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Research Article

Investigation of Antimicrobial Susceptibilities Among Bacteria Isolated from Blood Cultures in Hospitalized Patients, Tehran, Iran

Shadi Aghamohammad^{1, 2}, Maryam Nouri ¹, Fatemeh Ashrafian^{1, 3}, Mohtaram Sadat Kashi⁴, Mehrdad Gholami ¹, ⁵, Masoud Dadashi^{6, 7} and Hossein Dabiri^{1, *}

¹Department of Medical Microbiology, School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran

²Department of Bacteriology, Pasteur Institute of Iran, Tehran, Iran

³Microbiology Research Center (MRC), Pasteur Institute of Iran, Tehran, Iran

⁴Loghman General Hospital, Faculty of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran

⁵Department of Microbiology and Virology, Faculty of Medicine, Mazandaran University of Medical Sciences, Sari, Iran

⁶Department of Microbiology, School of Medicine, Alborz University of Medical Sciences, Karaj, Iran

⁷Non-communicable Diseases Research Center, Alborz University of Medical Sciences, Karaj, Iran

corresponding author: Department of Medical Microbiology, Faculty of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran. Email: hodabiri@gmail.com

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Abstract

Background: Bacteremia is the status, which is detected via a positive blood culture test with no contamination. Centers for Disease Control and Prevention (CDC) indicates that direct medical procedures and total costs are significantly high. Antibiotic resistance can play a major role in the costs, which are related to the long duration of treatment.

Objectives: The aim of this study was to investigate the rate and profiles of antimicrobial susceptibility of blood culture isolates from Tehran, Iran.

Methods: In the current cross-sectional study, a total of 5,000 blood culture samples were collected from patients hospitalized in the Loghman General Hospital, Tehran, Iran, with positive blood culture results from 2012 to 2013. Susceptibility to antimicrobial agents was analyzed using National Committee for Clinical Laboratory Standards guidelines.

Results: Coagulase-negative staphylococci (38.8%), *Staphylococcus aureus* (20.5%), *Acinetobacter* (11.9%), and *Escherichia coli* (11.7%) were the most frequent bacteria isolated from the blood cultures, collectively accounting for > 80% of the isolates. Of isolated microorganisms, 63.75% and 36.24% belonged to Gram-positive and Gram-negative bacteria, respectively. Moreover, 88% of the isolates were MRSA (oxacillin-/methicillin-resistant), and 7% were VRE (vancomycin-resistant).

Conclusions: The most frequent isolated organisms were Gram-positive bacteria, and the rate of MDR (multi-drug resistance) was high. The results of the current study obviously indicate the misuse of antibiotic in society. National surveillance studies in Iran will be useful for clinicians to choose the right empirical treatment and will help control and prevent infections caused by resistant organisms.

Keywords: Blood Cultures, Antibacterial Agents, Bacteria, Bacteremia

1. Background

Bacteremia is the status, which is detected via a positive blood culture test with no contamination. It can be ignored if the condition is transient without clinical manifestation (1). However, bloodstream infection (BSI) with symptoms like septic shock and multiple organ dysfunction syndromes can be more problematic (2). In fact, bacteremia can cause morbidity and mortality, especially in patients with underlying diseases e.g., patients on hemodialysis (3). Although blood culture is the gold standard method to detect bacteremia, using previous antibiotics can lead to false-negative results. Different classes of antibiotics are often used to treat bacteremia, specifically in ICU; however, studies revealed that more than 30% of antimicrobial agents are unsuitable. Increasing the rate of antimicrobial resistance (AMR) could increase the rate of morbidity and mortality or at least the cost of treatment (4). Centers for Disease Control and Prevention (CDC) indicates that US\$90 million in direct medical procedures and US\$230 in total costs are considered additional expenses. Most of the costs are related to the long duration of treatment due to using inappropriate antibiotics and hospitalization (5). The average hospital stays in ICU and wards for patients with bacteremia are 2 - 7 days and 2 - 3 weeks, respectively. As mentioned, AMR is challenging in treating patients with bacteremia, especially in microorgan-

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isms like Staphylococcus aureus (S. aureus). Antibiotic resistance in S. aureus has become a difficulty in the healthcare system since 1940. Besides, methicillin-resistance S. aureus (MRSA) has been detected since 1960 (6). Besides, MRSA isolates also can transfer antibiotic resistance to other genera (7, 8). Another antibacterial resistance, which has been significant since 1987, is vancomycin-resistant Enterococcus (VRE). Treatment of the bacteremia caused by VRE is complicated because the side effects of some antimicrobial agents such as chloramphenicol are problematic. Moreover, the rate of resistance to antibiotics, including ampicillin and aminoglycoside is high (9). Therefore, controlling the transmission of VRE isolates in hospitals is crucial (6). Different bacteria may be detected in blood culture (10). The bacteremia due to Enterobacteriaceae is associated with increased mortality compared with BSI caused by Gram-positive species (11). Among Gram-negative bacteria, Acinetobacter and Pseudomonas spp. can result in severe nosocomial bacteremia (12). On the other hand, in some regions, including Iran, CoNS (coagulase-negative staphylococci) and S. aureus are the most frequent organisms isolated from blood culture (13). The AMR is a critical consideration for physicians to choose a suitable regimen. This is particularly important for the treatment of systemic infections because initial antimicrobial chemotherapy is almost empiric, and it must be based on knowledge of their antimicrobial susceptibility patterns. Early initiation of appropriate antimicrobial treatment is critical to decreasing morbidity and mortality among patients with BSI (14).

2. Objectives

The aim of this study was to investigate the rate and profiles of antimicrobial susceptibility of blood culture isolates from Tehran, Iran.

3. Methods

3.1. Patients and Sample Collection

In the current cross-sectional study, a total of 5,000 blood culture samples were collected from patients hospitalized in the Loghman General Hospital, Tehran, Iran, with positive blood culture results from 2012 to 2013. Blood samples were collected through cleaning of the venous site with 70% alcohol and povidone-iodine. The blood sample was injected into brain heart infusion and sodium thioglycollate broths in the ratio of one part of a blood sample to five parts of the culture broth (15).

3.2. Bacteria Isolation and Identification

The blood culture broths were sent to the laboratory and incubated at 37°C for seven days. MacConkey, blood, and chocolate agar media were subcultured at 24, 72 hours and on the 7th day. Then they were incubated at the appropriate temperature and atmospheres according to standard procedures (16). Isolated organisms were identified by conventional biochemical methods.

3.3. Antibiotic Susceptibility Test

The susceptibilities of blood samples against 15 antibiotics: trimethoprim-sulfamethoxazole (SXT: 1.25/23.75 μ g), cefotaxime (CTX: 30 μ g), cefoperazone (CP: 75 μ g), ceftazidime (CAZ: 30 μ g), imipenem (IMP: 10 μ g), ampicillin (AMP: 10 μ g), gentamicin (GM: 30 μ g), chloramphenicol (C: 30 μ g), penicillin (P: 10 μ g), oxacillin (OX: 1 μ g), clindamycin (CC: 2 μ g), erythromycin (E: 15 μ g), vancomycin (V: 30 μ g), ofloxacin (OF: 5 μ g), and amikacin (AK: 30 μ g) (Mast Group Ltd., UK) were tested by agar disk-diffusion method according to clinical and laboratory standards institute (CLSI) guidelines (17). *E. coli* ATCC 25922 was used as a control for the disk diffusion method.

3.4. Statistical Analysis

Statistical analyses were performed using SPSS software (version 25; SPSS, Inc., Chicago, IL, USA). The Fisher's exact test or the chi-square test was applied to analyze categorical data. A P-values < 0.05 in all experiments were considered statistically significant.

4. Results

4.1. Results of Sample Collection

Of 5000 patients, 469 (9.3%) individuals had positive culture results. Also, 338 (72%) were male, and 131 (28%) were female. Of these 469 positive blood cultures, overall 469 isolates, 14 different species were detected. No mixed infection was observed. The most prevalent strains were CoNS (38.8%), *S. aureus* (20.5%), *Acinetobacter* (11.9%), and *E. coli* (11.7%). *Pseudomonas, Enterobacter, Citrobacter, Proteus, Klebsiella, Salmonella*, and *Aeromonas* spp. isolated in approximately 12% of cultures. Enterococcus and Streptococcus were found in 3% of positive cultures (Table 1). These results showed the predominance of bacteria, of which 63.75% and 36.24% were Gram-positive and Gram-negative bacteria, respectively.

Bacterial Species	No. of Isolates	% of Isolates
CoNS	182	38.8
Staphylococcus aureus	96	20.5
Acinetobacter spp.	56	11.9
E. coli	55	11.7
Klebsiella spp.	22	4.7
Pseudomonas spp.	17	3.6
Enterobacter spp.	13	2.8
Streptococcus spp.	11	2.3
Streptococcus pneumoniae	6	1.3
Enterococcus	4	0.9
Citrobacter spp.	2	0.4
Proteus spp.	2	0.4
Salmonella spp.	2	0.4
Aeromonas spp.	1	0.2
Total	469	100

Table 1. Comparison of Distribution of Isolated Bacteria

4.2. Results of AMR Among Gram-Positive Bacteria

Sixty-two percent and 69% of CoNS were susceptible to gentamycin and fluoroquinolones, respectively, while 69% were resistant to cephalosporin. In addition, 88% of *S. aureus* isolates were MRSA, and 7% were VRE. Among *Streptococcus* spp., rather than enterococci and pneumococci, cefoperazone was active against 81% of isolates. *Streptococcuspneumonia* was highly susceptible to penicillin (75%) and resistant to trimethoprim-sulfamethoxazole (84%) (Table 2).

4.3. Results of AMR Among Gram-Negative Bacteria

As mentioned above, *Acinetobacter* spp. was the third common isolate. Moreover, 72 and 82% of isolates were resistant to imipenem, and amikacin, respectively, while 43% of *Acinetobacter* isolates were susceptible to cefoperazone. Further, 58 and 43% of *E. coli* were susceptible to amikacin, and gentamycin, respectively, and the rate of resistance to ceftazidime was 60%. The isolates of *Klebsiella* spp. were extremely resistant to more than three classes of antimicrobial agents, including cefotaxime (55%) (Table 3).

5. Discussion

The rate of bacteremia has been increased over the past years (1). Antimicrobial resistance (AMR) among bacteria isolated from blood culture is a worrying issue since it can influence the rate of mortality and morbidity (18). In Iran, AMR is a common cause of treatment failure of BSI. Since

the determination of appropriate antibiotics is often not done in the best time, broad-spectrum antibiotics are used unnecessarily (19). Based on our results, Gram-positive organisms were the main pathogens (63.85%). Increasing rate of Gram-positive bacteria among blood cultures is seen in other regions (20). Bacteremia, which is caused by Gram-positive bacteria, is important because some microbial agents, including S. aureus, are often nosocomial and can make the situation of the patient more unfavorable. Besides, since some antimicrobial agents such as vancomycin has low tissue penetration, bacteremia caused by S. aureus often relapses (21). Increasing use of intravascular devices, as well as prolonged hospitalization, can result in raising the rate of the microorganisms, including CoNS and S. aureus (22). During the 1970s, BSI was most commonly associated with Gram-negative organisms, but recently, Gram-positive organisms began to emerge (23). This indicates that the organisms causing bacteremia are shifting toward Gram-positive in some regions (24). On the other hand, some studies reveal that Gram-negative bacteria remain predominant (25, 26). The critical issue is that Gram-negative bacteria can become problematic among patients with underlying diseases. Besides, previous use of antimicrobial agents in patients with underlying diseases can result in AMR among Gram-negative bacteria (27).

Besides the rates of MRSA, one of the most remarkable results in our study is high resistance to third-generation cephalosporins (3rd-GCs) among Gram-positive bacteria. Resistance to fluoroquinolone was almost variable among Gram-positive bacteria, which were 100% among S. aureus isolates and 0% among S. pneumonia. Furthermore, 3rd-GCs along with fluoroquinolones are generally used to treat BSI. As Lee et al. declared fluoroquinolones were more effective than 3rd-GCs for shortening the time of treatment of BSI (28); therefore, the high rate of susceptibility among S. pneumonia isolates can be considerable. Multidrug resistance (MDR) is another issue that is important. All Gram-positive isolates were MDR, and specifically, the emergence of MDR among MRSA isolates can make the treatment more complicated and affect infection control in healthcare settings (29).

In addition, AMR was previously reported among Gram-negative bacteria (30, 31), which can be discussed from several points of view. Susceptibility to aminogly-coside was almost high among some Gram-negative iso-lates, including *Salmonella*, *Aeromonas*, *Proteus*, and *Enterobacter*. The effectiveness of aminoglycoside to treat the carbapenem-resistant isolates was studied in Shields et al. investigation (32). Some of the isolates showed high resistance to imipenem, including *Enterobacter*, which was susceptible to amikacin and gentamicin; therefore, it can give more chance to treat the patients more successfully. The

able 2. Frequency/Percentage of Gram-Positive Isolates Susceptible to Antimicrobial Agents																
Microorganism	NO.	v	IPM	СС	GM	С	E	СР	SXT	AK	AM	CAZ	СТХ	ох	Р	OF
CoNS	182	89	46	43	62	36	2.5	69	32	1	9	0	31	2	1	1
S. aureus	96	86	63	60	62	39	49	69	62	2	4	1	50	12	0	0
Streptococcus	6	91	16	45	18	27	54	81	36	0	63	0	54	0	45	36
S. pneumoniae	22	83	66	33	0	66	16	83	16	16	50	0	66	0	75	100
Enterococcus	17	93	25	0	25	50	25	25	0	0	25	0	50	0	25	25

Abbreviations: SXT, trimethoprim-sulfamethoxazole; CTX, cefotaxime; CP, cefoperazone; CAZ, ceftazidime; IPM, imipenem; AK, amikacin; AM, ampicillin; GM, gentamicin; C, chloramphenicol; CC, clindamycin; E, erythromycin; V, vancomycin; P, penicillin; OX, oxacillin; OF, ofloxacin; CONS, coagulase-negative *Staphylococcus aureus*.

Table 3. Frequency/Percentage of Gram-Negative Isolates Susceptible to Antimicrobial Agents																
Microorganism	NO.	v	IPM	СС	GM	С	E	СР	SXT	AK	АМ	CAZ	СТХ	ox	Р	OF
Acinetobacter	56	0	28	0	23	0	0	43	28	18	0	17	20	0	0	0
E. coli	55	0	62	0	43	0	0	54	32	58	1	40	29	0	0	0
Klebsiella	4	0	68	0	45	0	0	64	45	81	4	63	45	0	0	0
Pseudomonas	2	0	59	0	29	0	0	65	29	35	5	29	29	0	0	0
Enterobacter	11	0	0	0	84	7	0	69	77	69	0	54	69	0	0	0
Citrobacter	2	0	100	0	50	0	0	50	100	50	0	50	100	0	0	0
Proteus	13	0	50	0	100	0	0	50	50	50	0	100	50	0	0	0
Salmonella	2	0	100	0	100	50	0	100	50	100	0	100	50	0	0	0
Aeromonas	1	0	100	0	100	0	0	100	100	100	0	0	50	0	0	0

Abbreviations: SXT, trimethoprim-sulfamethoxazole; CTX, cefotaxime; CP, cefoperazone; CAZ, ceftazidime; IPM, imipenem; AK, amikacin; AM, ampicillin; GM, gentamicin; C, chloramphenicol; CC, clindamycin; E, erythromycin; V, vancomycin; P, penicillin; OX, oxacillin; OF, ofloxacin.

rates of MDR isolates in Gram-negative bacteria are also impressive. Our Gram-negative isolates were totally resistant to ofloxacin, oxacillin, and clindamycin. The emergence of fluoroquinolone resistance along with beta-lactam drastically reduces the choice of treatment. Another important point is that previous use of antibiotics, including fluoroquinolone can influence increasing the rate of resistance in the future (33). Since the use of empirical antibiotics is a common way, previous use of antibiotics and subsequently the emergence of MDR can be threatening and challenging. The limitation of this study is the lack of molecular evaluation of antibiotics resistance-related genes, and it is suggested that minimal inhibitory concentration (MIC) and molecular tests should be performed for better assessment.

In conclusion, the results of current studies obviously indicate misuse of antibiotics in our society. National surveillance studies in Iran will be useful for clinicians to choose the right empirical treatment and will help control and prevent infections caused by resistant organisms.

Footnotes

Authors' Contribution: SA, lab working and writing the manuscript; MN, MSK, MG, and MD, lab working; FA, lab working and revision of the manuscript; HD, designed the study and reviewed/edited the manuscript.

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References

- Nielsen SL, Lassen AT, Gradel KO, Jensen TG, Kolmos HJ, Hallas J, et al. Bacteremia is associated with excess long-term mortality: A 12year population-based cohort study. J Infect. 2015;70(2):111–26. doi: 10.1016/j.jinf.2014.08.012. [PubMed: 25218427].
- 2. Smith DA, Nehring SM. Bacteremia. *StatPearls*. Treasure Island (FL): StatPearls Publishing; 2021.
- Fram D, Okuno MF, Taminato M, Ponzio V, Manfredi SR, Grothe C, et al. Risk factors for bloodstream infection in patients at a Brazilian hemodialysis center: A case-control study. *BMC Infect Dis.* 2015;**15**:158. doi: 10.1186/s12879-015-0907-y. [PubMed: 25879516]. [PubMed Central: PMC4377039].

- Daneman N, Rishu AH, Xiong W, Bagshaw SM, Cook DJ, Dodek P, et al. Bacteremia antibiotic length actually needed for clinical effectiveness (BALANCE): Study protocol for a pilot randomized controlled trial. *Trials*. 2015;**16**:173. doi: 10.1186/s13063-015-0688-z. [PubMed: 25903783]. [PubMed Central: PMC4407544].
- Pai S, Enoch DA, Aliyu SH. Bacteremia in children: Epidemiology, clinical diagnosis and antibiotic treatment. *Expert Rev Anti Infect Ther*. 2015;**13**(9):1073-88. doi: 10.1586/14787210.2015.1063418. [PubMed: 26143645].
- Friedman ND, Temkin E, Carmeli Y. The negative impact of antibiotic resistance. *Clin Microbiol Infect*. 2016;22(5):416–22. doi: 10.1016/j.cmi.2015.12.002. [PubMed: 26706614].
- Armin S, Fallah F, Navidinia M, Vosoghian S. Prevalence of blaOXA-1 and blaDHA-1 AmpC β-lactamase-producing and methicillinresistant Staphylococcus aureus in Iran. Arch Pediatr Infect Dis. 2017;5(4). e36778. doi: 10.5812/pedinfect.36778.
- Navidinia M, Fallah F, Lajevardi B, Shirdoost M, Jamali J. Epidemiology of methicillin-resistant Staphylococcus aureus isolated from health care providers in Mofid Children Hospital. Arch Pediatr Infect Dis. 2015;3(2). e16458. doi: 10.5812/pedinfect.16458.
- Patel R, Gallagher JC. Vancomycin-resistant enterococcal bacteremia pharmacotherapy. Ann Pharmacother. 2015;49(1):69–85. doi: 10.1177/1060028014556879. [PubMed: 25352037].
- Gandra S, Mojica N, Klein EY, Ashok A, Nerurkar V, Kumari M, et al. Trends in antibiotic resistance among major bacterial pathogens isolated from blood cultures tested at a large private laboratory network in India, 2008-2014. *Int J Infect Dis.* 2016;**50**:75–82. doi: 10.1016/J.ijid.2016.08.002. [PubMed: 27522002]. [PubMed Central: PMC5063511].
- Ma J, Li N, Liu Y, Wang C, Liu X, Chen S, et al. Antimicrobial resistance patterns, clinical features, and risk factors for septic shock and death of nosocomial E coli bacteremia in adult patients with hematological disease: A monocenter retrospective study in China. *Medicine*. 2017;96(21). e6959. doi: 10.1097/MD.0000000000006959. [PubMed: 28538389]. [PubMed Central: PMC5457869].
- Papadimitriou-Olivgeris M, Fligou F, Spiliopoulou A, Koutsileou K, Kolonitsiou F, Spyropoulou A, et al. Risk factors and predictors of carbapenem-resistant Pseudomonas aeruginosa and Acinetobacter baumannii mortality in critically ill bacteraemic patients over a 6-year period (2010-15): Antibiotics do matter. J Med Microbiol. 2017;66(8):1092-101. doi: 10.1099/jmm.0.000538. [PubMed: 28758623].
- Gitau W, Masika M, Musyoki M, Museve B, Mutwiri T. Antimicrobial susceptibility pattern of Staphylococcus aureus isolates from clinical specimens at Kenyatta National Hospital. *BMC Res Notes*. 2018;**11**(1):226. doi: 10.1186/s13104-018-3337-2. [PubMed: 29615129]. [PubMed Central: PMC5883409].
- Beganovic M, McCreary EK, Mahoney MV, Dionne B, Green DA, Timbrook TT. Interplay between rapid diagnostic tests and antimicrobial stewardship programs among patients with bloodstream and other severe infections. J Appl Lab Med. 2019;3(4):601-16. doi: 10.1373/jalm.2018.026450. [PubMed: 31639729].
- Snyder SR, Favoretto AM, Baetz RA, Derzon JH, Madison BM, Mass D, et al. Effectiveness of practices to reduce blood culture contamination: A Laboratory Medicine Best Practices systematic review and meta-analysis. *Clin Biochem*. 2012;45(13-14):999–1011. doi: 10.1016/j.clinbiochem.2012.06.007. [PubMed: 22709932]. [PubMed Central: PMC4518453].
- Garcia RA, Spitzer ED, Beaudry J, Beck C, Diblasi R, Gilleeny-Blabac M, et al. Multidisciplinary team review of best practices for collection and handling of blood cultures to determine effective interventions for increasing the yield of true-positive bacteremias, reducing contamination, and eliminating false-positive central line-associated bloodstream infections. *Am J Infect Control.* 2015;43(11):1222–37. doi: 10.1016/j.ajic.2015.06.030. [PubMed: 26298636].
- 17. CLSI. Performance standards for antimicrobial susceptibility testing. 27th ed. Pennsylvania, USA: Clinical and Laboratory Standards Institute;

2017.

- De Angelis G, Fiori B, Menchinelli G, D'Inzeo T, Liotti FM, Morandotti GA, et al. Incidence and antimicrobial resistance trends in bloodstream infections caused by ESKAPE and Escherichia coli at a large teaching hospital in Rome, a 9-year analysis (2007-2015). *Eur J Clin Microbiol Infect Dis.* 2018;**37**(9):1627–36. doi: 10.1007/s10096-018-3292-9. [PubMed: 29948360].
- Salehi Nobandegani A, Motamedifar M. Antibiotic sensitivity profile of the bacterial isolates from the blood samples of the patients in different wards of a major referral hospital, Shiraz, Iran 2015-2016. *Pharmacophores*. 2019;**10**(2):30–6.
- Otto M. Staphylococcus epidermidis: A major player in bacterial sepsis? *Future Microbiol.* 2017;**12**:1031–3. doi: 10.2217/fmb-2017-0143. [PubMed: 28748707]. [PubMed Central: PMC5627029].
- Corey GR, Rubinstein E, Stryjewski ME, Bassetti M, Barriere SL. Potential role for telavancin in bacteremic infections due to Grampositive pathogens: Focus on Staphylococcus aureus. *Clin Infect Dis.* 2015;**60**(5):787–96. doi: 10.1093/cid/ciu971. [PubMed: 25472944]. [PubMed Central: PMC4329924].
- Asaad AM, Ansar Qureshi M, Mujeeb Hasan S. Clinical significance of coagulase-negative staphylococci isolates from nosocomial bloodstream infections. *Infect Dis.* 2016;48(5):356–60. doi: 10.3109/23744235.2015.1122833. [PubMed: 26666168].
- Aamodt H, Mohn SC, Maselle S, Manji KP, Willems R, Jureen R, et al. Genetic relatedness and risk factor analysis of ampicillin-resistant and high-level gentamicin-resistant enterococci causing bloodstream infections in Tanzanian children. *BMC Infect Dis.* 2015;15:107. doi: 10.1186/s12879-015-0845-8. [PubMed: 25884316]. [PubMed Central: PMC4350950].
- Tang XJ, Sun B, Ding X, Li H, Feng X. Changing trends in the bacteriological profiles and antibiotic susceptibility in neonatal sepsis at a tertiary children's hospital of China. *Transl Pediatr.* 2020;9(6):734-42. doi: 10.21037/tp-20-115. [PubMed: 33457294]. [PubMed Central: PMC7804488].
- Pal N. Microbiological profile and antimicrobial resistant pattern of blood culture isolates, among septicaemia suspected patients. National Journal of Laboratory Medicine. 2016;5(1):17–21. doi: 10.7860/njlm/2016/17494.2090.
- Le Doare K, Bielicki J, Heath PT, Sharland M. Systematic review of antibiotic resistance rates among Gram-negative bacteria in children with sepsis in resource-limited countries. J Pediatric Infect Dis Soc. 2015;4(1):11–20. doi: 10.1093/jpids/piu014. [PubMed: 26407352].
- Moghnieh R, Estaitieh N, Mugharbil A, Jisr T, Abdallah DI, Ziade F, et al. Third generation cephalosporin resistant Enterobacteriaceae and multidrug resistant Gram-negative bacteria causing bacteremia in febrile neutropenia adult cancer patients in Lebanon, broad spectrum antibiotics use as a major risk factor, and correlation with poor prognosis. *Front Cell Infect Microbiol.* 2015;5:11. doi: 10.3389/fcimb.2015.00011. [PubMed: 25729741]. [PubMed Central: PMC4325930].
- Lee CC, Wang JL, Lee CH, Hsieh CC, Hung YP, Hong MY, et al. Clinical benefit of appropriate empirical fluoroquinolone therapy for adults with community-onset bacteremia in comparison with third-generation-cephalosporin therapy. *Antimicrob Agents Chemother*. 2017;61(2). doi: 10.1128/AAC.02174-16. [PubMed: 27855072]. [PubMed Central: PMC5278695].
- Goudarzi M, Seyedjavadi SS, Nasiri MJ, Goudarzi H, Sajadi Nia R, Dabiri H. Molecular characteristics of methicillin-resistant Staphylococcus aureus (MRSA) strains isolated from patients with bacteremia based on MLST, SCCmec, spa, and agr locus types analysis. *Microb Pathog.* 2017;104:328–35. doi: 10.1016/j.micpath.2017.01.055. [PubMed: 28159661].
- 30. Ashrafian F, Askari E, Kalamatizade E, Ghabouli-Shahroodi MJ, Naderi-Nasab M. The frequency of extended spectrum beta lactamase (ESBL) in Escherichia coli and Klebsiella pneumoniae: A report from Mash-

had, Iran. J Med Bacteriol. 2013;2(1-2):12-9.

- Ashrafian F, Fallah F, Hashemi A, Erfanimanesh S, Amraei S, Tarashi S. First detection of 16S rRNA methylase and blaCTX-M-15 genes among Klebsiella pneumoniae strains isolated from hospitalized patients in Iran. Res Mol Med. 2015;3(4):28–34. doi: 10.7508/rmm.2015.04.005.
- Shields RK, Clancy CJ, Press EG, Nguyen MH. Aminoglycosides for treatment of bacteremia due to carbapenem-resistant Klebsiella pneumoniae. Antimicrob Agents Chemother. 2016;60(5):3187–92.

doi: 10.1128/AAC.02638-15. [PubMed: 26926642]. [PubMed Central: PMC4862490].

 Dan S, Shah A, Justo JA, Bookstaver PB, Kohn J, Albrecht H, et al. Prediction of fluoroquinolone resistance in Gram-negative bacteria causing bloodstream infections. *Antimicrob Agents Chemother*. 2016;60(4):2265-72. doi: 10.1128/AAC.02728-15. [PubMed: 26833166]. [PubMed Central: PMC4808151].