Published online 2019 August 17.

Interleukin-6, Interleukin-17 and Transforming Growth Factor-Beta are Overexpressed in Newly Diagnosed Tuberculosis Patients; Potent Biomarkers of Mycobacterial Infection

Fakhri Sadat Seyedhosseini¹, Saeed Mohammadi ^{10, 2}, Mojtaba Zare Ebrahimabad³, Behnaz Khodabakhshi ¹⁰, Abdollah Abbasi¹ and Yaghoub Yazdani ¹⁰, ^{2,*}

¹Infectious Diseases Research Center, Golestan University of Medical Sciences, Gorgan, Iran

²Stem Cell Research Center, Golestan University of Medical Sciences, Gorgan, Iran

³Department of Biochemistry, Faculty of Medicine, Golestan University of Medical Sciences, Gorgan, Iran

corresponding author: Stem Cell Research Center, Golestan University of Medical Sciences, Gorgan, Iran. Email: yazdani@goums.ac.ir

Received 2018 March 07; Revised 2019 July 06; Accepted 2019 July 14.

Abstract

Background: Cytokines, as key regulators of immune responses, are imbalanced in tuberculosis (TB) which may reflect the status of the disease. Therefore, cytokines could be introduced as potential diagnostic and prognostic TB biomarkers.

Objectives: Here, we examined the plasma levels of IL-17, IL-6, and TGF- β among newly diagnosed patients with TB in comparison to normal subjects and analyzed the diagnostic utility of each cytokine to introduce a novel TB biomarker.

Methods: A total of 105 smear-positive, including 78 newly diagnosed (ND) and 27 under treatment (UT) patients with pulmonary TB and 111 age- and sex-matched healthy subjects were recruited. ELISA cytokine assay was used to determine the plasma levels of IL-17, IL-6, and TGF- β .

Results: IL-6 plasma level was higher in the ND patients than healthy subjects (P = 0.002) and the UT patients (P < 0.0001). Also, IL-17 and TGF- β were significantly overexpressed among both groups of patients with TB compared with healthy subjects. Receiver operating characteristic (ROC) curve analysis was used to assess the diagnostic utility of the evaluated cytokines. Area under the curve (AUC) for IL-6 was 0.5598 (P = 0.129). Regarding the diagnostic value of IL-6 to distinguish between the ND and UT patients, AUC for IL-6 was 0.8139 (P < 0.0001). Setting the optimal cut-off value at 23.75 gave a sensitivity of 77.78% and a specificity of 74.36%. AUC for IL-17 was 0.9148 (P < 0.0001), while AUC for TGF- β was 0.8877 (P < 0.0001). Setting the optimal cut-off value at 14.05 for IL-17 gave a sensitivity of 81.90% and a specificity of 79.28%, while setting the optimal cut-off value at 51.20 for TGF- β resulted in a sensitivity of 82.69% and a specificity of 72.73%.

Conclusions: All evaluated cytokines were overexpressed in newly diagnosed TB patients with different diagnostic utilities. IL-6 is more convenient in distinguishing the ND from UT patients. IL-17 is more specific in distinguishing TB infection and TGF- β represented a higher sensitivity. It is suggested that multiple cytokine assay should be conducted to have a better diagnostic performance rather than individual cytokine evaluation.

Keywords: Diagnosis, IL-17, IL-6, TB Biomarker, TGF- β , Tuberculosis

1. Background

Tuberculosis (TB) remains a major cause of morbidity and mortality and a public health burden in the developing countries, which is estimated to be responsible for nearly 1.4 million deaths with 8.7 million annual new cases (1). The causative agent of TB is *Mycobacterium tuberculosis* (MTB), which is a slow-growing facultative pathogen (2). According to the emergence of multidrug-resistant (MDR) and extensively drug-resistant (XDR) strains of MTB, as well as the growing spread of human-immunodeficiency virus (HIV) among patients with TB, the situation of TB is worsening worldwide (3, 4). Early diagnosis and consequent interventions are needed to effectively control TB and provide prognostic information (5, 6). T-cell-based assays such as highly sensitive and specific interferon-gamma release assay (IGRA) (7) are commonly used as gold standard tests for the detection of TB (8) and could be a potential diagnostic marker for other inflammatory diseases (9). However, this expensive and complicated assay is not capable of distinguishing active and latent TB infections (10). Therefore, there is an urgent need for more convenient and inexpensive TB biomarkers (11). Cytokines, as key regulators

Copyright © 2019, Archives of Clinical Infectious Diseases. This is an open-access article distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 International License (http://creativecommons.org/licenses/by-nc/4.0/) which permits copy and redistribute the material just in noncommercial usages, provided the original work is properly cited.

of immune responses, have a major role in determining the fate of host-pathogen interaction (12). The alterations of these molecules could reflect the status of disease (13). Thus cytokines could be introduced as potential diagnostic and prognostic TB biomarkers (14,15). Numerous cytokines have been introduced by Chegou et al. as potential diagnostic markers for distinguishing active TB from its latent form (16).

Interleukin-6 (IL-6) is a pro-inflammatory cytokine, which is secreted by T cells and macrophages (17). It is probably involved in the pathogenesis of TB by stimulating the secretion of IFN- γ , which plays a crucial role in the activation of MTB-infected macrophages (18). Interleukin-17 (IL-17) is also a pro-inflammatory cytokine, which is known for its crucial role in immune responses against extracellular pathogens (19). Despite the overexpression in the patients with active TB, IL-17 may play a role in protective immunity against MTB infection (20). Transforming growth factorbeta (TGF- β) is a multifunctional cytokine, which is produced by T cells, B cells, and myeloid cells with different functions (21). TGF- β may be involved in the suppression of T cell responses, macrophage deactivation, and tissue injury initiation and deterioration (22).

2. Objectives

In the present study, we examined the plasma expression of IL-17, IL-6, and TGF- β among newly diagnosed (ND) and under treatment (UT) patients with TB in comparison to the normal subjects and analyzed the diagnostic utility of each cytokine to introduce a novel TB biomarker.

3. Methods

3.1. Sample Collection

We recruited 105 smear positive, including 78 newly diagnosed (ND) and 27 under treatment (UT) patients with pulmonary TB from public health centers and educational hospitals of Golestan University of Medical Sciences, Gorgan, Iran. Patients with pregnancy and chronic inflammatory disorders, including diabetes, autoimmune diseases, cancer, allergic diseases, immunocompromised conditions, coinfections with any other infectious diseases such as viral hepatitis, and heart failure were not included in this study. Moreover, the patients receiving the treatment either self-medicated or prescribed by the physician were excluded from the ND group. Accordingly, a total of 111 age- and sex-matched healthy subjects were enrolled. The healthy controls were also evaluated for all mentioned inflammatory conditions and the individuals with a history of pulmonary disorders in their family were not included.

Written informed consent was taken and signed by all participants in compliance with the Declaration of Helsinki. Whole blood samples were taken from all subjects, plasma was separated and stored at -80°C until the measurement of cytokines.

3.2. ELISA Cytokine Assay

The commercially available ELISA kits were used to determine the plasma level of IL-17, IL-6 (Biolegend, CA, USA), and TGF- β (eBioscience, San Diego, USA) in patients with TB and healthy subjects, according to the manufacturer's instructions. The optical densities of all samples were obtained at the wavelength of 450 nm using Biotek ELISA reader ELX800 (Biotek, VT, USA), as previously described (23, 24). All samples were measured in duplicates and the results were reported as pg/mL.

3.3. Statistical Analysis

In order to analyze data statistically and prepare graphs, SPSS V. 22.0 and Graphpad Prism V. 5.04 software were used. Shapiro-Wilk test was used to address the normal distribution of variables in all groups. All data were demonstrated as mean \pm SE (standard Error). Significant differences were also assessed using independent samples Mann-Whitney U test for two-group comparisons. The nonparametric Kruskal-Wallis with Dunn-Bonferroni post hoc test was also used to compare the means of multiple samples. P values less than 0.05 were considered statistically significant.

4. Results

4.1. ELISA Expression of IL-6, IL-17 and TGF-eta

ELISA method was used to evaluate the plasma levels of IL-6 among the ND patients in comparison to the healthy subjects. The plasma expression of IL-6 among the ND patients was higher than the healthy subjects (P = 0.002) and the UT patients (P < 0.0001). The plasma level of IL-6 was decreased among the UT patients in comparison to the healthy subjects (P = 0.007) (Figure 1A). Moreover, IL-17 was significantly overexpressed among the ND (P < 0.0001) and UT (P < 0.0001) patients compared with the healthy subjects. However, no significant difference was observed between the two groups of the patients (P = 0.861) (Figure **1B**). Similarly, the expression of TGF- β was not different between the ND and UT patients (P = 0.795). However, it was significantly decreased among the healthy subjects compared with the ND (P < 0.0001) and UT (P < 0.0001) patients (Figure 1C).

4.2. The Segregation Value of Cytokines Between Patients and Normal Subjects

Receiver operating characteristic (ROC) curve analysis was used to assess the diagnostic utility of the evaluated cytokines to distinguish between infected and non-infected samples. Area under the curve (AUC) for IL-6 was 0.5598 (95% CI: 0.4823 - 0.6373; P = 0.129). Setting the optimal cutoff value at 31.95 (likelihood ratio = 1.14) gave a sensitivity of 51.43% (95% CI: 41.47% - 61.30%) and a specificity of 54.95% (95% CI: 45.22% - 64.41%) (Figure 2A). AUC for IL-17 was 0.9148 (95% CI: 0.8798 - 0.9497; P < 0.0001). Setting the optimal cut-off value at 14.05 (likelihood ratio = 3.95) resulted in a sensitivity of 81.90% (95% CI: 73.19% - 88.74%) and a specificity of 79.28% (95% CI: 70.55% - 86.39%) (Figure 2B). Moreover, AUC for TGF- β was 0.8877 (95% CI: 0.8440 - 0.9315; P < 0.0001). Setting the optimal cut-off value at 51.20 (likelihood ratio = 3.03) lead to a sensitivity of 82.69% (95% CI: 74.03% - 89.41%) and a specificity of 72.73% (95% CI: 63.41% -80.78%) (Figure 2C).

4.3. The Segregation Value of IL-6 Between Newly Diagnosed and Under Treatment Patients

We also used ROC curve analysis to investigate the diagnostic utility of IL-6 to distinguish between the ND and UT patients, which was the only altered cytokine following the treatment. Area under the curve (AUC) for IL-6 was 0.8139 (95% CI: 0.7147-0.9130; P < 0.0001). Setting the optimal cutoff value at 23.75 (likelihood ratio = 3.03) gave a sensitivity of 77.78% (95% CI: 57.74% - 91.38%) and a specificity of 74.36% (95% CI: 63.21% - 83.58%) (Figure 3).

5. Discussion

According to the growing burden of tuberculosis (TB) on public health in developing countries, early diagnosis and timely interventions are indispensable to effectively control TB (6, 25). Nowadays, conventional diagnostic tests, such as microscopic examination and culture of acidfast MTB organisms, are replaced by T-cell-based assays such as interferon-gamma release assay (IGRA) (7), which is also known as QuantiFERON-TB Gold test (8, 26). However, IGRA is an expensive and complicated assay, which is not also capable of distinguishing active TB infections from latent (10). Moreover, the evaluation of IFN- γ in the plasma (as a probable alternative for IGRA) is not trusted due to the scant amounts of detectable secreted IFN- γ (27). Therefore, there is an urgent need for more convenient and inexpensive TB biomarkers (11). Cytokines are major regulators of immune responses, which are secreted by immune cells and have definite roles in determining the fate of hostpathogen interaction (12, 13). Cytokine levels are altered in different states of the disease and could be of diagnostic value (13). Several studies have introduced cytokines and chemokines as potent TB biomarkers, such as IFN- γ (16), IP-10 (28), MCP-2 (29), TGF- β (14), IL-10 (30), IL-6 (31), and TNF- α (32). Therefore, cytokines can be introduced as potential diagnostic and prognostic TB biomarkers (14). However, previous studies have not thoroughly addressed the differences between active and latent TB infection, which is of great importance in determining the prescribed treatment strategies (33). Hence, we evaluated the plasma expression of IL-17, IL-6, and TGF- β among newly diagnosed TB patients in comparison to healthy subjects and analyzed the diagnostic utility of each cytokine to introduce a novel TB biomarker which may distinguish active TB infection.

The pro-inflammatory IL-6 is secreted by T cells and macrophages (34) and is involved in the pathogenesis of TB by stimulating the secretion of IFN- γ (18). It has been introduced in several studies as a potent TB biomarker (35). However, no clinical studies have confirmed the ability of IL-6 to distinguish the patients with active TB. In the current study, we showed that the plasma levels of IL-6 among newly diagnosed patients with TB were overexpressed but not statistically significant in comparison to the healthy subjects. We also used ROC curve analysis to assess the diagnostic utility of IL-6. In this regard, our findings reveal that the AUC for IL-6 has not an acceptable segregation value in distinguishing patients with TB from the heathy subjects. Moreover, the most suitable cut-off value for IL-6 did not give proper sensitivity and specificity. Therefore, IL-6 may not be introduced as a suitable TB biomarker to distinguish the disease. On the other hand, IL-6 was the only cytokine which was different between the two groups of the patients. ROC curve analysis to assess the diagnostic utility of IL-6 between the ND and UT patients gave an acceptable AUC with proper sensitivity and specificity. Therefore, IL-6 could be introduced as a suitable marker in distinguishing active state from latent and also monitoring the treatment efficiency.

IL-17 is a recently specified pro-inflammatory cytokine, which is known for its role in immune responses against extracellular pathogens (19). However, the role of IL-17 in MTB infection has been controversial (20). It has been reported that IL-17⁺CD4⁺ T cells constitute a higher population in healthy subjects compared to the patients with active and latent TB (36). On the other hand, IL-17 was reported to be downregulated in the PBMCs of vaccinated normal subjects in comparison to patients with active TB (37). However, no study has evaluated the diagnostic value of IL-17 plasma alterations in discrimination of patients with active TB. In the present study, we showed that IL-17 was significantly overexpressed among newly diagnosed patients with TB. ROC curve analysis results showed an AUC



Figure 1. ELISA cytokine assay: IL-6 plasma level was significantly higher in newly diagnosed patients with TB (A). IL-17 (B) and TGF- β (C) were significantly increased in both groups of patients with TB. The non-parametric Kruskal-Wallis with Dunn-Bonferroni post hoc test was also used to compare the means of multiple samples. Data of each bar demonstrates mean \pm SE. P values lower than 0.05 were considered statistically significant. SE, standard error; TB, Tuberculosis.



Figure 2. ROC curve analyses (patients with TB vs. healthy subjects); area under the curve (AUC) for IL-6 was 0.5598 (P = 0.129). Setting the optimal cut-off value at 31.95 gave a sensitivity of 51.43% and a specificity of 54.95% (A). AUC for IL-17 was 0.9148 (P < 0.0001). Setting the optimal cut-off value at 14.05 resulted in a sensitivity of 81.90% and a specificity of 79.28% (B). AUC for TGF- β was 0.8877 (P < 0.0001). Setting the optimal cut-off value at 51.20 lead in a sensitivity of 82.69% and a specificity of 72.73% (C).



Figure 3. ROC curve analysis of IL-6 diagnostic value (newly diagnosed vs under treatment patients); area under the curve (AUC) for IL-6 was 0.8139 (P < 0.0001). Setting the optimal cut-off value at 23.75 gave a sensitivity of 77.78% and a specificity of 74.36%.

of 0.9148, which is an optimum segregation value with acceptable sensitivity and quite high specificity.

TGF- β is a multifunctional cytokine, which is produced by T cells, B cells, and myeloid cells (21), and may be involved in the suppression of T cell responses, macrophage deactivation, and tissue injury initiation and propagation (22). It has been reported that the production of TGF- β was augmented by blood monocytes from patients with active TB (38). TGF- β is also increased in the HIV and pulmonary TB infections, which highlights the role of this cytokine in the propagation of TB-associated disorders (39). In the present study, we reported that TGF- β was significantly increased among newly diagnosed patients with TB. ROC curve analysis showed an optimum segregation value (AUC = 0.8877) with quite high sensitivity and acceptable specificity. Altogether, IL-17 and TGF- β plasma levels may be introduced as candidate biomarkers in the diagnosis of patients with active TB.

5.1. Limitations

The present study had several limitations. The most important limitation of our research was about compar-

ing the cytokine results of TB patients with normal subjects without any pulmonary diseases and not including patient groups suffering from other pulmonary disorders. Although we did not generalize the introduced biomarkers to distinguish TB from other diseases and we aimed to specifically segregate the patients with active TB from latent TB, we suggest designing further studies on patients suffering from non-TB pulmonary disorders to confirm or reject the specificity of our introduced biomarkers.

5.2. Conclusions

In conclusion, the expression of IL-6 was higher in newly diagnosed patients with TB. Therefore, IL-6 could be introduced as a suitable marker in distinguishing active TB state from latent and also monitoring the treatment efficiency. IL-17 and TGF- β cytokines were significantly overexpressed in newly diagnosed patients with TB. Although IL-17 is more specific in distinguishing active TB infection and TGF- β represented a higher sensitivity, it is suggested that multiple cytokine assay should be conducted to have a better diagnostic performance rather than individual cytokine evaluation.

Footnotes

Authors' Contribution: Study concept and design: Yaghoub Yazdani and Saeed Mohammadi; acquisition of data and experimental procedure: Fakhri Sadat Seyedhosseini, Mojtaba Zare Ebrahimabad, and Saeed Mohammadi; statistical analyses and interpretations of data: Behnaz Khodabakhshi, Abdollah Abbasi and Saeed Mohammadi; manuscript drafting, editing, and revising the manuscript: Saeed Mohammadi and Yaghoub Yazdani; final revision of the manuscript: Yaghoub Yazdani. All authors read and approved the final version of the manuscript.

Conflict of Interests: No conflict of interest was acclaimed by the authors.

Ethical Approval: The study was approved by the Ethics Committee of Golestan University of Medical Sciences (Ethics code: IR.GUOMS.REC.1394.48).

Funding/Support: This research was financially supported by the deputy of Research and Technology, Golestan University of Medical Sciences, Gorgan, Iran (Grant code: 940305036).

Patient Consent: Written informed consent was taken and signed by all participants.

References

 Sakai S, Kauffman KD, Schenkel JM, McBerry CC, Mayer-Barber KD, Masopust D, et al. Cutting edge: Control of Mycobacterium tuberculosis infection by a subset of lung parenchyma-homing CD4 T cells. J Immunol. 2014;**192**(7):2965–9. doi: 10.4049/jimmunol.1400019. [PubMed: 24591367]. [PubMed Central: PMC4010124].

- Javaheri-Kermani M, Farazmandfar T, Ajami A, Yazdani Y. Impact of hepcidin antimicrobial peptide on iron overload in tuberculosis patients. *Scand J Infect Dis.* 2014;**46**(10):693–6. doi: 10.3109/00365548.2014.929736. [PubMed: 25134646].
- Safdari H, Sadeghi A, Ghazvini K, Mohammadi S, Derakhshan M. Prevalence of Mycobacterium tuberculosis in the samples referred to the tuberculosis research laboratory in Mashhad Ghaem Hospital during 2005-2006. *Iran J Microbiol*. 2009:20–2.
- Gandhi NR, Andrews JR, Brust JC, Montreuil R, Weissman D, Heo M, et al. Risk factors for mortality among MDR- and XDR-TB patients in a high HIV prevalence setting. *Int J Tuberc Lung Dis.* 2012;**16**(1):90– 7. doi: 10.5588/ijtld.11.0153. [PubMed: 22236852]. [PubMed Central: PMC3302205].
- Falzon D, Jaramillo E, Schunemann HJ, Arentz M, Bauer M, Bayona J, et al. WHO guidelines for the programmatic management of drugresistant tuberculosis: 2011 update. *Eur Respir J.* 2011;38(3):516–28. doi: 10.1183/09031936.00073611.
- Storla DG, Yimer S, Bjune GA. A systematic review of delay in the diagnosis and treatment of tuberculosis. *BMC Public Health*. 2008;8(1):15. doi: 10.1186/1471-2458-8-15. [PubMed: 18194573]. [PubMed Central: PMC2265684].
- Belknap R, Daley CL. Interferon-gamma release assays. *Clin Lab Med.* 2014;**34**(2):337-49. doi: 10.1016/j.cll.2014.02.007. [PubMed: 24856531].
- Bianchi L, Galli L, Moriondo M, Veneruso G, Becciolini L, Azzari C, et al. Interferon-gamma release assay improves the diagnosis of tuberculosis in children. *Pediatr Infect Dis J.* 2009;28(6):510–4. [PubMed: 19504735].
- Mohammadi S, Sedighi S, Memarian A, Yazdani Y. Overexpression of interferon-γ and indoleamine 2, 3-dioxygenase in systemic lupus erythematosus: relationship with the disease activity. *LaboratoriumsMedizin*. 2017;41(1):41-7. doi: 10.1515/labmed-2016-0076.
- Zwerling A, van den Hof S, Scholten J, Cobelens F, Menzies D, Pai M. Interferon-gamma release assays for tuberculosis screening of healthcare workers: A systematic review. *Thorax*. 2012;67(1):62–70. doi: 10.1136/thx.2010.143180. [PubMed: 21228420].
- Yerlikaya S, Broger T, MacLean E, Pai M, Denkinger CM. A tuberculosis biomarker database: The key to novel TB diagnostics. *Int J Infect Dis.* 2017;56:253-7. doi: 10.1016/j.ijid.2017.01.025. [PubMed: 28159577].
- Cooper AM, Mayer-Barber KD, Sher A. Role of innate cytokines in mycobacterial infection. *Mucosal Immunol.* 2011;4(3):252–60. doi: 10.1038/mi.2011.13. [PubMed: 21430655]. [PubMed Central: PMC3294290].
- Sahiratmadja E, Alisjahbana B, de Boer T, Adnan I, Maya A, Danusantoso H, et al. Dynamic changes in pro- and anti-inflammatory cytokine profiles and gamma interferon receptor signaling integrity correlate with tuberculosis disease activity and response to curative treatment. *Infect Immun.* 2007;**75**(2):820–9. doi: 10.1128/IAI.00602-06. [PubMed: 17145950]. [PubMed Central: PMC1828524].
- Walzl G, Ronacher K, Hanekom W, Scriba TJ, Zumla A. Immunological biomarkers of tuberculosis. *Nat Rev Immunol.* 2011;11(5):343–54. doi: 10.1038/nri2960. [PubMed: 21475309].
- Salehi M, Dehghan Manshadi SA, Yassin Z, Hassannejad M. A simple approach for diagnosis of chronic tuberculous osteomyelitis: A case report. Arch Clin Infect Dis. 2017;12(2). e57281. doi: 10.5812/archcid.57281.
- Chegou NN, Heyckendorf J, Walzl G, Lange C, Ruhwald M. Beyond the IFN-gamma horizon: biomarkers for immunodiagnosis of infection with Mycobacterium tuberculosis. *Eur Respir J.* 2014;43(5):1472– 86. doi: 10.1183/09031936.00151413. [PubMed: 24311770].
- Goovaerts O, Jennes W, Massinga-Loembe M, Ceulemans A, Worodria W, Mayanja-Kizza H, et al. LPS-binding protein and IL-6 mark paradoxical tuberculosis immune reconstitution inflammatory syndrome in HIV patients. *PLoS One.* 2013;8(11). e81856. doi: 10.1371/journal.pone.0081856. [PubMed: 24312369]. [PubMed Central: PMC3842977].

- Singh PP. Interleukin-6: A potential biomarker of the success of tuberculosis treatment. Int J Infect Dis. 2016;45:413. doi: 10.1016/j.ijid.2016.02.881.
- Torrado E, Cooper AM. IL-17 and Th17 cells in tuberculosis. Cytokine Growth Factor Rev. 2010;21(6):455-62. doi: 10.1016/j.cytogfr.2010.10.004. [PubMed: 21075039]. [PubMed Central: PMC3032416].
- Peng MY, Wang ZH, Yao CY, Jiang LN, Jin QL, Wang J, et al. Interleukin 17-producing gamma delta T cells increased in patients with active pulmonary tuberculosis. *Cell Mol Immunol*. 2008;5(3):203-8. doi: 10.1038/cmi.2008.25. [PubMed: 18582402]. [PubMed Central: PMC4651291].
- DiFazio RM, Mattila JT, Klein EC, Cirrincione LR, Howard M, Wong EA, et al. Active transforming growth factor-beta is associated with phenotypic changes in granulomas after drug treatment in pulmonary tuberculosis. *Fibrogenesis Tissue Repair*. 2016;9:6. doi: 10.1186/s13069-016-0043-3. [PubMed: 27148404]. [PubMed Central: PMC4855369].
- Dheda K, Schwander SK, Zhu B, van Zyl-Smit RN, Zhang Y. The immunology of tuberculosis: From bench to bedside. *Respirol*ogy. 2010;15(3):433–50. doi: 10.1111/j.1440-1843.2010.01739.x. [PubMed: 20415982]. [PubMed Central: PMC5463744].
- Roohi A, Yazdani Y, Khoshnoodi J, Jazayeri SM, Carman WF, Chamankhah M, et al. Differential reactivity of mouse monoclonal anti-HBs antibodies with recombinant mutant HBs antigens. *World J Gastroenterol.* 2006;**12**(33):5368-74. doi: 10.3748/wjg.v12.i33.5368. [PubMed: 16981270]. [PubMed Central: PMC4088207].
- Yazdani Y, Roohi A, Khoshnoodi J, Shokri F. Development of a sensitive enzyme-linked immunosorbent assay for detection of hepatitis B surface antigen using novel monoclonal antibodies. *Avicenna J Med Biotechnol.* 2010;2(4):207–14. [PubMed: 23408744]. [PubMed Central: PMC3558164].
- Thomas T, Brighenti S, Andersson J, Sack D, Raqib R. A new potential biomarker for childhood tuberculosis. *Thorax*. 2011;66(8):727–9. doi: 10.1136/thx.2010.147363. [PubMed: 21228421].
- Mazurek GH, Jereb J, Lobue P, Iademarco MF, Metchock B, Vernon A, et al. Guidelines for using the QuantiFERON-TB Gold test for detecting Mycobacterium tuberculosis infection, United States. MMWR Recomm Rep. 2005;54(RR-15):49–55. [PubMed: 16357824].
- Arend SM, Thijsen SF, Leyten EM, Bouwman JJ, Franken WP, Koster BF, et al. Comparison of two interferon-gamma assays and tuberculin skin test for tracing tuberculosis contacts. *Am J Respir Crit Care Med.* 2007;**175**(6):618–27. doi: 10.1164/rccm.200608-1099OC. [PubMed: 17170386].
- Ruhwald M, Bodmer T, Maier C, Jepsen M, Haaland MB, Eugen-Olsen J, et al. Evaluating the potential of IP-10 and MCP-2 as biomarkers for the diagnosis of tuberculosis. *Eur Respir J.* 2008;**32**(6):1607–15. doi: 10.1183/09031936.00055508. [PubMed: 18684849].
- 29. Jacobsen M, Repsilber D, Gutschmidt A, Neher A, Feldmann K, Mollenkopf HJ, et al. Candidate biomarkers for discrimination between infection and disease caused by Mycobacterium tuberculo-

sis. J Mol Med (Berl). 2007;**85**(6):613–21. doi: 10.1007/s00109-007-0157-6. [PubMed: 17318616].

- Hasan Z, Jamil B, Khan J, Ali R, Khan MA, Nasir N, et al. Relationship between circulating levels of IFN-gamma, IL-10, CXCL9 and CCL2 in pulmonary and extrapulmonary tuberculosis is dependent on disease severity. Scand J Immunol. 2009;69(3):259–67. doi: 10.1111/j.1365-3083.2008.02217.x. [PubMed: 19281538].
- Il'yasova D, Colbert LH, Harris TB, Newman AB, Bauer DC, Satterfield S, et al. Circulating levels of inflammatory markers and cancer risk in the health aging and body composition cohort. *Cancer Epidemiol Biomarkers Prev*. 2005;14(10):2413-8. doi: 10.1158/1055-9965.EPI-05-0316. [PubMed: 16214925].
- Cavalcanti YV, Brelaz MC, Neves JK, Ferraz JC, Pereira VR. Role of TNF-Alpha, IFN-Gamma, and IL-10 in the development of pulmonary tuberculosis. *Pulm Med.* 2012;**2012**:745483. doi: 10.1155/2012/745483. [PubMed: 23251798]. [PubMed Central: PMC3515941].
- Yazdani Y, Keyhanvar N, Kalhor HR, Rezaei A. Functional analyses of recombinant mouse hepcidin-1 in cell culture and animal model. *Biotechnol Lett.* 2013;35(8):1191–7. doi: 10.1007/s10529-013-1198-2. [PubMed: 23553521].
- Riou C, Perez Peixoto B, Roberts L, Ronacher K, Walzl G, Manca C, et al. Effect of standard tuberculosis treatment on plasma cy-tokine levels in patients with active pulmonary tuberculosis. *PLoS One*. 2012;7(5). e36886. doi: 10.1371/journal.pone.0036886. [PubMed: 22606304]. [PubMed Central: PMC3351475].
- Singh PP, Goyal A. Interleukin-6: A potent biomarker of mycobacterial infection. *Springerplus*. 2013;2:686. doi: 10.1186/2193-1801-2-686. [PubMed: 24455461]. [PubMed Central: PMC3893321].
- Cowan J, Pandey S, Filion LG, Angel JB, Kumar A, Cameron DW. Comparison of interferon-gamma-, interleukin (IL)-17- and IL-22expressing CD4 T cells, IL-22-expressing granulocytes and proinflammatory cytokines during latent and active tuberculosis infection. *Clin Exp Immunol.* 2012;**167**(2):317–29. doi: 10.1111/j.1365-2249.2011.04520.x. [PubMed: 22236009]. [PubMed Central: PMC3278699].
- Savolainen LE, Kantele A, Knuuttila A, Pusa L, Karttunen R, Valleala H, et al. Combined expression of IFN-gamma, IL-17, and IL-4 mRNA by Recall PBMCs moderately discriminates active tuberculosis from latent mycobacterium tuberculosis infection in patients with miscellaneous inflammatory underlying conditions. *Front Immunol.* 2016;7:239. doi: 10.3389/fimmu.2016.00239. [PubMed: 27379100]. [PubMed Central: PMC4905973].
- Sasindran SJ, Torrelles JB. Mycobacterium tuberculosis infection and inflammation: What is beneficial for the host and for the bacterium? *Front Microbiol.* 2011;2:2. doi: 10.3389/fmicb.2011.00002. [PubMed: 21687401]. [PubMed Central: PMC3109289].
- Brighenti S, Andersson J. Induction and regulation of CD8+ cytolytic T cells in human tuberculosis and HIV infection. *Biochem Biophys Res Commun.* 2010;**396**(1):50–7. doi: 10.1016/j.bbrc.2010.02.141. [PubMed: 20494110].