



Evaluation of Plasma MMP-9 and VEGF-A mRNAs as Non-invasive Diagnostic Biomarkers in Women with Endometriosis

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Abstract

Background: Endometriosis is one of the common gynecological diseases and can lead to pelvic pain, dysmenorrhea, dyspareunia, and infertility in women. Thus, accurate and early diagnosis is a pivotal issue and an essential need for managing this disorder. At the present, the gold standard diagnostic method for endometriosis is laparoscopic surgery that is an invasive method and can lead to delay in diagnosis. Thus, there is an immediate necessity to search for non-invasive diagnostic biomarkers, such as blood-based ones.

Objectives: Matrix metalloproteinase-9 (MMP-9) and vascular endothelial growth factor-A (VEGF-A) have essential roles in the pathogenesis of endometriosis. Therefore, in this study, we evaluated the plasma mRNA levels of MMP-9 and VEGF-A, as potential non-invasive diagnostic biomarkers for endometriosis.

Methods: This study included 48 women (24 cases and 24 controls) who underwent laparoscopy for suspected endometriosis. Pre-operative plasma samples were collected, and after RNA extraction, the levels of MMP-9 and VEGF-A mRNAs were determined by reverse transcription-quantitative polymerase chain reaction (RT-qPCR).

Results: Plasma MMP-9 mRNA level was statistically higher in endometriosis patients compared with the control group (P value = 0.01). However, plasma VEGF-A mRNA level did not show a significant difference between the two groups (P value = 0.5).

Conclusions: It seems that the plasma level of MMP-9 mRNA in endometriosis patients is significantly higher than in non-endometriosis women. This finding can provide new insights regarding this mRNA's applicability as a non-invasive diagnostic biomarker for discovering new cases of endometriosis (newly diagnosed). According to our results, despite the suggested role of VEGF-A in endometriosis pathogenesis, it seems that the plasma level of VEGF-A mRNA does not have the potential to be used as a non-invasive diagnostic biomarker.

Keywords: Endometriosis, Matrix Metalloproteinase 9, Diagnosis, Biomarkers, VEGF-A, Plasma

1. Background

Endometriosis is a debilitating and chronic disease among women. In this disorder, the endometrial tissue grows in ectopic locations. It is diagnosed in approximately 10-15% of reproductive-aged women and 35-50% of infertile women. In addition, it is observed in 50-70% of women with pelvic pain (1). The common signs of endometriosis comprise pelvic pain, dysmenorrhea, dyspareunia, and infertility. Thus, it can influence the quality of life and impose health-related costs (2).

The gold standard diagnostic method for endometriosis is laparoscopic surgery and biopsy (3). Laparoscopic surgery is an invasive and costly procedure. In addition,

it harbors the risks associated with general anesthesia and post-procedure adhesions. Thus, a non-invasive diagnostic test for endometriosis can lead to timely diagnosis and better disease management (4). Blood biomarkers have been considered as non-invasive diagnostic parameters for this disease. Blood-based diagnostic markers for endometriosis can lead to cost-effective, less invasive, and easily acceptable procedures for patients and provide quick results (5). Despite many efforts, no non-invasive biomarker has been approved for the diagnosis of endometriosis yet.

Extracellular circulating RNAs in biological fluids such as plasma, urine, and saliva are becoming optimistic non-invasive and cost-effective diagnostic tools (6). A previous

study showed the capacity of some miRNA panels as blood-based non-invasive biomarkers for endometriosis (7), and some studies have suggested the usefulness of plasma mRNAs as biomarkers for different cancers (8-11). Accordingly, circulating mRNAs in serum may be helpful non-invasive tools for the diagnosis of endometriosis.

According to previous investigations, matrix metalloproteinase-9 (MMP-9) and vascular endothelial growth factor-A (VEGF-A) make an essential contribution to the pathogenesis of endometriosis (12, 13). Also, the mRNAs of these genes are overexpressed in endometrial lesions in women with endometriosis (14, 15). The MMP-9 gene encodes a matrix metalloproteinase involved in extracellular matrix (ECM) degradation (16). This feature is vital for the invasion of the endometrial tissue to ectopic locations (17). The VEGF-A gene encodes a factor essential for angiogenesis; a process that is necessary for the survival and propagation of ectopic endometrial lesions (18). Some studies have detected the mRNAs of these genes in the peripheral blood of endometriosis patients (19, 20). However, the diagnostic potential of these mRNAs circulating in the plasma of endometriosis patients is uncertain.

2. Objectives

The aim of this study was to quantify the mRNA levels of MMP-9 and VEGF-A in endometriosis patients' plasma samples by RT-qPCR.

3. Methods

3.1. Patients and Sample Collection

The study population consisted of 48 women with suspected endometriosis referred to a gynecologist with complications such as dysmenorrhea or pelvic pain. All the patients were scheduled to undergo laparoscopy and signed written informed consent before surgery. This study was approved by the Research Ethical Committee of Tehran University of Medical Sciences (IR.TUMS.MEDICINE.REC.1398.579). Before surgery, 5 mL of blood was obtained from each participant and transferred into a tube containing EDTA as an anticoagulant. Then blood samples, without delay, were transferred to the laboratory for subsequent processing and plasma separation. According to the gold standard for endometriosis diagnosis (direct visualization of lesions in laparoscopic surgery and confirmation by pathological assessment of the tissue), the patients were categorized into two groups: 24 patients with a confirmed diagnosis of endometriosis (as the case group) and 24 patients without endometriosis (as

the control group). Endometriosis stage was defined based on the guidelines of the American Society of Reproductive Medicine (ASRM).

The characteristics of study participants, including the menstrual cycle phase and medical background, were recorded in a clinical questionnaire based on their medical records. Moreover, none of the patients received pre-operative treatments, such as hormone therapy, which was regarded as one of important inclusion criteria due to the possible effects of drugs on gene expression. In addition, all the participants were in reproductive age and referred to a gynecologist with endometriosis symptoms. Patients with a previous diagnosis of diseases such as cancer, autoimmune diseases, and endocrine disturbances (polycystic ovarian syndrome, etc.) were excluded from this study.

3.2. Plasma Separation and RNA Extraction

By centrifugation (at 5000 rpm and then 1500 rpm for 15 min at 4 °C), plasma samples were initially separated from whole blood. Thermo Scientific Nanodrop 2000 was used to measure plasma quality at 414 nm for detecting hemolysis in samples. Then the plasma samples that had no hemolysis were stored at -80 °C. The TRIzol reagent (RiboEx LS, GeneAll, Korea) was used to isolate total RNA from 400 μ L plasma. Afterward, the RNA samples were subjected to standard ethanol precipitation, and the pellet was resuspended in 10 μ L of RNAase-free water. Finally, the quality and quantity of the extracted RNAs were appraised by a spectrophotometer.

3.3. Reverse Transcription-quantitative Polymerase Chain Reaction

The extracted RNA samples were converted to cDNA using a Reverse Transcription kit (Geneall, Korea) according to the manufacturer's protocol. The mRNA levels of MMP-9 and VEGF-A were measured by SYBR Green RealQ Plus 2x Master Mix Green (Ampliqon, Denmark) using Roche Light-Cycler® 96 System. Normalization was carried out employing β -Actin as a reference gene. The reactions included an initial step at 95°C for 30 sec, followed by 40 cycles of amplification; 95 °C for 10 s (denaturation), 60 °C for 15 s (annealing), and 72 °C for 30 s (extension). The qRT-PCR reaction mixture consisted of 5 μ L SYBR Green master mix, 1 μ L of each of forward and reverse primers (primer concentration: 5 pmol), 1 μ L of target cDNA, and 2 μ L sterile water in a total volume of 10 μ L. Each reaction was performed in duplicate. The specificity of RT-qPCR amplification was confirmed by melting curve analysis and observing a single peak. The $2^{-\Delta\Delta Cq}$ method was used for calculating the relative expression of each target mRNA. The sequences of the primers used in this study have been represented in [Table 1](#).

Table 1. The Sequences of the Primers Used for Gene Expression Analysis

Primers	Sequences
MMP-9	
Forward	5'-CAGGCAGCTGGCAGAGGAAT-3'
Reverse	5'-TTCGACTCTCCACGCATCTC-3'
VEGF-A	
Forward	5'-CTCCACCATGCCAAGTGGT-3'
Reverse	5'-TCTCGATTGGATGGCAGTAGC-3'
β-Actin	
Forward	5'-CCCAGCACAATGAAGATCAAGATCAT-3'
Reverse	5'-ATCTGCTGGAAGTGGACACGCA-3'

Abbreviations: MMP-9, matrix metalloproteinase-9; VEGF-A, vascular endothelial growth factor-A.

3.4. Statistical Analysis

The Kolmogorov-Smirnov test showed that the data had a non-normal distribution ($P < 0.05$). Thus, a non-parametric test, the Mann-Whitney U test, was used to compare the studied variables between the groups. All statistical tests were performed in IBM SPSS Statistics 22 (SPSS Inc., Chicago, IL, USA), and $P < 0.05$ denoted a statistically significant difference between the two groups.

4. Results

4.1. Participants' Descriptive Parameters

The participants' clinicopathological information has been briefed in Table 2. A total of 24 endometriosis patients with stage III ($n = 10$) and IV ($n = 14$) were enrolled in this study (mean age: 33.5 ± 5.1 years). The control group included 21 women with benign gynecological diseases and three healthy women (mean age: 37.5 ± 9.1 years). A definite diagnosis in all the patients was made by histological analysis. The menstrual cycle of all the participants was determined; there was no significant difference in the menstrual phase between patients with endometriosis and control subjects (P value > 0.05).

4.2. MMP-9 and VEGF-A mRNAs' Plasma Levels

The relative levels of the mRNAs of MMP-9 and VEGF-A were measured in plasma samples using RT-qPCR (Figure 1). The plasma mRNA level of MMP-9 was significantly higher in endometriosis patients compared with the control group (P value = 0.01). In contrast, no statistically significant difference was observed between the two groups regarding VEGF-A mRNA plasma level (P value = 0.5).

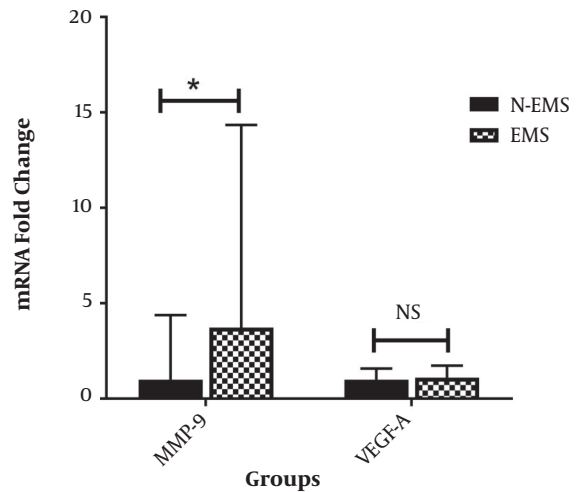


Figure 1. The mRNA fold-change of MMP-9 and VEGF-A in the plasma of endometriosis (EMS) patients compared with non-endometriosis (N-EMS) counterparts. (*: P value < 0.05 , NS: non-significant).

4.3. Association of mRNA Levels of MMP-9 and VEGF-A with Menstrual Cycle Phase and Disease Stage

There was no significant association between the mRNA level of MMP-9 and the menstrual cycle (P value > 0.05). However, there was a significant positive association between VEGF-A mRNA level and the proliferative phase of the menstrual cycle in the study participants (P value < 0.05). Moreover, the expression levels of both target mRNAs in this study were independent of disease stage (P value > 0.05).

5. Discussion

Endometriosis is a common and chronic gynecological disease that affects women's reproductive health and quality of life. Endometriosis can have complications such as pelvic pain, dysmenorrhea, dyspareunia, and infertility (21), and its diagnosis requires a surgical intervention that is accompanied by risks and complications for women (22). The lack of non-invasive diagnostic tests for endometriosis leads to a considerable delay in disease detection (3). So, developing non-invasive diagnostic biomarkers for endometriosis is an urgent need. In this context, blood-based biomarkers can be helpful. In recent years, most efforts for finding non-invasive diagnostic biomarkers for endometriosis have been directed towards blood miRNAs. Also, it seems that circulating mRNAs can be useful as potential biomarkers (5, 7). Recently, it has been indicated that diffusion RNAs are protected by lipoprotein complexes or phospholipids. Thus, these molecules

Table 2. Participants' Characteristics in This Study.

Characteristics	Endometriosis Women (n = 24)	Non-endometriosis Women (n = 24)
Age (y, mean \pm SD)	33.5 \pm 5.1	37.5 \pm 9.1
Cycle phase, No. (%)		
Follicular	20 (83)	19 (79)
Luteal	4 (17)	5 (21)
ARSM stage, No. (%)		
III	10 (42)	NA
IV	14 (58)	NA
Other diagnoses, No. (%)		
Healthy	NA	3 (13)
Ovarian cysts	NA	8 (33)
Myoma	NA	11 (46)
Other	NA	2 (8)

Abbreviations: ARSM, American Society of Reproductive Medicine; NA, Not Available.

are stable in circulation despite the high amounts of the RNases present in blood (23). It has been shown that various kinds of RNAs, including mRNAs and non-coding RNAs, can be extracted and detected in body fluids such as plasma and serum (24, 25). Indeed, several investigations have shown that cell-free mRNAs in plasma can be promising non-invasive diagnostic tools for cancer (26, 27). Hence, in this study, the plasma levels of MMP-9 and VEGF-A mRNAs, as the genes relevant to endometriosis pathogenesis, were evaluated by RT-qPCR.

In this study, for the first time, we measured plasma MMP-9 mRNA level via qRT-PCR in women with endometriosis. Our results showed a significant increase in serum MMP-9 mRNA in endometriosis compared with non-endometriosis women. Moreover, this study showed that the expression of MMP-9 was independent of the menstrual phase. Interestingly, endometriosis, as a benign disease, shares a common feature with malignancies, and that is its invasive behavior (28). It has been shown that MMP-9, as a matrix metalloproteinase, is an essential factor for the degradation of the extracellular matrix (ECM) and the basement membrane and a crucial factor for cells' invasive behaviors (29). Several studies have reported that MMP-9 can induce invasiveness in the endometrial tissue, leading to the degradation of ECM, invading other locations, and creating ectopic endometriosis lesions (30, 31). There are pieces of evidence suggesting that the MMP-9 gene is significantly overexpressed in the endometriosis tissue, which can be expected according to its role in endometriosis pathogenesis (13, 27). The elevation of MMP-9 mRNA in the plasma of endometriosis patients, observed in this study, was consistent with the results

of previous investigations on the expression MMP-9 in the endometriosis tissue, suggesting its potential as a non-invasive diagnostic biomarker for endometriosis.

This study showed no significant difference in plasma VEGF-A mRNA level between endometriosis and non-endometriosis women. Moreover, we showed that VEGF-A mRNA level positively correlated with the proliferative phase of the participants' menstrual cycles. This molecule is an essential factor for physiological and pathological angiogenesis and induces vascular endothelial cells' proliferation and migration (18). Angiogenesis plays a significant role in endometriosis progression because, similar to tumors' behaviors and their metastatic spread, endometriosis lesions require neovascularization to survive in ectopic locations. Indeed, the establishment, proliferation, and survival of endometriosis lesions depend on an adequate blood supply (12). Thus, regarding the role of VEGF-A in endometriosis pathogenesis, the overexpression of its gene in the endometriosis tissue can be expected, as shown in a previous study (12). According to our results, however, the plasma level of VEGF-A did not show a significant difference between endometriosis patients and controls. It seems that the plasma level of VEGF-A mRNA, in contrast to its expected role in endometriosis pathogenesis, cannot be a useful blood-based biomarker for discriminating endometriosis in women. Furthermore, our finding regarding the comparable levels of VEGF-A mRNA in plasma samples from endometriosis and non-endometriosis women was inconsistent with that of a previous study, indicating a significantly higher level of VEGF-A mRNA in endometriosis patients (32). This discrepancy may be due to the limited number of control subjects in the mentioned study (i.e.,

only 10 women, mostly infertile) and different samples in the two studies (serum vs. plasma).

We must address some limitations of the present study. Our sample size was relatively small, and the patients mainly presented stage III and IV endometriosis. Hence, large-scale studies should be performed to clarify the applicability of MMP-9 and VEGF-A mRNAs as non-invasive diagnostic biomarkers for endometriosis.

5.1. Conclusion

In the present study, the levels of MMP-9 and VEGF-A mRNAs, as non-invasive diagnostic biomarkers, were evaluated by RT-qPCR in the plasma samples of women endometriosis. A significant difference was detected in plasma MMP-9, but not VEGF-A, mRNA level between endometriosis and non-endometriosis women. In conclusion, it can be suggested that MMP-9 mRNA has the potential for being used as a diagnostic biomarker for endometriosis, and it can be considered for further investigations along with other mRNAs. However, VEGF-A seemed to be inapplicable as a non-invasive diagnostic biomarker despite its role in endometriosis pathogenesis.

Footnotes

Authors' Contribution: Concept and design: S. A. and P. I.; Collecting and interpreting clinical data and revising the manuscript: S. A., P. I., S. CH., and SH. N.; Statistical analysis: S. Y., S. A., and P. I.; Drafting the manuscript: S. A. and P. I. All authors read and approved the final manuscript.

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References

- Hirsch M, Dhillon-Smith R, Cutner AS, Yap M, Creighton SM. The Prevalence of Endometriosis in Adolescents with Pelvic Pain: A Systematic Review. *J Pediatr Adolesc Gynecol*. 2020;**33**(6):623–30. doi: [10.1016/j.jpog.2020.07.011](https://doi.org/10.1016/j.jpog.2020.07.011). [PubMed: 32736134].
- DiVasta AD, Vitonis AF, Laufer MR, Missmer SA. Spectrum of symptoms in women diagnosed with endometriosis during adolescence vs adulthood. *Am J Obstet Gynecol*. 2018;**218**(3):324 e1–324 e11. doi: [10.1016/j.ajog.2017.12.007](https://doi.org/10.1016/j.ajog.2017.12.007). [PubMed: 29247637].
- Saunders PTK, Horne AW. Endometriosis: Etiology, pathobiology, and therapeutic prospects. *Cell*. 2021;**184**(11):2807–24. doi: [10.1016/j.cell.2021.04.041](https://doi.org/10.1016/j.cell.2021.04.041). [PubMed: 34048704].
- Nisenblat V, Bossuyt PM, Farquhar C, Johnson N, Hull ML. Imaging modalities for the non-invasive diagnosis of endometriosis. *Cochrane Database Syst Rev*. 2016;**2**. CD009591. doi: [10.1002/14651858.CD009591.pub2](https://doi.org/10.1002/14651858.CD009591.pub2). [PubMed: 26919512]. [PubMed Central: PMC7100540].
- Nisenblat V, Bossuyt PM, Shaikh R, Farquhar C, Jordan V, Schefers CS, et al. Blood biomarkers for the non-invasive diagnosis of endometriosis. *Cochrane Database Syst Rev*. 2016;(5). CD012179. doi: [10.1002/14651858.CD012179](https://doi.org/10.1002/14651858.CD012179). [PubMed: 27132058]. [PubMed Central: PMC7076288].
- Tzimagiorgis G, Michailidou EZ, Kritis A, Markopoulos AK, Kouidou S. Recovering circulating extracellular or cell-free RNA from bodily fluids. *Cancer Epidemiol*. 2011;**35**(6):580–9. doi: [10.1016/j.canep.2011.02.016](https://doi.org/10.1016/j.canep.2011.02.016). [PubMed: 21514265].
- Zafari N, Tarafdari AM, Izadi P, Noruzinia M, Yekaninejad MS, Bahramy A, et al. A Panel of Plasma miRNAs 199b-3p, 224-5p and Let-7d-3p as Non-Invasive Diagnostic Biomarkers for Endometriosis. *Reprod Sci*. 2021;**28**(4):991–9. doi: [10.1007/s43032-020-00415-z](https://doi.org/10.1007/s43032-020-00415-z). [PubMed: 33398851].
- Ng EK, Tsui NB, Lam NY, Chiu RW, Yu SC, Wong SC, et al. Presence of filterable and nonfilterable mRNA in the plasma of cancer patients and healthy individuals. *Clin Chem*. 2002;**48**(8):1212–7. [PubMed: 12142376].
- Sueoka E, Sueoka N, Iwanaga K, Sato A, Suga K, Hayashi S, et al. Detection of plasma hnRNP B1 mRNA, a new cancer biomarker, in lung cancer patients by quantitative real-time polymerase chain reaction. *Lung Cancer*. 2005;**48**(1):77–83. doi: [10.1016/j.lungcan.2004.10.007](https://doi.org/10.1016/j.lungcan.2004.10.007). [PubMed: 15777973].
- Wang H, Zhang X, Wang L, Zheng G, Du L, Yang Y, et al. Investigation of cell free BIRC5 mRNA as a serum diagnostic and prognostic biomarker for colorectal cancer. *J Surg Oncol*. 2014;**109**(6):574–9. doi: [10.1002/jso.23526](https://doi.org/10.1002/jso.23526). [PubMed: 24338523].
- Dalle Carbonare L, Frigo A, Francia G, Davi MV, Donatelli L, Stranieri C, et al. Runx2 mRNA expression in the tissue, serum, and circulating non-hematopoietic cells of patients with thyroid cancer. *J Clin Endocrinol Metab*. 2012;**97**(7):E1249–56. doi: [10.1210/jc.2011-2624](https://doi.org/10.1210/jc.2011-2624). [PubMed: 22511796].
- Cardoso JV, Abrao MS, Vianna-Jorge R, Ferrari R, Berardo PT, Machado DE, et al. Combined effect of vascular endothelial growth factor and its receptor polymorphisms in endometriosis: a case-control study. *Eur J Obstet Gynecol Reprod Biol*. 2017;**209**:25–33. doi: [10.1016/j.ejogrb.2016.10.046](https://doi.org/10.1016/j.ejogrb.2016.10.046). [PubMed: 27836223].
- Yang M, Jiang C, Chen H, Nian Y, Bai Z, Ha C. The involvement of osteopontin and matrix metalloproteinase-9 in the migration of endometrial epithelial cells in patients with endometriosis. *Reprod Biol Endocrinol*. 2015;**13**:95. doi: [10.1186/s12958-015-0090-4](https://doi.org/10.1186/s12958-015-0090-4). [PubMed: 26289107]. [PubMed Central: PMC4545920].
- Danastas K, Miller EJ, Hey-Cunningham AJ, Murphy CR, Lindsay LA. Expression of vascular endothelial growth factor A isoforms is dysregulated in women with endometriosis. *Reprod Fertil Dev*. 2018;**30**(4):651–7. doi: [10.1071/RD17184](https://doi.org/10.1071/RD17184). [PubMed: 29017687].
- Zhang L, Xiong W, Xiong Y, Liu H, Li N, Du Y, et al. Intracellular Wnt/Beta-Catenin Signaling Underlying 17beta-Estradiol-Induced Matrix Metalloproteinase 9 Expression in Human Endometriosis. *Biol Reprod*. 2016;**94**(3):70. doi: [10.1095/biolreprod.115.135574](https://doi.org/10.1095/biolreprod.115.135574). [PubMed: 26888969].
- Mott JD, Werb Z. Regulation of matrix biology by matrix metalloproteinases. *Curr Opin Cell Biol*. 2004;**16**(5):558–64. doi: [10.1016/j.ccb.2004.07.010](https://doi.org/10.1016/j.ccb.2004.07.010). [PubMed: 15363807]. [PubMed Central: PMC2775446].
- Weigel MT, Kramer J, Schem C, Wenners A, Alkatout I, Jonat W, et al. Differential expression of MMP-2, MMP-9 and PCNA in endometriosis and endometrial carcinoma. *Eur J Obstet Gynecol Reprod Biol*. 2012;**160**(1):74–8. doi: [10.1016/j.ejogrb.2011.09.040](https://doi.org/10.1016/j.ejogrb.2011.09.040). [PubMed: 22056701].

18. May K, Becker CM. Endometriosis and angiogenesis. *Minerva Ginecol.* 2008;**60**(3):245–54. [PubMed: [18547987](#)].
19. Mabrouk M, Elmakky A, Caramelli E, Farina A, Mignemi G, Venturoli S, et al. Performance of peripheral (serum and molecular) blood markers for diagnosis of endometriosis. *Arch Gynecol Obstet.* 2012;**285**(5):1307–12. doi: [10.1007/s00404-011-2122-4](#). [PubMed: [22065163](#)].
20. De Sanctis P, Elmakky A, Farina A, Caramelli E, Seracchioli R, Mabrouk M, et al. Matrix metalloproteinase-3 mRNA: a promising peripheral blood marker for diagnosis of endometriosis. *Gynecol Obstet Invest.* 2011;**71**(2):118–23. doi: [10.1159/000320752](#). [PubMed: [21150162](#)].
21. Dun EC, Kho KA, Morozov VV, Kearney S, Zurawin JL, Nezhat CH. Endometriosis in adolescents. *JSLs.* 2015;**19**(2). doi: [10.4293/JSLs.2015.00019](#). [PubMed: [26005317](#)]. [PubMed Central: [PMC4432718](#)].
22. Lier MCI, Vlek SL, Ankersmit M, van de Ven PM, Dekker J, Bleeker MCG, et al. Comparison of enhanced laparoscopic imaging techniques in endometriosis surgery: a diagnostic accuracy study. *Surg Endosc.* 2020;**34**(1):96–104. doi: [10.1007/s00464-019-06736-8](#). [PubMed: [31028547](#)]. [PubMed Central: [PMC6946762](#)].
23. Mitchell PS, Parkin RK, Kroh EM, Fritz BR, Wyman SK, Pogosova-Agadjanyan EL, et al. Circulating microRNAs as stable blood-based markers for cancer detection. *Proc Natl Acad Sci U S A.* 2008;**105**(30):10513–8. doi: [10.1073/pnas.0804549105](#). [PubMed: [18663219](#)]. [PubMed Central: [PMC2492472](#)].
24. Schwarzenbach H, Hoon DS, Pantel K. Cell-free nucleic acids as biomarkers in cancer patients. *Nat Rev Cancer.* 2011;**11**(6):426–37. doi: [10.1038/nrc3066](#). [PubMed: [21562580](#)].
25. Tsui NB, Ng EK, Lo YM. Stability of endogenous and added RNA in blood specimens, serum, and plasma. *Clin Chem.* 2002;**48**(10):1647–53. [PubMed: [12324479](#)].
26. March-Villalba JA, Martinez-Jabaloyas JM, Herrero MJ, Santamaria J, Alino SF, Dasi F. Cell-free circulating plasma hTERT mRNA is a useful marker for prostate cancer diagnosis and is associated with poor prognosis tumor characteristics. *PLoS One.* 2012;**7**(8). e43470. doi: [10.1371/journal.pone.0043470](#). [PubMed: [22916267](#)]. [PubMed Central: [PMC3423343](#)].
27. Cao W, Zhou D, Tang W, An H, Zhang Y. Discovery of plasma messenger RNA as novel biomarker for gastric cancer identified through bioinformatics analysis and clinical validation. *PeerJ.* 2019;**7**:e7025. doi: [10.7717/peerj.7025](#). [PubMed: [31249732](#)]. [PubMed Central: [PMC6587939](#)].
28. Zeitvogel A, Baumann R, Starzinski-Powitz A. Identification of an invasive, N-cadherin-expressing epithelial cell type in endometriosis using a new cell culture model. *Am J Pathol.* 2001;**159**(5):1839–52. doi: [10.1016/S0002-9440\(10\)63030-1](#). [PubMed: [11696444](#)]. [PubMed Central: [PMC1867070](#)].
29. Chung HW, Wen Y, Chun SH, Nezhat C, Woo BH, Lake Polan M. Matrix metalloproteinase-9 and tissue inhibitor of metalloproteinase-3 mRNA expression in ectopic and eutopic endometrium in women with endometriosis: a rationale for endometriotic invasiveness. *Fertil Steril.* 2001;**75**(1):152–9. doi: [10.1016/S0015-0282\(00\)01670-8](#). [PubMed: [11163831](#)].
30. Shaco-Levy R, Sharabi S, Benharroch D, Piura B, Sion-Vardy N. Matrix metalloproteinases 2 and 9, E-cadherin, and beta-catenin expression in endometriosis, low-grade endometrial carcinoma and non-neoplastic eutopic endometrium. *Eur J Obstet Gynecol Reprod Biol.* 2008;**139**(2):226–32. doi: [10.1016/j.ejogrb.2008.01.004](#). [PubMed: [18295959](#)].
31. Mousazadeh S, Khaksar F. Gene Expression Levels of Gelatinases in Endometriosis. *Stud Med Sci.* 2018;**28**(12):795–804.
32. Acimovic M, Vidakovic S, Milic N, Jeremic K, Markovic M, Milosevic-Djeric A, et al. Survivin and VEGF as Novel Biomarkers in Diagnosis of Endometriosis. *J Med Biochem.* 2016;**35**(1):63–8. doi: [10.1515/jomb-2015-0005](#). [PubMed: [28356866](#)]. [PubMed Central: [PMC5346803](#)].