



The Association Between the Gut Microbiome and COVID-19 Severity: The Potential Role of TMAO Produced by the Gut Microbiome

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Abstract

Context: The COVID-19 pandemic has had profound impacts on public health, resulting in nearly 1 million deaths. Emerging evidence suggests an association between certain metabolites produced by gut microbiota and potential alterations in the severity of infection. Trimethylamine N-oxide (TMAO) is a waste metabolite generated by gut microbes from dietary choline and betaine.

Evidence Acquisition: Several investigations have indicated an association between serum TMAO concentrations and the development of inflammation and thrombosis. Trimethylamine N-oxide, produced by the gut microbiome in a state of dysbiosis, upregulates various molecular mechanisms, such as the nuclear factor kappa (NF- κ B) molecular pathway, and promotes the expression of scavenger receptors (SRs) on the surfaces of macrophages, leading to foam cell formation and inflammation. High levels of TMAO have been shown to induce the expression of pro-inflammatory cytokines like tumor necrosis factor-alpha (TNF- α) and interleukin 1 β (IL-1 β) while reducing the expression of anti-inflammatory cytokines such as interleukin-10 (IL-10). Additionally, gut-derived TMAO enhances platelet aggregation and adhesion to collagen, increasing the risk of thrombosis.

Conclusions: Understanding the association between gut microbiome compositions such as gut TMAO and their effects on SARS-CoV-19 infection progression helps to control disease severity. In this review, we presented a hypothesis that the gut TMAO has the potential to increase COVID-19 disease severity.

Keywords: COVID-19, TMAO-Gut Microbiota, Thrombosis, Inflammation, Probiotics

1. Context

There is a growing body of evidence indicating that microbial organisms in the gut, collectively referred to as the microbiota, play a crucial role in the metabolic processes of their host (1). These processes include the activation of vitamins (2), support for immune function, and maintenance of intestinal health (3). Recent studies have also highlighted the association between the microbiota and various diseases, such as cardiovascular diseases (4), insulin resistance, obesity, and autoimmune diseases (5, 6), among others.

Furthermore, research has shed light on the role of choline metabolism pathways carried out by the gut microbiome in contributing to the pathogenesis

of various disorders, as observed in both animal and human studies (7). Choline, a primary component of phosphatidylcholine, undergoes metabolism by the gut microbiome, resulting in the formation of an intermediate compound known as trimethylamine (TMA). Trimethylamine is subsequently oxidized within the intestine by microorganisms or transported to the liver, where hepatic flavin monooxygenases oxidize it further to produce trimethylamine N-oxide (TMAO). While TMAO was previously considered a waste metabolite with no significant biological effects, recent evidence strongly suggests an association between TMAO and inflammatory mechanisms (8, 9), atherosclerosis (10), thrombosis (11), and various pathological conditions.

Moreover, it has come to light that TMAO may increase the expression of scavenger receptors (SRs) (12). Among these receptors, SR-B1 is of particular interest as it has multifunctional roles, including facilitating the entry and efflux of cholesterol esters derived from high-density lipoproteins (HDL) into cells and tissues. Interestingly, SR-B1 has also been implicated in the entry of different viruses, including SARS-CoV-2, into host cells (13). A recent study by Wei et al. suggested that SR-B1 may enhance the uptake of SARS-CoV-2 and be associated with disease severity (12). These findings raise the possibility of an association between the presence of SR-B1 and the severity of COVID-19.

At the end of 2019, a novel coronavirus, severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), was identified in China, leading to the emergence of COVID-19 (14). The virus quickly spread globally, and in March 2020, the World Health Organization (WHO) declared it a pandemic. Since then, over 29.5 million individuals have contracted COVID-19 worldwide, resulting in approximately 1 million deaths (15). COVID-19 manifests with a range of symptoms, from mild to severe respiratory tract diseases, often accompanied by serious complications affecting various organs, potentially leading to organ failure and fatalities. Approximately 20% of COVID-19 patients develop severe complications, including acute immune responses characterized by the overproduction of inflammatory cytokines, commonly referred to as a cytokine storm, followed by respiratory distress syndrome (16). Other significant complications include hypercoagulation, increasing the risk of thrombosis, and digestive symptoms, such as diarrhea, which have been observed in some patients (17).

Numerous investigations have explored the relationship between gut microbiome metabolites and COVID-19. A recent study conducted in China revealed a positive correlation between the levels of *Lactobacillus* species in the gut and a more favorable prognosis in COVID-19 patients. This association was linked to an increase in the levels of interleukin-10 (IL-10) (18). Conversely, some pro-inflammatory species, such as *Klebsiella*, *Streptococcus*, and *Ruminococcus gnavus*, have been found to promote the production of pro-inflammatory cytokines, potentially exacerbating the severity of COVID-19 (19). Additionally, Esposito et al. documented changes in the gut microbiome of children with Kawasaki disease compared to healthy children. In their study, they observed higher levels of *Streptococcus* species relative to *Lactobacillus*, indicating dysbiosis in the gut microbiome of children with Kawasaki disease (20). These findings highlight the importance of understanding the relationship between specific

metabolites and compositions produced by the gut microbiome and the pathogenesis and progression of COVID-19. Such insights can identify potential targets for drug development and intervention strategies aimed at reducing morbidity and mortality.

The goal of this review was to review the different mechanisms by which TMAO produced by the gut microbiome is associated with the promotion of inflammation mechanisms, thrombosis, and expression of SR-B1 associated with COVID-19 infection development. We hypothesize that TMAO produced by the gut microbiome may increase the severity of COVID-19.

2. Evidence Acquisition

2.1. Metabolism of TMAO

The exact biological metabolism of TMAO in humans remains poorly understood. Trimethylamine N-oxide is a waste product resulting from the oxidation of TMA, which is produced by the gut microbiome when it metabolizes choline-containing compounds found in the diet, such as choline, L-carnitine, and betaine (21). Various enzymes, including betaine reductase, choline TMA lyase, and carnitine TMA lyase, play a role in converting these choline-containing products into TMA within the intestinal lumen (22). Trimethylamine is then either oxidized by microorganisms within the gut through TMA monooxygenase or transported to the liver, where it undergoes oxidation by flavin monooxygenase (23) (Figure 1). These findings suggest a potential link between dietary and chemical metabolisms mediated by the gut microbiome, influencing various aspects of biological pathways.

2.2. The Association Between TMAO Secreted by Gut Microbiome and Inflammation

Several studies have indicated an association between TMAO dependent on gut microbiota and systemic inflammation (8): Gut-derived TMAO upregulates various molecular mechanisms associated with inflammation. Some studies have demonstrated that TMAO, originating from the gut microbiome, can enhance the expression of heat shock proteins (HSPs), which may contribute to the abnormal activation of macrophages involved in foam cell formation (24). Trimethylamine N-oxide can induce the expression of different proteins, such as HSP60 and GRP78, responsible for endoplasmic reticulum stress induction (25). Furthermore, TMAO stimulates the expression of SRs on the surface of macrophages, thereby contributing to the uptake of oxidized low-density lipoprotein (ox-LDL) and foam cell formation (26, 27). Scavenger receptors

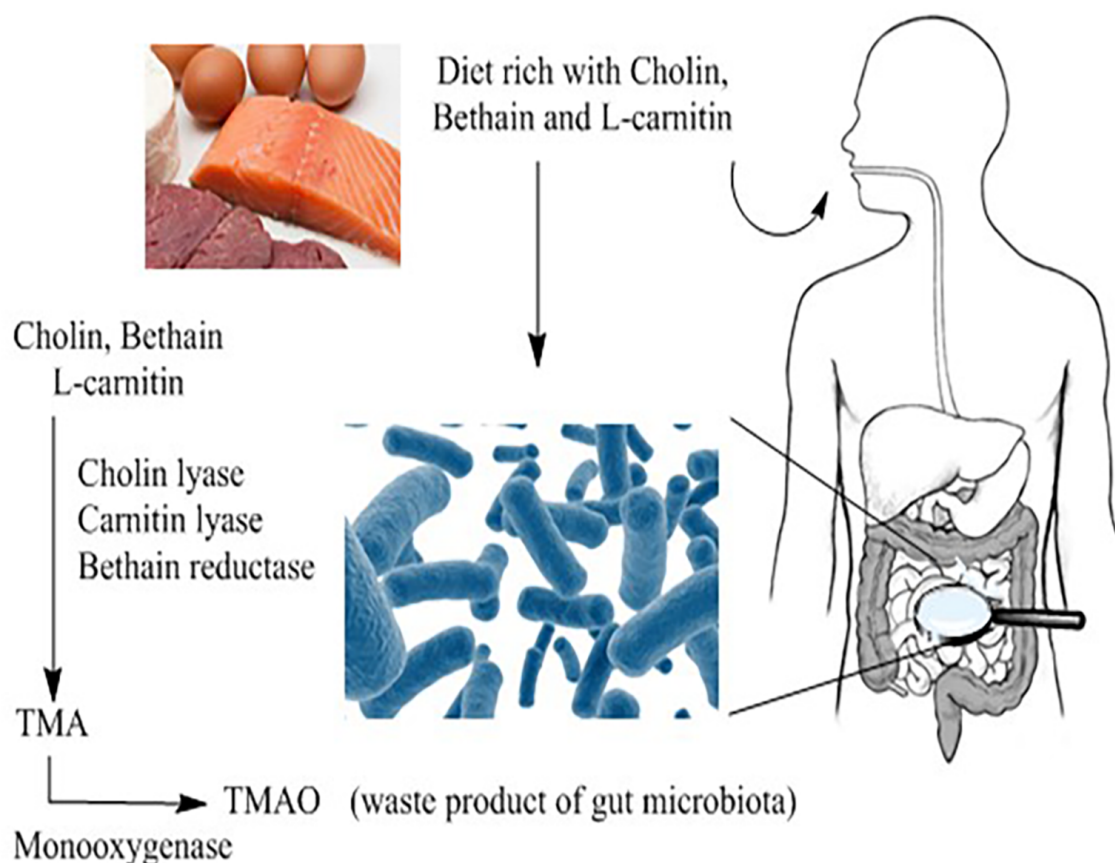


Figure 1. A schematic representation of the metabolic pathway for TMAO (trimethylamine N-oxide) formation by gut microbes from dietary sources

induce the expression of SR-A and CD36 on the surface of macrophages, which can identify ox-LDL molecules. Wei et al. have indicated that the inhibition of TMAO production through antibiotic therapy can reduce the number of macrophages and foam cell formation in mice (12). Moreover, several investigations have reported that TMAO increases CD36 expression and foam cell formation by inducing MAPK and c-Jun N-terminal kinase (JNK) signaling pathways (28, 29). Geng et al. have shown that using MAPK and JNK pathway inhibitors can reduce foam cell formation by decreasing CD36 expression (28).

In addition, multiple studies have demonstrated that higher levels of TMAO increase the expression of pro-inflammatory cytokines such as tumor necrosis factor- α (TNF- α) and interleukin 1beta (IL1 β) while downregulating the expression of the anti-inflammatory cytokine IL-10 (8). Chuo et al. have shown a positive association between TMAO levels, IL-1 β , and high-sensitivity C-reactive protein (hs-CRP) (30). In vitro studies conducted on cultured endothelial

progenitor cells (EPCs) have reported that gut-derived TMAO induces cellular inflammation and oxidative stress (30). The nuclear factor-kappa B (NF- κ B) pathway plays a crucial role in the expression of pro-inflammatory genes (31). Seldin et al. have demonstrated that increasing TMAO levels through a choline-rich diet in mice enhances the expression of inflammatory genes via an effect on the NF- κ B pathway (32). They found that gut-derived TMAO increases several pro-inflammatory molecules, including E-selectin, interleukin 6 (IL-6), cyclooxygenase 2, and intracellular adhesion molecule 1 (ICAM1), by activating NF- κ B (32). Ottinger et al. have reported that TMAO is related to poor outcomes in patients with acute inflammatory pneumonia. They measured the TMAO levels in blood samples from 317 patients with acute pneumonia and found a significant association between TMAO and mortality (33). This evidence suggests a link between TMAO produced by the gut microbiota and inflammation through various mechanisms, which could exacerbate the viral outcome of COVID-19 infection. The

association between gut-derived TMAO and molecular mechanisms related to inflammation is summarized in [Figure 2](#).

2.3. Relationship Between TMAO Produced by the Gut Microbiome, SR-B1 Expression, and COVID-19

SRs-B1 are cell-surface HDL receptors that mediate the uptake or influx of HDL-derived cholesteryl-esters into cells and tissues (34). Previous studies have reported that SR-B1 is an essential receptor that affects HCV entry (35). However, there is not enough information about the potential roles of SR-B1 in SARS-CoV-19 infection. Few studies have demonstrated that SR-B1 receptors facilitate SARS-CoV-2 cell entry. Henrich et al. have shown that human cells expressing SR-B1 are susceptible to SARS-CoV-2 infection (36). Major cell types targeted by SARS-CoV-2, such as hepatocytes, immune cells, fibroblasts, adipocytes, and type II pneumocytes, express SR-B1 to contribute to viruses' entry and virus effects (37-39). In another study, Palacios-Rapalo et al. have shown that the SARS-CoV-2 S protein may bind to the cholesterol of HDL and enter the target cells. They have found that SR-B1 mediates SARS-CoV-2 attachment and transfer to the cells (40). On the other hand, some investigations have reported that TMAO produced by gut microbiota upregulates the expression of SRs (12). Animal studies demonstrated that dietary supplementation with TMAO increases cholesterol deposition into the peripheral tissues in mice (41). Chen et al. have indicated that TMAO induces the expression of genes related to cholesterol metabolism, such as SR-B1. However, this effect was TMAO dose-dependent (42). More information is needed to better estimate gut TMAO influences on SR-B1 expression ([Figure 2](#)).

2.4. Trimethylamine N-oxide Secreted by the Gut Microbiome Increases the Risk of Thrombosis

Disseminated intravascular coagulation and thrombosis are common complications in COVID-19 infections. COVID-19 induces a pro-thrombotic state, and the high incidence of reported major thrombotic events raises concerns about unique pro-thrombotic pathophysiology (43, 44). Autopsies of patients have revealed diffuse alveolar damage and microthrombi not only in pulmonary vasculature but also in other organs (45). Studies have reported that thrombotic events occur in up to one-third of COVID-19 patients, primarily as pulmonary emboli, and are associated with more severe disease and increased mortality (46).

Pyrin domain-containing 3 (NLRP3) interacts with apoptosis-associated speck-like protein (ASC) to cleave caspase-1, leading to the maturation and secretion

of pro-inflammatory cytokines IL-18 and IL1 β , which trigger inflammatory responses and play a crucial role in promoting the development of lipid plaques and destabilizing atherosclerotic plaques and thrombosis (47). The up-regulation of cellular adhesion molecules, such as vascular cell adhesion molecule-1 (VCAM-1) and ICAM-1, plays an initial role in atheromatous plaque formation (48). This underscores the importance of effective thromboprophylaxis and the treatment of thrombotic complications in COVID-19 patients, especially those requiring intensive care.

Regarding the association between gut TMAO and the risk of thrombosis, Zhu et al. reported on the role of TMAO in platelet hyperreactivity (11). They found that in patients with an increased incidence of thrombotic events, TMAO concentrations were significantly elevated. Furthermore, in vitro studies have shown that TMAO increases platelet aggregation and adhesion to collagen. In the same experiment, the injection of TMAO in mice stimulated carotid thrombus formation with shortened occlusion time. Several animal studies have demonstrated the biological role of gut microbiota in thrombosis. Mice fed a choline-rich diet and supplemented with TMAO exhibited platelet aggregation and shortened occlusion time. Administration of antibiotics to mice receiving TMAO supplementation prevented platelet aggregation. In these findings, Zhu et al. reported that TMAO does not directly induce platelet activation but rather increases the release of Ca²⁺ and platelet activation in response to agonists (11).

In a human study, Zhu et al. demonstrated that dose-dependent choline supplementation increased platelet aggregation, which was correlated with TMAO levels. Aspirin treatment in combination with choline reduced both TMAO concentrations and platelet hyperactivity (49). The authors suggest that aspirin may alter gut microbiota composition, affecting its function and the production of metabolites. Altering gut microbiota with probiotic species has shown promise in the treatment of animal and human diseases. However, further research is needed to determine whether probiotic administration can specifically target TMAO produced by gut microbiomes.

Various studies have reported that probiotics can modulate innate and adaptive immune responses, regulate host-pathogen interactions, and stimulate the secretion of immunoglobulin A (50, 51). Several randomized clinical trials have demonstrated the potential benefits of probiotics in patient outcomes after surgery, reducing infectious and non-infectious complications (52, 53). Zeng's study reported the positive effects of probiotics in reducing ventilator-associated

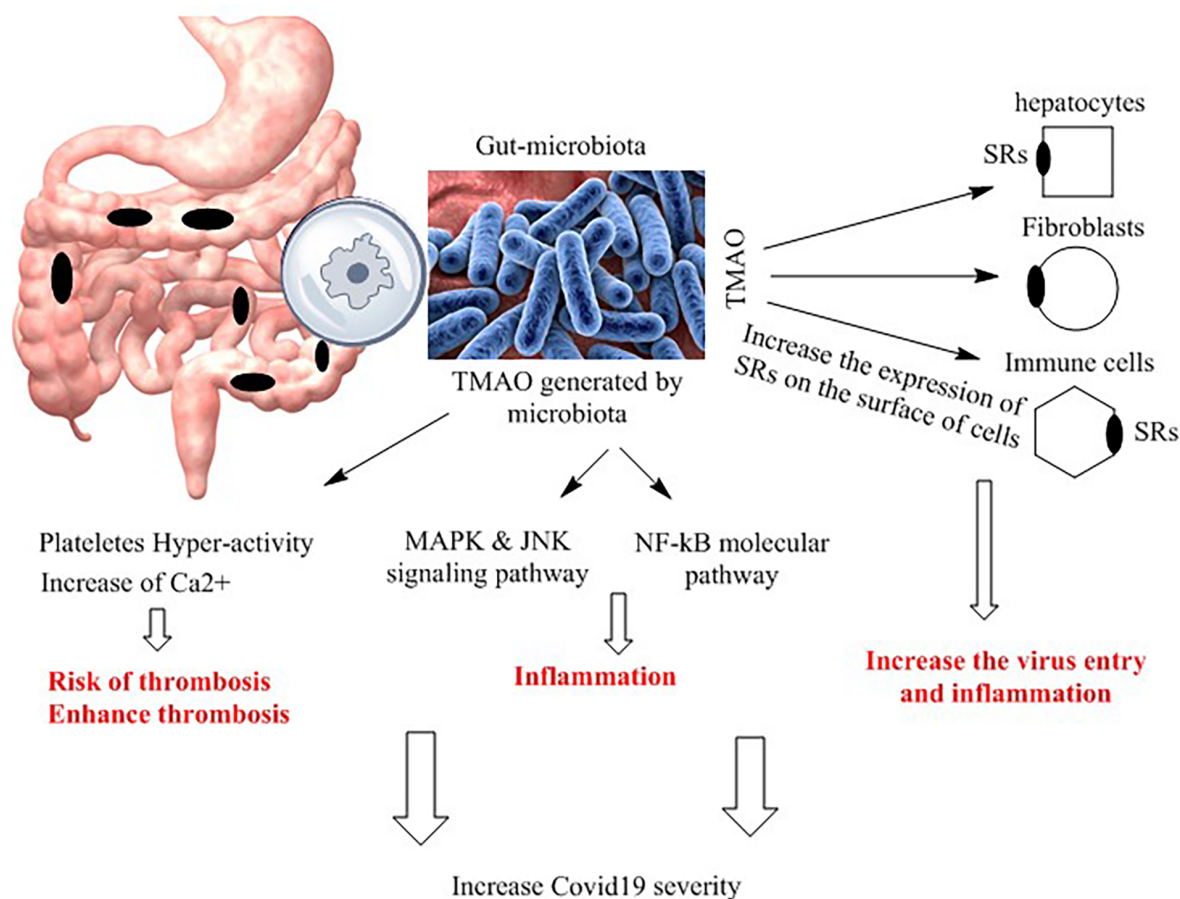


Figure 2. Mechanisms of trimethylamine N-oxide (TMAO) action. Trimethylamine N-oxide, through various molecular mechanisms, enhances inflammation, thrombosis, and virus entry into cells. It upregulates the expression of scavenger receptors (SRs) on the surfaces of different cells, facilitating virus entry. Furthermore, TMAO activates different molecular pathways, such as nuclear factor kappa (NF-kB), mitogen-activated protein kinase (MAPK), and c-Jun N-terminal kinase (JNK), promoting inflammation and exacerbating COVID-19 severity.

pneumonia in mechanically ventilated patients in the intensive care unit (54). The beneficial impact of probiotics on lung infections may be associated with the influence of gut microbiota on lung immunity, known as the gut-lung axis (55). Studies have indicated that microbiota can enhance resistance to viruses and pathogenic attacks on respiratory mucosa by improving the systemic immune response and preventing virus entry via the ACE2 receptor (56, 57).

To mitigate the production of TMAO by microorganisms, it is essential to block its synthesis pathway or reduce the population of responsible bacteria. This can be achieved through the use of broad-spectrum antibiotics or by introducing specific bacterial strains that limit or decrease the survival of other strains in the same niche, as in the case of probiotics.

3. Conclusions

Numerous studies have confirmed the significant role of the microbiota in synthesizing and releasing important metabolites with various biological properties, including TMAO. TMAO, produced by the gut microbiome, may potentially exacerbate the severity of COVID-19 through various mechanisms. This evidence underscores the importance of dysbiosis, which can lead to disease severity by producing different metabolites. The administration of prebiotics and probiotics to modulate the gut microbiome towards beneficial bacteria, thereby regulating pathways that generate proatherogenic metabolites, is proposed as a novel therapeutic strategy for COVID-19. However, additional studies, including animal models and clinical trials, are required to elucidate the underlying mechanisms and determine whether interventions

targeting the gut microbiota and TMAO could help prevent atherosclerosis in COVID-19.

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Footnotes

Authors' Contribution: Study concept and design, and editing the final version of the manuscript: MHA; Acquisition of data and writing the draft of the manuscript: MSSA; Interpreting the data for the study: RKH and MA; Interpreting the data for the study and revising the manuscript: FB. All the authors read and approved the final manuscript.

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References

- Jandhyala SM, Talukdar R, Subramanyam C, Vuyyuru H, Sasikala M, Nageshwar Reddy D. Role of the normal gut microbiota. *World J Gastroenterol.* 2015;**21**(29):8787–803. [PubMed ID: 26269668]. [PubMed Central ID: PMC4528021]. <https://doi.org/10.3748/wjg.v21.i29.8787>.
- O'Connor EM. The role of gut microbiota in nutritional status. *Curr Opin Clin Nutr Metab Care.* 2013;**16**(5):509–16. [PubMed ID: 23852088]. <https://doi.org/10.1097/MCO.0b013e3283638eb3>.
- Maslowski KM, Mackay CR. Diet, gut microbiota and immune responses. *Nat Immunol.* 2011;**12**(1):5–9. [PubMed ID: 21169997]. <https://doi.org/10.1038/nri0111-5>.
- Yang S, Li X, Yang F, Zhao R, Pan X, Liang J, et al. Gut Microbiota-Dependent Marker TMAO in Promoting Cardiovascular Disease: Inflammation Mechanism, Clinical Prognostic, and Potential as a Therapeutic Target. *Front Pharmacol.* 2019;**10**:1360. [PubMed ID: 31803054]. [PubMed Central ID: PMC6877687]. <https://doi.org/10.3389/fphar.2019.01360>.
- Turnbaugh PJ, Ley RE, Mahowald MA, Magrini V, Mardis ER, Gordon JI. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature.* 2006;**444**(7122):1027–31. [PubMed ID: 17183312]. <https://doi.org/10.1038/nature05414>.
- Turnbaugh PJ, Hamady M, Yatsunenko T, Cantarel BL, Duncan A, Ley RE, et al. A core gut microbiome in obese and lean twins. *Nature.* 2009;**457**(7228):480–4. [PubMed ID: 19043404]. [PubMed Central ID: PMC2677729]. <https://doi.org/10.1038/nature07540>.
- Wang Z, Klipfell E, Bennett BJ, Koeth R, Levison BS, Dugar B, et al. Gut flora metabolism of phosphatidylcholine promotes cardiovascular disease. *Nature.* 2011;**472**(7341):57–63. [PubMed ID: 21475195]. [PubMed Central ID: PMC3086762]. <https://doi.org/10.1038/nature09922>.
- Chen K, Zheng X, Feng M, Li D, Zhang H. Gut Microbiota-Dependent Metabolite Trimethylamine N-Oxide Contributes to Cardiac Dysfunction in Western Diet-Induced Obese Mice. *Front Physiol.* 2017;**8**:139. [PubMed ID: 28377725]. [PubMed Central ID: PMC5359299]. <https://doi.org/10.3389/fphys.2017.00139>.
- Chen ML, Zhu XH, Ran L, Lang HD, Yi L, Mi MT. Trimethylamine-N-Oxide Induces Vascular Inflammation by Activating the NLRP3 Inflammasome Through the SIRT3-SOD2-mtROS Signaling Pathway. *J Am Heart Assoc.* 2017;**6**(9). [PubMed ID: 28871042]. [PubMed Central ID: PMC5634285]. <https://doi.org/10.1161/JAHA.117.006347>.
- Zhang Y, Wang Y, Ke B, Du J. TMAO: how gut microbiota contributes to heart failure. *Transl Res.* 2021;**228**:109–25. [PubMed ID: 32841736]. <https://doi.org/10.1016/j.trsl.2020.08.007>.
- Zhu W, Gregory JC, Org E, Buffa JA, Gupta N, Wang Z, et al. Gut Microbial Metabolite TMAO Enhances Platelet Hyperreactivity and Thrombosis Risk. *Cell.* 2016;**165**(1):11–24. [PubMed ID: 26972052]. [PubMed Central ID: PMC4862743]. <https://doi.org/10.1016/j.cell.2016.02.011>.
- Wei C, Wan L, Yan Q, Wang X, Zhang J, Yang X, et al. HDL-scavenger receptor B type 1 facilitates SARS-CoV-2 entry. *Nat Metab.* 2020;**2**(12):1391–400. [PubMed ID: 33244168]. <https://doi.org/10.1038/s42255-020-00324-0>.
- Haissman JM, Haugaard AK, Ostrowski SR, Berge RK, Hov JR, Trosheid M, et al. Microbiota-dependent metabolite and cardiovascular disease marker trimethylamine-N-oxide (TMAO) is associated with monocyte activation but not platelet function in untreated HIV infection. *BMC Infect Dis.* 2017;**17**(1):445. [PubMed ID: 28645263]. [PubMed Central ID: PMC5481962]. <https://doi.org/10.1186/s12879-017-2547-x>.
- Zhang JJ, Dong X, Cao YY, Yuan YD, Yang YB, Yan YQ, et al. Clinical characteristics of 140 patients infected with SARS-CoV-2 in Wuhan, China. *Allergy.* 2020;**75**(7):1730–41. [PubMed ID: 32077115]. <https://doi.org/10.1111/all.14238>.
- Hamzah FAB, Lau CH, Nazri H, Ligo DV, Lee G, Tan CL, et al. Corona Tracker: worldwide COVID-19 outbreak data analysis and prediction. *Bull World Health Organ.* 2020. <https://doi.org/10.2471/blt.20.255695>.
- Mehta P, McAuley DF, Brown M, Sanchez E, Tattersall RS, Manson JJ, et al. COVID-19: consider cytokine storm syndromes and immunosuppression. *Lancet.* 2020;**395**(10229):1033–4. [PubMed ID: 32192578]. [PubMed Central ID: PMC7270045]. [https://doi.org/10.1016/S0140-6736\(20\)30628-0](https://doi.org/10.1016/S0140-6736(20)30628-0).
- Pan L, Mu M, Yang P, Sun Y, Wang R, Yan J, et al. Clinical Characteristics of COVID-19 Patients With Digestive Symptoms in Hubei, China: A Descriptive, Cross-Sectional, Multicenter Study. *Am J Gastroenterol.* 2020;**115**(5):766–73. [PubMed ID: 32287140]. [PubMed Central ID: PMC7172492]. <https://doi.org/10.14309/ajg.0000000000000620>.
- de Moreno de Leblanc A, Del Carmen S, Zurita-Turk M, Santos Rocha C, van de Guchte M, Azevedo V, et al. Importance of IL-10 modulation by probiotic microorganisms in gastrointestinal inflammatory diseases. *ISRN Gastroenterol.* 2011;**2011**:892971. [PubMed ID: 21991534]. [PubMed Central ID: PMC3168568]. <https://doi.org/10.5402/2011/892971>.
- Chhibber-Goel J, Gopinathan S, Sharma A. Interplay between severities of COVID-19 and the gut microbiome: implications of bacterial co-infections? *Gut Pathog.* 2021;**13**(1):14. [PubMed ID: 33632296]. [PubMed Central ID: PMC7906082]. <https://doi.org/10.1186/s13099-021-00407-7>.
- Esposito S, Polinori I, Rigante D. The Gut Microbiota-Host Partnership as a Potential Driver of Kawasaki Syndrome. *Front Pediatr.* 2019;**7**:124. [PubMed ID: 31024869]. [PubMed Central ID: PMC6460951]. <https://doi.org/10.3389/fped.2019.00124>.
- Liu J, Zhao M, Zhou J, Liu C, Zheng L, Yin Y. Simultaneous targeted analysis of trimethylamine-N-oxide, choline, betaine, and carnitine by high performance liquid chromatography tandem mass spectrometry. *J Chromatogr B Analyt Technol Biomed Life Sci.* 2016;**1035**:42–8. [PubMed ID: 27669507]. <https://doi.org/10.1016/j.jchromb.2016.09.026>.

22. Rath S, Heidrich B, Pieper DH, Vital M. Uncovering the trimethylamine-producing bacteria of the human gut microbiota. *Microbiome*. 2017;**5**(1):54. [PubMed ID: 28506279]. [PubMed Central ID: PMC5433236]. <https://doi.org/10.1186/s40168-017-0271-9>.
23. Warriar M, Shih DM, Burrows AC, Ferguson D, Gromovsky AD, Brown AL, et al. The TMAO-Generating Enzyme Flavin Monooxygenase 3 Is a Central Regulator of Cholesterol Balance. *Cell Rep*. 2015;**10**(3):326–38. [PubMed ID: 25600868]. [PubMed Central ID: PMC4501903]. <https://doi.org/10.1016/j.celrep.2014.12.036>.
24. Vickers NJ. Animal Communication: When I'm Calling You, Will You Answer Too? *Curr Biol*. 2017;**27**(14):R713–5. [PubMed ID: 28743020]. <https://doi.org/10.1016/j.cub.2017.05.064>.
25. Mohammadi A, Gholamhosseiniannajar A, Yaghoobi MM, Jahani Y, Vahabzadeh Z. Expression levels of heat shock protein 60 and glucose-regulated protein 78 in response to trimethylamine-N-oxide treatment in murine macrophage J774A.1 cell line. *Cell Mol Biol (Noisy-le-grand)*. 2015;**61**(4):94–100. [PubMed ID: 26429299].
26. Collot-Teixeira S, Martin J, McDermott-Roe C, Poston R, McGregor JL. CD36 and macrophages in atherosclerosis. *Cardiovasc Res*. 2007;**75**(3):468–77. [PubMed ID: 17442283]. <https://doi.org/10.1016/j.cardiores.2007.03.010>.
27. Getz GS. Thematic review series: the immune system and atherogenesis. Immune function in atherogenesis. *J Lipid Res*. 2005;**46**(1):1–10. [PubMed ID: 15547292]. <https://doi.org/10.1194/jlr.R400013-JLR200>.
28. Geng J, Yang C, Wang B, Zhang X, Hu T, Gu Y, et al. Trimethylamine N-oxide promotes atherosclerosis via CD36-dependent MAPK/JNK pathway. *Biomed Pharmacother*. 2018;**97**:941–7. [PubMed ID: 29136772]. <https://doi.org/10.1016/j.biopha.2017.11.016>.
29. Wu X, Chen L, Zeb F, Huang Y, An J, Ren J, et al. Regulation of circadian rhythms by NEAT1 mediated TMAO-induced endothelial proliferation: A protective role of asparagus extract. *Exp Cell Res*. 2019;**382**(1):111451. [PubMed ID: 31173767]. <https://doi.org/10.1016/j.yexcr.2019.05.032>.
30. Chou RH, Chen CY, Chen IC, Huang HL, Lu YW, Kuo CS, et al. Trimethylamine N-Oxide, Circulating Endothelial Progenitor Cells, and Endothelial Function in Patients with Stable Angina. *Sci Rep*. 2019;**9**(1):4249. [PubMed ID: 30862856]. [PubMed Central ID: PMC6414518]. <https://doi.org/10.1038/s41598-019-40638-y>.
31. Tak PP, Firestein GS. NF-kappaB: a key role in inflammatory diseases. *J Clin Invest*. 2001;**107**(17):7–11. [PubMed ID: 1134171]. [PubMed Central ID: PMC198552]. <https://doi.org/10.1172/JCI11830>.
32. Seldin MM, Meng Y, Qi H, Zhu W, Wang Z, Hazen SL, et al. Trimethylamine N-Oxide Promotes Vascular Inflammation Through Signaling of Mitogen-Activated Protein Kinase and Nuclear Factor-kappaB. *J Am Heart Assoc*. 2016;**5**(2). [PubMed ID: 26903003]. [PubMed Central ID: PMC4802459]. <https://doi.org/10.1161/JAHA.115.002767>.
33. Ottiger M, Nickler M, Steuer C, Odermatt J, Huber A, Christ-Crain M, et al. Trimethylamine-N-oxide (TMAO) predicts fatal outcomes in community-acquired pneumonia patients without evident coronary artery disease. *Eur J Intern Med*. 2016;**36**:67–73. [PubMed ID: 27567042]. <https://doi.org/10.1016/j.ejim.2016.08.017>.
34. Shen WJ, Azhar S, Kraemer FB. Lipid droplets and steroidogenic cells. *Exp Cell Res*. 2016;**340**(2):209–14. [PubMed ID: 26639173]. [PubMed Central ID: PMC4744538]. <https://doi.org/10.1016/j.yexcr.2015.11.024>.
35. Catanese MT, Ansuini H, Graziani R, Huby T, Moreau M, Ball JK, et al. Role of scavenger receptor class B type I in hepatitis C virus entry: kinetics and molecular determinants. *J Virol*. 2010;**84**(1):34–43. [PubMed ID: 19828610]. [PubMed Central ID: PMC2798406]. <https://doi.org/10.1128/JVI.02199-08>.
36. Henrich SE, McMahon KM, Palacio N, Bhalla P, Penalzoza-MacMaster P, Thaxton C. Targeting Scavenger Receptor Type B-1 (SR-B1) and Cholesterol Inhibits Entry of SARS-CoV-2 Pseudovirus in Cell Culture. *BioRxiv*. 2020. <https://doi.org/10.1101/2020.12.14.420133>.
37. Clayton RF, Rinaldi A, Kandyba EE, Edward M, Willberg C, Klenerman P, et al. Liver cell lines for the study of hepatocyte functions and immunological response. *Liver Int*. 2005;**25**(2):389–402. [PubMed ID: 15780065]. <https://doi.org/10.1111/j.1478-3231.2005.01017.x>.
38. Song GJ, Kim SM, Park KH, Kim J, Choi I, Cho KH. SR-B1 mediates high density lipoprotein (HDL)-induced anti-inflammatory effect in macrophages. *Biochem Biophys Res Commun*. 2015;**457**(1):112–8. [PubMed ID: 25528585]. <https://doi.org/10.1016/j.bbrc.2014.12.028>.
39. Rhainds D, Brissette L. The role of scavenger receptor class B type I (SR-B1) in lipid trafficking: defining the rules for lipid traders. *Int J Biochem Cell Biol*. 2004;**36**(1):39–77. [PubMed ID: 14592533]. [https://doi.org/10.1016/s1357-2725\(03\)00173-0](https://doi.org/10.1016/s1357-2725(03)00173-0).
40. Palacios-Rapalo SN, De Jesus-Gonzalez LA, Cordero-Rivera CD, Farfan-Morales CN, Osuna-Ramos JF, Martinez-Mier G, et al. Cholesterol-Rich Lipid Rafts as Platforms for SARS-CoV-2 Entry. *Front Immunol*. 2021;**12**:796855. [PubMed ID: 34975904]. [PubMed Central ID: PMC8719300]. <https://doi.org/10.3389/fimmu.2021.796855>.
41. Chistiakov DA, Bobryshev YV, Kozarov E, Sobenin IA, Orekhov AN. Role of gut microbiota in the modulation of atherosclerosis-associated immune response. *Front Microbiol*. 2015;**6**:671. [PubMed ID: 26175728]. [PubMed Central ID: PMC4485310]. <https://doi.org/10.3389/fmicb.2015.00671>.
42. Chen Y, Weng Z, Liu Q, Shao W, Guo W, Chen C, et al. FMO3 and its metabolite TMAO contribute to the formation of gallstones. *Biochim Biophys Acta Mol Basis Dis*. 2019;**1865**(10):2576–85. [PubMed ID: 31251986]. <https://doi.org/10.1016/j.bbadis.2019.06.016>.
43. Connors JM, Levy JH. COVID-19 and its implications for thrombosis and anticoagulation. *Blood*. 2020;**135**(23):2033–40. [PubMed ID: 32339221]. [PubMed Central ID: PMC7273827]. <https://doi.org/10.1182/blood.2020060600>.
44. Panigada M, Bottino N, Tagliabue P, Grasselli G, Novembrino C, Chantarangkul V, et al. Hypercoagulability of COVID-19 patients in intensive care unit: A report of thromboelastography findings and other parameters of hemostasis. *J Thromb Haemost*. 2020;**18**(7):1738–42. [PubMed ID: 32302438]. [PubMed Central ID: PMC9906150]. <https://doi.org/10.1111/jth.14850>.
45. Liu Q, Wang RS, Qu GQ, Wang YY, Liu P, Zhu YZ, et al. Gross examination report of a COVID-19 death autopsy. *Fa Yi Xue Za Zhi*. 2020;**36**(1):21–3. [PubMed ID: 32198987]. <https://doi.org/10.12116/j.issn.1004-5619.2020.01.005>.
46. Llitjos JF, Leclerc M, Chochois C, Monsallier JM, Ramakers M, Auvray M, et al. High incidence of venous thromboembolic events in anticoagulated severe COVID-19 patients. *J Thromb Haemost*. 2020;**18**(7):1743–6. [PubMed ID: 32320517]. [PubMed Central ID: PMC7264774]. <https://doi.org/10.1111/jth.14869>.
47. Janoudi A, Shamoun FE, Kalavakunta JK, Abela GS. Cholesterol crystal induced arterial inflammation and destabilization of atherosclerotic plaque. *Eur Heart J*. 2016;**37**(25):1959–67. [PubMed ID: 26705388]. <https://doi.org/10.1093/eurheartj/ehv653>.
48. Santos-Gallego CG, Picatoste B, Badimon JJ. Pathophysiology of acute coronary syndrome. *Curr Atheroscler Rep*. 2014;**16**(4):401. [PubMed ID: 24504549]. <https://doi.org/10.1007/s11883-014-0401-9>.
49. Zhu W, Wang Z, Tang WHW, Hazen SL. Gut Microbe-Generated Trimethylamine N-Oxide From Dietary Choline Is Prothrombotic in Subjects. *Circulation*. 2017;**135**(17):1671–3. [PubMed ID: 28438808]. [PubMed Central ID: PMC5460631]. <https://doi.org/10.1161/CIRCULATIONAHA.116.025338>.
50. Bermudez-Brito M, Plaza-Diaz J, Munoz-Quezada S, Gomez-Llorente C, Gil A. Probiotic mechanisms of action. *Ann Nutr Metab*. 2012;**61**(2):160–74. [PubMed ID: 23037511]. <https://doi.org/10.1159/000342079>.
51. Plaza-Diaz J, Ruiz-Ojeda FJ, Gil-Campos M, Gil A. Mechanisms of Action of Probiotics. *Adv Nutr*. 2019;**10**(suppl1):S49–66. [PubMed ID: 30721959]. [PubMed Central ID: PMC6363529]. <https://doi.org/10.1093/advances/nmy063>.
52. An S, Kim K, Kim MH, Jung JH, Kim Y. Perioperative Probiotics

- Application for Preventing Postoperative Complications in Patients with Colorectal Cancer: A Systematic Review and Meta-Analysis. *Medicina (Kaunas)*. 2022;**58**(11). [PubMed ID: 36422183]. [PubMed Central ID: PMC9699544]. <https://doi.org/10.3390/medicina58111644>.
53. Veziat J, Bonnet M, Océan BV, Dziri C, Pereira B, Slim K. Probiotics/Synbiotics to Reduce Infectious Complications after Colorectal Surgery: A Systematic Review and Meta-Analysis of Randomised Controlled Trials. *Nutrients*. 2022;**14**(15). [PubMed ID: 35893922]. [PubMed Central ID: PMC9332115]. <https://doi.org/10.3390/nui14153066>.
54. Zeng J, Wang CT, Zhang FS, Qi F, Wang SF, Ma S, et al. Effect of probiotics on the incidence of ventilator-associated pneumonia in critically ill patients: a randomized controlled multicenter trial. *Intensive Care Med*. 2016;**42**(6):1018–28. [PubMed ID: 27043237]. <https://doi.org/10.1007/s00134-016-4303-x>.
55. Dang AT, Marsland BJ. Microbes, metabolites, and the gut-lung axis. *Mucosal Immunol*. 2019;**12**(4):843–50. [PubMed ID: 30976087]. <https://doi.org/10.1038/s41385-019-0160-6>.
56. Patra S, Saxena S, Sahu N, Pradhan B, Roychowdhury A. Systematic Network and Meta-analysis on the Antiviral Mechanisms of Probiotics: A Preventive and Treatment Strategy to Mitigate SARS-CoV-2 Infection. *Probiotics Antimicrob Proteins*. 2021;**13**(4):1138–56. [PubMed ID: 33537958]. [PubMed Central ID: PMC7857647]. <https://doi.org/10.1007/s12602-021-09748-w>.
57. Racedo S, Villena J, Salva S, Alvarez S. Influence of yogurt consumption on the respiratory immune response. *Food Agric Immunol*. 2009;**20**(3):231–44. <https://doi.org/10.1080/09540100903061659>.