



Cloning and Expression of HP0242 Hypothetical Gene as a DNA Vaccine Candidate for *Helicobacter pylori* and Study of Its Immunoreactivity

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Abstract

Background: The HP0242 plays an important role in physiology of *H. pylori* in acidic condition.

Objectives: The aim of this study was cloning and expression of immunogenicity of HP0242 gene of *H. pylori*.

Methods: The whole sequence of the hypothetical gene was amplified with specific primers. The DNA sequence (716 bp) was cloned into *E. coli* TOP10F by utilizing PEGFP-C1 and PTZ vectors. The protein product in CHO cells was elucidated with the SDS-PAGE technique. Western blot analysis showed that HP0242 expressed by recombinant *E. coli* had favorable immune-reactivity. Western blot was conducted to assess the immune-reactivity.

Results: Presence of hypothetical gene in the expression PEGFP-C1 vector was confirmed with digestion by *sacII* and *smaI*. Finally, the expression of the hypothetical protein in CHO cells was confirmed by SDS-PAGE and thus, can be considered as a DNA vaccine candidate for *H. pylori*. Western blot analysis showed that HP0242 expressed by recombinant *E. coli* had a favorable immune-reactivity.

Conclusion: The expressed HP0242 gene showed the possibility that it can be used as a candidate DNA vaccine for *H. pylori* in the future research.

Keywords: DNA Vaccine, HP0242 hypothetical Gene, immunogenicity, *Helicobacter pylori*

1. Background

Prevention is the most effective way of interfering the medicine and synthesis of vaccines is a reliable approach (1, 2). DNA vaccines induce both humeral and cellular immunity and thus, have a high potential of eradication of pathogens compared to other vaccination routes (3, 4). The recombinant DNA enters the host and is expressed by host machinery in vivo. In addition, it is also presented by major histocompatibility complex (MHC) class I and II (5). In comparison with attenuated and inactivated vaccines, it has been discussed that DNA vaccines as "precious treasures" are appropriate for overcoming the infectious and genetic diseases (6). Several DNA vaccine candidates have been introduced and assessed such as heat shock proteins catalase, heparan sulphate-binding proteins, *vacA*, and lipoprotein. The hypothetical proteins HP0241 and HP0242 are encoded by an operon with 5 other functionally known proteins including HP0243 (neu-

trophil activating protein), HP0239 (glutamyl-tRNA reductase), HP0240 (octa prenyl-diphosphate synthase), HP0237 (porphobilinogen deaminase), and HP0238 (prolyl-tRNA synthetase). HP0242 is an acid-adaptive protein and plays an important role in physiology of *H. pylori* in acidic conditions. The ribbon like structure of HP0242 monomer shows 4 helices including H1, H2, H3, and H4. H2 helix plays an important role in dimer formation and thus, possibly carry out a key role in protein function (7). Therefore, this homodimeric protein attains a unique pseudo-knotted folding topology. It was shown that in the unfolding pathway, the N-terminal region is related to the first transition state, and the C-terminal region is the main sequence for contributing dimerization (8). The protein reportedly fulfills iron uptake and metabolism. There are numerous studies on the association of *H. pylori* in iron-mediated anemia in children and adults (9, 10).

2. Objectives

In this study, HP0242, as a DNA vaccine candidate, was cloned and the synthesis of the protein in Chinese hamster ovary (CHO) cells was assessed by the SDS-PAGE method.

3. Methods

3.1. Bacterial Strains

The standard strains of *H. pylori* ATCC 40504 and *E. coli* TOP10F were prepared from microbiology department of Pasteur institute of Iran and biotechnology research center of Shahrekord Islamic Azad University, respectively. *H. pylori* isolates were cultured on Brucella agar and LB- broth.

3.2. Antibiotics Preparation

Ampicillin and neomycin were prepared from the Merk company. Ampicillin 100 µg/ml was prepared and added to the Luria-Bertani (LB) medium.

3.3. DNA Extraction

For DNA extraction from *H. pylori*, the DNA isolation kit (Bioneer, Trefflab) was used according to the instructions of manufacturer.

3.4. Cloning of Hypothetical Gene

The PCR was used for hypothetical gene amplification by utilizing specific primers with linkers. The primer sequences including forward: 5'-AAACCGCGGAGAACAATAATGAAAGATTAC-3' and reverse: 5'-AACCCCGGGAGCCACCTTTTCACAACTAC-3' were used to generate a 716 bp fragment. The conditions and amplification thermal profile included initial denaturation at 95°C for 5 minutes and 30 cycles of denaturation at 94°C for 1 minute, annealing at 64°C, 72°C for 1 minute, and final extension for 10 minutes. The sequences "CCGCGG" and "CCCGGG" were designed and added for enzymatic digestion of *saclI* and *smal* (fermentase), respectively. An expression vector (fermentas) PEGFP-C1 was introduced with T4 ligase to construct the recombinant PEGFP-C1-hypothetical gene using a T/A cloning kit (DNA ligation kit mighty mix, TaKaRa). *Escherichia coli* TOP10F competent cells (Stratagene) was employed as the host for gene cloning and transformed cells were grown in LB medium with 100 µg/mL ampicillin. The detail protocol of cloning and transformation was followed as described previously (11, 12). All PEGFP-C1-Hypothetical cloned sequences were subsequently confirmed by direct sequencing (Bioneer, Gen Ray Company, South Korea).

3.5. Sodium Dodecyl Sulfate-Polyacrylamide Gel Electrophoresis (SDS-PAGE)

The PEGFP-C1-Hypothetical gene vectors were transformed into Chinese hamster ovary (CHO) cells and HP0242 protein synthesis was investigated with SDS-PAGE (13). HP0242 expression was induced by utilizing 100 mM IPTG.

Detection of the Immunoreactivity of HP0242 expressed by recombinant *E. coli* HP0242 expressed in recombinant *E. coli* was electrophoresed on a 15% SDS-PAGE and then transferred to the cellulose nitrate membrane. The membrane was probed with a primary polyclonal mice antiserum (dilution of 1:50) and next, it was incubated at 37°C for 2 hours and washed 3 times with TBS/Tween-20 (TBST) for 15 minutes, and then incubated with the horseradish peroxidase-linked secondary antibody (dilution of 1:1000). Detection of protein was visualized using the DAB western blot detection system.

4. Results

4.1. Hypothetical gene amplification

PCR amplification of HP0242 depicted a 716 bp product in 1% Agarose gel. The gene was also amplified after matrix (colonies on culture medium) DNA extraction and enzymatic digestion of PTZ-Hypothetical recombinant plasmids (Figure 1). The PTZ-Hypothetical recombination was confirmed by enzymatic digestion.

The PEGFP-C1-Hypothetical recombinant expression vector was also confirmed by digestion with *smal* and *saclI* enzymes. As shown in Figure 2, the hypothetical gene and PEGFP-C1-Hypothetical with 716 bp and 4731 bp sizes were obtained. Sequencing the final PEGFP-C1-Hypothetical recombinant and BLAST output in GenBank demonstrated the accuracy of sequences as well. Figure 2 depicts the schematic PEGFP-C1-Hypothetical recombinant plasmid.

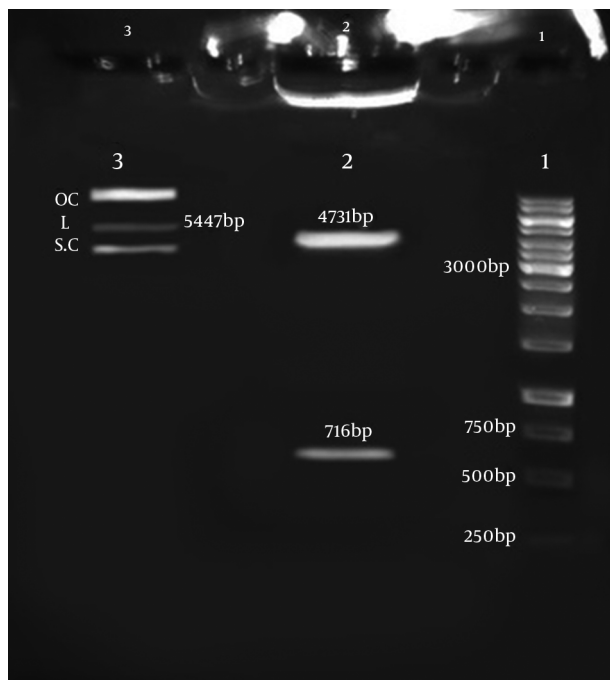
4.2. SDS-PAGE

The induced expression of HP0242 by utilizing 100 mM IPTG was depicted in the SDS-PAGE of protein product with a 30.60 kda size in CHO cells.

Identification of the Immunoreactivity of HP0242 was expressed by recombinant *E. coli*.

The HP0242 expressed by *E. coli* transformant was recognized by anti-Hp serum. This result indicated that the HP0242 protein expressed by *E. coli* transformant had favorable immunoreactivity.

Figure 1. Enzymatic Digestion of PEGFP-CI-Hypothetical, and PCR Amplification of HP0242 Gene with 1 kb DNA Marker



Column: 1 kb DNA size marker, column 2: hypothetical gene and PEGFP-CI-Hypothetical with 716 bp and 4731 bp, respectively. column3: open coiled (OC), linear (L) and supercoiled circular (SC) plasmids.

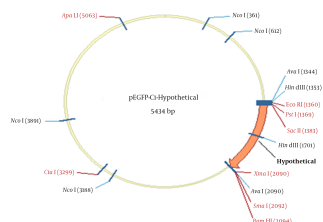


Figure 2. The Schematic Representation of PEGFP-CI-Hypothetical Recombinant Plasmid and Restriction Enzyme Cut Sites

5. Discussion

In the current study, the hypothetical gene (HP0242) was cloned in PEGFP-CI and PTZ vectors and next the protein was translated/ expressed in the CHO cells. Moreover, the immunoreactivity of HP0242 was shown in the western blot analysis. Several other proteins have been introduced as DNA vaccine candidates and induction of immune responses and have been assessed to some extent (14). However, the potential of specific immunogenicity has not been fully examined. In a previous study by

Tsai, the HP0242 was cloned into *E. coli* BL21 (DE3) and digested with BamHI and XhoI (7). This protein is supposedly involved in the persistence of *H. pylori* in the acidic medium of gut and iron uptake. In a study by Chen, the outer membrane protein A (oipA) was administered intradermally ('gene gun' immunization) in C57BL/6 mice and promoted a strong Th2 and with adjuvants a Th1 immune responses (15). In the present study, the immune responses and adjuvants were not investigated. In another study, Chen et al. showed that oipA DNA vaccine administered orally to C57BL/6 mice could significantly enhance levels of IgG2a/IgG1 antibodies and IFN- γ /IL-4 cytokines (mixed Th1/Th2 immune response) (14). In several other studies, hapA, cagA, and alpA DNA vaccine candidates have been cloned in different vectors and the immune responses have demonstrated to be suitable in case of them, however, hspA results have shown that this protein cannot be considered as an appropriate vaccine candidate (16-19). Study on bacterial vectors such as *Lactococcus lactis* (*L. lactis*) carrying UreB subunit of *H. pylori* fused with human interleukin 2 has also shown effective for oral route of prevention and induction of secretion of antibody and IFN- γ , IL-4, and IL-17 cytokines (20). Moreover, DNA fragments of *H. pylori* ureI-ureB and *Vibrio cholera* ctb subunits were fused and injected into the BALB/C mice and led to significant protection against *H. pylori* via secretion of IgA and a mixed response of Th1/Th2/Th17 cells (21). In this study, the animal model and potential of immunogenicity due to HP0242 was not examined and it is proposed that this experiment be fulfilled in future research. Development of efficient vaccines against *H. pylori* relies on the exact study over the host-pathogen interactions and also considering molecular structures, epitope mapping, and antigenic regions. In vivo studies and induction of multi-component responses is a more efficient approach (22).

5.1. Conclusions

The PEGFP-CI-Hypothetical recombinant plasmid was expressed and then recognized by serum that contained polyclonal antibodies and therefore, it can be considered as a DNA vaccine candidate in the future for prevention of *H. pylori* infection. Development of efficient vaccines against *H. pylori* relies on the exact study over host-pathogen interactions and also considering molecular structures, epitope mapping, antigenic regions, and work on multi-component vaccine is more efficient way.

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Footnote

Conflict of Interest: None to declare.

References

1. Stewart AJ, Devlin PM. The history of the smallpox vaccine. *J Infect*. 2006;**52**(5):329-34. doi: [10.1016/j.jinf.2005.07.021](https://doi.org/10.1016/j.jinf.2005.07.021). [PubMed: [16176833](https://pubmed.ncbi.nlm.nih.gov/16176833/)].
2. Salehi M, Ghasemian A, Shokouhi Mostafavi SK, Najafi S, Rajabi Vardanjani H. Sero-prevalence of Helicobacter pylori Infection in Neyshabur, Iran, During 2010-2015. *Iran J Pathol*. 2017;**12**(2):183-8.
3. Todoroki I, Joh T, Watanabe K, Miyashita M, Seno K, Nomura T, et al. Suppressing effects of DNA vaccines encoding heat shock protein on Helicobacter pylori-induced gastritis in mice. *Biochem Biophys Res Commun*. 2000;**277**(1):159-63. doi: [10.1006/bbrc.2000.3632](https://doi.org/10.1006/bbrc.2000.3632). [PubMed: [11027657](https://pubmed.ncbi.nlm.nih.gov/11027657/)].
4. Wilson KT, Crabtree JE. Immunology of Helicobacter pylori: insights into the failure of the immune response and perspectives on vaccine studies. *Gastroenterology*. 2007;**133**(1):288-308. doi: [10.1053/j.gastro.2007.05.008](https://doi.org/10.1053/j.gastro.2007.05.008). [PubMed: [17631150](https://pubmed.ncbi.nlm.nih.gov/17631150/)].
5. Flingai S, Czerwonko M, Goodman J, Kudchodkar SB, Muthumani K, Weiner DB. Synthetic DNA vaccines: improved vaccine potency by electroporation and co-delivered genetic adjuvants. *Front Immunol*. 2013;**4**:354. doi: [10.3389/fimmu.2013.00354](https://doi.org/10.3389/fimmu.2013.00354). [PubMed: [24204366](https://pubmed.ncbi.nlm.nih.gov/24204366/)].
6. Kennedy RB, Ovsyannikova IG, Lambert ND, Haralambieva IH, Poland GA. The personal touch: strategies toward personalized vaccines and predicting immune responses to them. *Expert Rev Vaccines*. 2014;**13**(5):657-69. doi: [10.1586/14760584.2014.905744](https://doi.org/10.1586/14760584.2014.905744). [PubMed: [24702429](https://pubmed.ncbi.nlm.nih.gov/24702429/)].
7. Tsai JY, Chen BT, Cheng HC, Chen HY, Hsiao NW, Lyu PC, et al. Crystal structure of HP0242, a hypothetical protein from Helicobacter pylori with a novel fold. *Proteins*. 2006;**62**(4):1138-43. doi: [10.1002/prot.20864](https://doi.org/10.1002/prot.20864). [PubMed: [16395670](https://pubmed.ncbi.nlm.nih.gov/16395670/)].
8. Wang LW, Shih P, Liu YN, Hsu BD, Lyu PC, Hsu STD. *Multiparametric Characterization of the Three-state Folding Equilibrium of a Pseudo-knotted Protein from H. pylori*. 2014.
9. Sapmaz F, Basyigit S, Kalkan IH, Kisa U, Kavak EE, Guliter S. The impact of Helicobacter pylori eradication on serum hepcidin-25 level and iron parameters in patients with iron deficiency anemia. *Wien Klin Wochenschr*. 2016;**128**(9-10):335-40. doi: [10.1007/s00508-016-0961-5](https://doi.org/10.1007/s00508-016-0961-5). [PubMed: [26932797](https://pubmed.ncbi.nlm.nih.gov/26932797/)].
10. Choe YH. *Extraintestinal Manifestations of H. pylori Infection: H. pylori-Associated Iron-Deficiency Anemia*. Springer; 2016.
11. Sambrook J, Fritsch EF, Maniatis T. *Molecular cloning: a laboratory manual*. Cold spring harbor laboratory press; 1989.
12. Zimmerman SA, Tomb JF, Ferry JG. Characterization of CamH from Methanosarcina thermophila, founding member of a subclass of the {gamma} class of carbonic anhydrases. *J Bacteriol*. 2010;**192**(5):1353-60. doi: [10.1128/JB.01164-09](https://doi.org/10.1128/JB.01164-09). [PubMed: [20023030](https://pubmed.ncbi.nlm.nih.gov/20023030/)].
13. Legendre N. *Purification of Proteins and Peptides by SDS-PAGE-A Practical Guide to Protein and Peptide Purification for Microsequencing (Edited by Matsudaira PT)*. Academic Press, New York; 1989.
14. Chen J, Li N, She F. Helicobacter pylori outer inflammatory protein DNA vaccine-loaded bacterial ghost enhances immune protective efficacy in C57BL/6 mice. *Vaccine*. 2014;**32**(46):6054-60. doi: [10.1016/j.vaccine.2014.09.014](https://doi.org/10.1016/j.vaccine.2014.09.014). [PubMed: [25236588](https://pubmed.ncbi.nlm.nih.gov/25236588/)].
15. Chen J, Lin L, Li N, She F. Enhancement of Helicobacter pylori outer inflammatory protein DNA vaccine efficacy by co-delivery of interleukin-2 and B subunit heat-labile toxin gene encoded plasmids. *Microbiol Immunol*. 2012;**56**(2):85-92. doi: [10.1111/j.1348-0421.2011.00409.x](https://doi.org/10.1111/j.1348-0421.2011.00409.x). [PubMed: [22150716](https://pubmed.ncbi.nlm.nih.gov/22150716/)].
16. Soleimani N, Mohabati Mobarez A, Farhangi B. Cloning, expression and purification flagellar sheath adhesion of Helicobacter pylori in Escherichia coli host as a vaccination target. *Clin Exp Vaccine Res*. 2016;**5**(1):19-25. doi: [10.7774/cevr.2016.5.1.19](https://doi.org/10.7774/cevr.2016.5.1.19). [PubMed: [26866020](https://pubmed.ncbi.nlm.nih.gov/26866020/)].
17. Wang KX, Wang XF. Cloning and sequencing of cagA gene fragment of Helicobacter pylori with coccoid form. *World J Gastroenterol*. 2004;**10**(23):3511-3. [PubMed: [15526375](https://pubmed.ncbi.nlm.nih.gov/15526375/)].
18. Xu C, Li ZS, Du YQ, Tu ZX, Gong YF, Jin J, et al. Construction of a recombinant attenuated Salmonella typhimurium DNA vaccine carrying Helicobacter pylori hpaA. *World J Gastroenterol*. 2005;**11**(1):114-7. [PubMed: [15609408](https://pubmed.ncbi.nlm.nih.gov/15609408/)].
19. Zhang XJ, Feng SY, Li ZT, Feng YM. Expression of Helicobacter pylori hspA gene in Lactococcus lactis NICE system and experimental study on its immunoreactivity. *Gastroenterol Res Pract*. 2015;**2015**.
20. Zhang HX, Qiu YY, Zhao YH, Liu XT, Liu M, Yu AL. Immunogenicity of oral vaccination with Lactococcus lactis derived vaccine candidate antigen (UreB) of Helicobacter pylori fused with the human interleukin 2 as adjuvant. *Mol Cell Probes*. 2014;**28**(1):25-30. doi: [10.1016/j.mcp.2013.08.003](https://doi.org/10.1016/j.mcp.2013.08.003). [PubMed: [24036137](https://pubmed.ncbi.nlm.nih.gov/24036137/)].
21. Yang J, Dai LX, Pan X, Wang H, Li B, Zhu J, et al. Protection against Helicobacter pylori infection in BALB/c mice by oral administration of multi-epitope vaccine of CTB-UreI-UreB. *Pathog Dis*. 2015;**73**(5). doi: [10.1093/femspd/ftv026](https://doi.org/10.1093/femspd/ftv026). [PubMed: [25846576](https://pubmed.ncbi.nlm.nih.gov/25846576/)].
22. Mirzaei N, Poursina F, Moghim S, Rashidi N, Ghasemian Safaei H. The study of H. pylori putative candidate factors for single- and multi-component vaccine development. *Crit Rev Microbiol*. 2017;**43**(5):631-50. doi: [10.1080/1040841x.2017.1291578](https://doi.org/10.1080/1040841x.2017.1291578).