Published online 2020 July 25.

Research Article

Detection of R282w P53 Gene Mutation on Exon 8 in Gastric Cancer Patients in Southwest Iran

Sajad Afrouz¹, Mohammad Amin Ghatee^{2,*}, Amroallah Roozbehi² and Mohammad Hossein Sangtarash¹

¹Department of Biological Sciences, University of Sistan and Baluchistan, Zahedan, Iran ²Cellular and Molecular Research Center, Yasuj University of Medical Sciences, Yasuj, Iran

Corresponding author: Ph.D., Cellular and Molecular Research Center, Yasuj University of Medical Sciences, Yasuj, Iran. Email: ghateea1980@gmail.com

Received 2020 April 04; Revised 2020 June 15; Accepted 2020 June 27.

Abstract

Background: Gastric adenocarcinoma is the most common type of gastric cancer all around the world. The epithelial cells of stomach tissue are influenced by environmental factors and genetic disorders. P53 is the most remarkable gene that controls the growth of cells. Mutation in some nucleotides of the P53 gene increases the genetic instability and is assumed as an important prognostic factor in gastric cancer. More than 90% of mutations occur in the exons 5-8 of p53.

Objectives: This study was conducted in Kohgiluyeh and Boyer Ahmad (K&B) province (Southwest Iran) to determine the rate of R282W P53 gene mutation of exon 8 in gastric cancer.

Methods: This case-control study was conducted on 90 subjects that were divided into two groups (each including 45 patients and 45 controls). The samples were randomly collected from the tissue bank of the pathology laboratory in Yasuj city and then were transferred to the Cellular and Molecular Research Center. DNA extraction was performed by the DNA extraction kit. Molecular analysis on exon 8 was performed by the PCR-RFLP method and using the *Mspl* restricting enzyme. Data were analyzed by descriptive statistics and bivariate correlation tests.

Results: No difference was found between the two groups concerning age, gender, and education level. The prevalence of R282W P53 gene mutation on exon 8 in the cancer group was 17.8% (8/45), while no mutation was found in the control group.

Conclusions: According to the results, the R282W P53 gene mutation on exon 8 may play an important role in the development of gastric cancer in Yasuj district, Southwest Iran.

Keywords: P53, PCR, RFLP, Exon 8, R282W

1. Background

Gastric cancer (GC) is the second leading cause of cancer-related mortality (1). Its incidence shows a wide geographical variation. About half of the total gastric cancer load occurs in East Asia, particularly in China and Japan (2). The low-risk areas include Southern Asia, and North and East Africa (3). Although the incidence of GC is gradually decreasing in many parts of the world, it is still the most common malignancy in Iran (4). There are several intermediate and low-risk populations in geographical areas, while the northern and northwestern regions are highrisk areas for gastric cancer (4). The mean incidence rate of stomach cancer is 10.5 - 12 (5). Based on the previous studies, the incidence of stomach cancer is rising in K&B province, including in Yasuj district (6).

Regarding the marked variation of gastric cancer risk in different geographical areas and striking differences in frequency of possible environmental and ethnic risk factors, research on the gastric cancer etiology in each population should be considered as a priority (3).

GC is a multifactorial disease that develops due to continuous cell damage caused by life-long exposure to different predisposing factors, including carcinogens (7-9). Epigenetic alteration involving tumor suppressor genes mutations, DNA repair genes, and loss of heterozygosis (LOH) A may cause cancerous cell mutation (10, 11). Recent studies have revealed that p53 mutations are biologically and clinically distinct. Genetic alterations in the TP53 gene are fundamental events in both early-stage and advanced stomach tumors (12, 13). Approximately, over 50% of human cancers carry a loss of function mutations in the p53 gene (14) in which, 95% of TP53 mutations occur within the genomic region encoding the sequence-specific DNAbinding domain of TP53 protein (exons 4-9) (15-17). A mis-

Copyright © 2020, Gene, Cell and Tissue. This is an open-access article distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 International License (http://creativecommons.org/licenses/by-nc/4.0/) which permits copy and redistribute the material just in noncommercial usages, provided the original work is properly cited.

sense mutation results in a single amino acid change, and this type of point mutation in the DNA-binding domain of p53 can encode a protein that is transcriptionally inactive or that displays altered transcriptional activity compared to the wild-type p53 (18). The majority of p53 mutations in human cancer are missense mutations, sitting within the DNA-binding domain with hot spots at codons G245, R273, and R282 (19). Although hotspot mutants of p53 are frequently investigated, few studies are conducted on R282W. Codon 282 encodes arginine amino acid on the P53 binding site in the central domain during gene expression and makes the minor groove contact (20-22). Mutation in the aforementioned site leads to the exclusion of arginine and breaking the conjunction of central domain with DNA major groove and following a decrease in P53 gene expression (R282W designates an arginine mutated to tryptophan at position 282) (18, 23). R282W P53 mutant can alter the behavior and fate of the tumor cell and is thought to promote the progression of many types of cancer (18). According to a study conducted by Zhang and colleagues, R282W mutation can lead to loss of some wild-type P53 tumorsuppressive activity (24).

Although the incidence of GC is gradually decreasing in many regions of Iran, but it is still the most common malignancy in K&B province (6). Based on the official statistics of the Ministry of Health (2015) the number of patients with GC registered in the K&B province has been increasing (25). However, no study is performed at the molecular level to identify the mutation rate of effective genes in GC, such as the R282W P53 mutation in the patients and healthy populations in K&B province.

2. Objectives

Thus, in the current study, samples were subjected to PCR-RFLP to investigate the R282W P53 gene mutation on exon 8 in GC patients in this province of Southwest Iran.

3. Methods

3.1. Study Area

K&B Province (in the Southwest of Iran) is located at an average altitude of 1200 meters high above the sea level in the close proximity of the Dena Mountain. The weather varies from semi-arid and warm in western areas to cold and humid in the north. Its population is estimated to be 750,000, in which 42 and 58% of individuals settle in urban and rural/nomadic areas, respectively.

2

3.2. Samples Data

This case-control study was conducted on 90 subjects that were divided into two groups (i.e., 45 patients and 45 controls). The samples were randomly collected from the tissue bank of the pathology laboratory of Beheshti hospital in Yasuj city. Demographic data (including age, gender, and etc.) were collected from the medical records of the hospital. The cancerous tissue (based on ICD-10 Coding System: code C16, stages III and IV of the disease) and control samples (no code c16) were transferred to the Cellular and Molecular Research Center and were subjected to DNA extraction.

3.3. DNA Extraction

DNA extraction from Formaldehyde-Fixed, Paraffin-Embedded (FFPE) biopsy samples was carried out using QI-Aamp DNA FFPE Tissue Kit (Qiagen, cat: 56404) according to manufacturer's instruction.

3.4. PCR Reaction

The PCR was carried out on the extracted DNA samples. The reaction mixture was provided by adding 12.5 μ L Taq 2X Master Mix (Ampliqon), 20 pmol of each primer, 2 μ L of DNA template, and deionized molecular grade water up to 25 μ L. Primers sequences were forward, 5-TGGTAATCTACTGGGACGGA-3 (Tm:58.4); reverse, 5-CTGCTTGCTTACCTCGCTTA-3 (Tm: 58.4)(26). The thermal cycle program consisted of a preincubation step of 5min at 95°C for complete denaturation of DNA followed by 35 cycles at 94°C for 30s, 55°C for 30s and 72°C for 30 s and final elongation at 72°C for 5 min. The DNA product of exon8 was 149 bp.

3.5. RFLP Reaction

After PCR, 16μ L of the PCR product was subjected to enzymatic digestion with 2μ L of the endonuclease *Mspl* (Fermentas) and 2μ L of enzyme-specific buffer overnight at the recommended temperature (37°C). *Mspl* was used to recognize the sequence CC/GG and cut it at the same site even if the internal cytosine was methylated.

MspI cleaves CC/GG at codon 282, generating 87bp and 62bp from the 149 bp the purified DNA product. Mutation at codon 282 resulted in an uncleaved (c.844C>T (CGG \rightarrow TGG); Arg282Trp), 149 bp fragment, and this feature had been distinguished from that of normal samples on 10% polyacrylamide gel. The presence of the uncleaved 149 bp fragment indicated that there are mutations in the corresponding samples.

3.6. Statistical Analysis

Data were analyzed by descriptive statistics, including frequency and percentage. Analytical evaluations were performed using the chi-square test. All data were analyzed using the SPSS software version 17. Statistically significant was defined at P-value < 0.05.

4. Results

The demographic and clinical characteristics of the patient and normal groups are described in Table 1. There were 27 (60.0%) males and 22 (48.9%) females with a mean age of 52.07 \pm 15.4 years for GC patients. For the control group, 18 (40.0%) were males and 23 (51.1%) females, with a mean age of 47.13 \pm 15.12 years. The results showed that 26/45 (57.8%) of the patient group and 24/45 (53.3%) of the control group were illiterate. According to the results, there was no significant difference concerning the education levels, age, and gender between the two groups (Table 1).

Fable 1. The Population Analysis Among Case and Normal Groups				
Variables	Case, No. (%)	Control, No. (%)	P-Value	
Gender			0.397	
Male	27(60.0)	22 (48.9)		
Female	18 (40.0)	23 (51.1)		
Age, y			0.399	
< 40	12 (26.7)	18 (40.0)		
40 - 60	19 (42.2)	18 (40.0)		
61 - 80	13 (28.9)	9 (20.0)		
> 80	1(2.2)	0 (0.0)		
Education			0.906	
Illiterate	26 (57.8)	24 (53.3)		
Literally	19 (42.3)	21(46.7)		

The frequency of R282W P53 mutation in the control and tumor samples is shown in Table 2. The prevalence of R282W P53 on exon 8 in the cancer group was 17.8% (8/45), while no mutation was found in the control group (0/45) (P = 0.006).

Table 2. Prevalence of R282W Mutation on Exon 8 of P53 in the Normal and Tumo Samples				
Group	Mutation, No. (%)	No Mutation, No. (%)	P-Value	
Case	8 (17.8)	37 (82.2)	0.006	
Control	0 (0.0)	45 (100.0)		
Total	8 (8.9)	82 (91.1)		

Detection of the mutation of R282W P53 by the *Mspl* restriction enzyme is shown in Figure 1. Besides, we arranged positive (with 282 mutations) and negative (wild-type) controls in Figure 1.



Figure 1. Detection of R282WP53 gene mutation by *Mspl* restriction enzyme on 10% polyacrylamide gel. M; Marker 50 bp, Patient; P, h; health, nc; negative control. Normal samples (87 and 62 bp): 10P, 25P, 18P, 17P, 4P and 20h, 18h, 13h, 5h. Mutant samples with R282W mutation (149 bp): 8P, 9P, 28P 44P and 33p.

5. Discussion

P53 gene mutations are one of the most frequent alterations in human cancers (27). Mutation at codon 282 of the P53 gene is identified as a hotspot mutation for GC (28). The R282W mutant is associated with earlier onset of the familial cancers and poorer outcomes of cancer patients (24, 29).

According to the results, there was a significant association between GC and R282W mutation in the studied population in the Southwest of Iran. Abdullah and colleagues showed that 20% of gastric patients in Kashmir Valley harbor R282W mutation of P53 gene (30), which is in accordance with the present study. Fischer and colleagues found a strong association between R282W P53 mutation and GC in Toronto, where P53 mutation was identified in 40% of cases (31). According to the AACR (American association cancer research) project, P53 was altered in 42.67% of GC patients with P53 Codon 282 Missense present in 1.78% of all GC patients (32). Also, the results of Juvan and colleagues found R282W mutation in 23 nucleotide changes at the P53 gene on exon8 in Slovenian patients with GC (33). This study in accordance with the previous studies, which reported a significant association between the loss of heterozygosity (LOH) at R282W loci and GC. It is well known that certain types of mutations (LOH) do not produce stable proteins, and protein overexpression may be due to the results of the stress environment in GC (28).

Exon8 is an evolutionarily conserved region of P53, and the previous studies showed that 80% - 90% of all P53 mutations in a variety of human malignancies occur here (34). The higher frequency of R282W P53 mutation reported in GC is also reported for some other malignancies. Rashid and colleagues found a significantly higher frequency of R282W P53 mutation (58%) between Chronic myeloid leukemia patients in the Indian population (23). Javid and colleagues found that 62% of patients with Nonsmall cell lung cancer were positive for R282W P53 mutation in India (35). Besides, point mutations at codon 282 are mutational hotspots reported in hematologic diseases, including Burkitt's lymphoma (36), Myelodysplastic syndrome (37), T-cell leukemia (38), Lymphoid leukemia (39) and etc.

Analyzing mutation of the P53 in our neighbors and other regions of Iran and other prevalent cancers could be done to compile an inclusive databank on the mutation of P53 and its association with poor prognoses of the disease (40). Karim and colleagues found a significant correlation at exon8 (21.04%) with the P53 alterations in adenocarcinoma of GC (41). The Saffari-Chaleshtori and colleagues study found no mutation in exon8 of the P53 gene in gastric patients Shahrekord city (42). Abbasi and colleagues reported a mutation rate of 6.7% in p53 in the bladder of patients with cancer in Kermanshah city (western are of Iran) (43). Lohrasbi Nejad and colleagues found 4 mutations at the P53 gene (codons: 140, 142, 184 and 248) in colorectal cancer patients in Kerman province (40). However, these studies mentioned to many differences in the incidence of gastrointestinal cancers in various cities of the country (44), but did not provide clear information about R282W P53 mutations. The current study is the first study that specifically examined the R282W P53 gene mutation on exon 8 in GC patients, in K&B Province. The differences in the prevalence of P53R282W gene mutation in various reports may reflect the multitude of factors, including environmental factors, geographic region, race and ethnicity, detection methods, sample size, stage of cancer, etc. (27, 40, 45, 46).

Many studies investigated P53 gene mutations in neighboring provinces, however, no systematic study is performed to distinguish mutants with R282W alterations in GC. The present data are the first report on the R282W P53 abnormalities in GC patients from K&B province. We suppose that a high prevalence of p53 mutation might be related to genomic profiles, environmental factors, or lifestyle-related factors such as H-pylori infection, diet, sunlight, etc. in K&B province, Southwest Iran with nomadic populations and mountainous weather (47-49). Recently, some studies reported that high altitude and increased exposure to ultraviolet (UV) radiations of sunlight, (level of ultraviolet radiation increase by about 10% with every 300 m), may be related to the higher incidence rate of cancers (50, 51). UV radiation has been shown to induce the expression of DNA damage and is known to produce signature mutations in the p53 gene in human (52, 53). Also, there are more than 138 medicinal plants, as the most important sources of herbal food that have been used ethnomedicaly by local people in K&B province for medical and food purposes (54, 55), but anti-cancer or cancerous effects of these plants in humans have not been well studied yet (56). For example, Dorema aucheri (Bilhar) grows in the Southwest of Iran and is routinely consumed by the people in K&B province (57, 58). The biochemical analysis showed that D. aucheri compounds might have carcinogenic and genotoxicity effects on the human cell lines (57, 59).

This study encountered some limitations. Sampling was restricted to those patients on stages III and IV, so it was not possible to increase the sample size. Furthermore, samples were FFPE, and a number of them did not give an amplifiable DNA. Some samples did not show any mutation, which may be because they had not passed stage IIIA or they may have had a mutation in other exons, except exons 5-8. Although, passing this stage did not guarantee the mutation of the P53 gene.

In conclusion, the results of this study indicated that the R282W P53 gene mutation in exon 8 may play an important role in the development of GC in K&B province (Southwest Iran). It is advised to find more probable mutations through investigating other exons of P53 sequences and by using more sensitive methods such as the sequencing approach. Also, we suggest using fixatives other than formalin to preserve the quality of DNA in pathology laboratories for future analysis of molecular investigations. The P53 mutation is of crucial importance in the success of the treatment plan of cancer patients. The current chemotherapy or radiotherapy methods for cancer are completely dependent on the P53 function because they induce the intrinsic pathway of apoptosis only when P53 is normal (60).

Acknowledgments

The authors thank the Deputy of Research and Technology of Yasuj University of Medical Science and University of Sistan & Baluchestan for providing the financial support for this study (P/23/2/1406).

Footnotes

Authors' Contribution: Sajad Afrouz: conceptualization and designing the study; Mohammad Amin Ghatee: collecting, analysis, and interpretation of data; Amroallah Roozbehi: analyzed the data and co-wrote the paper; Mohammad Hossain Sangtarash: drafted the manuscript and provided final approval of the final version.

Conflict of Interests: The authors report no conflict of interest. The authors are responsible for the content of this article.

Ethical Approval: P/23/2/1406

Funding/Support: The authors thank the Deputy for Research and Technology of Yasuj University of Medical Science and University of Sistan & Baluchestan for their financial support (P/23/2/1406).

References

- Sitarz R, Skierucha M, Mielko J, Offerhaus GJA, Maciejewski R, Polkowski WP. Gastric cancer: epidemiology, prevention, classification, and treatment. *Cancer management and research*. 2018;10:239.
- Fock KM, Ang TL. Epidemiology of Helicobacter pylori infection and gastric cancer in Asia. *Journal of gastroenterology and hepatology*. 2010;25(3):479-86.
- 3. Stock M, Otto F. Gene deregulation in gastric cancer. *Gene*. 2005;**360**(1):1-19.
- 4. Malekzadeh R, Derakhshan MH, Malekzadeh Z. Gastric cancer in Iran: epidemiology and risk factors. *Archives of Iranian Medicine*. 2009.
- Mehrabian A, Esna-Ashari F, Zham H, Hadizadeh M, Bohlooli M, Khayamzadeh M, et al. Gastric cancer prevalence, according to survival data in iran (national study-2007). *Iran J Public Health.* 2010;**39**(3):27–31. [PubMed: 23113019]. [PubMed Central: PMCPmc3481621].
- Iravani S. Gastric cancer as a multifactorial disease. Annals of military and health sciences research. 2013.
- 7. Hartgrink HH, Jansen EP, van Grieken NC, van de Velde CJ. Gastric cancer. *The Lancet*. 2009;**374**(9688):477-90.
- Oliveira C, Seruca R, Carneiro F. Hereditary gastric cancer. Best Practice *δ* Research Clinical Gastroenterology. 2009;23(2):147-57.
- Parkin DM. The global health burden of infection-associated cancers in the year 2002. International journal of cancer. 2006;118(12):3030–44.
- 10. Bunz F. *Principles of cancer genetics*. **1**. Springer; 2008.
- 11. Sonnenschein C, Soto AM. Theories of carcinogenesis: an emerging perspective. *Seminars in cancer biology*. Elsevier; 2008. p. 372–7.
- Longo DL, Fauci AS, Kasper DL, Hauser S, Jameson JL, Loscalzo J. Harrison's principles of internal medicine. *NEW YORK: McGraw-Hill CO*. 2012;6(1):312.
- Smith MG, Hold GL, Tahara E, El-Omar EM. Cellular and molecular aspects of gastric cancer. World journal of gastroenterology: WJG. 2006;12(19):2979.
- 14. Ozaki T, Nakagawara A. p53: the attractive tumor suppressor in the cancer research field. *BioMed Research International*. 2010;**2011**.
- Lee KE, Khoi PN, Xia Y, Park JS, Joo YE, Kim KK, et al. Helicobacter pylori and interleukin-8 in gastric cancer. World Journal of Gastroenterology: WJG. 2013;19(45):8192.
- Benchimol S, Lamb P, Crawford LV, Sheer D, Shows TB, Bruns GAP, et al. Transformation associated p53 protein is encoded by a gene on human chromosome 17. Somatic cell and molecular genetics. 1985;11(5):505–10.
- McBride OW, Merry D, Givol D. The gene for human p53 cellular tumor antigen is located on chromosome 17 short arm (17p13). Proceedings of the National Academy of Sciences. 1986;83(1):130–4.
- Guru S, Ahmad I, Mir R, Javid J, Farooq S, Yadav P, et al. Biological and Clinical Implications of Exon 8 P53 (R282W) Gene Mutation in Relation to Development and Progression of Chronic Myeloid

Leukemia Patients in India Population. *Clinical Lymphoma, Myeloma and Leukemia*. 2014;**14**. S140.

- Muller PA, Vousden KH. p53 mutations in cancer. Nature cell biology. 2013;15(1):2-8.
- Robles AI, Harris CC. Clinical outcomes and correlates of TP53 mutations and cancer. Cold Spring Harbor perspectives in biology. 2010;2(3). a001016.
- 21. Mello SS, Attardi LD. Not all p53 gain-of-function mutants are created equal. Nature Publishing Group; 2013.
- Cho Y, Gorina S, Jeffrey PD, Pavletich NP. Crystal structure of a p53 tumor suppressor-DNA complex: understanding tumorigenic mutations. *Science*. 1994;**265**(5170):346–55. doi: 10.1126/science.8023157. [PubMed: 8023157].
- 23. Mir R, Zuberi M, Ahmad I, Javid J, Yadav P, Farooq S, et al. Biological and clinical implications of exon 8 P53 (R282W) gene mutation in relation to development and progression of chronic myeloid leukaemia patients in India population. *J Cell Sci Ther*. 2013;**4**(2).
- 24. Zhang Y, Coillie SV, Fang JY, Xu J. Gain of function of mutant p53: R282W on the peak? *Oncogenesis*. 2016;**5**(2):e196.
- Ostovar R, Eghdami A, Jafari A, Ravangard R. Burden of gastric cancer: a case study of iran. World Family Medicine Journal: Incorporating the Middle East Journal of Family Medicine. 2018;99(5897):1–5.
- Shiao Y, Rugge MASSIMO, Correa PELAYO, Lehmann HP, Scheer WD. p53 alteration in gastric precancerous lesions. *The American journal of pathology*. 1994;**144**(3):511.
- Olivier M, Hollstein M, Hainaut P. TP53 mutations in human cancers: origins, consequences, and clinical use. *Cold Spring Harbor perspectives in biology*. 2010;2(1). a001008.
- Fenoglio-Preiser CM, Wang J, Stemmermann GN, Noffsinger A. TP53 and gastric carcinoma: a review. *Human mutation*. 2003;21(3):258-70.
- 29. Xu J, Qian J, Hu Y, Wang J, Zhou X, Chen H, et al. Heterogeneity of Li-Fraumeni syndrome links to unequal gain-of-function effects of p53 mutations. *Scientific reports*. 2014;**4**:4223.
- 30. Abdullah S, Sameer SA, Dil-Afroze SN, Das BC. P53-The Molecular Guardian Crashes in Gastric Adenocarcinomas-A Study in an Ethnic Kashmiri Population. *J Carcinogene Mutagene*. 2010;1(106):2.
- Fischer NW, Prodeus A, Gariépy J. Survival in males with glioma and gastric adenocarcinoma correlates with mutant p53 residual transcriptional activity. JCl insight. 2018;3(15).
- AACR Project GENIE Consortium. AACR Project GENIE: powering precision medicine through an international consortium. *Cancer discov*ery. 2017;7(8):818–31.
- 33. Juvan R, Hudler P, Gazvoda B, Repse S, Bracko M, Komel R. Significance of genetic abnormalities of p53 protein in Slovenian patients with gastric carcinoma. *Croatian medical journal*. 2007;**48**(2):207–17.
- Ahrendt SA, Halachmi S, Chow JT, Wu L, Halachmi N, Yang SC, et al. Rapid p53 sequence analysis in primary lung cancer using an oligonucleotide probe array. *Proceedings of the National Academy of Sciences*. 1999;**96**(13):7382-7.
- Javid J, Masroor M, Mir AR, Ahamad I, Farooq S, Yadav P, et al. Clinical and prognostic significance of R282W p53 gene mutation in north India patients with non small cell lung cancer. *Transl Med.* 2012;2(110):2161-1025.100011.
- Bhatia K, Gutierrez MI, Magrath IT. A novel mutation in the p53 gene in a Burkitt's lymphoma cell line. *Human molecular genetics*. 1992;1(3):207–8.
- 37. Ludwig L, Schulz AS, Janssen JW, Grünewald K, Bartram CR. P53 mutations in myelodysplastic syndromes. *Leukemia*. 1992;**6**(12):1302–4.
- Cheng JIAN, Haas MARTIN. Frequent mutations in the p53 tumor suppressor gene in human leukemia T-cell lines. *Molecular and cellular biology*. 1990;10(10):5502–9.
- Sugimoto K, Toyoshima H, Sakai R, Miyagawa K, Hagiwara K, Hirai H, et al. Mutations of the p53 gene in lymphoid leukemia. *Blood.* 1991.
- 40. Lohrasbi Nejad A, Yaghoobi MM. Mutation analysis of TP53 tumor suppressor gene in colorectal cancer in patients from Iran (Kerman Province). *Iranian journal of basic medical sciences*. 2012;**15**(1):683.

- 41. Karim S, Ali A. Correlation of p53 over-expression and alteration in p53 gene detected by polymerase chain reaction-single strand conformation polymorphism in adenocarcinoma of gastric cancer patients from India. World Journal of Gastroenterology: WJG. 2009;15(11):1381.
- 42. Saffari-Chaleshtori J, Tabatabaiefar MA, Ghasemi-Dehkordi P, Farokhi E, Moradi M, Hashemzadeh-Chaleshtori M. The lack of correlation between TP53 mutations and gastric cancer: a report from a province of Iran. *GENETIKA-BELGRADE*. 2017;**49**(1):235–46.
- 43. Abbasi M, Mirmomeni MH, Khazaei S, Ranjbaran R. Detection of p53 exon 9 gene mutation in bladder cancer by polymerase chain reaction-single-strand conformation polymorphism method. *Indian Journal of Medical Sciences*. 2017;**69**(1):47–51.
- Almasi SZ, Salehiniya H. Trends in colorectal cancer incidence in Iran. Journal of Mazandaran University of Medical Sciences. 2015;24(122):391–6.
- Rivlin N, Brosh R, Oren M, Rotter V. Mutations in the p53 tumor suppressor gene: important milestones at the various steps of tumorigenesis. *Genes & cancer.* 2011;2(4):466–74.
- Boyd MT, Vlatkovic N. p53: a molecular marker for the detection of cancer. Expert opinion on medical diagnostics. 2008;2(9):1013–24.
- Salih BA, Gucin Z, Bayyurt N. A study on the effect of Helicobacter pylori infection on p53 expression in gastric cancer and gastritis tissues. *The Journal of Infection in Developing Countries*. 2013;7(9):651–7.
- Pucułek M, Machlowska J, Wierzbicki R, Baj J, Maciejewski R, Sitarz R. Helicobacter pylori associated factors in the development of gastric cancer with special reference to the early-onset subtype. *Oncotarget*. 2018;9(57):31146.
- Rastaghi S, Jafari-Koshki T, Mahaki B, Bashiri Y, Mehrabani K, Soleimani A. Trends and Risk Factors of Gastric Cancer in Iran (2005-2010). *Int J Prev Med.* 2019;**10**:79. doi: 10.4103/ijpvm.IJPVM_188_17. [PubMed: 31198514]. [PubMed Central: PMCPmc6547778].
- International Agency for Research on Cancer. IARC working group on the evaluation of carcinogenic risks to humans. *IARC Monogr Eval Carcinog Risks Hum.* 1994;61:45–119.

- De Gruijl FR. Skin cancer and solar UV radiation. European Journal of Cancer. 1999;35(14):2003-9.
- 52. Soehnge H, Ouhtit A, Ananthaswamy ON. Mechanisms of induction of skin cancer by UV radiation. *Front Biosci*. 1997;**2**:D538–51.
- Benjamin CL, Ananthaswamy HN. p53 and the pathogenesis of skin cancer. *Toxicology and applied pharmacology*. 2007;**224**(3):241–8. doi: 10.1016/j.taap.2006.12.006. [PubMed: 17270229].
- Bahmani M, Zargaran A, Rafieian-Kopaei M. Identification of medicinal plants of Urmia for treatment of gastrointestinal disorders. *Revista Brasileira de Farmacognosia*. 2014;24(4):468–80.
- 55. Jahantab E. Ethnobotanical study of medicinal plants of Boyer Ahmad and Dena regions in Kohgiluyeh and Boyer Ahmad province, Iran. Advanced Herbal Medicine. 2018;4(4):12–22.
- Desai AG, Qazi GN, Ganju RK, El-Tamer M, Singh J, Saxena AK, et al. Medicinal plants and cancer chemoprevention. *Curr Drug Metab.* 2008;9(7):581–91. doi: 10.2174/138920008785821657. [PubMed: 18781909]. [PubMed Central: PMCPmc4160808].
- 57. Mostafavi SH, Fazilati M, Mostafavi S, Vahhabi MR, Mostafavi F, Omidvarinia S, et al. Hepatotoxicity of Dorema aucheri (Bilhar) in albino mice. *Archives of Iranian Medicine (AIM)*. 2013;**16**(9).
- Mosaddegh M, Naghibi F, Moazzeni H, Pirani A, Esmaeili S. Ethnobotanical survey of herbal remedies traditionally used in Kohghiluyeh va Boyer Ahmad province of Iran. *Journal of ethnopharmacology*. 2012;141(1):80–95.
- Etebari M, Sajjadi SE, Jafarian-Dehkordi A, Nazmakanipour S. Genotoxicity evaluation of hydroalcoholic and aqueous extracts of Dorema aucheri by the comet assay. *Adv Biomed Res.* 2016;5:199. doi: 10.4103/2277-9175.190993. [PubMed: 28217637]. [PubMed Central: PMCPmc5220686].
- Webley KM, Shorthouse AJ, Royds JA. Effect of mutation and conformation on the function of p53 in colorectal cancer. *The Journal of pathology*. 2000;**191**(4):361–7.