

Resistance Rate and Minimum Inhibitory Concentration of Metronidazole Among *Helicobacter pylori* Strains in Tehran, Iran

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Abstract

Background: Constant monitoring of *Helicobacter pylori* resistance is necessary for selection of the best treatment regimen for eradication of the resistant strains.

Objectives: The aim of this study was to investigate diversity of resistance and minimum inhibitory concentration (MIC) of *H. pylori* strains against metronidazole in Tehran, Iran.

Patients and Methods: This cross-sectional study was performed among 96 patients, who had undergone gastric endoscopy at Imam Khomeini hospital in Tehran, during years 2013 to 2014. *Helicobacter pylori* isolates were obtained from gastric biopsy samples on selective culture media after characterization by conventional biochemical tests and polymerase chain reaction. Minimum inhibitory concentration of metronidazole was determined by the agar dilution method.

Results: *Helicobacter pylori* infection was detected in 22 patients (22.92%). Identity of the isolates was confirmed by PCR using *glmM* primers. Chronic gastritis, duodenitis, intestinal metaplasia, dysplasia and cancer were detected among 70.1%, 6.25%, 1.04%, 1.4% and 10.41% of the patients, respectively. Smoking showed a negative relationship with *H. pylori* infection. The rate of antibiotic resistance was 81.8% (18/22) and MIC ranges of 8 to 512 $\mu\text{g}/\text{mL}$ were detected. Furthermore, MIC₅₀ and MIC₉₀ were determined as 256 and 512 $\mu\text{g}/\text{mL}$, respectively.

Conclusions: In conclusion, these results suggested a need for a switch to second line therapy regimens for treatment of infected patients in the Iranian population.

Keywords: Microbial Sensitivity Tests, Metronidazole, *Helicobacter pylori* Infection

1. Background

Helicobacter pylori are microaerophilic gram-negative bacteria that infect more than half of the world's population. This pathogen has been categorized as a group I carcinogen by the international agency for research on cancer and its infection occurs more frequently in developing countries than in industrialized countries (1). Colonization in the human stomach commonly leads to gastritis that could progress to ulcer in the duodenum or the stomach. In some cases, the disease can finally cause mucosa associated lymphoid tissue (MALT) lymphomas or gastric carcinoma in nearly 1% of the patients (2). Although infection with *H. pylori* is a global health problem, socioeconomic class, age, and health status affects its prevalence in

different countries (3). In Iran, it was estimated that 80% of the adult population have a history of infection with these bacteria (4). However, recent studies revealed that this rate has decreased up to 40% in some cities with higher socioeconomic status (5).

Efforts to improve health status and administration of appropriate doses of antibiotic regimens for eradication therapy could be effective in the control of *H. pylori* infection in developing countries. The eradication can be achieved by using combination therapies consisting of a proton pump inhibitor or bismuth citrate and two antibiotics, such as amoxicillin (AML), clarithromycin (CLR) or metronidazole (MTZ). Failure of eradication therapy in these patients is mainly due to the resistance property of the responsible strains to the prescribed antibiotics (6).

Eradication failure in these patients is commonly associated with recurrence of the infection by those strains that are resistant to the prescribed drugs. Since resistance to antibiotics is the main reason for treatment failure, knowledge about minimal inhibitory concentration (MIC) values of the prescribing antibiotics for bacterial strains could help clinicians select the best choice of medication in each geographic region (7).

Metronidazole is one of the main drugs commonly prescribed for this infection worldwide. Resistance of *H. pylori* to metronidazole is a growing medical problem in Iran and various parts of the world. Previous studies illustrated that metronidazole resistance is very high in Africa (80% - 100%) and Asia (50% - 95%) (8). Different rates of resistance to metronidazole were reported in the United States, ranging from about 20% to more than 50%. Similarly, the collected data showed an increasing trend of resistance to metronidazole in the recent years in Iran (94%) (9). Eradication rate of triple therapy regimens was found as 97% in metronidazole-susceptible strains, which was much higher than the value for the metronidazole-resistant strains (72.6%) (10). According to the latest guideline for *H. pylori* medication, the prescribed regimens should be selected based on resistance of *H. pylori* strains to metronidazole and clarithromycin in each geographic region (11). Therefore, constant monitoring of infection rate and resistance property of the responsible strains is necessary to choose the best treatment option for eradication of resistant cases in each country. While there are some data about this resistance in different cities of Iran, lack of data exists for their MIC levels at a broad range of antibiotic concentrations. In this study, we provided these data to help physicians with better management of *H. pylori* infection in Iran.

2. Objectives

The present study aimed to investigate the susceptibility of *H. pylori* strains and their MIC values among infected patients with different gastric disorders in Tehran, Iran.

3. Patients and Methods

3.1. Patients and Bacterial Strains

In this cross-sectional study, *H. pylori* infection was investigated among 96 dyspeptic patients, who had referred to the Endoscopy Unit of a governmental hospital (Imam Khomeini hospital) in Tehran, Iran, during year 2013. Simple random sampling was used for the patients and data were collected using a standardized questionnaire. All the

patients signed an informed consent form. Exclusion criteria for the study included usage of antibiotics and proton pump inhibitors within the last two weeks prior to the endoscopy, and bleeding from the gastrointestinal tract. Endoscopy was requested and biopsy sampling was performed for the patients, based on the standards of practice committee of the American society for gastrointestinal endoscopy (12, 13). The biopsies were transported to the laboratory in a sterile tube containing semi solid Thioglycolate medium (Merck, Homburg, Germany) supplemented with 3% yeast extract (Oxoid Ltd., Basingstoke, UK) for culture in a selective medium as described below. This study was approved by the ethics committee of Shahid Beheshti University of Medical Sciences (SBMU 721).

3.2. Isolation and Identification of *Helicobacter pylori*

The biopsy samples were gently homogenized and cultured on Brucella agar medium (Merck, Homburg, Germany) supplemented with 5% defibrinated sheep blood, 7% fetal bovine serum (FCS), amphotericin B (10 mg, Sigma-Aldrich, USA) and *Campylobacter* selective supplement consisting of vancomycin 2.0 mg, polymyxin 0.05 mg and trimethoprim 1.0 mg (Merck, Homburg, Germany), as a selective medium for *H. pylori* strains. The inoculated plates were incubated at 37°C under microaerobic conditions (5% O₂, 10% CO₂, and 85% N₂) in a CO₂ incubator (Innova-Co 170; USA). The cultures were examined after three to seven days for observation of pinpoint (1 - 2 mm) glistening colonies. The bacterial strains were identified as *H. pylori* on the basis of data regarding their spiral microscopic appearance, as well as positive activities of urease, oxidase, catalase tests and polymerase chain reaction results that were obtained using species specific primers (Section 3.2; 10). Individual cultures representing colonies from each patient was frozen at -70°C in brain heart infusion broth (Merck Co, Germany) containing 15% glycerol (Merck Co, Germany) and FCS (20%) for further studies.

3.3. DNA Extraction and Polymerase Chain Reaction Amplification Analysis

To confirm identity of the bacterial isolates, DNA extraction was performed on freshly grown colonies of the isolates using QIAamp tissue DNA extraction kit, according to the manufacturer's instructions (Qiagen, Hilden, Germany). The PCR was done by primers targeting *glmM* (*ureC*), which amplify a 296-bp fragment. The PCR was performed in a final volume of 25 µL containing 2.5 µL PCR buffer (10 X), 0.3 µL of each primer, forward primer: GlmM2-F 5[U+02B9]-GGATAAGCTTTTAGGGGTGTTAGGGG and reverse primer: GlmM1-R 5[U+02B9]-GCTTACTTTCTAACACTAACGCGC, 1

mM MgCl₂; 0.3 μL each deoxyribonucleotide triphosphate (dNTP), 0.2 μL Taq DNA polymerase, and 3 μL DNA sample. It was implemented in a thermocycler (AG 22331; Eppendorf, Hamburg, Germany) under the following conditions: initial denaturation for four minutes at 94°C followed by 30 cycles at 94°C for one minute, 57°C for 45 seconds and 72°C for one minute. After a final extension at 72°C for ten minutes, electrophoresis was done on 1.2% agarose gel, according to standard procedures. DNA of the *H. pylori* strain RIGLD 245 (GenBank accession number JQ765441.1) was used as an internal control for the PCR assay.

3.4. Antimicrobial Susceptibility Testing

Susceptibility of the bacterial strains was determined based on guidelines of the European committee on antimicrobial susceptibility testing (EUCAST) (14). Antimicrobial susceptibility testing was performed using the agar dilution method. Accordingly, a stock solution of metronidazole (MAST, London, United Kingdom) was prepared in accordance with the manufacturer's instructions and the working solutions were made in distilled water at appropriate concentrations immediately before incorporation into sterile Muller Hinton agar medium (MHA, 45°C) in ranges of 8 to 512 μg/mL at two-fold dilutions (6). To inoculate the plates, an inoculum of freshly prepared *H. pylori* suspension was adjusted to a density corresponding to 2 McFarland's turbidity standard tube in normal saline and then 5 μL of bacterial suspension was inoculated on Muller-Hinton agar (Merck, Germany). The inoculated plates were left at room temperature for 10 minutes for drying and incubated at 37°C in 5% CO₂ for 48-72 hours. The EUCAST clinical breakpoint of > 8 μg/mL was defined for evaluation of the resistant strains and MIC values were recorded in each case (14). *Helicobacter pylori* strain RIGLD 245 was used as a reference strain for all the experiments.

3.5. Data Analysis

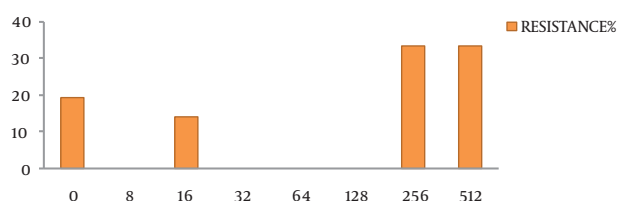
Fisher's exact test was used for analysis of the categorical data. Bootstrap confidence interval was measured using the StatKey software. Analyses were done using statistical analysis for windows V2.03 (SPSS, Chicago, IL, USA). P values of less than 0.05 were considered statically significant.

4. Results

Out of the 96 studied patients, consisting of 52 males (54.1%) and 44 females (45.8%), *H. pylori* infection were detected in 22 patients (22.92%). The isolates indicated positive results for the common identification tests and *H. pylori*-specific PCR (*glmM*). The ages of the patients ranged

between 15 and 88 years. Most of the infected patients suffered from chronic gastritis (70.1%), while the others showed duodenitis (6.25%), intestinal metaplasia (1.04%), dysplasia (1.4%) and cancer (10.41%). In this study, smoking was negatively related to *H. pylori* infection. In other words, non-smokers had a 2.9-fold higher risk of *H. pylori* infection than smokers. The rate of antibiotic resistance in the current study was 81.8% (18/22). Minimum inhibitory concentration ranges of 8 to 512 μg/mL were detected for the 22 *H. pylori* strains against metronidazole (Figure 1). On the basis of our findings, resistance to metronidazole was equal in the female and male patients. An MIC of ≥ 512 μg/mL was detected among 40.9% of the metronidazole-resistant strains. Furthermore, MIC₅₀ and MIC₉₀ of all the isolates were determined as 256 and 512 μg/mL, respectively (Table 1). In our study, no significant correlation was found between resistance to metronidazole and gender or owning a pet (P > 0.05, CI, 0.003) (Table 2).

Figure 1. Resistance Rates of *Helicobacter pylori* Strains to Metronidazole in Iran



X axis: Concentration of metronidazole (μg/mL); Y axis: The percentage of resistance strains.

5. Discussion

H. pylori are the main pathogens for gastroduodenal diseases. It is now well-known that late stages of *H. pylori* infection may cause gastric atrophy with intestinal metaplasia, and that this injury probably predispose an individual to gastric cancer. Metronidazole is the main antibiotic used in numerous eradication therapy regimens for treatment of *H. pylori* infection; however increasing resistance to this antibiotic is a worldwide problem. It appears that the main resistance to metronidazole has been attributed to frequent use of this drug, which is regularly prescribed for other diseases and infections. For this reason, metronidazole has been omitted from first-line experiential therapy strategies in some countries (15). Prolonged amoxicillin-based standard triple therapy was recommended by the European guidelines for countries with > 40% metronidazole resistant *H. pylori* strains (16). In the present study, a high rate of resistance to metronidazole was characterized among the Iranian *H. pylori* strains,

Table 1. Susceptibility of *Helicobacter pylori* Strains to Metronidazole at Different Concentrations

Gender	MTZ Resistance No. (%)	MIC \leq 8	MIC = 16 - 128	MIC = 256	MIC = 512	MIC ₅₀ /MIC ₉₀
Males, n = 12	9 (40.90)	-	-	-	-	-
Females, n = 10	9 (40.90)	-	-	-	-	-
Total, n = 22	18 (81.80)	4 (18.1)	3 (13.6)	7 (31.8)	8 (36.3)	256/512

Abbreviations: MTZ, metronidazole; MIC, minimum inhibitory concentration ($\mu\text{g}/\text{mL}$); MIC₅₀, the MIC value at which 50% of the tested strains were inhibited; MIC₉₀, the MIC value at which 90% of the tested strains were inhibited.

Table 2. Clinicopathologic Characteristics of the Patients With *Helicobacter pylori* Infection and Diversity of Minimum Inhibitory Concentrations Values

Variable	Frequency, No. (%)	Positive Culture, No. (%)	Negative Culture, No. (%)	Resistance to Metronidazole, MIC, $\mu\text{g}/\text{mL}$
Gender				
Female	44 (45.8)	10 (10.41)	34 (35.41)	$\leq 8 - \geq 512$
Male	52 (54.1)	12 (12.5)	40 (41.66)	$\leq 8 - \geq 512$
Atrophy	7 (7.2)	1 (1.04)	6 (6.25)	256
Chronic gastritis	46 (47.91)	12 (12.5)	34 (35.41)	≥ 512
Severe active gastritis	2 (2.08)	NA	2 (2.08)	NA
Intestinal metaplasia	2 (2.08)	NA	2 (2.08)	NA
Antibiotic usage	9 (9.37)	0	9 (9.37)	NA
PPI usage	7 (7.29)	2 (2.08)	5 (5.2)	$\leq 8 - \geq 256$
Cancer	9 (9.37)	2 (2.08)	7 (7.2)	256 - ≥ 512
Peptic ulcer	28 (29.16)	4 (4.1)	24 (25)	256 - ≥ 512
Moderate gastritis	1 (1.04)	NA	1 (1.04)	256 - ≥ 512

Abbreviation: NA, not available.

which is higher than those reported previously from Iran. There are variable resistance rates in different countries. While a higher resistance rate was reported in India (85%), a lower rate was determined in the USA (33.9%) during years 2000 to 2012 (8, 10). These differences were also seen among different cities of Iran. In the north of Iran, resistance to metronidazole was reported as 73.4% during 2007 to 2010 (17); however the rate of resistance in south of Iran (Shiraz) decreased from 72.6% in 2007 to 44% in 2010 (14, 18). Mirzaei et al. (19) reported a resistance rate of 56.3% to metronidazole in Isfahan in June 2013, yet the resistance rate was higher in Tehran, Iran (78% and 61.3%) (6). Comparison of data from different studies is difficult, as in many instances a different methodology has been used (20-22). Further studies are needed to confirm these results in a higher sample collection from different provinces of Iran. Since practical treatment is ordinarily used in clinical therapies without any care about resistance rate of *H. pylori* strains in Iran, there is an increasing risk of drug resistance. Results of our study confirmed increased levels of resistance rate and MIC₅₀ and MIC₉₀ values among these

strains that was higher than those reported previously in Tehran during year 2011 (16 and $> 32 \mu\text{g}/\text{mL}$, respectively) (23). Therefore, monitoring of antibiotic resistance seems to be essential to present appropriate treatment regimens. Prescription of treatment regimens based on the susceptibility tests will reduce chronic infection that consequently could limit emergence of new resistant variants of *H. pylori* in the infected patients. Although this will make management more expensive, it will be cost effective since it will provide better eradication of the organism.

In conclusion, these results showed an increased rate of resistance to metronidazole and MIC values among the *H. pylori* strains in Tehran, which proposed a switch to second line therapy for treatment of this infection by physicians in Iran. Better management of the infection depends on detection of *H. pylori* eradication rates based on the prescribed regimens and performance of antimicrobial susceptibility tests.

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Footnotes

Authors' Contribution: Study concept and design: Masoud Alebouyeh and Arash Mahboubi; experiment: Mahnaz Shahabi Mehr and Nastaran Farzi; acquisition of data: Mahnaz Shahabi Mehr; drafting of the manuscript: Mahnaz Shahabi Mehr and Nastaran Farzi; critical revision of the manuscript: Masoud Alebouyeh; statistical analysis: Mahnaz Shahabi Mehr and Masoud Alebouyeh; study supervision: Masoud Alebouyeh; material support and specimen collect: Mahnaz Shahabi Mehr, Mohammad Reza Zali, Arash Mahboubi and Reza Taslimi.

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References

- Vaziri F, Najar Peerayeh S, Alebouyeh M, Mirzaei T, Yamaoka Y, Molaie M, et al. Diversity of *Helicobacter pylori* genotypes in Iranian patients with different gastroduodenal disorders. *World J Gastroenterol*. 2013;**19**(34):5685-92. doi: [10.3748/wjg.v19.i34.5685](#).
- Yadegar A, Mobarez Mohabati A, Alebouyeh M, Mirzaei T, Kwok T, Zali MR. Clinical relevance of *cagL* gene and virulence genotypes with disease outcomes in a *Helicobacter pylori* infected population from Iran. *World J Microbiol Biotechnol*. 2014;**30**(9):2481-90. doi: [10.1007/s11274-014-1673-5](#).
- Mirzaei T, Alebouyeh M, Shokrzadeh L, Aghdaei HA, Farzi N, Zojaji H, et al. Smoking, proton pump inhibitors and antibiotic administration as factors affecting direct screening of *Helicobacter pylori* infection among patients with dyspepsia. *Arch Clin Infect Dis*. 2014;**9**(2) doi: [10.5812/archcid.15774](#).
- Daryani Ebrahimi N, Taher M, Shirzad S. *Helicobacter pylori* infection: A review. *Arch Clin Infect Dis*. 2011;**6**(1):56-64.
- Farzi N, Malekian T, Alebouyeh M, Vaziri F, Zali MR. Genotype Diversity and Quasispecies Development of *Helicobacter pylori* in a Single Host. *Jpn J Infect Dis*. 2015;**68**(3):176-80. doi: [10.7883/yoken.JJID.2014.165](#). [PubMed: [25672355](#)].
- Shokrzadeh L, Alebouyeh M, Mirzaei T, Farzi N, Zali MR. Prevalence of multiple drug-resistant *Helicobacter pylori* strains among patients with different gastric disorders in Iran. *Microb Drug Resist*. 2015;**21**(1):105-10. doi: [10.1089/mdr.2014.0081](#).
- Toracchio S, Cellini L, Di Campi E, Cappello G, Malatesta MG, Ferri A, et al. Role of antimicrobial susceptibility testing on efficacy of triple therapy in *Helicobacter pylori* eradication. *Aliment Pharmacol Ther*. 2000;**14**(12):1639-43. [PubMed: [11121913](#)].
- De Francesco V, Giorgio F, Hassan C, Manes G, Vannella L, Panella C, et al. Worldwide *H. pylori* antibiotic resistance: a systematic review. *J Gastrointest Liver Dis*. 2010;**19**(4):409-14. [PubMed: [21188333](#)].
- Sirous M, Mehrabadi JF, Daryani NE, Eshraghi S, Hajikhani S, Shirazi MH. Prevalence of antimicrobial resistance in *Helicobacter pylori* isolates from Iran. *Afr J Biotechnol*. 2013;**9**(36).
- Yang JC, Lu CW, Lin CJ. Treatment of *Helicobacter pylori* infection: current status and future concepts. *World J Gastroenterol*. 2014;**20**(18):5283-93. doi: [10.3748/wjg.v20.i18.5283](#). [PubMed: [24833858](#)].
- Alebouyeh M, Yadegar A, Farzi N, Miri M, Zojaji H, Gharibi S, et al. Impacts of *H. pylori* mixed-infection and heteroresistance on clinical outcomes. *Gastroenterol Hepatol Bed Bench*. 2015;**8**(Suppl 1):S1-5. [PubMed: [26171132](#)].
- Asge Standards of Practice Committee, Early DS, Ben-Menachem T, Decker GA, Evans JA, Fanelli RD, et al. Appropriate use of GI endoscopy. *Gastrointest Endosc*. 2012;**75**(6):1127-31. doi: [10.1016/j.gie.2012.01.011](#). [PubMed: [22624807](#)].
- Asge Standards of Practice Committee, Sharaf RN, Shergill AK, Odze RD, Krinsky ML, Fukami N, et al. Endoscopic mucosal tissue sampling. *Gastrointest Endosc*. 2013;**78**(2):216-24. doi: [10.1016/j.gie.2013.04.167](#). [PubMed: [23867371](#)].
- Kahlmeter G, Brown DF, Goldstein FW, MacGowan AP, Mouton JW, Odenholt I, et al. European Committee on Antimicrobial Susceptibility Testing (EUCAST) Technical Notes on antimicrobial susceptibility testing. *Clin Microbiol Infect*. 2006;**12**(6):501-3. doi: [10.1111/j.1469-0691.2006.01454.x](#). [PubMed: [16700696](#)].
- Torres-Debat ME, Perez-Perez G, Olivares A, Fernandez L, Raisler K, Gonzalez N, et al. Antimicrobial susceptibility of *Helicobacter pylori* and mechanisms of clarithromycin resistance in strains isolated from patients in Uruguay. *Rev Esp Enferm Dig*. 2009;**101**(11):757-62. [PubMed: [20001152](#)].
- Malferteiner P, Megraud F, O'Morain C, Bazzoli F, El-Omar E, Graham D, et al. Current concepts in the management of *Helicobacter pylori* infection: the Maastricht III Consensus Report. *Gut*. 2007;**56**(6):772-81. doi: [10.1136/gut.2006.101634](#). [PubMed: [17170018](#)].
- Haghi Tomatari F, Mohabbati Mobarez A, Amini M, Hosseini D, Talebi Bezhmin Abadi A. *Helicobacter pylori* resistance to metronidazole and clarithromycin in dyspeptic patients in Iran. *Iran Red Crescent Med J*. 2010;**12**(4):409.
- Farshad S, Alborzi A, Japoni A, Ranjbar R, Hosseini Asl K, Badiie P, et al. Antimicrobial susceptibility of *Helicobacter pylori* strains isolated from patients in Shiraz, Southern Iran. *World J Gastroenterol*. 2010;**16**(45):5746-51. [PubMed: [21128326](#)].
- Mirzaei N, Poursina F, Faghri J, Talebi M, Khataminezhad MR, Hasan-zadeh A, et al. Prevalence of resistance of *Helicobacter pylori* strains to selected antibiotics in Isfahan, Iran. *Jundishapur J Microbiol*. 2013;**6**(5) doi: [10.5812/jjm.6342](#).
- Kohanteb J, Bazargani A, Saberi-Firooz M, Mobasser A. Antimicrobial susceptibility testing of *Helicobacter pylori* to selected agents by agar dilution method in Shiraz-Iran. *Indian J Med Microbiol*. 2007;**25**(4):374-7. [PubMed: [18087088](#)].
- Falsafi T, Mobasher F, Nariman F, Najafi M. Susceptibilities to different antibiotics of *Helicobacter pylori* strains isolated from patients at the pediatric medical center of Tehran, Iran. *J Clin Microbiol*. 2004;**42**(1):387-9. [PubMed: [14715786](#)].
- Mohammadi M, Doroud D, Mohajerani N, Massarrat S. *Helicobacter pylori* antibiotic resistance in Iran. *World J Gastroenterol*. 2005;**11**(38):6009-13. [PubMed: [16273615](#)].
- Sayadi S, Darboue M, Dabiri H, Shokrzadeh L, Mirzaee T, Alebouyeh M, et al. Study of antibiotic resistant *H. pylori* isolated from Iranian patients during 2009-2010. *Healthmed*. 2011;**5**(6):1970-6.