in antimicrobial treatment worldwide.

In this study, the prevalence of *Salmonella* spp. was 7%, which is similar to that reported by Awole et al. from Ethiopia (7.2%) (13) and lower than those found in other studies as 11.5% (14) and 15.4% (15).

The prevalence of *Shigella* spp., in the current investigation, was 8.9%, which is similar to that reported by studies from Ethiopia (8.7%) (16) and South Africa (8.5%) (17), but higher than those reported in other similar studies in Iran as (5.1%) and (3.8%) (18, 19). The prevalence of shigellosis reported in other developing countries such as Cameroon (4.5%) (20), India (5%) (21), and Ghana (5%) (22) was also lower than that observed in the current study.

This discrepancy may be due to the levels of public health and the availability of safe drinking water among populations, the source of samples, sample size, geographical distribution, the season in which samples were collected, differences in personal hygiene and the age of the target group.

Based on the results of the present study, low levels of *Salmonella* resistance was observed to ampicillin (12%), which is lower than that reported by Jimma (59.3%)(15) and Harar (71.4%) (14). *Salmonella* isolates were highly suscepti-

Organisms	Antibiotics						
	Ampicillin (10 $\mu$ g)	Co-trimoxazole (25 $\mu$ g)	Cefotaxime (30 $\mu$ g)	Nalidixic Acid (30 $\mu {f g}$ )			
S. sonnei	105 (36.6)	280 (97.6)	96 (33.4)	165 (57.5)			
S. flexneri	178 (96.2)	158 (85.4)	73 (39.5)	69 (37.3)			
S. serogroup B	6 (9.4)	11 (17.2)	0(0)	11 (17.2)			
S. serogroup C	23 (17.6)	64 (48.8)	19 (14.5)	90 (68.7)			
S. serogroup D	16 (9.1)	9 (5.1)	6(3.4)	129 (73.3)			

Abbreviations: S. flexneri, Shigella flexneri; S. serogroup B, Salmonella serogroup B; S. serogroup C, Salmonella serogroup C; S. serogroup D, Salmonella serogroup D; S. sonnei, Shigella sonnei.

<sup>a</sup>Values are expressed as No. (%).

ble to cefotaxime (94.1%), which was similar to that found by Hamze et al. (23) but dissimilar to the findings of Bialvaei et al. (9).

Our results showed that among the *Salmonella* serogroups, group C had the highest level of antibiotic resistance to nalidixic acid (68.7%) and co-trimoxazole (48.8%). Various studies have reported different levels of resistance to these antibiotics; for instance, Anvarinejad et al. found 31.5% and 15.79% resistance to nalidixic acid and co-trimoxazole, respectively (3). In another study carried out by Mamuye et al., resistance to nalidixic acid and co-trimoxazole was reported, respectively, as 20% and 60% (8). Moreover, 93.4% of *Salmonella* spp. was resistant to co-trimoxazole as reported by Bialvaei et al. (9).

In this study, *S. sonnei* (60.8%) was the most common *Shigella* species followed by *S. flexneri* (39.2%), which is in contrast with the results of studies by Jomezadeh et al. (18) and MoezArdalan et al. (24) that showed *S. flexneri* as the most prevalent *Shigella* species in Abadan and Karaj, respectively. Our results, however, are in agreement with the results of studies from Shiraz and Tehran (25, 26), confirming recent reports claiming a change over the last years in the prevalence of *Shigella* spp. in Tehran (5).

In this study, most of the *Shigella* isolates were resistant to ampicillin and co-trimoxa-zole, which is in accordance with the previous studies from Iran (18, 27) and other countries such as Nepal (28) and India (29). The wide use of co-trimoxazole and ampicillin, as broad spectrum and low priced empirical treatments for diarrheal diseases (30), has led to the advent of resistance of *Shigella* strains to these antibiotics. According to the results of the present study, these two antibiotics are no longer suitable choices for the treatment of *Shigella* infections. It seems that resistance to nalidixic acid, as the first line of treatment for diarrheal infections, is on the rise in Iran. A study carried out in 200 - 2003 by MoezArdalan in Iran showed that only 1% of the *Shigella* isolates were resistant to nalidixic acid (24), whilst our study showed a rise of 47.4% resistance to this antibi-

otic. In other countries, like India, the resistance of *S. sonnei* to nalidixic acid was reported as 94% - 100% (31, 32).

In this study, 39.5% of *S. flexneri* and 33.4% of *S. sonnei* isolates were resistant to cefotaxime, which is consistent with the results of Bialvaei et al. (9) but much higher than the prevalence found by Barari Savadkoohi et al. (33) and Mamuye et al. (8). Moreover, 35.7% of *S. flexneri* and 31% of *S. sonnei* isolates produced ESBL in the present study, which is in agreement with the results of Bialvaei et al. (9) but higher than those reported by Nath et al. (2.8%) (21), Ranjbar et al. (7.3%) (7), and Sakhaei et al. (11.4%) (34).

ESBL producing *Shigella* species can be problematic in near future and therefore, treatment guidelines should be designed to monitor the emerging ESBL producing isolates at a national level (21).

High rates of antibiotic resistance may result from the uncontrolled or inappropriate use of antibiotics. This can cause antibiotic-resistant bacteria to spread easily between hospital wards, leading to protracted outbreaks with high mortality and morbidity rates (35). The epidemiological information and monitoring system are, therefore, necessary to control *Salmonella* and *Shigella* infections in public health sectors.

## 5.1. Conclusion

This study provided information on the prevalence and antimicrobial susceptibility patterns of *Salmonella* and *Shigella* infections in Tehran, Iran. The antimicrobial resistance patterns suggest the widespread resistance of *Shigella* to ampicillin and co-trimoxazole, which requires specific attention because co-trimoxazole is considered the drug of choice for the treatment of patients with inflammatory diarrhea. Due to the continuous change in antibiotic resistance profiles, *Salmonella* and *Shigella* strains should be under-scrutinized surveillance in order to monitor local susceptibility patterns for the administration of appropriate antibiotics.

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## References

- Abbasi P, Kargar M, Doosti A, Mardaneh J, Ghorbani-Dalini S, Dehyadegari MA. Characterization of Shiga-toxin producing E.coli (STEC) and enteropathogenic E.coli (EPEC) using multiplex Real-Time PCR assays for stx1, stx2, eaeA. *Iran J Microbiol.* 2014;6(3):169–74. [PubMed: 25870750]. [PubMed Central: PMC4393493].
- Ahs JW, Tao W, Löfgren J, Forsberg BC. Diarrheal diseases in low- and middle-income countries: Incidence, prevention and management. *Open Infect Dis J.* 2010;4(1):113–24. doi: 10.2174/1874279301004010113.
- Anvarinejad M, Pouladfar GR, Pourabbas B, Amin Shahidi M, Rafaatpour N, Dehyadegari MA, et al. Detection of Salmonella spp. with the BACTEC 9240 Automated Blood Culture System in 2008 - 2014 in Southern Iran (Shiraz): Biogrouping, MIC, and Antimicrobial Susceptibility Profiles of Isolates. *Jundishapur J Microbiol*. 2016;9(4). e26505. doi: 10.5812/jjm.26505. [PubMed: 27284396]. [PubMed Central: PMC4897598].
- Pazhani GP, Niyogi SK, Singh AK, Sen B, Taneja N, Kundu M, et al. Molecular characterization of multidrug-resistant Shigella species isolated from epidemic and endemic cases of shigellosis in India. J Med Microbiol. 2008;57(Pt 7):856–63. doi: 10.1099/jmm.0.2008/000521-0. [PubMed: 18566144].
- Hosseini Nave H, Mansouri S, Sadeghi A, Moradi M. Molecular diagnosis and anti-microbial resistance patterns among Shigella spp. isolated from patients with diarrhea. *Gastroenterol Hepatol Bed Bench*. 2016;9(3):205–10. [PubMed: 27458513]. [PubMed Central: PMC4947135].
- Mardaneh J, Abbas Poor S, Afrugh P. Prevalence of Shigella species and antimicrobial resistance patterns of isolated strains from infected pediatrics in Tehran. *Int J Enteric Pathog.* 2013;1(1):28–31. doi: 10.17795/ijep10705.
- Ranjbar R, Ghazi FM, Farshad S, Giammanco GM, Aleo A, Owlia P, et al. The occurrence of extended-spectrum beta-lactamase producing Shigella spp. in Tehran, Iran. *Iran J Microbiol.* 2013;5(2):108–12. [PubMed: 23825726]. [PubMed Central: PMC3696844].
- Mamuye Y, Metaferia G, Birhanu A, Desta K, Fantaw S. Isolation and antibiotic susceptibility patterns of shigella and salmonella among under 5 children with acute diarrhoea: A cross-sectional study at selected public health facilities in Addis Ababa, Ethiopia. *Clin Microbiol.* 2015;4(1). doi: 10.4172/2327-5073.1000186.
- Bialvaei AZ, Pourlak T, Aghamali M, Asgharzadeh M, Gholizadeh P, Kafil HS. The Prevalence of CTX-M-15 Extended-spectrum beta-Lactamases Among Salmonella spp. and Shigella spp. Isolated from three Iranian Hospitals. *Eur J Microbiol Immunol (Bp)*. 2017;7(2):133-7. doi:10.1556/1886.2017.00004. [PubMed: 28690880]. [PubMed Central: PMC5495085].
- Abbasi P, Kargar M, Doosti A, Mardaneh J, Ghorbani-Dalini S, Dehyadegari MA. Molecular detection of diffusely adherent escherichia coli strains associated with diarrhea in Shiraz, Iran. Arch Pediatr Infect Dis. 2016;5(2). doi: 10.5812/pedinfect.37629.
- Tille P. Bailey & Scott's Diagnostic Microbiology-E-Book. Elsevier Health Sciences; 2013.
- Clinical and Laboratory Standards Institute . Performance standards for antimicrobial susceptibility testing. 26th Informational Supplement. CLSI document M100-S26. Wayne PA: CLSI; 2016.
- 13. Awole M, Gebre-Selassie S, Kassa T, Kibru G. Isolation of potential bacterial pathogens from the stool of HIV-infected and HIV-non-infected

patients and their antmicrobial susceptibility patterns in Jimma Hospital, south west Ethiopia. *Ethiop Med J.* 2002;**40**(4):353-64. [PubMed: 12596655].

- Reda AA, Seyoum B, Yimam J, Fiseha S, Jean-Michel V. Antibiotic susceptibility patterns of Salmonella and Shigella isolates in Harar, Eastern Ethiopia. J Infect Dis Immun. 2011;3(8):134–9.
- Mache A. Salmonella serogroups and their antibiotic resistance patterns isolated from diarrhoeal stools of pediatric out-patients in Jimma Hospital and Jimma health center, South West Ethiopia. *Ethiop* J Health Sci. 2002;12(1):37–46.
- Tiruneh M. Serodiversity and antimicrobial resistance pattern of Shigella isolates at Gondar University teaching hospital, Northwest Ethiopia. Jpn J Infect Dis. 2009;62(2):93–7. [PubMed: 19305047].
- Samie A, Guerrant RL, Barrett L, Bessong PO, Igumbor EO, Obi CL. Prevalence of intestinal parasitic and bacterial pathogens in diarrhoeal and non-diarroeal human stools from Vhembe district, South Africa. J Health Popul Nutr. 2009;27(6):739–45. [PubMed: 20099757]. [PubMed Central: PMC2928113].
- Jomezadeh N, Babamoradi S, Kalantar E, Javaherizadeh H. Isolation and antibiotic susceptibility of Shigella species from stool samples among hospitalized children in Abadan, Iran. *Gastroenterol Hepatol Bed Bench.* 2014;7(4):218-23. [PubMed: 25289136]. [PubMed Central: PMC4185876].
- Talebreza A, Memariani M, Memariani H, Shirazi MH, Eghbali Shamsabad P, Bakhtiari M. Prevalence and antibiotic susceptibility of shigella species isolated from pediatric patients in Tehran. *Arch Pediatr Infect Dis.* 2015;4(1). doi: 10.5812/pedinfect.32395.
- Njunda AL, Assob JC, Nsagha DS, Kamga HL, Awafong MP, Weledji EP. Epidemiological, clinical features and susceptibility pattern of shigellosis in the buea health district, Cameroon. *BMC Res Notes*. 2012;5:54. doi: 10.1186/1756-0500-5-54. [PubMed: 22264300]. [PubMed Central: PMC3285523].
- Nath R, Saikia L, Choudhury G, Sharma D. Drug resistant Shigella flexneri in & around Dibrugarh, north-east India. *Indian J Med Res.* 2013;137(1):183-6. [PubMed: 23481070]. [PubMed Central: PMC3657885].
- Opintan J, Newman MJ. Distribution of serogroups and serotypes of multiple drug resistant Shigella isolates. *Ghana Med J.* 2007;**41**(1):8–29. [PubMed: 17622331]. [PubMed Central: PMC1890532].
- 23. Hamze M, Osman M, Mallat H, Achkar M. Antibiotic susceptibility of Salmonella spp., Shigella spp. and enteropathogenic Escherichia coli strains isolated from diarrheic children in Tripoli, North Lebanon. *Int Arabic J Antimicrob Agents*. 2016. doi: 10.3823/787.
- MoezArdalan K, Zali MR, Dallal MM, Hemami MR, Salmanzadeh-Ahrabi S. Prevalence and pattern of antimicrobial resistance of Shigella species among patients with acute diarrhoea in Karaj, Tehran, Iran. J Health Popul Nutr. 2003;21(2):96–102. [PubMed: 13677436].
- Tajbakhsh M, Garcia Migura L, Rahbar M, Svendsen CA, Mohammadzadeh M, Zali MR, et al. Antimicrobial-resistant Shigella infections from Iran: an overlooked problem? *J Antimicrob Chemother*. 2012;67(5):1128–33. doi: 10.1093/jac/dks023. [PubMed: 22345385].
- Farshad S, Sheikhi R, Japoni A, Basiri E, Alborzi A. Characterization of Shigella strains in Iran by plasmid profile analysis and PCR amplification of ipa genes. *J Clin Microbiol*. 2006;**44**(8):2879– 83. doi: 10.1128/JCM.00310-06. [PubMed: 16891506]. [PubMed Central: PMC1594633].
- 27. Eftekhari N, Bakhshi B, Pourshafie MR, Zarbakhsh B, Rahbar M, Hajia M, et al. Genetic diversity of Shigella spp. and their integron content. *Foodborne Pathog Dis.* 2013;**10**(3):237-42. doi: 10.1089/fpd.2012.1250. [PubMed: 23489046].
- Ansari S, Sherchand JB, Parajuli K, Mishra SK, Dahal RK, Shrestha S, et al. Bacterial etiology of acute diarrhea in children under five years of age. J Nepal Health Res Counc. 2012;10(22):218–23. [PubMed: 23281455].
- 29. Pazhani GP, Ramamurthy T, Mitra U, Bhattacharya SK, Niyogi SK. Species diversity and antimicrobial resistance of Shigella spp. iso-

lated between 2001 and 2004 from hospitalized children with diarrhoea in Kolkata (Calcutta), India. *Epidemiol Infect.* 2005;**133**(6):1089– 95. doi: 10.1017/S0950268805004498. [PubMed: 16274506]. [PubMed Central: PMC2870343].

- Pourakbari B, Mamishi S, Mashoori N, Mahboobi N, Ashtiani MH, Afsharpaiman S, et al. Frequency and antimicrobial susceptibility of Shigella species isolated in Children Medical Center Hospital, Tehran, Iran, 2001-2006. *Braz J Infect Dis*. 2010;14(2):153-7. doi: 10.1016/S1413-8670(10)70029-5. [PubMed: 20563441].
- Ghosh AR, Sugunan AP, Sehgal SC, Bharadwaj AP. Emergence of nalidixic acid-resistant Shigella sonnei in acute-diarrhea patients on Andaman and Nicobar Islands, India. *Antimicrob Agents Chemother*. 2003;47(4):1483. doi: 10.1128/AAC.47.4.1483.2003. [PubMed: 12654701].

[PubMed Central: PMC152529].

- 32. Jesudason MV. Shigella isolation in Vellore, south India (1997-2001). Indian J Med Res. 2002;115:11-3. [PubMed: 12424931].
- Barari Savadkoohi R, Ahmadpour-Kacho M. Prevalence of Shigella species and their antimicrobial resistance patterns at Amirkola children hospital, North of Iran. *Iran J Pediatr.* 2007;17(2):118–22.
- 34. Sakhaei A, Savari M, Shokoohizadeh L, Hadian M, Ekrami A. Characterization of shigella strains by plasmid profile analysis and antibiotic susceptibility patterns in a pediatric Hospital in Ahvaz. Int J Enteric Pathog. 2015;3(4). doi: 10.17795/ijep29924.
- 35. Mardaneh J, Soltan Dallal MM. Study of cronobacter sakazakii strains isolated from powdered milk infant formula by phenotypic and molecular methods in Iran. *Arch Pediatr Infect Dis.* 2017;**5**(1).