

Bacterial infection in neutropenic cancer patients: an overview

Kenneth V.I. Rolston

Department of Infectious Diseases, Infection Control and Employee Health, The University of Texas M.D. Anderson Cancer Center, Houston, Texas, USA

INTRODUCTION

Infection remains the most common complication of myelosuppressive antineoplastic therapy, and is associated with substantial morbidity and mortality despite major advances in supportive care (1).

Bacterial infections are predominant during the early stages of neutropenia, whereas fungal infections are more common in patients with prolonged and severe neutropenia (2). The spectrum of bacterial and fungal infection undergoes periodic change and is impacted upon by several factors including the use of antibacterial/antifungal prophylaxis, the use of foreign medical devices (e.g. various catheters), the nature and intensity of the antineoplastic regimen, surgical procedures, and local epidemiological factors.

The standard of care for the treatment of febrile neutropenic patients is the administration of broad-spectrum antibiotic therapy with the intention to “cover” the majority of bacterial pathogens encountered in this setting. In order to achieve the best possible coverage with the initial empiric regimen, it is essential to monitor changes in the epidemiology of infections in this setting, and take

into consideration local susceptibility/ resistance patterns (3).

Etiology of fever

Fever is the most common, and sometimes the only manifestation of infection in neutropenic patients, as the usual signs/symptoms of inflammation may be blunted in this setting. Approximately 20-25% of such episodes will be due to a microbiologically documented infection (i.e. positive cultures from a normally sterile site). A similar proportion (20-25%) will be due to clinically documented infections (i.e. a clinical site of infection such as cellulitis or pneumonia but with no microbiologic documentation). The majority of episodes (45-50%) have neither a microbiologically nor a clinically documented infection. Such episodes are termed “unexplained fever” and are presumed to be caused by infection, since the majority respond to anti-infective therapy. A small proportion of febrile episodes (approximately 5%) are due to non-infectious causes (e.g. tumor fever, drug fever).

Sites of infection

The most common sites of infection and the frequency of infection at these sites are listed in table 1. These include bloodstream infections, respiratory tract infections (both upper and lower respiratory tract), urinary tract infections, and sites

Received: 19 June 2009 *Accepted:* 29 June 2009

Reprint or Correspondence: Kenneth V. I. Rolston, MD.
The University of Texas, M.D. Anderson Cancer Center,
Department of Infectious Diseases, Infection Control and
Employee Health, 1515 Holcombe Blvd. (Unit 1460)
Houston, Texas 77030 U. S. A.

E-mail: krolston@mdanderson.org

116 Bacterial infection in neutropenic cancer patients

along the gastro-intestinal tract. Most bacteremias, urinary tract infections, and some skin and skin structure infections are microbiologically documented, whereas most infections at other sites are clinically documented. The majority of these are diagnosed using a combination of clinical features and information from radiographic imaging, ultrasonography, serologic testing, or other laboratory diagnostic techniques.

Table 1. Common sites of infection in neutropenic patients

Site of infection	Frequency (%)
Bloodstream*	20-25
Respiratory tract	25-30
Urinary tract	10-15
Skin/Skin structure	10-15
Gastrointestinal tract†	5-10
Other sites#	1-5

* Including catheter-related infections

† Esophagitis, neutropenic enterocolitis, peri-rectal sites, biliary tract infections

Meningitis, septic arthritis and other uncommon infections

Despite the fact that bloodstream infections account for only 20-25% of microbiologically documented infections in patients with neutropenia, most surveys describing the etiology of bacterial infections in such patients provide detailed information only on bloodstream infections caused by single organisms (monomicrobial infections), and exclude or provide very little information about infections at other site, and about polymicrobial infections (4,5). This paints an incomplete and inaccurate picture since blood stream infections are caused predominantly by gram-positive pathogens whereas infections at many other sites are predominantly gram-negative or polymicrobial (6). For example, the EORTC and SCOPE data indicate that 75-70% of bacterial infections are caused by gram-positive pathogens although information about monomicrobial bacteremias only, was provided. This type of information led to the widespread use (misuse?) of agents such as vancomycin and teicoplanin as part of the initial empiric regimen in neutropenic

patients. Increased glycopeptide usage has been associated with increased costs, increased toxicity, and reduced susceptibility (MIC creep) or overt resistance (VISA, VRSA, VRE) among gram-positive pathogens, without significant improvement in overall outcome (mortality) of gram-positive infections (7-10).

When data from non-bacteremic sites of infection and polymicrobial infection are presented, a substantially different picture emerges (2,6). Gram-positive organisms account for <50% of documented infections, gram-negative pathogens for 20-25%, and polymicrobial infections for 25-30%. Several studies have documented that approximately 80% of polymicrobial infections have a gram-negative component, and approximately 35% are caused by multiple species of gram-negative pathogens (11,12). This changes the approach that needs to be taken when selecting agents/regimens for initial empiric therapy in febrile neutropenic patients.

Common bacterial pathogens

Gram-positive organisms: The most commonly isolated gram-positive pathogens from neutropenic patients are Coagulase-negative staphylococci (CoNS) followed by *Staphylococcus aureus*, *Enterococcus* species, and viridans group streptococci (VGE) (4,13). Organisms colonizing the skin also cause infections frequently including catheter-related bacteremias. These include *Bacillus* species and *Corynebacterium* species. Some recent reports have focused on the increasing frequency of infections caused by *Stomatococcus mucilaginosus*, particularly in patients who develop severe oral mucositis (14,15). Although *Listeria monocytogenes* and *Rhodococcus equi* are encountered more frequently in patients with impaired cellular immunity, they need to be considered when such patients are rendered neutropenic (16,17). *Streptococcus* species including *Streptococcus pneumoniae* and beta-

haemolytic streptococci (Lance- field group A, B, C, G and F) are also important pathogens in neutropenic patients (2,18,19). (Table 2)

Table 2. Common causes of infection in neutropenic patients

Gram-positive bacteria

Coagulase-negative staphylococci
Staphylococcus aureus
Enterococcus species
Viridans group streptococci
Bacillus species
Corynebacterium species
Streptococcus pneumoniae
Beta-hemolytic streptococci (Groups A, B, C, G, F)
Stomatococcus mucilaginosus

Gram-negative bacteria

Escherichia coli
Klebsiella species
Other Enterobacteriaceae
Pseudomonas aeruginosa
Pseudomonas (non-aeruginosa) species
Acinetobacter species
Stenotrophomonas maltophilia

Anaerobes

Bacteroids species
Clostridium species

Many gram-positive pathogens have developed resistance to agents commonly used for prophylaxis (the fluoroquinolones) and/or empiric therapy (beta-lactams) of febrile episodes in neutropenic patients. At most cancer treatment centers more than 90% of CoNS and >50% of *S. aureus* isolates are methicillin-resistant. Approximately 17-20% of *Enterococcus* species are glycopeptide resistant (20). Non-susceptibility to penicillin among VGS approaches 60% and 20% of these isolates have high level penicillin resistance (MIC \geq 2.0 μ g/ml) (18). Similar non-susceptibility and resistance rates have been documented for *S. pneumoniae* isolates (21). Even among susceptible gram-positive organisms, increased MIC's (MIC creep) and widespread tolerance (MCB \geq from 32 times the MIC) have been documented. Increasing levels of resistance are of great concern since the pipeline for new drugs is relatively empty (22). This also highlights

the critical role of antimicrobial stewardship and infection control in the overall management of febrile episodes in neutropenic patients (23).

Gram-negative organisms: *Escherichia coli*, *Klebsiella* species, and *Pseudomonas aeruginosa* are the most common gram-negative pathogens isolated from neutropenic patients and collectively account for 60-65% of documented bacterial infections (1,24). Other Enterobacteriaceae, *Acinetobacter* species, *Stenotrophomonas maltophilia* and non-aeruginosa *Pseudomonas* species are also encountered frequently (25,26). As with gram-positive pathogens, resistance levels among gram-negative pathogens have risen to alarming levels, and some organisms have developed unique and/or multiple mechanisms of resistance, rendering them multi-drug-resistant defined as resistance to at least 3 classes of antibiotics (27,28). Organisms of particular concern include ESBL producers, *Acinetobacter* species, *Stenotrophomonas maltophilia*, *Pseudomonas aeruginosa*, and *Klebsiella* spp. producing carbapenemases (KPC).

Gram-negative infections have traditionally been associated with greater morbidity and mortality than gram-positive infections with a few notable exceptions (MRSA, VRE). Consequently, antimicrobial prophylaxis in neutropenic patients has been targeted primarily against these organisms. The agents used most often for this indication are the fluoroquinolones (ciprofloxacin, levofloxacin). The use of prophylactic agents has reduced the frequency of febrile episodes in neutropenic patients, and the frequency of documented gram-negative infections as well (29,30). However, most studies have not shown a decrease in mortality as a result of this strategy, and many have documented either no impact on, or an increase in the frequency of gram-positive infections. However, one recent meta-analysis has pooled data from several studies indicating a reduction in overall mortality (31). This strategy has resulted in a substantial increase in the level of

118 Bacterial infection in neutropenic cancer patients

fluoroquinolone resistance in common gram-negative pathogens (e.g. *E. coli* and *P. aeruginosa*) and most societies/guidelines caution against the routine use of prophylaxis in neutropenic patient (1,32).

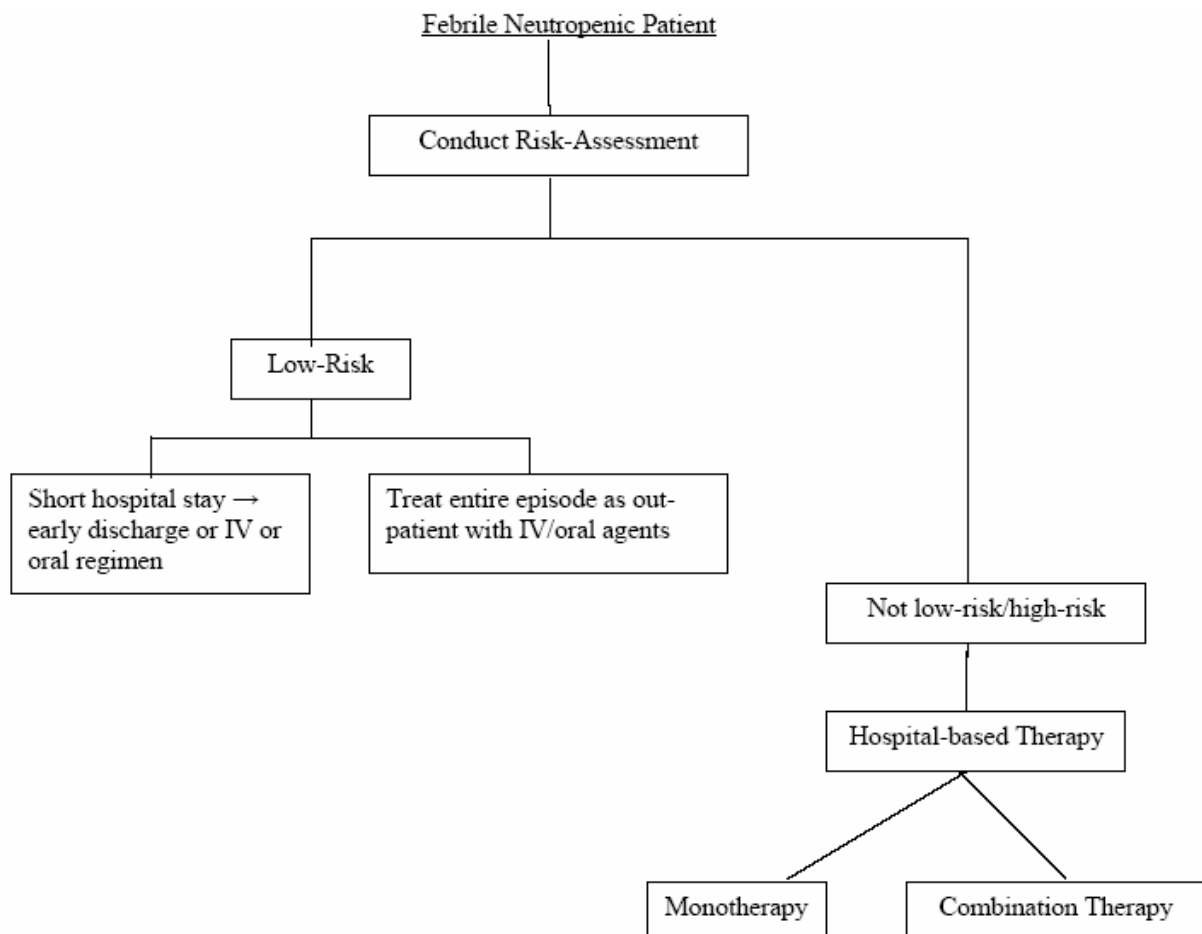
Polymicrobial Infection

As already indicated, polymicrobial infections are at least as frequent, if not more so, than single organism gram-negative infections (11,12). The majority of these infections are deep tissue infections such as pneumonia, neutropenic enterocolitis, perirectal infection, biliary-tract infections. Approximately 10-15% of bacteremias and urinary tract infections are polymicrobial as well (33).

P. aeruginosa is the most common organism isolated from such infections (45-55% of cases), perhaps indicating its ability to invade tissues more so than other organisms. One large study of 507 episodes of polymicrobial infections demonstrated that approximately 80% had a gram-negative component, and approximately 33% had multiple species of gram-negative isolates (11).

Polymicrobial infections are associated with greater morbidity and mortality than single organism infections. One study of the outcomes of bacteremia in neutropenic cancer patients suggested that these infections respond more often to combination antibacterial regimens than to monotherapy (34).

Figure 1. Management strategies for febrile episodes in neutropenic patients



Data regarding polymicrobial infections are still quite limited, as many centers fail to report them either in epidemiologic surveys or therapeutic trials. We encourage all investigators taking care of neutropenic patients to include polymicrobial infections in their reports.

Antimicrobial Therapy

The accepted standard of care is to provide broad-spectrum, empiric coverage to febrile neutropenic patients, based on local epidemiology and susceptibility/resistance patterns (Figure 1) (1,2,32). Until recently, such treatment was always administered in the hospital. It has now become possible to identify a “low-risk” subset among febrile neutropenic patients at the onset of a febrile episode (35,36).

This has made it possible to move the treatment setting from the hospital to the outpatient clinic/home environment (37,38). Consequently, the first step in the management of a febrile neutropenic patient is to conduct a risk assessment using either statistically derived risk assessment tools (e.g. the MASCC risk-index) or simple clinical criteria (35).

If the patient is classified as low-risk, a short period of stabilization in the hospital (4-48 hours) followed by outpatient antibiotic therapy, or treatment of the entire episode in the outpatient setting is appropriate (38). This strategy is associated with a high success rate, a low-rate (<3%) of complications or readmission for any reason, better resource utilization, reduced costs, and an improved quality of life for patients and their caregivers (39). If the patient is not in the low-risk category, standard, hospital-based, parenteral therapy is recommended so that closer monitoring of the patient for response, toxicity, and superinfections or other complications can be achieved (1,2). As previously mentioned, institutional differences in epidemiology, and susceptibility/resistance patterns are not

uncommon. Consequently, the specific agent(s) used for therapy will depend on local data.

Antimicrobial stewardship

The frequency and duration of antibiotic usage in neutropenic patients is probably greater than in any other patient population. Antibiotics are used for a number of indications including prophylaxis, pre-emptive therapy, empiric therapy, specific (targeted) therapy, and maintenance or suppressive therapy. All this creates significant selection pressure for the emergence of organisms that are resistant to the most commonly used antibiotics in this setting. One of the traditional methods for overcoming this problem has been the development of novel agents. However, for the last decade or so, new drug development has almost come to a standstill (22,40). This situation has forced clinicians to take a closer look at their antimicrobial usage habits, and devise methods to improve the appropriate use of these agents. This is now termed “antimicrobial stewardship”, and leading societies have published comprehensive guidelines dealing with the establishment and implementation of antimicrobial stewardship programs (23). Various stewardship strategies are outlined in table 3. These strategies, along with strict adherence to infection control policies and practices go a long way in reducing the selection and spread of resistant organisms.

In conclusion, bacterial infections occur frequently in cancer patients especially in the setting of severe neutropenia. The most important aspects of management of these patients are:

- thorough evaluation
- knowledge of local epidemiology and susceptibility/ resistance patterns
- prompt administration of empiric antibiotic therapy based on risk- group
- close monitoring and follow-up
- antimicrobial stewardship and infection control

Table 3. Recommendations for antimicrobial stewardship

Baseline data/infrastructure

- Determine local epidemiology and resistance patterns
- Know institutional formulary and prescribing habits
- Develop multidisciplinary antimicrobial stewardship team (MAST)

Recommendations for antimicrobial usage

- Limit antibacterial prophylaxis
- Encourage targeted/specific therapy
- Consider formulary restriction and/or pre-authorization
- Create guidelines and clinical pathways
- Consider antimicrobial heterogeneity
- Consider de-escalation (streamlining) of empiric regimen
- Dose optimization
- Parenteral to oral conversion
- Optimization of duration of therapy

Other strategies

- Prospective audits of antimicrobial usage with feedback to prescribers
- Educational activities (Grand Rounds, in-services)
- Strict adherence to infection control policies

REFERENCES

1. Hughes WT, Armstrong D, Bodey GP, Bow EJ, Brown AE, Calandra T, et al. 2002 guidelines for the use of antimicrobial agents in neutropenic patients with cancer. *Clin Infect Dis* 2002;34(6):730-51.
2. Rolston KVI, Bodey GP. Infections in patients with cancer. In: Kufe DW, Bast Jr, RC, Hait WN, Hong WK, Pollock RE, Weichselbaum RR, Holland JF, Frei, III E, eds. *Cancer medicine*. 7th edition, Hamilton, Ontario: BC Decker, Hamilton, Ontario. 2006;p:2222-45.
3. Rolston KVI. Challenges in the treatment of infections caused by gram-positive and gram-negative bacteria in patients with cancer and neutropenia. *Clin Infect Dis* 2005;40:S246-52.
4. Wisplinghoff H, Seifert H, Wenzel RP, Edmond MB. Current trends in the epidemiology of nosocomial bloodstream infections in patients with hematological malignancies and solid neoplasms in hospitals in the United States. *Clin Infect Dis* 2003;36:1103-10.
5. Zinner SH. Changing epidemiology of infections in patients with neutropenia and cancer: emphasis on gram-positive and resistant bacteria. *Clin Infect Dis* 1999;29:490-94.
6. Yadegarynia D, Tarrand J, Raad I, Rolston K. Current spectrum of bacterial infections in cancer patients. *Clin Infect Dis* 2003;37:1144-45.

7. Sakoulas G, Moise-Broder PA, Schentag J, Forrest A, Moellering RC Jr., Eliopoulos GM. Relationship of MIC and bactericidal activity to efficacy of vancomycin for treatment of methicillin-resistant *Staphylococcus aureus* bacteremia. *J Clin Microbiol* 2004;42:2398-402.
8. Kapadia M, Coyle E, Prince R, Rolston KVI. Declining in-vitro activity of vancomycin against *Staphylococcus aureus* isolates from cancer patients. (Abstract # E-807) 45th Annual Interscience Conference on Antimicrobial Agents and Chemotherapy. Washington, D.C., Dec. 16-19, 2005.
9. Moise-Broder PA, Sakoulas G, Eliopoulos GM. Accessory gene regulator group II polymorphism in methicillin-resistant *Staphylococcus aureus* is predictive of failure of vancomycin therapy. *Clin Infect Dis* 2004;38:1700-5.
10. Safdar A, Rolston KVI. Vancomycin, tolerance, a potential mechanism for refractory gram-positive bacteremia observational study in patients with cancer. *Cancer* 2006; 106:1815-20.
11. Elting LS, Bodey GP, Fainstein V. Polymicrobial septicemia in the cancer patient. *Medicine* 1986;65:218-25.
12. Rolston KVI, Bodey GP, Safdar A. Polymicrobial infection in patients with cancer: An underappreciated and underreported entity. *Clin Infect Dis* 2007;45:228-33.
13. Rolston KVI, Yadegarynia D, Kontoyiannis DP, Raad II, Ho DH. The spectrum of gram-positive bloodstream infections in patients with hematologic malignancies, and the in-vitro activity of various quinolones against gram-positive bacteria isolated from cancer patients. *Int J Infect Dis* 2006;10:223-30.
14. Goldman M, Chaudhary UB, Greist A, Fausel CA. Central nervous system infections due to *Stomatococcus mucilaginosus* in immunocompromised hosts. *Clin Infect Dis* 1998;27:1241-46.
15. Fanourgiakis P, Georgala A, Vekemans M, Daneau D, Heymans C, Aoun M. Bacteremia due to *Stomatococcus mucilaginosus* in neutropenic patients in the setting of a cancer institute. *Clin Microbiol Infect* 2003;9:1068-72.
16. Rivero GA, Torres HA, Rolston KVI, Kontoyiannis DP. *Listeria monocytogenes* infection in patient with cancer. *Diag Microbiol Infect Dis* 2003;47: 393-98.
17. Weinstock DM, Brown AE. *Rhodococcus equi*: an emerging pathogen. *Clin Infect Dis* 2002;15:1379-85.
18. Han XY, Kamana M, Rolston KVI. Viridans streptococci isolated by culture from blood of cancer

- patients: Clinical and microbiologic analysis of 50 cases. *J Clin Microbiol* 2006;44:160-65.
19. Kumashi P, Girgawy E, Tarrand J, Rolston KV, Raad II, Safdar A. Streptococcus pneumoniae bacteremia in patients with cancer: Disease characteristics and outcomes in the era of escalating drug resistance (1998-2002). *Medicine* 2005; 84:303-12.
 20. Matar MJ, Tarrand J, Raad I, Rolston KVI. Colonization and infection with vancomycin-resistant Enterococcus among patients with cancer. *Am J Infect Control* 2006;34:534-36.
 21. Youssef S, Rodriguez G, Rolston KV, Champlin RE, Raad II, Safdar A. Streptococcus pneumoniae infections in 47 hematopoietic stem cell transplantation recipients. Clinical characteristics of infections and vaccine-breakthrough infections, 1989-2005. *Medicine* 2007;86:69-77.
 22. Bad bugs, no drugs. Infectious Diseases Society of America. July 2004. <http://www.idsociety.org/pa/IDSA-Paper4-final-web.pdf>.
 23. Dellit TH, Owens RC, McGowan JE Jr, Gerding DN, Weinstein RA, Burke JP, et al. Infectious Diseases Society of America; Society for Healthcare Epidemiology of America. Infectious Diseases Society of America and the Society for Healthcare Epidemiology of America guidelines for developing an institutional program to enhance antimicrobial stewardship. *Clin Infect Dis* 2007;44(2):159-77.
 24. Rolston KVI, Tarrand JJ. Pseudomonas aeruginosa; Still a frequent pathogen in patients with cancer: 11-year experience from a comprehensive cancer center. *Clin Infect Dis* 1999;29:463-64.
 25. Rolston KVI, Kontoyiannis DP, Yadegarynia D, Raad I. Non-fermentative gram-negative bacilli in cancer patients: increasing frequency of infection and antimicrobial susceptibility of clinical isolates to fluoroquinolones. *Diag Microbiol Infect Dis* 2005;51:215-18.
 26. Safdar A, Rolston KV. Stenotrophomonas maltophilia: Changing spectrum of a serious bacterial pathogen in patients with cancer. *Clin Infect Dis* 2007;45:1602-9.
 27. Toleman MA, Rolston K, Jones RN, Walsh TR. blaVIM-7, an evolutionarily distinct metallo-β-lactamase gene in a Pseudomonas aeruginosa isolate from the United States. *Antimicrob Agents Chemother* 2004;48:329-32.
 28. Aboufaycal H, Sader HS, Rolston K, Deshpande LM, Toleman M, Bodey G, et al. blaVIM-2 and blaVIM-7 carbapenemase-producing Pseudomonas aeruginosa isolates detected in a tertiary care medical center in the United States: report from the MYSTIC program. *J Clin Microbiol* 2007;45(2):614-5.
 29. Bucaneve G, Micozzi A, Menichetti F, Martino P, Dionisi MS, Martinelli G, et al.; Gruppo Italiano Malattie Ematologiche dell'Adulto (GIMEMA) Infection Program. Levofloxacin to prevent bacterial infection in patients with cancer and neutropenia. *N Engl J Med* 2005;353(10):977-87.
 30. Cullen M, Steven N, Billingham L, Gaunt C, Hastings M, Simmonds P, et al; Simple Investigation in Neutropenic Individuals of the Frequency of Infection after Chemotherapy +/- Antibiotic in a Number of Tumors (SIGNIFICANT) Trial Group. Antibacterial prophylaxis after chemotherapy for solid tumors and lymphomas. *N Engl J Med* 2005;353(10):988-98.
 31. Gafter-Gvill A, Fraser A, Paul M, Liebovici L. Meta-analysis: antibiotic prophylaxis reduces mortality in neutropenic patients. *Ann Intern Med* 2005;142:979-95.
 32. Segal BH, Freifeld AG, Baden LR. Prevention and treatment of cancer-related infections. *J Natl Compr Canc Netw* 2008;6:122-74.
 33. Klastersky J, Ameye L, Maertens J, Georgala A, Muanza F, Aoun M, et al. Bacteraemia in febrile neutropenic cancer patients. *Int J Antimicrob Agents* 2007;30 Suppl 1:S51-9.
 34. Elting LS, Rubenstein EB, Rolston KVI, Bodey GP. Outcomes of bacteremia in patients with cancer and neutropenia: observations from two decades of epidemiological and clinical trials. *Clin Infect Dis* 1997;25:247-59.
 35. Klastersky J, Paesmans M, Rubenstein EB, Boyer M, Elting L, Feld R, et al. The Multinational Association for Supportive Care in Cancer risk index: A multinational scoring system for identifying low-risk febrile neutropenic cancer patients. *J Clin Oncol* 2000;18(16):3038-51.
 36. Feld R, Paesmans M, Freifeld AG, Klastersky J, Pizzo PA, Rolston KV, et al.; Immunocompromised Host Society; Multinational Association for Supportive Care in Cancer. Methodology for clinical trials involving patients with cancer who have febrile neutropenia: updated guidelines of the Immunocompromised Host Society/Multinational Association for Supportive Care in Cancer, with emphasis on outpatient studies. *Clin Infect Dis* 2002;35(12):1463-8.
 37. Kern KV. Risk assessment and treatment of low-risk patients with febrile neutropenia. *Clin Infect Dis* 2006;15:533-40.

122 Bacterial infection in neutropenic cancer patients

38. Rolston KVI. Oral antibiotic administration and early hospital discharge is a safe and effective alternative for treatment of low-risk neutropenic fever. *Cancer Treatment Rev* 2003;29:551-54.

39. Rolston K. New trends in patient management: Risk-based therapy for febrile patients with neutropenia. *Clin Infect Dis* 1999;29:515-21.

40. Talbot GH, Bradley J, Edwards JE, Jr., Gilbert D, Scheld M, Bartlett JG. Bad bugs need drugs: An update on the development pipeline from the antimicrobial availability task force of the Infectious Diseases Society of America. *Clin Infect Dis* 2006;42:657-68.