Published online 2017 April 30.

Brief Report

Characterization of 5 Episodes of Vancomycin Nonsusceptible *Streptococcus pneumoniae* From Clinical Isolates in Tehran, Iran

Ali Nazari Alam,¹ Sedighe Rafiei Tabatabaii,² Ali Hashemi,³ Masoud Yousefi,⁴ and Seyedeh Mahsan

Hoseini Alfatemi^{2,*}

¹Department of Microbiology, School of Medicine, Kashan University of Medical Sciences, Kashan, IR Iran
²Pediatric Infections Research Center, Research Institute for Children Health, Shahid Beheshti University of Medical Sciences, Tehran, Iran
³Department of Microbiology, School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, IR Iran
⁴Department of Microbiology, School of Medicine, Birjand University of Medical Sciences, Birjand, IR Iran

^{*} Corresponding author: Seyedeh Mahsan Hoseini Alfatemi, Pediatric Infections Research Center, Mofid Children's Hospital, Shahid Beheshti University of Medical Sciences, 15468-15514, Tehran, Iran. Tel/Fax: +98-22226941, E-mail: mahsan.hoseinialfatemi@gmail.com

Received 2016 June 19; Revised 2017 February 08; Accepted 2017 February 12.

Abstract

Background: *Streptococcus pneumoniae* is an important bacterial pathogen in children and older adults. It is associated with the highest case fatality rates in patients with pneumonia and meningitis.

Objectives: In the present study, we determined the phenotypic and genotypic properties of a vancomycin nonsusceptible *S. pneumoniae* isolates from clinical specimens in Tehran, Iran.

Methods: Five pneumococcal isolates were included in the study from different clinical specimen in 2 hospitals in Tehran (Milad and Sina), Iran. Antimicrobial susceptibility tests of pneumococcal isolates were performed by the broth microdilution method according to CLSI guideline, and the presence of *vanA* gene was detected by PCR.

Results: During the study period, 5 *S. pneumoniae* isolates resistance to vancomycin were identified. Vancomycin minimum inhibitory concentrations (MIC) ranged from 2 μ g/mL to 16 μ g/mL. All the isolates were resistant to cefotaxime and erythromycin. Moreover, only 1 isolate was susceptible to penicillin. PCR results showed that all isolates were negative for *vanA* gene.

Conclusions: Our results demonstrated that an alarming rate of vancomycin resistant pneumococci may result from the uncontrolled use of vancomycin and self-medication. Moreover, multiple drug resistant pneumococci were observed especially in cefotaxime and erythromycin, which may be a major health problem in Iranian patients.

Keywords: Antibiotic Resistance, Vancomycin, vanA Gene

1. Background

Streptococcus pneumoniae is an important bacterial pathogen in children and older adults (1). The *S. pneumoniae* is a leading cause of bacterial meningitis, pneumonia, bacteremia, and acute otitis media. It is associated with the highest case fatality rate in patients with pneumonia and meningitis (2). In the recent years, penicillin nonsusceptible and multiple antibiotic resistance clinical isolates have emerged in many parts of the world (3-5). Alterations in the penicillin-binding proteins lead to a decrease in the bacteria's affinity for the antibiotics (6). Therefore, increasing resistance rate may cause serious problems in the treatment of pneumococcal infections (6).

Antibiotics such as cephalosporins and vancomycin serve as alternative therapeutic agents in infections with

clinical isolates resistance to other antibiotics such as penicillin (7). In addition, emerging resistance to beta-lactam, resistance to macrolides, tetracycline and trimethoprimsulfamethoxazole were reported (8). Increase in the use of vancomycin for invasive pneumococcal infections may help develop resistance to this important drug; moreover, vancomycin tolerated strains of *S. pneumoniae* have been characterized in the recent studies. To date, vancomycinresistant strains of pneumococci have not been seen, but strains of S pneumoniae tolerant to vancomycin have been reported (9).

2. Objectives

In the present study, we aimed at determining the phenotypic and genotypic properties of vancomycin nonsus-

Copyright © 2017, Archives of Clinical Infectious Diseases. This is an open-access article distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 International License (http://creativecommons.org/licenses/by-nc/4.0/) which permits copy and redistribute the material just in noncommercial usages, provided the original work is properly cited.

ceptible *S. pneumoniae* isolates obtained from clinical isolates in Tehran, Iran.

3. Methods

3.1. Bacterial Isolation and Identification

This was a cross-sectional descriptive study conducted during November 2014 and March 2016 on 5 pneumococcal isolates (isolates selected from a total of 73 S. pneumoniae and collected during the study period), which were obtained from different clinical specimens in 2 hospitals (Milad and Sina) in Tehran, Iran. The isolates were transported to department of microbiology affiliated to Shahid Beheshti University of Medical Sciences for further analysis. All suspected S. pneumoniae isolates were streaked for single colonies on blood agar, supplemented with 5% sheep's blood (Merck, Germany, Lot No. VM1287860091). All plates were incubated for 24 to 48 hours at 37°C in candle jar. All isolates were identified as S. pneumoniae based on colonial morphology, hemolysis, gram staining, bile solubility, and susceptibility to optochin disc (1 μ g, Mast, UK, Lot No.303005). S. pneumoniae ATCC 49619 was used for quality control. Identification of the isolates as S. pneumoniae was confirmed by PCR for the cpsA gene (coding for capsular polysaccharide biosynthesis) by speciesspecific primers (8). A medium containing skim milk, tryptone, glucose, and glycerin (STGG, Merck, Germany, Lot No. VL101263934) was used for the storage of bacteria at -70°C.

3.2. Antimicrobial Susceptibility Testing

Antimicrobial susceptibility tests of pneumococcal isolates were performed by the broth microdilution method according to guidelines of the clinical and laboratory standards institute (CLSI) (10) for vancomycin, penicillin, cefotaxime, ceftriaxone, erythromycin, trimethoprim-sulfamethoxazol, ofloxacin, and meropenem. The bacterial isolates were incubated overnight on 5% sheep blood agar plates. Inoculants were prepared from the bacterial colonies that formed on the 5% sheep blood agar plates, and then the colonies were suspended in 0.9% saline with a turbidity equivalent of 0.5 McFarland units. The final inoculum density was of 5×10^5 CFU/mL in each test tube. Antimicrobials agents had been diluted in cation-adjusted Mueller-Hinton broth (BD; USA), with 5% lysed horse blood. The following concentrations for the antimicrobials were used: 32 μ g/mL, 16 μ g/mL, 8 μ g/mL, 4 μ g/mL, 2 μ g/mL, 1 μ g/mL, 0.5 μ g/mL, 0.25 μ g/mL, 0.12 μ g/mL, and 0.06 μ g/mL. The microplates were incubated at 35° C in ambient air for 20 to 24 hours.

3.3. DNA Extraction and PCR Assay

Bacterial DNA extraction was done by high pure PCR template preparation kit (Roche, Germany). The extracted DNA suspension was kept frozen at -70 °C until further use. The polymerase chain reaction (PCR) was performed to detect *vanA* gene, which was described previously (9).

3.4. Statistical Analysis

Analysis was performed using SPSSTM software, version 21.0 (IBM Corp., USA). The results are presented as descriptive statistics in relative frequency. Values were expressed as the mean \pm standard deviation (continuous variables) or percentages of the group (categorical variables).

4. Results

During the study period, 5 *S. pneumoniae* clinical isolates, which were nonsusceptible to vancomycin, were identified. Table 1 demonstrates the full antibiotic susceptibility pattern and clinical source of pneumococcal isolates. Vancomycin minimum inhibitory concentrations (MIC) for tested isolates ranged from 2 μ g/mL to 16 μ g/mL. In total, 80%, 60%, and 40% of the isolates were susceptible to ofloxacin, penicillin, and trimethoprimsulfamethoxazol, respectively. The only penicillin resistant isolate was susceptible to trimethoprim-sulfamethoxazol and resistant to other tested antibiotics. Moreover, all the isolates were resistant to cefotaxime and erythromycin. Finally, our findings showed that all vancomycin nonsusceptible isolates with MIC $\geq 1 \mu$ g/mL were PCR negative for *vanA* gene (Figure 1).

5. Discussion

Antimicrobial resistant pneumococcal isolates are becoming more common throughout the world, especially in developing countries (11). Although some studies indicated that penicillin and cephalosporin are often included in the initial empirical antibiotics regimen for patients with pneumococcal infections, penicillin and cephalosporin-resistant S. pneumoniae isolate are being isolated with an increasing level and become a serious health problem around the world (12). Insufficient information on distribution of antibiotic resistant pneumococci in our geographical area necessitates a serious attention to estimate burden of resistant isolates. In the present study, we investigated a total of 5 S. pneumoniae clinical isolates, which showed resistance to vancomycin with high level of MIC. Our results demonstrated an alarming rate of vancomycin resistant pneumococci, which might have

Isolates	Gender	Age	Sample	VanMIC, μ g/mL	PEN	стх	CRO	ERY	SXT	OFX	MER
No. 1	F	17 <	BAL	2	Ι	R	R	R	I	S	R
No. 2	F	17 >	BAL	2	S	R	I	R	S	S	R
No. 3	F	17 >	Eye	16	R	R	R	R	s	R	R
No. 4	М	17 <	Blood	16	S	R	R	R	R	S	I
No. 5	М	17 >	CSF	2	S	R	R	R	I	S	I

Table 1. Antibiotic Susceptibility Pattern and Characteristics of Five Non-Susceptible Vancomycin S. pneumonia

Abbreviations: BAL, bronchoalveolar lavage; CTX, cefotaxime; CRO, ceftriaxone; CSF, Cerebrospinal fluid; ERY: erythromycin; F, female; I, Intermediate; M, male; MER, meropenem; OFX, ofloxacin; PEN, penicillin; R, Resistant; S, Susceptible; SXT, trimethoorim-sulfamethoxazol; Van, vancomvcin.

1000 bp 500 bp

Figure 1. A Representative Gel Image of vanA Gene Detection by PCR

Lane 1: positive control; lane 2: a negative clinical isolates; lane 3: negative control (distilled water); M: 100 bp ladder.

resulted from the uncontrolled and frequent use of vancomycin and self-medication (13).

In this study, 2 phenotypic (broth microdilution) and genotypic (PCR) methods were used to determine susceptible and resistant isolates to vancomycin, and our results demonstrated that all isolates with MIC $\geq 1 \mu g/mL$ were negative for the presence of *vanA* gene.

In previous Iranian studies, range of resistance to penicillin was 0% to 70% among clinical isolates (1, 14). Compared with other reports in Iran, our results showed only 1 pneumococcal isolate resistant to penicillin (20%), therefore, the rate of resistance to penicillin in our isolates was lower than those reported from other studies (15, 16). Our study also found that both penicillin-resistant and penicillin-susceptible strains showed resistance to erythromycin and cefotaxime, which was similar to reports from Zahedan, Iran (15).

In this study, it was observed that isolate No. 3 was not only resistant to vancomycin, but was also resistant to other antibiotics such as ceftriaxone, cefotaxime, ofloxacin, meropenem, erythromycin, and penicillin. Four of the isolates in the present study remained susceptible to ofloxacin, which probably can be recommended as an alternative treatment option for penicillin and macrolides resistant isolates. However, we found no significant differences between source of infection and antibiotic resistance pattern among the studied isolates.

5.1. Conclusions

In summary, it seems that the increasing use of penicillin and vancomycin in the recent years created an opportunity for the emergence of penicillin and vancomycinresistant clinical isolates. The increase in antibiotic resistance among *S. pneumoniae* strains in Iran seems to be alarming, and the treatment of infections should be further analyzed and investigated.

Acknowledgments

We thank all persons at the laboratory of Mofid Children hospital, Tehran, IR Iran for their technical assistance.

References

 Mamishi S, Moradkhani S, Mahmoudi S, Hosseinpour-Sadeghi R, Pourakbari B. Penicillin-Resistant trend of Streptococcus pneumoniae in Asia: A systematic review. *Iran J Microbiol.* 2014;6(4):198–210. [PubMed: 25802701].

- Tan TQ. Pediatric invasive pneumococcal disease in the United States in the era of pneumococcal conjugate vaccines. *Clin Microbiol Rev.* 2012;25(3):409-19. doi: 10.1128/CMR.00018-12. [PubMed: 22763632].
- Chiu SS, Ho PL, Chow FK, Yuen KY, Lau YL. Nasopharyngeal carriage of antimicrobial-resistant Streptococcus pneumoniae among young children attending 79 kindergartens and day care centers in Hong Kong. Antimicrob Agents Chemother. 2001;45(10):2765-70. doi: 10.1128/AAC.45.10.2765-2770.2001. [PubMed: 11557466].
- 4. Khashei R, Sedigh HSE, Alfatemi MH, Zomorodian K. Antimicrobial resistance patterns of colonizing microflora on the personnel hands and noses working in the Neonatal Intensive Care Unit (NICU). *World Appl Sci J.* 2014;**30**(10):1232–7.
- Hoseini Alfatemi SM, Motamedifar M, Hadi N, Sedigh Ebrahim Saraie H. Analysis of Virulence Genes Among Methicillin Resistant Staphylococcus aureus (MRSA) Strains. Jundishapur J Microbiol. 2014;7(6):e10741. doi: 10.5812/jjm.10741. [PubMed: 25371805].
- Temime L, Boelle PY, Courvalin P, Guillemot D. Bacterial resistance to penicillin G by decreased affinity of penicillin-binding proteins: a mathematical model. *Emerg Infect Dis.* 2003;9(4):411–7. doi: 10.3201/eid0904.020213. [PubMed: 12702219].
- Kaplan SL, Mason EJ. Management of infections due to antibiotic-resistant Streptococcus pneumoniae. *Clin Microbiol Rev.* 1998;11(4):628–44. [PubMed: 9767060].
- Johnson DM, Stilwell MG, Fritsche TR, Jones RN. Emergence of multidrug-resistant Streptococcus pneumoniae: report from the SENTRY Antimicrobial Surveillance Program (1999-2003). Diagn Microbiol Infect Dis. 2006;56(1):69–74. doi: 10.1016/j.diagmicrobio.2005.12.008. [PubMed: 16546341].
- 9. McCullers JA, English BK, Novak R. Isolation and characterization

of vancomycin-tolerant Streptococcus pneumoniae from the cerebrospinal fluid of a patient who developed recrudescent meningitis. *J Infect Dis.* 2000;**181**(1):369–73. doi: 10.1086/315216. [PubMed: 10608791].

- CLSI . Performance Standards for Antimicrobial Susceptibility Testing; 25th Informational Supplement. M100-S25. Wayne: Clinical and Laboratory Standards Institute; 2015.
- Xue L, Yao K, Xie G, Zheng Y, Wang C, Shang Y, et al. Serotype distribution and antimicrobial resistance of Streptococcus pneumoniae isolates that cause invasive disease among Chinese children. *Clin Infect Dis.* 2010;**50**(5):741–4. doi: 10.1086/650534. [PubMed: 20113175].
- Rafiei Tabatabaei S, Rahbar M, Nazari Alam A, Fallah F, Hashemi A, Yousefi M, et al. Detection of pbp2b Gene and Antimicrobial Susceptibility Pattern of Streptococcus Pneumoniae Isolates in Tehran Hospitals, Iran. Arch Pediatr Infect Dis. 2016;5(1) doi: 10.5812/pedinfect.38891.
- Nateghian AR, Robinson JL, Samadi B, Abdi N. Appropriate use of vancomycin in an educational tertiary care hospital in Tehran, Iran. *Med J Islamic Republic Iran*. 2007;21(1):43–9.
- Najafi Mosleh M, Gharibi M, Alikhani MY, Saidijam M, Kalantarian G. Antimicrobial Susceptibilities and Distribution of Resistance Genes for beta-Lactams in Streptococcus pneumoniae Isolated in Hamadan. *Jundishapur J Microbiol.* 2014;7(10):e12714. doi: 10.5812/jjm.12714. [PubMed: 25632328].
- Bokaeian M, Khazaei HA, Javadimehr M. Nasopharyngeal Carriage, Antibiotic Resistance and Serotype Distribution of Streptococcus Pneumoniae among Healthy Adolescents in Zahedan. *Iran Red Crescent Med* J. 2011;13(5):328–33. [PubMed: 22737489].
- Kohanteb J, Sadeghi E. Penicillin-resistant Streptococcus pneumoniae in Iran. *Med Princ Pract.* 2007;**16**(1):29–33. doi: 10.1159/000096137. [PubMed: 17159361].