



# Transplacental Transfer Efficiency of SARS-CoV-2-Specific IgG Antibodies in Iranian Seropositive Mothers: A Monocentric Study in Karaj, Iran

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## Abstract

**Background:** Passively acquired maternal antibodies through the placenta and/or breastfeeding may protect neonates against SARS-CoV-2 before they are considered eligible for active immunization.

**Objectives:** This study aimed to determine the concentration and correlation of maternal and cord blood anti-SARS-CoV-2 IgG antibodies in Iranian seropositive women. In this preliminary study, attempts were also made to assess the effects of some variables on the transplacental transfer efficiency.

**Methods:** From September 2021 to November 2021, pregnant women presenting to Kamali Hospital in Karaj, Iran, were enrolled in this monocentric study. The maternal and neonatal demographic characteristics were collected. The maternal and cord blood spike protein-specific antibody levels, measured at delivery (119 paired samples), were evaluated by the enzyme-linked immunosorbent assay (ELISA).

**Results:** Based on the transplacental antibody transfer ratios, the participants were divided into 2 groups: Transfer ratio  $\leq 1$  ( $n = 53$ ) and transfer ratio  $> 1$  ( $n = 66$ ). The results revealed that anti-SARS-CoV-2 IgG antibodies were transferred across the placenta in all seropositive pregnant Iranian women, and a significant correlation was found between maternal and cord blood-specific IgG antibody levels ( $r = 0.9646$ ;  $P < 0.05$ ). A relatively efficient transplacental transfer ratio was observed in Iranian seropositive women (transfer ratio  $> 1$  in 66 out of 119 individuals; 55.46%). Although there were significant differences in the transplacental transfer ratio regarding the neonatal birth weight and maternal body mass index (BMI), more extensive investigations with a larger sample size are needed to confirm the significant association of these variables with the transplacental transfer efficiency of specific SARS-CoV-2 IgG antibodies.

**Conclusions:** The present study highlighted the importance of vaccination during pregnancy to improve neonatal immune responses against SARS-CoV-2.

**Keywords:** COVID-19, Immunization, Maternal-fetal exchange, Neonate, SARS-CoV-2

## 1. Background

Severe acute respiratory syndrome-coronavirus 2 (SARS-CoV-2), the etiological agent of COVID-19, is responsible for one of the most widespread and deadliest pandemics in human history. This pandemic has affected different aspects of human health and has imposed significant burdens on the global economy. Severe acute respiratory syndrome-coronavirus 2 is an enveloped RNA virus that belongs to the subgenus *Sarbecovirus*, genus

*Betacoronavirus*, and the family *Coronaviridae* (1). Several SARS-CoV-2 variants have been characterized during this pandemic (2). It seems that the hospitalization rate of young children and infants infected with the last dominant and most heavily mutated variant was higher than that of individuals infected with other variants (3), probably due to its higher transmissibility rate. In addition, the risk of COVID-19 mortality is higher in pregnant women (4).

The transplacental transfer of maternal IgG antibodies to an unborn baby is a specific adaptive mechanism for providing protection against some pathogens (5), especially viruses and bacterial toxins. Although, according to several studies, either maternal COVID-19 infection or vaccination can result in the transplacental transfer of SARS-CoV-2-specific antibodies (6-17), different parameters, such as the altered glycosylation profile of antibodies and timing of vaccination/infection during pregnancy, may affect the efficiency of transplacentally acquired immunity (13, 15, 16). Additionally, the lower transplacental transfer ratio of specific and/or total IgG has been reported in certain areas around the world (18-20).

Moreover, previous studies have indicated the differential efficiency of the transplacental transfer ratio in distinct IgG subclasses (21). It may be attributed to the binding affinity to the neonatal Fc receptor (FcRn), which is strongly expressed in the endosomal vesicles of placental syncytiotrophoblasts (22). On the other hand, reports suggest that the serum IgG subclass concentrations are influenced by ethnicity (23). Thus, ethnicity may affect the transplacental antibody transfer efficiency.

## 2. Objectives

This study aimed to assess the placental transfer ratio of antibodies in Iranian seropositive women. Further attempts were also made to assess the effects of some maternal and neonatal variables on the transplacental antibody transfer efficiency.

## 3. Methods

### 3.1. Study Population

From September 2021 to November 2021, pregnant women presenting to Kamali Hospital in Karaj, Iran, were enrolled in this monocentric study. Maternal venous blood and cord blood samples were collected directly after delivery. The census method was used, and all 119 paired samples were collected from Iranian seropositive pregnant women. The most salient inclusion criteria for eligible pregnant women were a SARS-CoV-2 seropositive result and the ability to understand oral information (in Persian). The participants' demographic characteristics, including maternal age at delivery, maternal education, maternal weight (body mass index [BMI]), maternal blood group, neonatal sex, and neonatal birth weight, are reported in Table 1.

**Table 1.** The Demographics and Characteristics of Women and Their Neonates Included in the Study

Demographics/Characteristics	Values
<b>Mothers (n=112)</b>	
Maternal age at delivery (y); mean $\pm$ SD (range)	30.9 $\pm$ 6.7 (13 - 47)
<b>Maternal education; No. (%)</b>	
Illiterate	10 (8.9)
< Grade 12	44 (39.3)
High school	42 (37.5)
Academic education	16 (14.3)
Maternal BMI, mean $\pm$ SD (range)	26.6 $\pm$ 4.4 (17.1 - 41.5)
<b>Maternal blood group; No. (%) (missing, n = 21)</b>	
A	35 (38.5)
B	19 (20.9)
AB	6 (6.6)
O	31 (34)
<b>Neonates (n = 119)</b>	
Neonatal birth weight (g); mean $\pm$ SD (range)	3023.4 $\pm$ 562.8 (900 - 42,000)
<b>Neonatal sex, No. (%)</b>	
Male	56 (47)
Female	63 (53)

Abbreviation: BMI, body mass index.

### 3.2. Antibody Measurements

The concentration of anti-SARS-CoV-2 IgG in the maternal and cord blood samples was measured using the EUROIMMUN Anti-SARS-CoV-2 ELISA (IgG) kit, according to the manufacturer's instructions. The results were considered protective if the OD ratio exceeded 0.8. If the corresponding ratio exceeded the upper limit of the analytical measurement range, the result was reported as  $>10$ . Based on the transplacental antibody transfer ratios, the participants were divided into 2 groups: transfer ratio  $\leq 1$  (n = 53) and transfer ratio  $> 1$  (n = 66).

### 3.3. Statistical Analysis

Correlations between the maternal and cord blood antibody concentrations were evaluated using Pearson's correlation test, with the corresponding r and P-values. The transplacental antibody transfer ratio was calculated as the cord blood antibody concentration divided by the maternal antibody concentration. The chi-square test and t-test were used as appropriate to compare the demographic characteristics, antibody levels, and transplacental transfer ratios. The level of statistical

significance was set at  $P < 0.05$ . Analysis was performed using STATA version 16.1 (StataCorp, College Station, TX, USA).

#### 4. Results

As shown in Figure 1, strong positive correlations were found regarding the concentration of SARS-CoV-2-specific IgG antibodies between the maternal plasma and cord blood samples in all pairs ( $r = 0.9646$ ;  $P < 0.05$ ), indicating the adequate level of transplacental transfer antibodies.

The antibody concentrations in neonates born to mothers with high antibody titer levels also increased. Among all maternal-cord blood pairs of Iranian seropositive women, only 5% had a very low transfer ratio ( $< 0.5$ ). Efficient transplacental transfer ratios (transfer ratio  $> 1$ ) were observed in 55.46% of cases.

As reported earlier, the participants were divided into 2 groups based on the transplacental antibody transfer ratio (ratio  $\leq 1$  [ $n = 53$ ] and ratio  $> 1$  [ $n = 66$ ]). Some variables, including maternal age at delivery, maternal BMI, neonatal birth weight, and neonatal sex, were compared between the 2 groups (Table 2).

**Table 2.** The Association of Placental Transfer Ratio and the Characteristics of the Participants ( $n = 119$ )

Characteristics	Transfer Ratio		P-Value <sup>a</sup>
	$\leq 1$ ( $n = 53$ )	$> 1$ ( $n = 66$ )	
Maternal age at delivery (y); mean $\pm$ SD	30.9 $\pm$ 6.7	30.9 $\pm$ 6.6	0.95
Maternal BMI; mean $\pm$ SD	27.7 $\pm$ 5.1	25.7 $\pm$ 4.3	0.02 <sup>b</sup>
Neonatal birth weight (g); mean $\pm$ SD	2874.5 $\pm$ 826.4	3138.5 $\pm$ 573.8	0.04 <sup>b</sup>
Neonatal sex; No. (%)			0.30
Male	28 (52.8)	28 (42.5)	
Female	25 (47.2)	38 (57.5)	

Abbreviation: BMI, body mass index.

<sup>a</sup> P-values for continuous variables are determined by the *t*-test, and P-values for categorical variables are determined by the chi-square test.

<sup>b</sup>  $P < 0.05$ .

There was a significant difference in the transplacental transfer ratio regarding neonatal birth weight and maternal BMI ( $P < 0.05$ ). Conversely, maternal age and neonatal sex did not affect the placental antibody transfer.

#### 5. Discussion

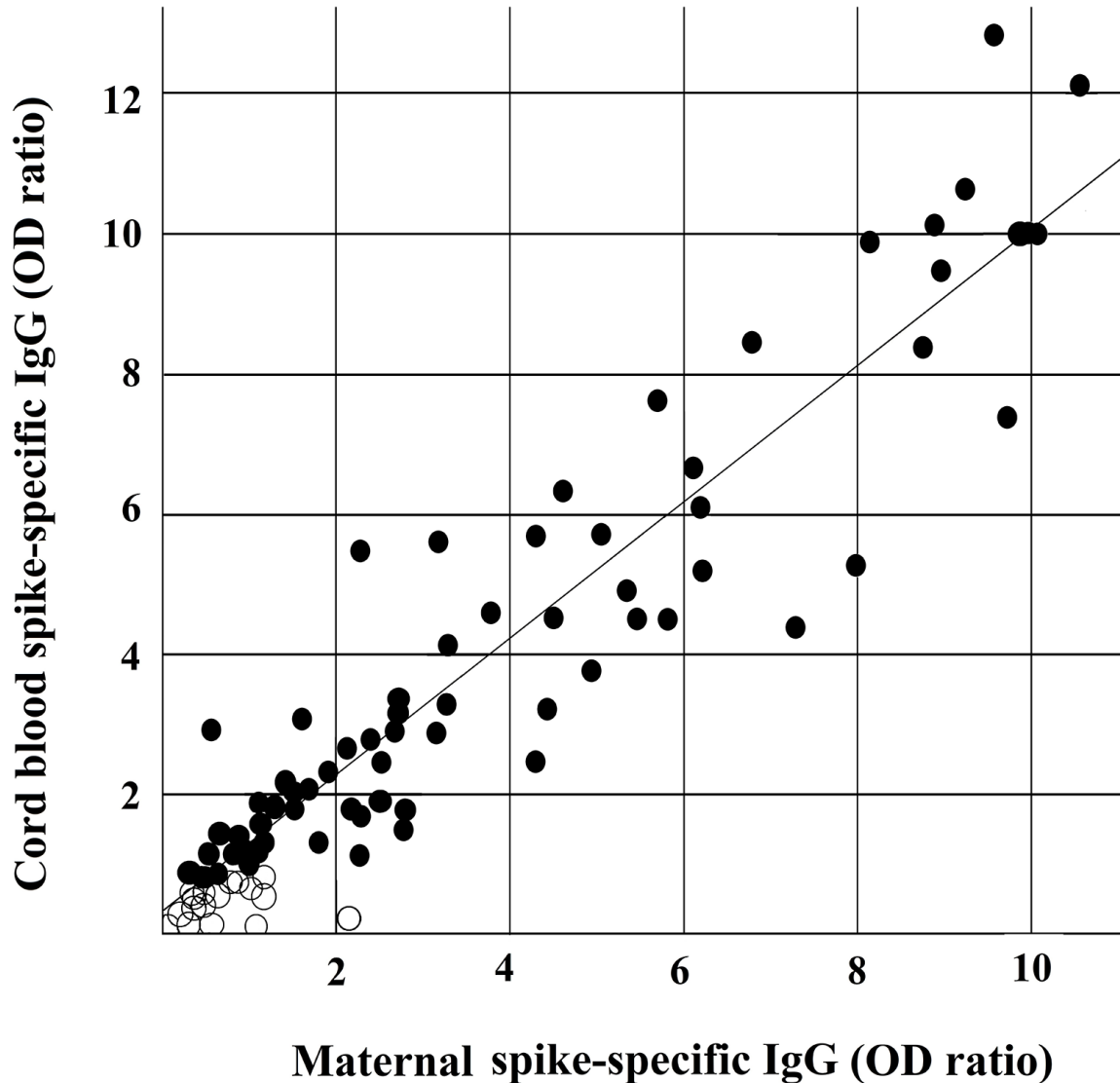
The EUROIMMUN Anti-SARS-CoV-2 ELISA (IgG) assay was used to analyze spike-specific antibodies collected from maternal and cord blood samples upon delivery. Previous studies, based on comparisons with the gold

standard assay (microneutralization), have shown that this serological kit is suitable for evaluating protective immunity following infection or vaccination (24, 25). This standardized, commercially available kit is based on a serological test to detect specific anti-spike (S1) antibodies. It is known that the spike protein mediates viral attachment and entry into target cells (1). It is a viral fusion protein that contributes to the merging of the viral envelope with the cell membranes, allowing for the release of viral genomes into the cell cytoplasm (1).

In this study, spike-specific SARS-CoV-2 antibodies were measured in 119 maternal-cord blood pairs. The results showed that the level of protective SARS-CoV-2-specific antibodies in newborns depended on the concentration of maternal antibodies, as reinforced by the results of several other studies in different parts of the world (6-17). Besides, relatively efficient transplacental transfer ratios (transfer ratio  $> 1$  in 66 out of 119 subjects [55.46%]) were observed in Iranian seropositive women. Similarly, a previous study examining 72 seropositive mother-infant pairs from 4 other race/ethnicity groups reported an efficient transplacental IgG transfer (transfer ratio  $\geq 1$ ) in 55.5% of cases (8). Notably, the transplacental transfer ratios (range, 0.8-1.7) were similar to those reported for vaccine-induced antibodies to other pathogens, such as influenza and pertussis (26, 27). The present findings highlighted the importance of vaccination during pregnancy to improve neonatal immune responses to SARS-CoV-2.

Currently, several other vaccines, such as tetanus toxoid, tetanus diphtheria acellular pertussis diphtheria (Tdap) vaccine, inactivated influenza vaccine, inactivated polio vaccine (IPV), pneumococcal polysaccharide vaccine, meningococcal polysaccharide vaccine, hepatitis A vaccine, hepatitis B vaccine, and inactivated rabies vaccine, are administered for routine use or under special circumstances to protect women and their offspring by placental transmission of maternal antibodies. Alternatively, infected/vaccinated pregnant women may transfer protective antibodies to newborn infants via breastfeeding after birth. Specific SARS-CoV-2 IgA antibodies have also been detected in the breast milk of seropositive mothers (28). Moreover, the detection of specific antibodies in the neonates' blood confirmed the acquired passive immunity (29-32).

The results of this preliminary study showed significant differences in the transplacental transfer ratio with respect to the neonatal birth weight and maternal BMI. It has been previously observed that low-birth-weight neonates have lower passive total and/or specific IgG levels because of an impaired transplacental transfer (33, 34). Moreover, a previous study showed that maternal obesity could affect the turnover of placental



**Figure 1.** Correlations between maternal and matched cord blood IgG concentrations. Each dot represents data from a single pair of maternal and cord blood antibodies to the anti-spike domain of the S protein of SARA-CoV-2 ( $r = 0.9646$ ;  $P < 0.05$ ). Protective levels ( $> 0.8$ ) are represented by filled circles, and insufficient protective levels ( $\leq 0.80$ ) are represented by open circles.

cells, such as syncytiotrophoblasts (35). Overall, the IgG transfer from the mother to the offspring occurs across syncytiotrophoblasts (5). Accordingly, maternal BMI may affect the efficiency of transplacental antibody transfer. However, further extensive studies with a larger sample size are needed to confirm the significant association between these variables and the transplacental transfer efficiency of specific SARS-CoV-2 IgG antibodies.

To the best of our knowledge, only 1 recently published study in Iran has examined the levels of maternal and

cord-blood antibodies. However, this study only involved 23 pregnant women who received single or 2 doses of the BBIBP-CorV (Sinopharm) vaccine (36). Based on reports, Iran has reported millions of confirmed COVID-19 (98% recovered cases and 2% deaths). Until today, millions of COVID-19 vaccine doses have been administered in Iran, and approximately most of the population is fully vaccinated. Additionally, pregnant women were not excluded from the vaccination program in Iran. Therefore, many newborns in Iran, especially those born to infected

and/or vaccinated mothers, may acquire some maternal antibodies. However, future research needs to concentrate on the durability of passively acquired immunity in Iranian neonates.

### 5.1. Conclusions

Although there were significant differences in the transplacental transfer ratio regarding the neonatal birth weight and maternal BMI, more extensive investigations with a larger sample size are needed to confirm the significant association of these variables with the transplacental transfer efficiency of specific SARS-CoV-2 IgG antibodies. The present study supports previous studies conducted worldwide, demonstrating that vaccination during pregnancy is highly effective in enhancing neonatal immune responses against SARS-CoV-2.

### 5.2. Limitations

This study has several limitations. The most significant limitation is its small sample size. Therefore, further studies with a larger sample size are needed to confirm the significant association between variables and the transplacental transfer efficiency of specific SARS-CoV-2 IgG antibodies. Additionally, no demographically similar seronegative pregnant women were included as a control cohort. Last but not least, performing assays at different times might have affected the measurement of antibody levels.

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### Footnotes

**Authors' Contribution:** Study conception/design and scientific supervision were provided by AM and MF. Material preparation, experimental work, and data collection were performed by DB, RS, FE, and BA. Data and statistical analysis was carried out by MP. The first draft of the manuscript and preparation of figures were performed by BA. All authors read, commented, and approved the final manuscript.

**Conflict of Interests:** There is no conflict of interest in this study to declare.

**Data Reproducibility:** The dataset presented in the study is available on request from the corresponding author during submission or after publication.

**Ethical Approval:** This study was approved by the Research Ethics Committee of Alborz University of Medical Sciences under the ethical code of [IR.ABZUMS.REC.1400.294](https://doi.org/10.1038/s41579-020-00459-7).

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### References

- Hu B, Guo H, Zhou P, Shi ZL. Characteristics of SARS-CoV-2 and COVID-19. *Nat Rev Microbiol*. 2021;**19**(3):141–54. [PubMed ID: 33024307]. [PubMed Central ID: PMC7537588]. <https://doi.org/10.1038/s41579-020-00459-7>.
- Rathinasamy M, Kandhasamy S. An exploratory study on the propagation of SARS-CoV-2 variants: Omicron is the most predominant variant. *J Med Virol*. 2022;**94**(6):2414–21. [PubMed ID: 35112734]. [PubMed Central ID: PMC9015234]. <https://doi.org/10.1002/jmv.27634>.
- Cloete J, Kruger A, Masha M, du Plessis NM, Mawela D, Tshukudu M, et al. Paediatric hospitalisations due to COVID-19 during the first SARS-CoV-2 omicron (B.1.1.529) variant wave in South Africa: a multicentre observational study. *Lancet Child Adolesc Health*. 2022;**6**(5):294–302. [PubMed ID: 35189083]. [PubMed Central ID: PMC8566663]. [https://doi.org/10.1016/S2352-4642\(22\)00027-X](https://doi.org/10.1016/S2352-4642(22)00027-X).
- Mardani M, Pourkaveh B. COVID-19 Vaccination Considerations in Pregnant Women. *Arch Clin Infect Dis*. 2021;**16**(1). <https://doi.org/10.5812/archcid.115218>.
- Fouda GG, Martinez DR, Swamy GK, Permar SR. The Impact of IgG transplacental transfer on early life immunity. *Immunohorizons*. 2018;**2**(1):14–25. [PubMed ID: 29457151]. [PubMed Central ID: PMC5812294]. <https://doi.org/10.4049/jimmunohorizons.1700057>.
- Atyeo C, Pullen KM, Bordt EA, Fischinger S, Burke J, Michell A, et al. Compromised SARS-CoV-2-specific placental antibody transfer. *Cell*. 2021;**184**(3):628–642 e10. [PubMed ID: 33476549]. [PubMed Central ID: PMC7755577]. <https://doi.org/10.1016/j.cell.2020.12.027>.
- Paul G, Chad R. Newborn antibodies to SARS-CoV-2 detected in cord blood after maternal vaccination - a case report. *BMC Pediatr*. 2021;**21**(1):138. [PubMed ID: 33752624]. [PubMed Central ID: PMC7982334]. <https://doi.org/10.1186/s12887-021-02618-y>.
- Flannery DD, Gouma S, Dhudasia MB, Mukhopadhyay S, Pfeifer MR, Woodford EC, et al. Assessment of Maternal and Neonatal Cord Blood SARS-CoV-2 Antibodies and Placental Transfer Ratios. *JAMA Pediatr*. 2021;**175**(6):594–600. [PubMed ID: 33512440]. [PubMed Central ID: PMC7846944]. <https://doi.org/10.1001/jamapediatrics.2021.0038>.
- Wang X, Yang P, Zheng J, Liu P, Wei C, Guo J, et al. Dynamic changes of acquired maternal SARS-CoV-2 IgG in infants. *Sci Rep*. 2021;**11**(1):8021. [PubMed ID: 33850202]. [PubMed Central ID: PMC8044122]. <https://doi.org/10.1038/s41598-021-87535-x>.
- Joseph NT, Dude CM, Verkerke HP, Irby LS, Dunlop AL, Patel RM, et al. Maternal Antibody Response, Neutralizing Potency, and Placental Antibody Transfer After Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) Infection. *Obstet Gynecol*. 2021;**138**(2):189–97. [PubMed ID: 33910220]. [PubMed Central ID: PMC8288196]. <https://doi.org/10.1097/AOG.0000000000004440>.
- Shen CJ, Fu YC, Lin YP, Shen CF, Sun DJ, Chen HY, et al. Evaluation of Transplacental Antibody Transfer in SARS-CoV-2-Immunized Pregnant Women. *Vaccines (Basel)*. 2022;**10**(1). [PubMed ID: 35062762]. [PubMed Central ID: PMC8778956]. <https://doi.org/10.3390/vaccines10010101>.



12. Beharier O, Plitman Mayo R, Raz T, Nahum Sacks K, Schreiber L, Suissa-Cohen Y, et al. Efficient maternal to neonatal transfer of antibodies against SARS-CoV-2 and BNT162b2 mRNA COVID-19 vaccine. *J Clin Invest*. 2021;**131**(13). [PubMed ID: 34014840]. [PubMed Central ID: PMC8245182]. <https://doi.org/10.1172/JCI150319>.
13. Song D, Prahl M, Gaw SL, Narasimhan SR, Rai DS, Huang A, et al. Passive and active immunity in infants born to mothers with SARS-CoV-2 infection during pregnancy: prospective cohort study. *BMJ Open*. 2021;**11**(7). e053036. [PubMed ID: 34234001]. [PubMed Central ID: PMC8264915]. <https://doi.org/10.1136/bmjopen-2021-053036>.
14. Kashani-Ligumsky L, Lopian M, Cohen R, Senderovich H, Czeiger S, Halperin A, et al. Titers of SARS CoV-2 antibodies in cord blood of neonates whose mothers contracted SARS CoV-2 (COVID-19) during pregnancy and in those whose mothers were vaccinated with mRNA to SARS CoV-2 during pregnancy. *J Perinatol*. 2021;**41**(11):2621-4. [PubMed ID: 34564695]. [PubMed Central ID: PMC8475451]. <https://doi.org/10.1038/s41372-021-01216-1>.
15. Rottenstreich A, Zarbiv G, Oiknine-Djian E, Zigran R, Wolf DG, Porat S. Efficient Maternofetal Transplacental Transfer of Anti-Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) Spike Antibodies After Antenatal SARS-CoV-2 BNT162b2 Messenger RNA Vaccination. *Clin Infect Dis*. 2021;**73**(10):1909-12. [PubMed ID: 33822014]. [PubMed Central ID: PMC8083549]. <https://doi.org/10.1093/cid/ciab266>.
16. Rottenstreich A, Zarbiv G, Oiknine-Djian E, Vorontsov O, Zigran R, Kleinstern G, et al. Timing of SARS-CoV-2 vaccination during the third trimester of pregnancy and transplacental antibody transfer: a prospective cohort study. *Clin Microbiol Infect*. 2022;**28**(3):419-25. [PubMed ID: 34740773]. [PubMed Central ID: PMC8563509]. <https://doi.org/10.1016/j.cmi.2021.10.003>.
17. Trostle ME, Agüero-Rosenfeld ME, Roman AS, Lighter JL. High antibody levels in cord blood from pregnant women vaccinated against COVID-19. *Am J Obstet Gynecol*. 2021;**3**(6):100481. [PubMed ID: 34562636]. [PubMed Central ID: PMC8455300]. <https://doi.org/10.1016/j.ajogmf.2021.10.0481>.
18. Hartter HK, Oyedele OI, Dietz K, Kreis S, Hoffman JP, Muller CP. Placental transfer and decay of maternally acquired antimeasles antibodies in Nigerian children. *Pediatr Infect Dis J*. 2000;**19**(7):635-41. [PubMed ID: 10917222]. <https://doi.org/10.1097/00006454-200007000-00010>.
19. Englund JA. The influence of maternal immunization on infant immune responses. *J Comp Pathol*. 2007;**137** Suppl 1:S16-9. [PubMed ID: 17553516]. <https://doi.org/10.1016/j.jcpa.2007.04.006>.
20. Okoko BJ, Wesumperuma LH, Ota MO, Pinder M, Banya W, Gomez SF, et al. The influence of placental malaria infection and maternal hypergammaglobulinemia on transplacental transfer of antibodies and IgG subclasses in a rural West African population. *J Infect Dis*. 2001;**184**(5):627-32. [PubMed ID: 11494168]. <https://doi.org/10.1086/322808>.
21. Clements T, Rice TF, Vamvakas G, Barnett S, Barnes M, Donaldson B, et al. Update on Transplacental Transfer of IgG Subclasses: Impact of Maternal and Fetal Factors. *Front Immunol*. 2020;**11**:1920. [PubMed ID: 33013843]. [PubMed Central ID: PMC7516031]. <https://doi.org/10.3389/fimmu.2020.01920>.
22. Abdiche YN, Yeung YA, Chaparro-Riggers J, Barman I, Strop P, Chin SM, et al. The neonatal Fc receptor (FcRn) binds independently to both sites of the IgG homodimer with identical affinity. *MAbs*. 2015;**7**(2):331-43. [PubMed ID: 25658443]. [PubMed Central ID: PMC4622529]. <https://doi.org/10.1080/19420862.2015.1008353>.
23. Gunsolley JC, Pandey JP, Quinn SM, Tew J, Schenkein HA. The effect of race, smoking and immunoglobulin allotypes on IgG subclass concentrations. *J Periodontol Res*. 1997;**32**(4):381-7. [PubMed ID: 9210092]. <https://doi.org/10.1111/j.1600-0765.1997.tb00548.x>.
24. Weidner L, Gansdorfer S, Unterweger S, Weseslindtner L, Drexler C, Farcet M, et al. Quantification of SARS-CoV-2 antibodies with eight commercially available immunoassays. *J Clin Virol*. 2020;**129**:104540. [PubMed ID: 32652475]. [PubMed Central ID: PMC7336952]. <https://doi.org/10.1016/j.jcv.2020.104540>.
25. Harvala H, Robb ML, Watkins N, Ijaz S, Dicks S, Patel M, et al. Convalescent plasma therapy for the treatment of patients with COVID-19: Assessment of methods available for antibody detection and their correlation with neutralising antibody levels. *Transfus Med*. 2021;**31**(3):167-75. [PubMed ID: 33333627]. [PubMed Central ID: PMC8246874]. <https://doi.org/10.1111/tme.12746>.
26. Munoz FM, Patel SM, Jackson LA, Swamy GK, Edwards KM, Frey SE, et al. Safety and immunogenicity of three seasonal inactivated influenza vaccines among pregnant women and antibody persistence in their infants. *Vaccine*. 2020;**38**(33):5355-63. [PubMed ID: 32571718]. <https://doi.org/10.1016/j.vaccine.2020.05.059>.
27. Post AL, Li SH, Berry M, Itell H, Martinez DR, Xie G, et al. Efficiency of placental transfer of vaccine-elicited antibodies relative to prenatal Tdap vaccination status. *Vaccine*. 2020;**38**(31):4869-76. [PubMed ID: 32482459]. [PubMed Central ID: PMC7388058]. <https://doi.org/10.1016/j.vaccine.2020.05.036>.
28. Low JM, Low YW, Zhong Y, Lee CYC, Chan M, Ng NBH, et al. Titres and neutralising capacity of SARS-CoV-2-specific antibodies in human milk: a systematic review. *Arch Dis Child Fetal Neonatal Ed*. 2022;**107**(2):174-80. [PubMed ID: 34257103]. <https://doi.org/10.1136/archdischild-2021-322156>.
29. Zeng H, Xu C, Fan J, Tang Y, Deng Q, Zhang W, et al. Antibodies in Infants Born to Mothers With COVID-19 Pneumonia. *JAMA*. 2020;**323**(18):1848-9. [PubMed ID: 32215589]. [PubMed Central ID: PMC7099444]. <https://doi.org/10.1001/jama.2020.4861>.
30. Dong L, Tian J, He S, Zhu C, Wang J, Liu C, et al. Possible Vertical Transmission of SARS-CoV-2 From an Infected Mother to Her Newborn. *JAMA*. 2020;**323**(18):1846-8. [PubMed ID: 32215581]. [PubMed Central ID: PMC7099527]. <https://doi.org/10.1001/jama.2020.4621>.
31. Cavaliere AF, Marchi L, Aquilini D, Brunelli T, Vasarri PL. Passive immunity in newborn from SARS-CoV-2-infected mother. *J Med Virol*. 2021;**93**(3):1810-3. [PubMed ID: 33073377]. <https://doi.org/10.1002/jmv.26609>.
32. Mo H, Wang M, Wang M, Han Y, Zhang Y, Hu K. Detectable antibodies against SARS-CoV-2 in newborns from mothers infected with COVID-19 at different gestational ages. *Pediatr Neonatol*. 2021;**62**(3):321-3. [PubMed ID: 33836982]. [PubMed Central ID: PMC7997304]. <https://doi.org/10.1016/j.pedneo.2021.03.011>.
33. Okoko BJ, Wesumperuma HL, Fern J, Yamuah LK, Hart CA. The transplacental transfer of IgG subclasses: influence of prematurity and low birthweight in the Gambian population. *Ann Trop Paediatr*. 2002;**22**(4):325-32. [PubMed ID: 12530282]. <https://doi.org/10.1179/027249302125001985>.
34. Costa-Carvalho BT, Vieria HM, Dimantas RB, Arslanian C, Naspiat CK, Sole D, et al. Transfer of IgG subclasses across placenta in term and preterm newborns. *Braz J Med Biol Res*. 1996;**29**(2):201-4. [PubMed ID: 8731349].
35. Higgins L, Mills TA, Greenwood SL, Cowley EJ, Sibley CP, Jones RL. Maternal obesity and its effect on placental cell turnover. *J Matern Fetal Neonatal Med*. 2013;**26**(8):783-8. [PubMed ID: 23270521]. <https://doi.org/10.3109/14767058.2012.760539>.
36. Hantoushzadeh S, Eshraghi N, Younesi S, Salehi M, Rezaei N, Hasheminejad MM, et al. Cord blood antibodies following BBIBP-CorV (Sinopharm) vaccination during pregnancy. *Immun Inflamm Dis*. 2023;**11**(6). e874. [PubMed ID: 37382259]. [PubMed Central ID: PMC1026641]. <https://doi.org/10.1002/iid3.874>.