




# Evaluation of Antimicrobial Activity of Some Medicinal Plants on Human Standard Bacteria and *Candida albicans*

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## Abstract

**Background:** Continuous and indiscriminate use of chemical drugs causes resistance to microorganisms, which in turn weakens the effect of drugs. This adverse event is associated with an increased number of drugs used by patients and the tendency to use compounds with newer and stronger formulations. Furthermore, the essential oils of several plants contain a significant inhibitory effect on pathogenic microorganisms.

**Objectives:** Hence, the current study intended to evaluate the antimicrobial activity of some medicinal plants on some standard human pathogenic bacteria and *Candida albicans* fungi isolated from women.

**Methods:** Leaves of *Cichorium intybus* L., *Hypericum perforatum* L., *Lavandula angustifolia*, *Thymus vulgaris* L., and *Taxus baccata* L. were collected and analyzed in the botanical laboratory of the University of Zabol. Then, the ethanolic extract was prepared using 40 g of dried leaves in 400 cc of ethanol. Standard bacteria and fungi were obtained from the center for genetic and biological resources of Iran. To determine the activity of free radical trapping, diphenylpicryl hydrazyl was used, and then the antimicrobial effects were investigated by diffusion method in Müller-Hinton agar medium using 6 mm paper disks according to the Bauer and Kirby instructions. Statistical calculations were administered using Statistx Ver10. Mean comparisons were performed using the LSD at the 1% level, and Excel was also used to draw the shapes.

**Results:** The diameter of the inhibitory zone of plant extracts against standard bacteria and clinical fungi of *Candida albicans* at a dilution of 100 ppm was analyzed, which revealed different effects ( $P < 0.01$ ). *Taxus baccata* L., with a 15 mm diameter growth zone, showed the highest effect on inhibiting the growth of *Bacillus cereus* and *Pseudomonas aeruginosa*. *Hypericum perforatum* L. with a diameter of 15 mm was found as the most useful plant in inhibiting *Shigella dysenteriae*, with a diameter of 10 mm, was the most useful plant in inhibiting the growth of *Escherichia coli*. The *Taxus baccata* L., with a maximum growth inhibition zone diameter (20 mm), has been the most effective plant against *Candida albicans*.

**Conclusions:** Considering the side effects of chemical drugs and antibiotics as well as the significant effect of medicinal plant extracts used in this study, the *Taxus baccata* L. was the most useful plant on inhibiting *Candida albicans*, *Bacillus cereus*, and *Pseudomonas aeruginosa*. Moreover, *Hypericum perforatum* L. was found as the most useful plant to control the growth of *Escherichia coli*.

**Keywords:** *Taxus baccata* L., *Lavandula angustifolia*, Medicinal Plants

## 1. Background

Investigating the antimicrobial effects of secondary metabolites of medicinal plants, such as essential oils and plant extracts, showed that at least one-third of all medicinal products are of plant origin or have been modified after extraction from the plant. In addition to preventing the growth of bacteria and food contaminated mold, these substances are used to increase the shelf life

of processed foods as well as fruits and vegetables (1-7). Chicory (*Cichorium intybus* L.) belongs to the genus Asteraceae. Its roots, leaves, and seeds contain several medicinal compounds such as inulin, sesquiterpene, Lactone, Kumarin, flavonoids, and vitamins that are used as anti-hepatitis, anti-nephrotic, anti-cancer, appetizer, and anti-inflammatory. In addition, their antibacterial properties against chicory root extract have been reported (8-10).

*Hypericum perforatum* L. belongs to the genus Clasiac and contains a wide range of secondary metabolites such as flavonoids, proanthocyanidins, and naphthodiandrons (ipricin and pseudoeprycin (acyl fluoroglucinols (hyperforin and adheiphorphin)). It has many therapeutic properties like hypothyroidism, anti-inflammatory, anti-viral, antioxidant, anti-tumor, and antibacterial, but its most significant effect and application has been reported in the treatment of depression (11-13).

*Lavandula angustifolia* not only affects most organs and cells of the body but also has analgesic and anti-inflammatory effects and can reduce morphine tolerance and dependence. *Lavandula angustifolia* also affects cellular mechanisms such as oxidation reactions (reduction of oxidative reactions), programmed cell death (anti-apoptosis), and nitric oxide production (reduction of NO) and can affect cell genetic health. *Lavandula angustifolia* seems to be exerted by calmodulin calcium and its related kinases (14, 15).

*Thymus vulgaris* L. is a perennial plant of the genus *Lamiaceae* (*Labiatae*) with a cushion or clumpy structure. This plant is native to the countries of the Mediterranean region and grows in arid areas and between the boulders of the Mediterranean region, especially in France, Spain, Portugal, and some parts of Asia. *Thymus vulgaris* L. contains essential oils and compounds such as flavonoids, saponins, and bitter substances. The most critical components of *Thymus vulgaris* L. essential oil are phenolic compounds like thymol and carvacrol (5, 7).

The *Taxus baccata* L. is a coniferous leaf belonging to the Taxaceae family. Its medicinal value roots in the presence of Paclitaxel under the brand name Taxol in its needle leaves. Taxol, by forming an abnormal division spindle, can cease DNA transcription at the G2/M stage of mitotic division, thereby killing proliferating cells. The Food and Drug Administration has approved Taxol in 1977 for the treatment of uterine and breast cancers. Despite the invention of new methods of preparation of Taxol, such as cell culture, extraction from plant sources still retains its importance and place in supplying this valuable drug (16, 17).

As different medicinal plants pose various effects on microbes and the other hand the use of antibiotics is common (18).

## 2. Objectives

The current study aimed to investigate the effects of *Cichorium intybus* L., *Hypericum perforatum* L., *Lavandula angustifolia*, *Thymus vulgaris* L., and *Taxus baccata* on the stunting and non-growth of *Bacillus cereus* standard bacteria (i.e. *Escherichia coli*, *Shigella dysentery*, and *Pseudomonas aeruginosa*) as well as the clinical fungi (i.e. *Candida albicans* 1,

*Candida albicans* 2, *Candida albicans* 3 and *Candida albicans* 4).

## 3. Methods

### 3.1. Herbal Materials

*Cichorium intybus* L., *Hypericum perforatum* L., *Lavandula angustifolia*, and *Thymus vulgaris* L. from Shahrekord (Coordinates: 32°19'32"N 50°51'52 E) and the leaves of the yew tree (*Taxus baccata* L.) were collected from Behshahr, Mazandaran province (36°41'32 36 N°53°33 53 09'E), and the species were determined in the botanical laboratory of the University of Zabol.

### 3.2. Method of Preparing Ethanolic Extract

The leaves of the plants used were dried in the shade and in the vicinity of air. Then 40 grams of leaves were ground (IKA company model A11 basic in Germany). The dried leaves were then soaked in 400 cc of ethanol for 96%. Shake for 48 hours at room temperature on a shaker (UniEquip SKIR-601L Germany). Then, the extracts were filtered, and the solvent was evaporated at a temperature of less than 40°C by a rotary device (Pars Azma Company, model RO02, Iran). The weighted extracts were weighed, and then 100 mg of the extract powder was dissolved in 1 cc of DMSO solvent. The extracts were stored in the refrigerator at 4°C until used in antimicrobial experiments (5).

### 3.3. Bacterial and Fungal Strains

Standard strains of bacteria such as *Bacillus cereus* (ATCC117718), *Escherichia coli* (ATCC700728), *Shigella dysentery* (ATCC13313), and *Pseudomonas aeruginosa* (ATCC27853) were obtained from Patten Teb. Also, clinical fungi of *Candida albicans* 1, *Candida albicans* 2, *Candida albicans* 3, and *Candida albicans* 4 were prepared from female patients in Zabol City.

### 3.4. Investigation of Antimicrobial Effects

In this section, the antimicrobial effects of the above-mentioned plant extracts on human pathogens were investigated by diffusion method in Müller Hinton agar culture medium (made by Merck Germany) using paper discs (6 mm). In addition, microbial susceptibility was determined following the method proposed by Bauer et al.

### 3.5. Determination of Minimum Inhibitory Concentration and Minimum Bactericidal Concentration

To determine the minimum inhibitory concentration (MIC) of essential oils of plants used by eye method, the first 100 microliters of Mueller Hinton broth (made by Merk-Germany) was added to each well of the titer plate (19, 20). Then, 100  $\mu$ L of dilution of 20 mg/mL of essential oil was added to the first well, followed by mixing. In the next step, 100  $\mu$ L of the first well was removed and added to the second well. Similarly, the construction of double dilutions in other wells was continued. It should be noted that in this case, the first well contained 20 mg per microliter of essential oil, and, thus, in subsequent wells, its concentration was half of the previous well. Eventually, 10  $\mu$ L of each bacterial suspension (cfu = 108 11.5 per mL, half McFarland) was added to the wells. DMSO was added to the negative control well (without essential oil), followed by incubation at 37°C for 37 hours.

The MIC was defined as the lowest concentration required to stop the bacterial growth after 24 hours of incubation. To determine the minimum bactericidal concentration (MBC), 10 microliters of the wells was secondarily cultured on a nutrient agar medium (manufactured by Merk-Germany) after 24 h of incubation, and plates were examined for bacterial growth for 24 hours. The lowest concentration of essential oil in which 99.9% of bacteria did not grow was considered as MBC (19, 20). All antimicrobial tests were performed in triplet.

### 3.6. Data Analysis

Statistical calculations were administered using Statistix software version 10. Mean comparisons were performed using the least significant difference (LSD) at the 1% level, and Excel was also used to draw the shapes.

## 4. Results

### 4.1. Diameter of Inhibition Zone of Plant Extracts on the Bacteria Used

The inhibitory diameter zone of plant extracts against different bacteria was diluted to 100 ppm, and it was found that different extracts had different effects on growth inhibition of *Bacillus cereus*, *Escherichia coli*, *Shigella dysentery*, and *Pseudomonas aeruginosa* ( $P < 0.01$ ) (Figure 1 and Table 1). According to the LSD post hoc test, the yew tree, with a 15 mm diameter, growth inhibition zone was the most useful plant concerning inhabitation of *Bacillus cereus* and *Pseudomonas aeruginosa* growth. *Thymus vulgaris* L., with a diameter of 15 mm, was not the most useful plant in inhibiting the growth of *Shigella dysentery*. With a diameter of 10 mm, it was not the most useful plant in inhibiting the growth of *Escherichia coli* (Figure 2).

### 4.2. Diameter of Inhibition Zone of Plant Extracts on *Candida albicans*

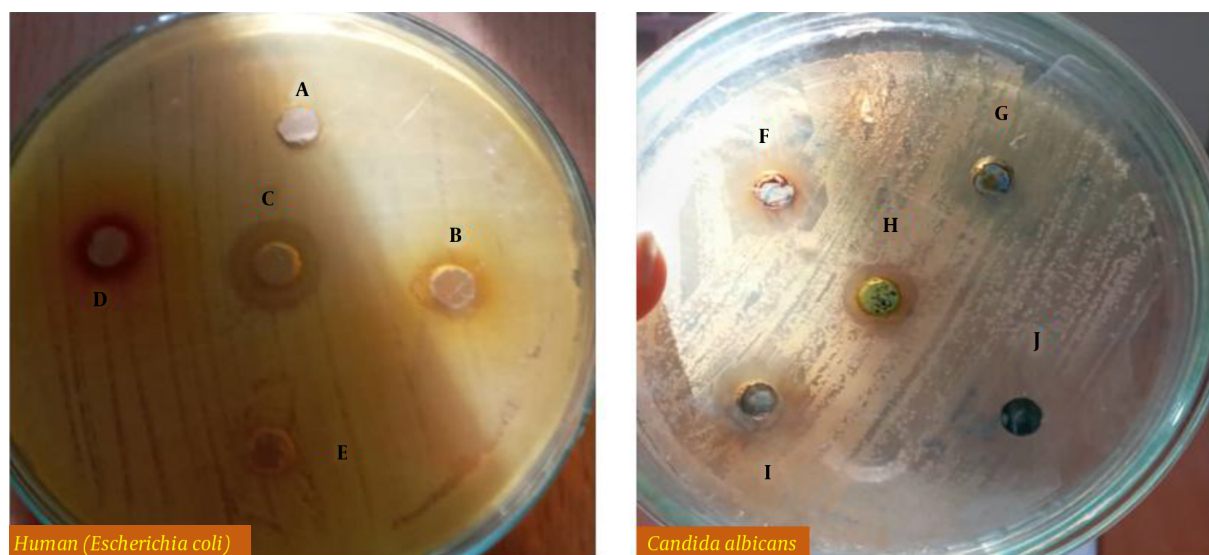
The diameter of the inhibitory zone diameter of plant extracts against *Candida albicans* at 100 ppm was investigated and it was found that different extracts had different effects on inhibiting the growth of *Candida albicans* ( $P < 0.01$ ) (Figure 1 and Table 2). LSD post hoc test showed that the yew tree with 20-, 10-, and 14-mm diameter growth inhibition zone was the most useful plant on *Candida albicans* 1, 3, and 4, respectively. Herring flower was the most useful plant against *Candida albicans* 2 (a diameter of 15), followed by the yew tree with yew and chicory (diameters of 12 and 12, respectively) (Figure 3).

### 4.3. Minimum Inhibitory Concentration and Minimum Bactericidal Concentration of Plant Extracts

The MIC of ethanolic extract of yew on *Bacillus cereus*, *Escherichia coli*, and *Pseudomonas aeruginosa* was 12.5, 12.5, and 25 ppm, respectively; in contrast, *Escherichia coli* was inhibited in all concentrations of yew and the lowest lethal concentration on yew *Bacillus cereus*, *Escherichia coli*, and *Pseudomonas aeruginosa* was 25 - 25 and 50 ppm, respectively (Table 3). The MIC of chicory ethanolic extract against *Bacillus cereus*, *Escherichia coli*, *Shigella dysentery*, and *Pseudomonas aeruginosa* was 25 - 25 - 50 and 25 ppm, respectively, and the highest lethal concentration was 100 ppm when *Shigella dysentery* was killed (Table 3). The MIC of ethanolic extract of *Thymus vulgaris* L. against *Bacillus cereus*, *Escherichia coli*, *Shigella dysentery*, and *Pseudomonas aeruginosa* was 25 - 50 - 25 and 12.5 ppm, respectively, and the highest lethal concentration against *Bacillus cereus*, *Shigella dysentery*, *Escherichia coli*, and *Pseudomonas aeruginosa* was equal to 50, 100, 50, and 25 ppm (Table 3).

The MIC of *Hypericum perforatum* L. extract against *Bacillus cereus*, *Escherichia coli*, *Shigella dysentery*, and *Pseudomonas aeruginosa* was 12.5, 50, 50, and 50 ppm, and the MBC was 25 - 100 - 100 - 100. The minimum inhibitory concentration of *Lavandula angustifolia* extract against bacteria was 12.5 - 25, and 12.5 ppm, and the MBC was 25 - 50 - 50 and 25 ppm (Table 3).

Investigating the antifungal activity of plant extracts showed that the lowest inhibitory concentration of the yew extract was equal to 12.5 ppm, which was inhibited by both parties, in comparison, the lowest inhibitory concentration of chicory extract was 25 ppm, which all strains were inhibited, and the lowest inhibitory concentration of *Thymus vulgaris* L. was 12.5 ppm, of which 3 strains were inhibited (Table 4). The lowest and highest inhibitory concentrations of basil ethanolic extract against *Candida albicans* were 25 and 100 ppm, respectively. The MIC of *Lavandula angustifolia* extract against *Candida albicans* was 25 ppm, in which all strains were inhibited (Table 4).



**Figure 1.** Diameter of plant growth inhibition zone on human *E. coli* (A, chicory; B, fennel; C, yew; D, lavender; E, thyme) and *Candida albicans* (F, chicory; G, sage; H, yew; I, lavender; J, garden thyme).

**Table 1.** Diameter Variance Analysis of Inhibitory Zone of Extracts of different Plants Against Different Bacterial Agents at Dilution of 100 ppm

Source	DF	SS	MS	F
Plant	4	207.60	51.9	51.9 <sup>a</sup>
Error	10	10	1.00	
Total	14	217.60		

<sup>a</sup>Significant at the level of one percent.

**Table 2.** Analysis of Variance Diameter Inhibitory Zone of Different Plant Extracts Against *Candida albicans* at 100 ppm

Source	DF	SS	MS	F
Plant	4	417.6	104.4	163.12 <sup>a</sup>
Error	10	6.4	0.64	
Total	14	424.00		

<sup>a</sup>Significant at 1%

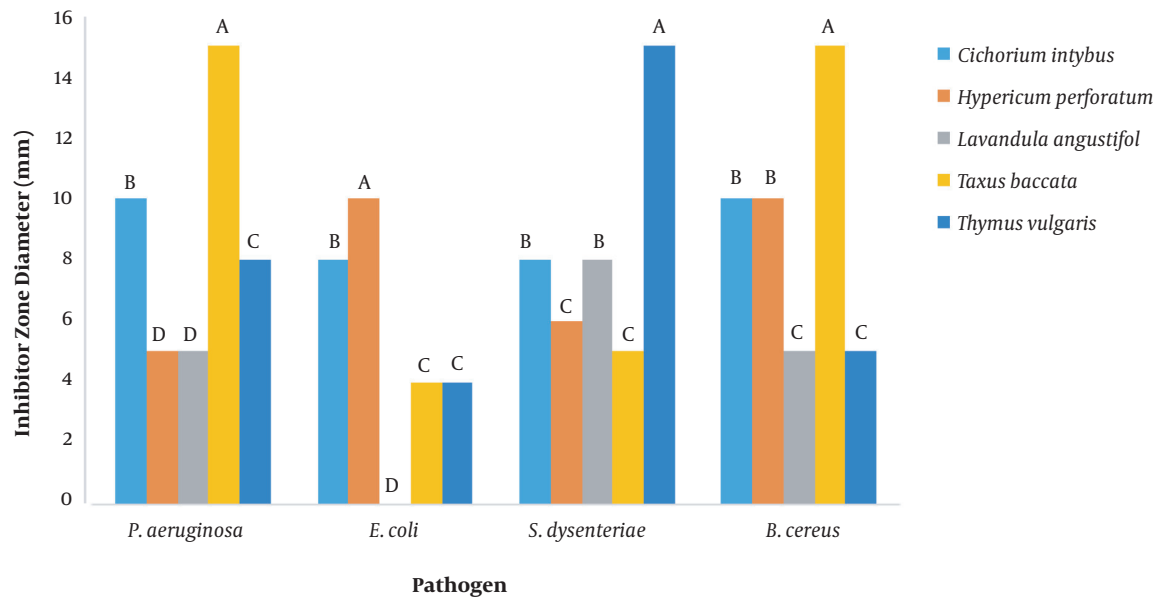
**Table 3.** Investigation of Antimicrobial Activity of Plant Extracts on Pathogenic Bacteria

Bacteria	<i>Cichorium intybus</i> L. MIC/MBC	<i>Thymus vulgaris</i> L. MIC/MBC	<i>Hypericum perforatum</i> L. MIC/MBC	<i>Nepeta binaludensis</i> Jamzad MIC/MBC	<i>Taxus baccata</i> L. MIC/MBC
<i>Bacillus cereus</i>	25 - 50	12.5 - 25	25 - 50	25 - 50	12.5 - 25
<i>E. coli</i>	25 - 50	50 - 100	50 - 100	25 - 50	No growth
<i>Shigella dysenteriae</i>	12.5 - 25	50 - 100	25 - 50	50 - 100	12.5 - 25
<i>Pseudomonas aeruginosa</i>	12.5 - 25	50 - 100	12.5 - 25	25 - 50	25 - 50

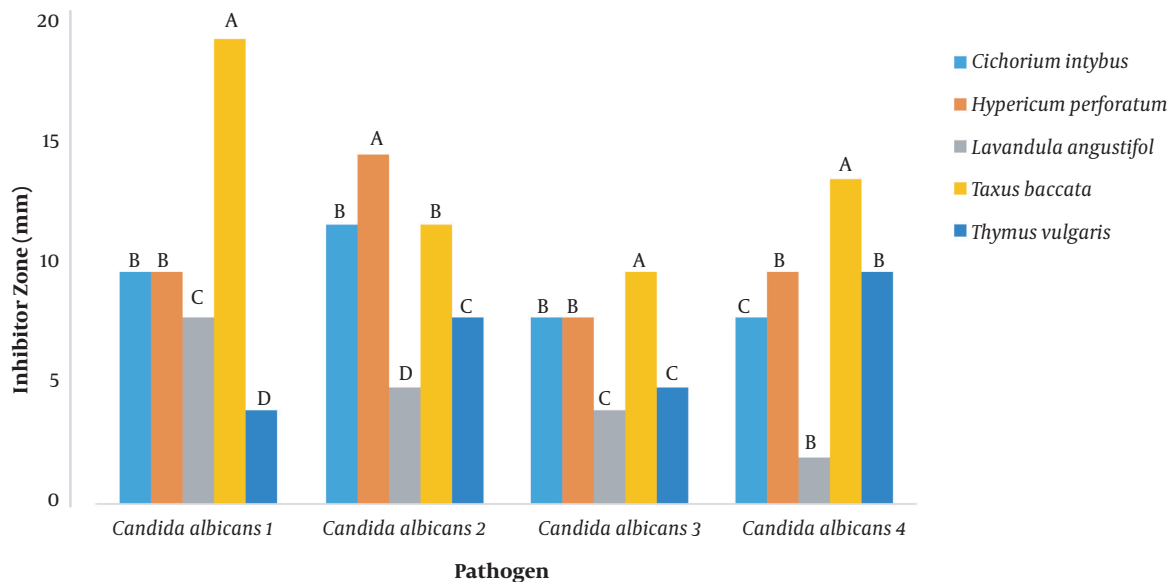
## 5. Discussion

According to the findings, the yew tree with a growth inhibition zone of a 15 mm diameter was the most use-

ful plant to inhibit the growth of *Bacillus cereus* and *Pseudomonas aeruginosa*. *Thymus vulgaris* L., with a diameter of 15 mm, was not the most useful plant in inhibiting the



**Figure 2.** The diameter of non-growth zone of different plants extracts against bacteria used in a dilution of 100 ppm. Similar letters indicate no significant difference



**Figure 3.** The diameter of growth inhibition zone of different plants extracts against *Candida albicans* at 100 ppm dilution. Similar letters indicate no significant difference

growth of *Shigella dysenteriae*. With a diameter of 10 mm, this herb was the most useful plant in inhibiting *Escherichia coli*. The yew tree with a growth inhibition zone of 20-, 10-, and 14-mm was the most useful plant on *Candida albicans* 1, 3, and 4, respectively. Herring flower was the most useful plant against *Candida albicans* 2 (a diameter of 15), followed

by the yew tree with yew and chicory (diameters of 12 and 12, respectively).

A study on the chemical composition and antibacterial activity of Iranian *Lavandula* × hybrid concluded that the diameter of the growth inhibition zone against *S. aureus* and *E. coli* was 9.36 and 23.3 mm, respectively. A sig-



**Table 4.** Investigation of Antifungal Effects of Medicinal Plants on *Candida albicans*

Spices	<i>Cichorium intybus</i> L. MIC/MBC	<i>Thymus vulgaris</i> L. MIC/MBC	<i>Hypericum perforatum</i> L. MIC/MBC	<i>Nepeta binaludensis</i> Jamzad MIC/MBC	<i>Taxus baccata</i> L. MIC/MBC
<i>Candida albicans</i> 1	25 - 50	50 - 100	12.5 - 25	25 - 50	25 - 50
<i>Candida albicans</i> 2	25 - 50	100 - 200	12.5 - 25	25 - 50	12.5 - 25
<i>Candida albicans</i> 3	25 - 50	50 - 100	25 - 50	25 - 50	25 - 50
<i>Candida albicans</i> 4	25 - 50	25 - 50	12.5 - 25	25 - 50	12.5 - 25

nificant association between the composition of essential oil and the level of antibacterial effect, expressed as inhibition areas, has been reported (21). In the present study, no *E. coli* was grown in a medium containing *N. binaludensis* Jamzad extracts.

A study on antibacterial properties of essential oils and hydrosols and aqueous extracts of *Lavandula* spp. grown in Australia reported that the hydrosols and aqueous extracts of the leaves of the plant had no antibacterial activity. This study also and concluded that different species of *Lavandula* spp. may have different antibacterial properties (22). In the present study, *N. binaludensis* Jamzad was effective against *E. coli* and no *E. coli* was grown, which confirms the different effects of various species of lavender against bacteria. They have interesting microbes and maybe a new potential source of natural antimicrobial as well as a new wound healing product (23), which confirms the effect of plant species on antimicrobial properties. The synthesis of antibacterial silver nanoparticles has been investigated using the extract of yew (*Taxus baccata* L.). It was concluded that the lowest MIC was obtained for *S. pyogenes* at a concentration of 50 µg/mL and for *E. coli* and *S. aureus* at a concentration of 25 µg/mL. Finally, they suggested that the use of biological compounds, mainly plant extracts, instead of toxic and hazardous chemicals, to synthesis silver nanoparticles, could reduce the environmental concerns regarding these nanoparticles (24). Similar to the findings of the present study, the MIC of ethanolic extract of yew on *Bacillus cereus*, *Escherichia coli*, and *Pseudomonas aeruginosa* was 12.5, 12.5, and 25 ppm.

The antimicrobial effects of watercress extract and the derived micro-emulsion on gram-negative clinical strains were investigated, and the highest zone and micro-emulsion were obtained at a concentration of 8 mg/mL with a diameter of 42 mm. This composition formed a zone with a diameter of 30 mm on the same bacterium at a concentration of 0.1 mg/mL; however, no zone formation was observed on *Pseudomonas*. The highest effect of the extract, which resulted in the MIC, was observed at 0.01563 mg/mL on *Salmonella* and for *Shigella* 0.00625 mg/mL. The MIC of the extract was 0.5 mg/mL for *Escherichia coli* and *Proteus* and 0.025 mg/mL for micro-emulsion of both bacteria (25).

In the present study, the largest diameter of the growth inhibition zone against *Candida albicans* was 20 mm.

Medicinal plants have special place in the field of allochemicals, mainly due to their secondary metabolites. In addition, the demand for medicinal compounds has increased, but some of these plants have limited natural habitats, and depending on the environmental and geographical conditions, their collection may be difficult. Researchers have focused on the use of biotechnology techniques to increase the production and productivity of medicinal plants as follows; Low concentration of phytochemicals in plants, limitation of natural resources, increasing degradation of forests, pastures, and green space, extinction of diverse plant species, problems related to domestication and crop cultivation. Biotechnology can provide solutions to increase the efficiency of producing medicinal plants, as renewable sources for drug production, using various sciences such as biology, biochemistry, genetics, etc., and using cell, organ, and porcine culture methods as well as genetic engineering, and molecular markers (8).

Overuse of antibiotics has often led to the growing resistance of bacteria to these drugs. On the other hand, overuse of antibiotics is often associated with side effects in the human body. Because some plants have antimicrobial properties, they can be used to fight specific pathogenic microbes and find harmless alternatives to antibiotics.

On the other hand, in recent years, the effectiveness of antibiotics has decreased as a result of microbial resistance that led to the growth of research intended to develop new antibiotic compounds or to enhance the performance of antibiotics, among which increased attention has been paid to medicinal plants (26-28). Hence, the plants used in the present study can be examined against other microbes to obtain a comprehensive result. However, further studies are needed to determine the best-uncomplicated dose with the highest effectiveness as well as the active ingredient.

### 5.1. Conclusions

Despite their widespread benefits, chemical drugs, including antibiotics, also have side effects. According to the findings of the present study, the yew tree (*Taxus baccata* L.) was the most useful plant against *Candida albicans*, *Bacillus cereus*, and *Pseudomonas aeruginosa*. *Hypericum perforatum* L. was also the most useful plant in controlling the growth of *Escherichia coli*.

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### Footnotes

**Authors' Contribution:** All authors had an equal role in study design, statistical analysis, and manuscript writing.

**Conflict of Interests:** None of the authors have any conflict of interest to declare.

**Ethical Approval:** No human or animals were used in the present research. The study protocol was approved by the Ethics Committee of University of Zabol (code: IR.UOZ.REC.1399.006)

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