Published online 2019 August 7.

**Research Article** 

# Detection of Enteroviruses in Children with Acute Diarrhea

Morteza Fazelipour<sup>1</sup>, Manoochehr Makvandi<sup>1,2,\*</sup>, Alireza Samarbafzadeh<sup>1</sup>, Niloofar Nisi<sup>1,2</sup>, Azarakhsh Azaran<sup>1,2</sup>, Shahram Jalilian<sup>2</sup>, Roya Pirmoradi<sup>2</sup>, Roya Nikfar<sup>3</sup>, Ahmad Shamsizadeh<sup>3</sup> and Kambiz Ahmadi Angali<sup>4</sup>

<sup>1</sup>Infectious and Tropical Diseases Research Center, Health Research Institute, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran

<sup>2</sup>Virology Department, School of Medicine, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran

<sup>3</sup>Infectious Department, Abozar Hospital, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran

<sup>4</sup>Biostatistic Department, School of Health, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran

<sup>\*</sup> Corresponding author: Infectious and Tropical Disease Research Center, Health Research Institute, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran. Email: manoochehrmakvandi299@gmail.com

Received 2018 September 04; Revised 2019 May 25; Accepted 2019 June 16.

#### Abstract

**Background:** Diarrhea is one of the most significant diseases in children, causing morbidity and mortality worldwide. Diarrhea is caused by viruses, bacteria, and parasites. There are several viruses that can cause diarrhea, including some groups of enteroviruses that have a significant role in acute diarrhea in children.

Objectives: This study was conducted to evaluate the presence of enteroviruses in the stool of children with diarrhea.

**Methods:** We collected 85 stool samples including 50 (58.82%) from males and 35 (41.17%) from females with acute diarrhea. All the stool samples proved negative for bacterial and parasitic pathogens. The RNA was extracted from the stool samples and cDNA was prepared. The semi-nested PCR was carried out for the detection of the 5'-UTR region of enteroviruses. To determine the enterovirus serotypes, the sequences of semi-nested PCR product was performed using conserved primers for the 5'-UTR region.

**Results:** Overall, 21/85 (24.7%) patients including 12/50 (24%) males and 9/35 (25.71%) females showed positive results for enteroviruses (P = 0.3). Based on the results of sequencing, one of the isolated serotypes was identified as coxsackievirus A6 and the other isolated serotype was echovirus 9.

**Conclusions:** Overall, 21/85 (24.7%) children with acute diarrhea were infected with enteroviruses. The distribution of enteroviruses was not significantly different between male and female patients. The results of sequencing indicated that one of the isolated serotypes was coxsackievirus A6 and the other isolated serotype was echovirus 9. The remaining 64/85(75.29%) isolates were negative for enteroviruses. The role of other viral gastroenteritis agents including rotaviruses, noroviruses, calicivirus, astrovirus, and adenoviruses was not explored that needs further investigation.

Keywords: Diarrhea, Enterovirus, Polymerase Chain Reaction

## 1. Background

Diarrhea is caused by viruses broadly circulating in low-income countries and is the third leading cause of death in children younger than five years of age. Its highest incidence occurs in developing countries where there are 2 - 2.5 million deaths annually worldwide (1, 2). Several viruses including rotavirus (RV), norovirus (NoV), astrovirus (AstV), and adenovirus (AdV) account for a significant number of cases of acute gastroenteritis (3-6).

Recently, the association of enteroviruses with acute gastroenteritis has been reported (5, 6). Enteroviruses (EVs) are small, non-enveloped, single-stranded RNA viruses that belong to the Picornaviridae family (7, 8).

Enteroviruses are classified into poliovirus (PV) consisting of three serotypes, coxsackievirus type A (CV-A) with 23 serotypes, coxsackievirus type B (CV-B) with 6 serotypes, and echovirus (E) with 28 serotypes (7), besides some more recently discovered numbered serotypes. EVs show both endemic and epidemic patterns (8). Epidemics are typically caused by certain EV types such as E-30 (causing aseptic meningitis), EV-D70 (causing acute hemorrhagic conjunctivitis), EV-A71 (causing severe central nervous system), CV-A6 and CV-A10 (causing severe HFMD), and EV-D68 (causing flu-like symptoms) (8, 9). They are responsible for major public health concerns (9). EVs are considered a significant cause of many illnesses, including gastroenteritis, paralysis, meningitis, cardiomyopathy, respiratory tract infection, and the central nervous system infection (10-12). Although most infections are asymptomatic or cause less severe conditions, such as colds and fever, about 1.0% of

Copyright © 2019, Archives of Clinical Infectious Diseases. This is an open-access article distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 International License (http://creativecommons.org/licenses/by-nc/4.0/) which permits copy and redistribute the material just in noncommercial usages, provided the original work is properly cited.

EV infections result in severe diseases with high morbidity and mortality in young children and infants (10). EVs are transmitted through fecal-oral and respiratory routes.

The association of EV infection with diarrheal diseases is not well confirmed. However, the role of EV infection in acute diarrhea has been reported by a number of studies (5-10). Some EVs including E11, CVA6, CVB2, PV3, CVB4, E18, and CVA2 were detected in the stool of children with acute diarrhea in Hebei province, China (13). Moreover, the outbreak of gastroenteritis caused by echovirus type 6 was reported among children in Japan (14). E11 was similarly reported to be responsible for outbreaks of diarrhea in Malaysia (15) and southern India (16). Moreover, acute gastroenteritis caused by Coxsackie B-6 virus was confirmed in infants and children in Brasilia (17).

Another genus of picornaviridae named human parechovirus (HPeVs) has been found to be implicated in acute gastroenteritis (18). The role of EVs in acute diarrhea in children has not been yet investigated in Iran. However, the distribution of echovirus 30 and coxsackievirus A9 was formerly investigated among young neonates with sepsis in Ahvaz city, Iran (19). The complete sequence of echovirus 30 isolated from patients with meningitis has been detected and analyzed in Ahvaz (20). The high prevalence (59.09%) of EVs was also approved in children with aseptic meningitis in Ahvaz city, Iran (21).

#### 2. Objectives

The detection of EVs is often not included in the pathogen surveillance of diarrhea, particularly in Iran. Nevertheless, based on the reported circulation of EVs in Ahvaz city, this study was conducted to evaluate the epidemiology and molecular characteristics of EVs circulating in outpatient children with acute diarrhea under < 5 years of age referring to Abuzar Hospital, Ahvaz, Iran.

## 3. Methods

At the initial stage of the study, about 300 liquidly fecal stool samples were collected from under-five children with acute diarrhea. All the patients were admitted to Abuzar Children Hospital, Ahvaz, Iran, during January - December 2013. Abuzar Hospital is the only referral children hospital in Ahvaz city. The inclusion criteria were samples negative for bacterial and parasitic pathogens, children younger than five years of age, and patients with acute diarrhea for less than 48 hours. The exclusion criteria were stool samples positive for bacterial and parasitic pathogens, the presence of WBC in the stool specimen, patients with mild diarrhea, and patients aged > 5 years. Based on the inclusion/exclusion criteria, 215 stool samples were excluded and the remaining 85 stool samples were kept at -70°C for the prospective detection of EVs. Diarrhea is defined as  $\geq$  3 times/day defecation for less than 14 days (22). We collected data regarding the clinical characteristics of diarrhea, clinical history, physical examination results encompassing frequency and duration of diarrhea, vomiting, electrolyte disturbance, dehydration status, and the epidemiologic data including age, gender and treatment type. The data were recorded according to the WHO criteria (23).

All children presented clinical signs of gastroenteritis, i.e. fever and/or abdominal pain and/or diarrhea and/or vomiting. Diarrhea was the most frequently reported symptom (85/85: 100%), followed by vomiting (59/85: 69.41%), and fever (48/85: 56.47%). Abdominal pain was observed in 28 cases (32.94%).

#### 3.1. Extraction of RNA and Preparation of cDNA

The viral RNA was extracted from the sample using a High-pure Viral Nucleic Acid Kit (Roche, Germany) following the manufacturer's instructions. The cDNA was prepared (Fermentas, USA) according to the manufacturer's instructions. The identification of the conserved 5'-UTR and/or the viral capsid sequence could differentiate between different EV species and types (24).

#### 3.2. Semi-Nested PCR

Numerous EVs do not grow simply in cell culture. Hence, their diagnosis is based on PCR, which is more sensitive than traditional methods. In addition, PCR is more sensitive, reliable, and faster than culture. At present, PCR is the "gold standard" for EVs detection (25).

The semi-nested PCR was carried out for the detection of 5'-UTR region of EVs. The PCR reaction mixture contained 2.5  $\mu$ L of PCR buffer, 0.5  $\mu$ L of dNTP, 0.75  $\mu$ L of each of EV-F1 and EV-R primers (Table 1), 0.75  $\mu$ L of MgCl<sub>2</sub>, 0.12  $\mu$ L of Tag polymerase, 2.5  $\mu$ L of template, and up to 25  $\mu$ L of distilled water (DW). The reaction mixture was transferred to a thermocycler (TC-512, Techne, UK) for 35 cycles of amplification (94°C for 45 seconds, 54°C for 45 seconds, 72°C for 45 seconds, and 72°C for 10 minutes). Then, 1  $\mu$ L of the product was used as the template for the second round that contained 2.5  $\mu$ L of PCR buffer, 0.5  $\mu$ L of dNTP, 0.75  $\mu$ L of each of EV-F2 and EV-R primers, 0.75  $\mu$ L of MgCl<sub>2</sub>, 1  $\mu$ L of template, 0.12  $\mu$ L of Tag polymerase, and 18.62  $\mu$ L of DW. The mixture was subjected to 35 cycles of amplification (94°C for 30 seconds, 58°C for 30 seconds, 72°C for 30 seconds, and 72°C for 5 minutes). The final product of 155 bp electrophoresed on the 2% gel indicated a positive result (21). The Sabin vaccine was used as a positive control.

Lable I. Universal Primers of Semi-Nested PCR					
Primer	Genome Region	Sequence	Product Size	Reference	
First step	5' UTR	EV-1 (F1): CAAGCACTTCTGTTTCCCCGG	- 440 bp	(25)	
		EV-R: ATTGTCACCATAAGCAGCCA			
Second step	5' UTR	EV2 (F2): TCCTCCGGCCCCTGAATGCG	- 155 bp	(25)	
		EV-R: ATTGTCACCATAAGCAGCCA			

Table 1. Universal Primers of Semi-Nested PCF

## 3.3. Sequencing

To confirm the results of PCR and determine the EV serotypes, two positive PCR products were randomly selected and sequenced (Bioneer Company, South Korea). The sequences were blasted using available databases. A phylogenic tree was constructed with the neighborjoining method using the partial nucleotide sequences of 5'-UTR region of EV positive samples. The reference sequences were retrieved from GenBank using their accession numbers.

## 3.4. Statistical Analysis

Statistical analysis was carried out for variables including gender, age groups, and season. The variables were recorded as means and standard deviation and analyzed by the chi-square test to determine the association of EVs with gender and season. Statistical analysis was performed using SPSS version 17.0 software. P values of < 0.05 were considered statistically significant.

## 4. Results

The patients' age ranged between < 12 months and 60 months. The distribution of EVs among different age groups is presented in Table 2.

Overall, 21/85 (24.7%) children including 12/50 (24%) males and 9/35 (25.7%) females showed positive results for EVs (P=0.3). The remaining 64/85 (75.29%) showed negative results for EVs. Figure 1 shows the results of the PCR assay.



Figure 1. 5' UTR PCR. N: Line 1, L, 100-bp DNA ladder, line 2N, negative control; line 1 - 5, unknown samples line; line 6, positive control.

Based on the results of sequencing, one of the isolated samples from a two-year-old male was associated with coxsackievirus A6 with accession number BankIt2145449 Ahvaz MH813958 and the other one was echovirus 9 isolated from a one-year-old female patient with accession number BankIt2145451 Ahvaz MH813959. The results of the phylogenetic tree are demonstrated in Figures 2 and 3.

Table 2 shows that the frequency of EVs was not found to be significantly different between males and females (P = 0.3). The distribution of EVs among different age groups was not significant (P=0.468). The frequency of EVs proved to be the highest in winter (P = 0.001).

#### 5. Discussion

The association between human enterovirus (EV) infection and acute gastroenteritis is well documented (5-18). In our study, 21/85 (24.7%) patients with acute diarrhea were positive for EVs. The prevalence of EVs in children with gastroenteritis has been reported in different parts of the world. Alcala et al. (Venezuela, 2018) detected EVs in 37.9% of children with gastroenteritis (5). Chitambar et al. (Mumbay, 2012) observed EVs in 40% of children

Table 2. The Profile of Patients with Acute Diarrhea Associated with EVs					
Characteristics	<b>EV Positive</b>	Percentage	P Value		
No. of patients (n = 85)	21	24.7	0.3		
Female	12/35	14.11			
Male	12/50	14.11			
Age (months)			0.468		
> 12	10/45	11.76			
13 - 24	3/16	3.52			
25 - 36	2/6	2.35			
37-48	0/3	-			
49 - 60	6/15	7.05			
Season			0.001		
Spring	0/10	-			
Summer	0/11	-			
Autumn	7/29	8.23			
Winter	17/35	20			



Figure 2. Neighbor-joining phylogenetic tree drawn with MEGA version 6 software based on the nucleotide sequences in the 5'-UTRregion of coxsackievirus A6 strain with the accession number BankIt2145449Ahvaz MH813958 labeled by black solid circle isolated from a patient with sepsis and comparison with other coxsackievirus A6 isolated from other regions of the world. Scale bar = 0.02.

with gastroenteritis, which was higher than the prevalence found in this study (26). Bubba et al. (Milan, Italy, 2015) found that 28/111 (25.2%) patients with gastroenteritis were EV-positive (27); this finding is in concordance with our results (24.7%). The frequency of EVs was not significantly different between males and females (14.11% vs. 10.58%; P > 0.05). In the present study, the obtained results of sequencing and blasting of the 5'-UTR region showed that one of the isolated serotypes from a two-year-old male showed 99% nucleotide identity to coxsackievirus A6 responsible for hand, foot, and mouth disease in China and Vietnam (28, 29), and 98% nucleotide identity to coxsackievirus A6 isolated in China (30). The role of coxsackievirus A6 in gastroenteritis in children was also reported by Chansaenroj et al. in Thailand (8). Another coxsackievirus A strain was also associated with gastroenteritis in children. Shobha et al. (Mumbay, 2012) reported coxsackievirus A13, 17, 19, and 21 in patients with gastroenteritis (26). The results of sequencing and blasting of the 5'-UTR region of the other EV strain isolated from a one-year-old female patient showed 96% nucleotide identity to echovirus 9 isolated from a small child at the clinical onset of type 1 diabetes in Finland and 94% nucleotide identity to echovirus 9 isolated from patients with aseptic meningitis in South Korea (31, 32). Other echovirus serotypes have been also associated with gastroenteritis in children. Shobha et al. (Mumbay, 2012) found echovirus serotypes 21 and 32 in patients with gastroenteritis (26).

The results of the phylogenetic tree exhibited that isolated coxsackievirus A6 was clustered with isolated coxsackievirus A6 causing hand, foot, and mouth disease in China (KX064302.1) and Vietnam (KX430802.1). The results of sequencing and blasting of the 5'-UTR region showed that isolated echovirus 9 showed 96% nucleotide identity to isolated echovirus 9 in Finland (KC238667.1).

In this study, EVs associated with acute diarrhea was detected in all age groups except for 37- 48-month age group. The highest rates of EVs were detected in patients' age group of < 12 months (10/85; 11.76%) and 49 - 60 months (6/85; 7.05%).

The current study investigated the frequency of EVs associated with acute diarrhea and observed its occurrence only in autumn and winter seasons. The rate of EVs was found higher in winter than in autumn (P=0.001). Shobha et al. (Mumbay, 2012) reported the high prevalence of EVs in autumn (40%) and winter 30% (26). Bubba et al. (Milan, Italy, 2015) documented the high prevalence of EVs in winter and autumn (27). To the best of our knowledge, it is the



Figure 3. Neighbor-joining phylogenetic tree of the nucleotide sequences in the 5'-UTRregion of Echovirus 9 strain with accession number Banklt2145451Ahvaz MH813959 isolated from a patient with sepsis in Ahwaz, Iran, labeled by the black square. It was compared with other echovirus 9 strains isolated from other regions of the world. Numbers in the branch are reproducibility after 1000 bootstraps. Scale bar = 0.005.

first report of EVs to cause acute diarrhea in Iran.

Viral intestinal infections are the most common cause of acute infectious diarrhea in children. Over the past decade, there have been major advances in the understanding of viral gastroenteritis etiology. Group A rotavirus is responsible for the majority of acute diarrhea cases in young children worldwide (28). Furthermore, other viruses like norovirus, adenovirus, enterovirus, bocavirus, sapovirus, astrovirus, calicivirus and more recently, torovirus and parechovirus have been associated with acute infectious diarrhea in children (33, 34).

# 5.1. Conclusions

In this study, a high prevalence of EVs (21/85; 24.7%) was detected in children with acute diarrhea, calling attention to the potential importance of EVs in diarrhea development. Based on sequencing results, one of the isolates in this study was identified as coxsackievirus A6 and the other isolate was echovirus 9. The remaining 64/85 (75.29%) showed negative results for EVs. Therefore, more studies

are required to elucidate the distribution and characteristics of EV serotypes and their implications in this region. Based on the aforementioned results, it is suggested that the screening of enteroviruses be implemented by molecular methods such as PCR or real-time PCR in children with acute diarrhea.

## Acknowledgments

This study was conducted as a research project with registration number 92117 carried out by Morteza Fazeli Pour (M.Sc., Virology). The authors appreciate all efforts made by the staff of the laboratory of Abuzar Children Hospital for providing the samples, as well as the Chief of the Infectious and Tropical diseases Research Center for approval and financial support of this study.

## Footnotes

Authors' Contribution: Experimental work: Morteza Fazelipour; study design: Manoochehr Makvandi; study

supervision: Manoochehr Makvandi and Alireza Samarbafzadeh; coordination: Niloofar Nisi and Shahram Jalilian; sample collection: Azarakhsh Azaran and Roya Pirmoradi; children specialist (patient recruitment): Roya Nikfar and Ahmad Shamsizadeh; statistical analysis: Kambiz Ahmadi Angali.

**Conflict of Interests:** Upon the completion of the study, all authors had no conflict of interest.

**Ethical Approval:** This study was approved by the Ethics Committee of Deputy of Research, Ahvaz Jundishapur University of Medical Sciences, with approval No. ajums.REC,1392.55.

**Funding/Support:** This research project, with registration number 92117, was financially supported by the Infectious and Tropical Diseases Research Center, Health Research Institute, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran.

**Patient Consent:** Signed informed consent was obtained from parents.

#### References

- Liu L, Oza S, Hogan D, Chu Y, Perin J, Zhu J, et al. Global, regional, and national causes of under-5 mortality in 2000-15: An updated systematic analysis with implications for the sustainable development goals. *Lancet*. 2016;**388**(10063):3027-35. doi: 10.1016/S0140-6736(16)31593-8. [PubMed: 27839855]. [PubMed Central: PMC5161777].
- G. B. D. Diarrhoeal Diseases Collaborators. Estimates of global, regional, and national morbidity, mortality, and aetiologies of diarrhoeal diseases: A systematic analysis for the Global Burden of Disease Study 2015. *Lancet Infect Dis.* 2017;17(9):909–48. doi: 10.1016/S1473-3099(17)30276-1. [PubMed: 28579426]. [PubMed Central: PMC5589208].
- Monavari SHR, Hadifar S, Mostafaei S, Miri A, Keshavarz M, Babaei F, et al. Epidemiology of rotavirus in the Iranian children: A systematic review and meta-analysis. J Glob Infect Dis. 2017;9(2):66-72. doi:10.4103/0974-777X.205173. [PubMed: 28584458]. [PubMed Central: PMC5452554].
- Sriwanna P, Chieochansin T, Vuthitanachot C, Vuthitanachot V, Theamboonlers A, Poovorawan Y. Molecular characterization of human adenovirus infection in Thailand, 2009-2012. *Virol J.* 2013;10:193. doi: 10.1186/1743-422X-10-193. [PubMed: 23758792]. [PubMed Central: PMC3693972].
- Alcala AC, Perez K, Blanco R, Gonzalez R, Ludert JE, Liprandi F, et al. Molecular detection of human enteric viruses circulating among children with acute gastroenteritis in Valencia, Venezuela, before rotavirus vaccine implementation. *Gut Pathog.* 2018;10:6. doi: 10.1186/s13099-018-0232-2. [PubMed: 29483944]. [PubMed Central: PMC5822563].
- Jain S, Thakur N, Grover N, Vashistt J, Changotra H. Prevalence of rotavirus, norovirus and enterovirus in diarrheal diseases in Himachal Pradesh, India. Virusdisease. 2016;27(1):77–83. doi: 10.1007/s13337-016-0303-2. [PubMed: 26925447]. [PubMed Central: PMC4758311].
- Ouedraogo N, Kaplon J, Bonkoungou IJ, Traore AS, Pothier P, Barro N, et al. Prevalence and genetic diversity of enteric viruses in children with diarrhea in Ouagadougou, Burkina Faso. *PLoS One*. 2016;11(4). e0153652. doi: 10.1371/journal.pone.0153652. [PubMed: 27092779]. [PubMed Central: PMC4836733].

- Chansaenroj J, Tuanthap S, Thanusuwannasak T, Duang-In A, Klinfueng S, Thaneskongtong N, et al. Human enteroviruses associated with and without diarrhea in Thailand between 2010 and 2016. *PLoS One*. 2017;**12**(7). e0182078. doi: 10.1371/journal.pone.0182078. [PubMed: 28750058]. [PubMed Central: PMC5531555].
- Tan Y, Hassan F, Schuster JE, Simenauer A, Selvarangan R, Halpin RA, et al. Molecular evolution and intraclade recombination of enterovirus D68 during the 2014 outbreak in the United States. J Virol. 2016;90(4):1997–2007. doi: 10.1128/JVI.02418-15. [PubMed: 26656685]. [PubMed Central: PMC4733988].
- Zhou HT, Yi HS, Guo YH, Pan YX, Tao SH, Wang B, et al. Enterovirusrelated diarrhoea in Guangdong, China: Clinical features and implications in hand, foot and mouth disease and herpangina. *BMC Infect Dis.* 2016;16:128. doi: 10.1186/s12879-016-1463-9. [PubMed: 26983856]. [PubMed Central: PMC4794821].
- Lee MK, Chan PK, Ho II, Lai WM. Enterovirus infection among patients admitted to hospital in Hong Kong in 2010: Epidemiology, clinical characteristics, and importance of molecular diagnosis. *J Med Virol.* 2013;85(10):1811-7. doi: 10.1002/jmv.23663. [PubMed: 23824683].
- Macaya A, Felipe-Rucian A. Enterovirus and neurological complications. Anales de Pediatria (English Edition). 2017;86(3):107–9. doi: 10.1016/j.anpede.2017.01.001.
- Shen XX, Qiu FZ, Li GX, Zhao MC, Wang J, Chen C, et al. A case control study on the prevalence of enterovirus in children samples and its association with diarrhea. *Arch Virol.* 2018;164(1):63–8. doi: 10.1007/s00705-018-4021-5.
- Abe O, Kimura H, Minakami H, Akami M, Inoue M, Saito A, et al. Outbreak of gastroenteritis caused by echovirus type 6 in an orphanage in Japan. *J Infect.* 2000;**41**(3):285–6. doi: 10.1053/jinf.2000.0748. [PubMed: 11120625].
- Bina Rai S, Wan Mansor H, Vasantha T, Norizah I, Chua KB. An outbreak of echovirus 11 amongst neonates in a confinement home in Penang, Malaysia. *Med J Malaysia*. 2007;62(3):223–6. [PubMed: 18246912].
- Patel JR, Daniel J, Mathan VI. An epidemic of acute diarrhoea in rural southern India associated with echovirus type 11 infection. *J Hyg* (*Lond*). 1985;**95**(2):483–92. doi: 10.1017/s0022172400062902. [PubMed: 4067300]. [PubMed Central: PMC2129531].
- Carvalho RPS, Kirchner E, Moura RA, Murahovschi J. Acute gastroenteritis associated with coxsackie B-6 virus in infants and children, in Sao Paulo, Brazil. *Rev Inst Med Trop Sao Paulo*. 1977;19(1):73–6.
- Chen X, Shi T, Huang J, Xiao G, Huang J, Xiong Y, et al. Molecular detection and phylogenetic analysis of human parechovirus in individuals with acute diarrhea and healthy controls in Guangzhou, China. *J Med Virol.* 2018;**90**(9):1444–52. doi: 10.1002/jmv.25222. [PubMed: 29738080].
- Shabani A, Makvandi M, Samarbafzadeh A, Teimoori A, Rasti M, Karami C, et al. Echovirus 30 and coxsackievirus A9 infection among young neonates with sepsis in Iran. *Iran J Microbiol*. 2018;**10**(4):258–65. [PubMed: 30483379]. [PubMed Central: PMC6243150].
- Abbasi S, Makvandi M, Teimoori A, Samarbaf-Zadeh A. Complete genome sequence of a multi-recombinant echovirus 6 strain isolated from CSF in Ahvaz, Southwestern Iran. J Chin Med Assoc. 2018;81(4):340-7. doi: 10.1016/j.jcma.2017.06.026. [PubMed: 29475814].
- Rasti M, Makvandi M, Neisi N, Azaran A, Rastegarvand N, Khalafkhany D, et al. Three cases of mumps virus and enterovirus coinfection in children with enteroviral meningitis. *Medicine (Baltimore)*. 2016;**95**(49). e5610. doi: 10.1097/MD.000000000005610. [PubMed: 27930588]. [PubMed Central: PMC5266060].
- Shane AL, Mody RK, Crump JA, Tarr PI, Steiner TS, Kotloff K, et al. 2017 Infectious Diseases Society of America clinical practice guidelines for the diagnosis and management of infectious diarrhea. *Clin Infect Dis*. 2017;**65**(12):e45–80. doi: 10.1093/cid/cix669. [PubMed: 29053792]. [PubMed Central: PMC5850553].
- 23. Kirk MD, Pires SM, Black RE, Caipo M, Crump JA, Devleesschauwer B, et al. World Health Organization estimates of the global and regional disease burden of 22 foodborne bacterial, protozoal, and viral

diseases, 2010: A data synthesis. *PLoS Med*. 2015;**12**(12). e1001921. doi: 10.1371/journal.pmed.1001921. [PubMed: 26633831]. [PubMed Central: PMC4668831].

- Oberste MS, Maher K, Kilpatrick DR, Flemister MR, Brown BA, Pallansch MA. Typing of human enteroviruses by partial sequencing of VP1. *J Clin Microbiol*. 1999;**37**(5):1288–93. [PubMed: 10203472]. [PubMed Central: PMC84754].
- Robinson CC, Willis M, Meagher A, Gieseker KE, Rotbart H, Glode MP. Impact of rapid polymerase chain reaction results on management of pediatric patients with enteroviral meningitis. *Pediatr Infect Dis J.* 2002;**21**(4):283-6. [PubMed: 12075757].
- Chitambar S, Gopalkrishna V, Chhabra P, Patil P, Verma H, Lahon A, et al. Diversity in the enteric viruses detected in outbreaks of gastroenteritis from Mumbai, Western India. *Int J Environ Res Public Health*. 2012;9(3):895–915. doi: 10.3390/ijerph9030895. [PubMed: 22690171]. [PubMed Central: PMC3367285].
- 27. Bubba L, Pellegrinelli L, Pariani E, Primache V, Amendola A, Binda S. A novel multiplex one-step real-time RT-PCR assay for the simultaneous identification of enterovirus and parechovirus in clinical fecal samples. *J Prev Med Hyg.* 2015;**56**(2):E57–60. [PubMed: 26789989]. [PubMed Central: PMC4718355].
- Guo WP, Lin XD, Chen YP, Liu Q, Wang W, Wang CQ, et al. Fourteen types of co-circulating recombinant enterovirus were associated with hand, foot, and mouth disease in children from Wenzhou, China. J Clin Virol. 2015;70:29–38. doi: 10.1016/j.jcv.2015.06.093. [PubMed: 26305816].
- 29. Nguyen AT, Tran TT, Hoang VM, Nghiem NM, Le NN, Le TT, et al. Development and evaluation of a non-ribosomal random PCR and

next-generation sequencing based assay for detection and sequencing of hand, foot and mouth disease pathogens. *Virol J.* 2016;**13**:125. doi: 10.1186/s12985-016-0580-9. [PubMed: 27388326]. [PubMed Central: PMC4937578].

- Gaunt E, Harvala H, Osterback R, Sreenu VB, Thomson E, Waris M, et al. Genetic characterization of human coxsackievirus A6 variants associated with atypical hand, foot and mouth disease: A potential role of recombination in emergence and pathogenicity. *J Gen Virol.* 2015;**96**(Pt 5):1067-79. doi: 10.1099/vir.0.000062. [PubMed: 25614593]. [PubMed Central: PMC4631059].
- Paananen A, Ylipaasto P, Smura T, Lempinen M, Galama J, Roivainen M. A single amino acid substitution in viral VP1 protein alters the lytic potential of clone-derived variants of echovirus 9 DM strain in human pancreatic islets. *J Med Virol*. 2013;**85**(7):1267–73. doi: 10.1002/jmv.23574. [PubMed: 23595636].
- Lee ST, Ki CS, Lee NY. Molecular characterization of enteroviruses isolated from patients with aseptic meningitis in Korea, 2005. Arch Virol. 2007;152(5):963-70. doi: 10.1007/s00705-006-0901-1. [PubMed: 17238012].
- Pham NT, Trinh QD, Chan-It W, Khamrin P, Shimizu H, Okitsu S, et al. A novel RT-multiplex PCR for detection of Aichi virus, human parechovirus, enteroviruses, and human bocavirus among infants and children with acute gastroenteritis. *J Virol Methods*. 2010;**169**(1):193–7. doi: 10.1016/j.jviromet.2010.07.038. [PubMed: 20691209].
- Holtz LR, Finkbeiner SR, Zhao G, Kirkwood CD, Girones R, Pipas JM, et al. Klassevirus 1, a previously undescribed member of the family Picornaviridae, is globally widespread. *Virol J.* 2009;6:86. doi: 10.1186/1743-422X-6-86. [PubMed: 19552824]. [PubMed Central: PMC2706813].