Arch Clin Infect Dis. 2015 October; 10(4): e28544.

Published online 2015 October 24.

Research Article

Association of Fusobacterium Isolation From Periodontal Pockets With Halitosis and the Related Risk Factors in Shiraz, Iran

Hadi Sedigh Ebrahim-Saraie,^{1,2} Mohammad Motamedifar,^{1,3,*} Davood Mansury,^{1,2} Hooman Ebrahimi,⁴ Sara Pourshahidi,⁵ Mehrdad Halaji,^{1,2} and Hadi Raeisi Shahraki⁶

¹Department of Bacteriology and Virology, School of Medicine, Shiraz University of Medical Sciences, Shiraz, IR Iran

Student Research Committee, Shiraz University of Medical Sciences, Shiraz, IR Iran Shiraz HIV/AIDS Research Center, Shiraz University of Medical Sciences, Shiraz, IR Iran ⁴Department of Oral Medicine, Dental School, Islamic Azad University, Tehran, IR Iran ⁹Department of Oral and Maxillofacial Medicine, Tehran University of Medical Sciences, Tehran, IR Iran

⁶Department of Biostatistics, School of Medicine, Shiraz University of Medical Sciences, Shiraz, IR Iran

*Corresponding author: Mohammad Motamedifar, Department of Bacteriology and Virology, School of Medicine, Shiraz University of Medical Sciences, Shiraz, IR Iran. Tel/Fax: +98-7132304356, E-mail: motamedm@sums.ac.ir

Received: March 8, 2015; Revised: July 1, 2015; Accepted: July 11, 2015

Background: Over recent decades, halitosis has become a priority in oral hygiene maintenance. Bad breath is one of the primary reasons for referral to dentists in Iran. Although halitosis is mainly caused by endogenous factors such as microbial metabolism, it is a multifactorial condition.

Objectives: This study aimed to identify the probable relationship of the presence of Fusobacterium species in periodontal pockets with halitosis and determine the risk factors for this condition.

Patients and Methods: This case-control study included patients referred to a polyclinic in Shiraz, which is located in Fars province in the southwest of Iran. In total, 50 patients with halitosis confirmed by an organoleptic test and 50 patients without oral malodor were recruited. Samples were obtained from their periodontal pockets using absorbent paper points and cultured for characterization by biochemical tests.

Results: In total, 26% (n = 13) and 8% (n = 4) samples were positive for *Fusobacterium* species in the halitosis and control groups, respectively, with F. nucleatum present in the greatest proportion in both groups. Halitophobia was significantly more frequent in the halitosis group than in the control group (P < 0.001). Sinusitis was the most common systemic disease. Moreover, the halitosis group patients exhibited a greater tendency to include curry powder, chili, and sausage in their diet compared with the control subjects (P < 0.05). Conclusions: The results of the present study suggest that the presence of Fusobacterium species in periodontal pockets is an important risk factor for halitosis.

Keywords: Anaerobic Bacteria; Periodontal Pocket; Halitosis; Fusobacterium

1. Background

Over recent decades, halitosis has become a priority in oral hygiene maintenance worldwide (1) and significantly affects social relationships (1). Currently, bad breath is one of the main reasons for referral to dentists in Iran, requiring clinicians and manufacturers to make large investments in cosmetic products for halitosis improvement (1). However, the primary underlying reason for halitosis is often unclear, resulting in only temporary relief after the use of these products (2).

Volatile sulfur compounds (VSCs) are considered the main causative substances for halitosis. The majority of VSCs reportedly comprise hydrogen sulfide and methyl mercaptan (3, 4) and can cause various types of mouth odor (4). It has been shown that the main origin of oral

malodor is in the oral cavity, e.g., periodontal pockets (5, 6). This phenomenon mainly results from the putrefaction of amino acids by bacteria under anaerobic conditions (1, 6). This anaerobic microenvironment houses different bacterial species that majorly include a range of gram-negative proteolytic strains (1, 3). VSC is the main product of proteolysis by these organisms, which utilizes sulfur-containing amino acid resources such as cysteine and methionine (7).

Several anaerobic gram-negative bacteria have been mentioned for their role in VSC production, with Fuso*bacterium* species being one of the most common (5, 7, 8). Previously, it was shown that the plaque biofilm is a major source of halitosis-inducing VSCs produced by

Copyright @ 2015, Infectious Diseases and Tropical Medicine Research Center. This is an open-access article distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 International License (http://creativecommons.org/licenses/by-nc/4.0/) which permits copy and redistribute the material just in noncommercial usages, provided the original work is properly cited.

anaerobic gram-negative bacteria. *Fusobacterium* species play a critical role in this phenomenon, along with other bacteria in plaque biofilms (7). With regard to this issue, it was previously indicated that FomA, which is one of the major outer membrane proteins of *Fusobacterium nucleatum*, could be an agent for recruiting other anaerobic gram-negative bacteria into periodontal pockets, thus facilitating the formation of plaque biofilms (7).

Although halitosis is primarily the result of endogenous factors such as microbial metabolism, it is a multifactorial condition (7) that can be affected by exogenous factors such as diet and smoking or, occasionally, physiological factors such as a low salivary flow due to insufficient water intake (1, 3).

2. Objectives

Because of the reported role of *Fusobacterium* species in production of bad smell and the lack of a study showing an obvious association between halitosis and any specific bacterial isolate, we aimed to determine any probable relationship between the presence of *Fusobacterium* species and halitosis, as well as the relationship between some exogenous and physiological factors and halitosis.

3. Patients and Methods

3.1. Subjects and Study Design

In this case-control study conducted from October 2012 to March 2013, 50 patients with confirmed halitosis (27 women and 23 men; mean age, 37 ± 11 years; range, 19 - 54 years) and 50 patients (33 women and 17 men; mean age, 38 ± 12 years; range, 20 - 62 years) without oral malodor were included. All subjects were patients referred to Emam Reza polyclinic, which is affiliated to Shiraz university of medical sciences, Shiraz, Iran. Emam Reza

polyclinic is a government-owned medical center with 11 floors and several clinical facilities, including dentistry, and it serves approximately 600 thousand patients in Fars province every month. All individuals who had not brushed, rinsed, smoked, eaten, or consumed any beverage for at least 2 h prior to assessment were sampling and included for analysis. Furthermore, none of the subjects had a history of antibiotic therapy for at least a month. The Ethics Committee of Shiraz University of Medical Sciences approved the study design (EC-91-6329). Informed consent was obtained from all participants before study initiation. A standardized checklist was designed by the authors and subjected to pilot tests before the survey. Self-reported information regarding medical and dental histories and a few dietary habits was obtained from all participants to evaluate the presence of potential risk factors for halitosis (Table 1).

3.2. Oral Malodor Assessment

The degree of oral malodor in each subject was assessed by two dentists (kappa = 0.82, P < 0.05) using an organoleptic test (OLT). Scores were estimated on a scale of 0 - 5: 0, no malodor; 1, very slight; 2, slight; 3, moderate; 4, strong; and 5, very strong. Subjects with a score of > 2 were included in the halitosis group, while those with a score of \leq 2 were included in the control group (4).

3.3. Clinical Sampling

Samples were obtained from the periodontal pockets of subjects under aseptic conditions using absorbent paper points. After sampling, the paper points were pooled in capped plastic tubes containing 1 mL of anaerobic transport medium composed of thioglycollate broth (Merck, Germany), 0.05% Tween 80 (Merck, Germany), 5 μ g of hemin (Sigma, UK) per mL, and 0.5 μ g of menadione (Sigma, UK) per mL. All samples were immediately transported to the laboratory and cultured.

Table 1. Potential Risk Factors for Halitosis in the Included Participants

| Oral Disease ^a | Drug ^b | Systemic Disease ^a | Foods with Sulfur-Containing Amino Acids ^C | References |
|---|---|---|---|------------|
| Periodontal disease, dental cavity, cracked tooth, and ulcers, etc. | Antidepressive, antihistaminic, antihypertensive, antiparkinson, anxiolytic, diuretic, and anorexigenic agents | Sinusitis, diabetes, gastrointestinal disease, renal failure, respiratory infection, urinary tract infection, lactose intolerance, decreased salivary flow | Curry powder, chili, coffee, tea, carbohydrates (sugar), garlic, onion, yogurt, lentils, sausage, and chicken meat | (1,3) |

^a Self-reported disease was considered a positive risk factor.

^b Self-reported consumption of any of the mentioned drugs was considered a positive risk factor.

^C Self-reported high consumption of any of the mentioned foods was considered a positive risk factor.

3.4. Culture and Identification

The samples were mixed using a vortex for 1 min to allow the dispersal of bacteria. Then, 20 µL of the solution was taken from each tube and spread onto an anaerobic agar plate modified for the selection of *Fusobacteria*. The agar comprised columbia agar (Merck, Germany), 0.05% Tween 80 (Merck, Germany), 0.5 µg of menadione (Sigma, UK) per mL, and 5% fresh defibrinated blood. This medium was supplemented with vancomycin (5 µg/mL, Sigma), neomycin (100 μ g/mL, Sigma), and erythromycin (5 μ g/mL, Sigma) (9, 10). The plates were incubated in anaerobic jars using Gas-Pak A (Merck, Germany) for up to 5 days at 37°C. The primary identification of Fusobacterium was based on the presence of typical pleomorphic, gram-negative, rod-shaped or fusiform bacilli on gram staining and positive susceptibility of Fusobacterium species to kanamycin (1 mg) and colistin (10 μ g) and resistance to vancomycin (5 μ g) on susceptibility testing (MAST, UK). The species were characterized through standard biochemical tests (11). A simplified flow chart of the study is illustrated in Figure 1.

3.5. Statistical Analysis

All statistical analyses were performed using SPSS software version 19.0 (IBM Corp., Armonk, NY, USA). The frequency of bacterial isolation and risk factors are presented as descriptive statistics in terms of relative frequency. Chi-square and Fisher's exact tests were used to estimate any statistical associations as appropriate. A P value < 0.05 was considered statistically significant.

4. Results

Statistical analysis revealed no significant association of sex and age with halitosis. In total, 26% (n = 13) and 8% (n = 4) samples from the halitosis and control groups, respectively, were positive for *Fusobacterium* species, with *F. nucleatum* present in the largest proportion in both groups (Table 2).

The frequency of halitophobia was significantly higher in the halitosis group (68%) than that in the control group (26%). Sinusitis was the major systemic disease in the halitosis [70% (14/20)] and control groups [61.5% (8/13)] and in subjects with positivity for *Fusobacteria* species.

Curry powder, chili, and sausage consumption was more common (P < 0.05) in the diet of subjects with hali-

tosis than in those without. There were no significant differences in other diet patterns between the two groups. Correlation analyses for risk factors in the studied groups are presented in Table 3.

Although the majority of subjects positive for *Fusobacterium* species in the halitosis group (53.8%) exhibited some form of oral disease (periodontal disease, dental caries, cracked tooth, ulcers, etc.), there were no significant differences in the disease incidence between these subjects and those negative for *Fusobacterium* species (Table 4). Furthermore, job-related dryness of mouth was more frequent in subjects positive for *Fusobacterium* species than in their negative counterparts in the halitosis group (P < 0.001). In addition, the rates of flossing and consumption of chili, coffee, tea, carbohydrates, garlic, and onion were significantly different (P < 0.05) between the positive and negative subjects in the halitosis group.



Figure 1. Simplified Flow Chart of the Present Study

| Table 2. Fusobacterium Isolation Rate in the Studied Groups $(n = 50)^a$ | | | | |
|---|-------------------------------|----------------------------------|--------------------------------|--|
| Isolates - | Groups | | | |
| | Individuals With Oral Malodor | Individuals Without Oral Malodor | Level of Significance, P-Value | |
| F. nucleatum | 10 (20.0) | 4 (8.0) | 0.148 | |
| F. necrophorum | 3(6.0) | 0(0) | 0.242 | |
| Total | 13 (26.0) | 4 (8.0) | 0.05 | |

^a Data are presented as No. (%).

| Table 3. Proportion and Correlation Analyses for Risk Factors in the Studied Groups $(n = 50)^a$ | | | | |
|---|----------------------------|-------------------------|-----------------------|--|
| | Groups | | | |
| Risk Factors | Individuals With Halitosis | Individual Without Oral | Level of Significance | |
| | | Malodor | (P-Value) | |
| Fusobacteria isolation | 13 (26.0) | 4(8.0) | 0.05 | |
| Halitophobia | 34 (68.0) | 13 (26.0) | 0.001 | |
| Active or passive smoking | 8 (16.0) | 10 (20.0) | 0.79 | |
| Job-related dryness of mouth | 13 (26.0) | 5(10.0) | 0.05 | |
| Drug usage | 9 (18.0) | 8 (16.0) | 0.79 | |
| Oral disease | 19 (38.0) | 18 (36.0) | 0.84 | |
| Sinusitis | 14 (28.0) | 8 (16.0) | 0.23 | |
| Systemic disease | 20(40.0) | 13 (26.0) | 0.14 | |
| Tooth brushing | 48 (96.0) | 49 (98.0) | 0.56 | |
| Flossing | 20 (40.0) | 23 (46.0) | 0.55 | |
| Rinsing | 9 (18.0) | 14 (28.0) | 0.24 | |
| Curry powder | 37(74.0) | 27 (54.0) | 0.05 | |
| Chili | 27 (54.0) | 17 (34.0) | 0.05 | |
| Coffee | 9 (18.0) | 10 (20.0) | 0.8 | |
| Tea | 32(64.0) | 30 (60.0) | 0.68 | |
| Carbohydrates | 32(64.0) | 27 (54.0) | 0.31 | |
| Garlic | 14 (28.0) | 12 (24.0) | 0.65 | |
| Onion | 26 (52.0) | 25 (50.0) | 0.84 | |
| Yogurt | 42 (84.0) | 42 (84.0) | 1.0 | |
| Lentil | 30 (60.0) | 23 (46.0) | 0.16 | |
| Sausage | 12 (24.0) | 2(4.0) | 0.05 | |
| Chicken meat | 43 (86.0) | 40 (80.0) | 0.42 | |

Sedigh Ebrahim-Saraie H et al.

^a Data are presented as No. (%).

| * | Groups | | | |
|------------------------------|--|--|------------------------------------|--|
| Risk Factors | Fusobacteria positive/With Oral Malodor | Fusobacteria Negative/With Oral Malodor | Level of Significance (P-Value) | |
| Halitophobia | 9 (69.2) | 25 (67.6) | 0.89 | |
| Active or passive smoking | 3 (23.1) | 5 (13.5) | 0.39 | |
| Job-related dryness of mouth | 8 (61.5) | 5 (13.5) | 0.0006 | |
| Drug usage | 4 (30.8) | 5 (13.5) | 0.16 | |
| Oral disease | 7 (53.8) | 12 (32.4) | 0.17 | |
| Sinusitis | 3 (23.1) | 11 (29.7) | 0.67 | |
| Systemic disease | 5 (38.5) | 15 (40.5) | 0.89 | |
| Tooth brushing | 12 (92.3) | 36 (97.3) | 0.44 | |
| Flossing | 10 (76.9) | 10 (27.0) | 0.0019 | |
| Rinsing | 4 (30.8) | 5 (13.5) | 0.16 | |
| Curry powder | 11 (84.6) | 26 (70.3) | 0.32 | |
| Chili | 6(46.2) | 21 (56.8) | 0.53 | |
| Coffee | 0(0) | 9 (24.3) | 0.05 | |
| Tea | 5 (38.5) | 27(73) | 0.028 | |
| Carbohydrates | 4 (30.8) | 28 (75.7) | 0.0038 | |
| Garlic | 1 (7.7) | 13 (35.1) | 0.05 | |
| Onion | 3 (23.1) | 23 (62.2) | 0.015 | |
| Yogurt | 2 (15.4) | 31 (83.8) | 0.93 | |
| Lentil | 9 (69.2) | 21 (56.8) | 0.41 | |
| Sausage | 4 (30.8) | 8 (21.6) | 0.50 | |
| Chicken meat | 12 (92.3) | 31 (83.8) | 0.43 | |

Table 4. Proportion and Correlation Analyses for Risk Factors in Subjects Positive for Fusobacterium Species $(n = 50)^{a}$

^a Data are presented as No. (%).

5. Discussion

Halitosis is associated with not only severe personal problems and social embarrassment but also periodontal disease (5). Therefore, accurate diagnosis of the underlying cause is important for effective therapy (12).

The relationship between the presence of Fusobacterium species in periodontal pockets and halitosis in our study is consistent with that observed in the study by Donaldson et al. who also isolated Fusobacterium species from patients with and without halitosis. However, the rate of bacterial isolation was slightly higher in the halitosis group in our study (P < 0.05)(13). The majority of isolates (82.3%, 14/17) in our study were those of F. nucleatum. F. nucleatum causes halitosis through the production of large amounts of VSCs and the accumulation of other VCS-producing bacteria (7, 14). In addition, it is often associated with periodontal diseases (14). Fusobacterium species with the ability to cause oral malodor belong to endogenous flora (7). Of the total Fusobacterium isolates in our study, 23.1% were those of F. *necrophorum*, a proportion close to that in the study by Gomes et al. (15). Although not confirmed, F. necrophorum has been predicted to be a part of the commensal flora in the oral cavity, considering its close association with infections in the head and neck region (16, 17).

In our study, there was a significant correlation between halitosis and job-related dryness of mouth (lack of water intake for several hours during work). A decrease in the salivary flow rate is generally considered a risk factor for bad breath because of the decreased cleansing action normally provided by the flow of saliva, with a concomitant change in the pattern of the oral flora (18). However, in our study, the presence of dry mouth was self-reported by subjects.

It was previously mentioned that oral hygiene maintenance (e.g., tooth brushing, flossing, and rinsing) could be an influential factor in the elimination or amelioration of halitosis, although there were no significant differences in this regard between subjects with and without halitosis in our study (Table 3) (19).

The correlation between systemic diseases and oral malodor, particularly sinusitis, was not strong in our study, although some authors have reported systemic diseases as possible risk factors (7, 20).

Several studies have reported strong correlations between halitosis and periodontal diseases, which are characterized by deep pockets that are considered sites for bacterial interactions and, consequently, greater VSC production (12, 21). However, there was no significant difference in the rate periodontal diseases between the halitosis and control groups in our study. Oral diseases were observed in up to 40% subjects in both groups, with an incidence of >50% among subjects positive for *Fusobacterium* species.

Halitophobia is a psychological problem used to describe apparently healthy individuals with a delusional fear of halitosis, even though it is not actually present (22). In our study, the majority (68%) of individuals with an organoleptic score of >2 before examination by a physician complained of bad breath. It can be concluded that the fear of halitosis is more likely to come true in those with the actual condition.

There are reports of a consistent association between halitosis and the consumption of volatile foods such as onion, garlic, and spices giving rise to transient changes in breath odor (1, 3). One of the main influential factors in the present study was the consumption of foods containing cysteine and methionine residues, which are potential nutrient sources for proteolytic anaerobic bacteria and subsequent VCS production (1, 7). Increased consumption of curry powder, chili, and sausage in individuals with halitosis is indicated to be a potential etiological factor (3).

The proportion of isolated anaerobic bacteria was reported to be significantly higher in individuals with active periodontal sites characterized by probing depths of ≥ 6 mm than in those with shallow and noninflamed sites (23). This may explain the significant difference in the Fusobacterium isolation rate between subjects who flossed and those who did not in the halitosis group in our study (Table 4).

In our study, there was no significant association between carbohydrate consumption and halitosis (Table 3). However, the consumption of carbohydrates was significantly higher in subjects negative for *Fusobacterium* species than in those positive for the same in the halitosis group (Table 4). Previously, it was documented that pH reduction by the production of acid compounds from carbohydrates, the main nutrients for oral bacteria, may inhibit the growth of proteolytic bacteria such as *Fusobacterium* (24-26). Moreover, Han et al. showed that the attachment of *F. nucleatum* to epithelial cells involves a lectin-like adhesin that can be inhibited by galactose-containing sugars (27). However, it has been demonstrated that the presence of fimbriae play a key role in the attachment of *F. necrophorum* to host cells (28).

Also, another notable finding in the halitosis group in our study was a significantly (P < 0.05) higher consumption of chili-containing foods, coffee, tea, garlic, and onion by subjects negative for *Fusobacterium* species than by those positive for the same. Such eating habits could be another cause of oral malodor regardless of the presence of *Fusobacterium*, because volatile foods such as onion and garlic can affect breath odor (3). This finding highlights the possible role of *Fusobacterium* species in the development of halitosis in the other group as well.

The present study has some limitations. First, we used conventional methods for the detection of bacterial isolates. Because of the fastidious nature of anaerobic bacteria, if molecular methods were also employed, they could improve our detection rates. Second, our sampling spot was limited to periodontal pockets, while anaerobic bacteria could colonize in other spots of the human oral cavity as well (23, 29). Finally, and most importantly, our sample size was small and the results cannot be generalized to the entire community. In summary, within the limitations, the results of this study suggest that the presence of *Fusobacterium* species in periodontal pockets is an important risk factor for halitosis and may be associated with some background factors that can contribute to halitosis. Therefore, the development of treatment strategies focused on *Fusobacterium* eradication may effectively prevent the progression of bad breath. However, until the optimal treatment is established, further studies should work toward determining the specific role of anaerobic bacteria other than *Fusobacterium* species.

Acknowledgements

We thank all the participants for their friendly cooperation in this study. This work was funded by the student research committee, Shiraz university of medical sciences, Iran (grant No. 91-6329).

References

- 1. Lee PP, Mak WY, Newsome P. The aetiology and treatment of oral halitosis: an update. *Hong Kong Med J.* 2004;**10**(6):414–8.
- 2. Krespi YP, Shrime MG, Kacker A. The relationship between oral malodor and volatile sulfur compound-producing bacteria. *Otolaryngol Head Neck Surg.* 2006;**135**(5):671–6.
- Porter SR. Diet and halitosis. Curr Opin Clin Nutr Metab Care. 2011;14(5):463-8.
- Loesche WJ, Kazor C. Microbiology and treatment of halitosis. Periodontol 2000. 2002;28:256–79.
- Nakano Y, Yoshimura M, Koga T. Correlation between oral malodor and periodontal bacteria. *Microbes Infect.* 2002;4(6):679–83.
- 6. Sahebjam Atabaki M, Moradi Haghgoo J, Khoshhal M, Arabi R, Khodadoostan A, Gholami L. Clinical Effect of Periodontal Pocket Irrigation with H2O2. *Avicenna J Dent Res.* 2011;**3**(1):53–9.
- Liu PF, Haake SK, Gallo RL, Huang CM. A novel vaccine targeting Fusobacterium nucleatum against abscesses and halitosis. *Vaccine*. 2009;27(10):1589–95.
- Amel Y, Bouziane D, Ahmed MLB. Microbiological Study of Periodontitis in the West of Algeria. World J Med Sci. 2010;5(1):7-12.
- Morgenstein AA, Citron DM, Finegold SM. New medium selective for Fusobacterium species and differential for Fusobacterium necrophorum. J Clin Microbiol. 1981;13(4):666–9.
- 10. Walker CB, Ratliff D, Muller D, Mandell R, Socransky SS. Medium for selective isolation of Fusobacterium nucleatum from human periodontal pockets. *J Clin Microbiol.* 1979;**10**(6):844–9.
- Church D. Aerobic Bacteriology. In: Isenberg H, editor. Clinical Microbiology Procedures Handbook. 2th ed. Garcia L. ASM Press; 2007.
- Cortelli JR, Barbosa MD, Westphal MA. Halitosis: a review of associated factors and therapeutic approach. *Braz Oral Res.* 2008;22

Suppl 1:44-54.

- Donaldson AC, McKenzie D, Riggio MP, Hodge PJ, Rolph H, Flanagan A, et al. Microbiological culture analysis of the tongue anaerobic microflora in subjects with and without halitosis. Oral Dis. 2005;11 Suppl 1:61–3.
- Signat B, Roques C, Poulet P, Duffaut D. Fusobacterium nucleatum in periodontal health and disease. *Curr Issues Mol Biol.* 2011;13(2):25–36.
- Gomes BP, Pinheiro ET, Gade-Neto CR, Sousa EL, Ferraz CC, Zaia AA, et al. Microbiological examination of infected dental root canals. Oral Microbiol Immunol. 2004;19(2):71–6.
- Batty A, Wren MW, Gal M. Fusobacterium necrophorum as the cause of recurrent sore throat: comparison of isolates from persistent sore throat syndrome and Lemierre's disease. J Infect. 2005;51(4):299–306.
- Falkler WJ, Enwonwu CO, Idigbe EO. Isolation of Fusobacterium necrophorum from cancrum oris (noma). Am J Trop Med Hyg. 1999;60(1):150–6.
- Rad M, Kakoie S, NiliyeBrojeni F, Pourdamghan N. Effect of Longterm Smoking on Whole-mouth Salivary Flow Rate and Oral Health. J Dent Res Dent Clin Dent. 2010;4(4):110–4.
- Aylikci BU, Colak H. Halitosis: From diagnosis to management. J Nat Sci Biol Med. 2013;4(1):14–23.
- 20. Washio J, Sato T, Koseki T, Takahashi N. Hydrogen sulfide-producing bacteria in tongue biofilm and their relationship with oral malodour. *J Med Microbiol*. 2005;**54**(Pt 9):889–95.
- Takeshita T, Suzuki N, Nakano Y, Shimazaki Y, Yoneda M, Hirofuji T, et al. Relationship between oral malodor and the global composition of indigenous bacterial populations in saliva. *Appl Envi*ron Microbiol. 2010;**76**(9):2806–14.
- Akpata O, Omoregie OF, Akhigbe K, Ehikhamenor EE. Evaluation of oral and extra-oral factors predisposing to delusional halitosis. *Ghana Med J.* 2009;43(2):61–4.
- Tanaka M, Yamamoto Y, Kuboniwa M, Nonaka A, Nishida N, Maeda K, et al. Contribution of periodontal pathogens on tongue dorsa analyzed with real-time PCR to oral malodor. *Microbes Infect.* 2004;6(12):1078–83.
- 24. Gnanasekhar JD. Aetiology, diagnosis and management of halitosis: a review. *Periodontal Pract Today*. 2007;4(3)
- 25. Bradshaw DJ, McKee AS, Marsh PD. Effects of carbohydrate pulses and pH on population shifts within oral microbial communities in vitro. *J Dent Res.* 1989;**68**(9):1298–302.
- 26. Doran A, Kneist S, Verran J. Ecological control: in vitro inhibition of anaerobic bacteria by oral streptococci. *Microb Ecol Health Dis.* 2004;**16**(1):23-7.
- 27. Han YW, Shi W, Huang GT, Kinder Haake S, Park NH, Kuramitsu H, et al. Interactions between periodontal bacteria and human oral epithelial cells: Fusobacterium nucleatum adheres to and invades epithelial cells. *Infect Immun.* 2000;**68**(6):3140–6.
- 28. Riordan T. Human infection with Fusobacterium necrophorum (Necrobacillosis), with a focus on Lemierre's syndrome. *Clin Microbiol Rev.* 2007;**20**(4):622–59.
- 29. Rani A, Chopra A. Isolation and identification of root canal bacteria from symptomatic non-vital teeth with periapical pathosis. *Endodontol.* 2006;1:12-7.