



Stepwise Regression Analysis of Citrus Genotype Under Cold Stress

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Abstract

Background: Citrus is among the cold temperature (freezing) susceptible plants. The performance of storage and breeding of horticultural crops is mainly dependent on the correlation among the traits.

Objectives: The present study was conducted to identify the physiological and biochemical traits of some citrus cultivars with the greatest influence on stress tolerance and determine the direct and indirect effects of these traits on the median Lethal Time (LT₅₀) using integrated pathway analysis (i.e., stepwise regression and causality coefficient).

Methods: Pathway analysis (i.e., stepwise regression and causality coefficient) based on additive models, which is among the robust statistical methods to describe the relationship among the traits, was used to investigate the physiological and biochemical characteristics of four citrus cultivars (i.e., Japanese mandarin, Thomson orange, Ruby Star grapefruit, and Lisbon lemon) at temperature levels (-8, -4, and +4°C) in a completely randomized design with three replications in 2021.

Results: The results showed that Japanese mandarin and Thomson orange cultivars had the highest stress tolerance (LT₅₀). The results of the correlation coefficient demonstrated that the highest positive and significant correlations belonged to the LT₅₀ with total flavonoid (0.443**), chlorophyll a with chlorophyll b (0.613**), carotenoid with chlorophyll a (0.929**), chlorophyll b (0.573**), and total chlorophyll (0.849**), relative moisture content with malondialdehyde (0.559**), glycine-betaine with catalase (0.919**), hydrogen peroxide with total flavonoid (0.405**), and catalase (0.611**).

Conclusions: The results of stepwise regression for all traits indicated the importance and critical role of total flavonoid, proline, and LT₅₀ characteristics.

Keywords: Citrus, LT₅₀, Stepwise Regression, Stress Tolerance

1. Background

The world population is proliferating, and it is predicted to reach 9.5 billion individuals by 2050. On the other hand, food production diminishes due to various abiotic stresses (1). Therefore, the facilitation of the yield loss in crops is among the global concerns for the countries; therefore, they can supply human food requirements (2). Crops are mainly affected by stresses, such as cold temperature (3), salinity (4), drought (5), flooding (6), oxidative stress (7), pathogens (8), and heavy metal toxicity (9). Human activities have exacerbated the present stress-inducing factors. All these stresses are menaces to plants and prevent them from achieving their maximum genetic potential. They have also limited crop production all over the world.

Citrus is among the three significant fruits in the world (10). Citrus cultivation, especially orange, is a major industry and an essential part of the United States, Brazil, Mex-

ico, China, India, Iran, and numerous Mediterranean countries, such as Spain and Greece (11). Citrus is cultivated in more than 100 countries, mainly situated in tropical and subtropical regions with suitable soil and climate (12). Citrus is valuable both in terms of fresh consumption and in the processing industry. The citrus processing industry has focused on juice and essence production for years. It is estimated that 33% of citrus harvested globally is used for juice production (11). Iran is ranked 5th regarding citrus production worldwide, and climatic diversity and production of the best citrus varieties have given the country a good advantage. Citrus is produced in seven provinces of Iran. Mazandaran province is regarded as the main center of citrus production with 1.5 - 1.7 million tons annually, followed by Golestan, Guilan, Hormozgan, and Kerman provinces.

Weather fluctuation is one of the significant obstacles to the production of crops, especially citrus (13). Low temperature is a challenging factor for commercial citrus production, as citrus species are sensitive to low temperatures

and harvest in winter (14). Various citrus species are susceptible to -2.2°C and lower temperatures. However, some commercial citrus species are cultivated in regions with an increased risk of freezing (15). The high risk of freezing in winter during citrus growth is a big issue in the Caspian Seabank, along the coasts of the Black Sea, coastal regions of France, the Adriatic Sea in Yugoslavia, some parts of Turkey, Spain, Israel, and, Greece. A sudden decline in the below-freezing temperature threshold renders the trees susceptible to freezing. Although citrus growers mitigate the freezing risk using various methods, such as sprinkler irrigation, protective materials, and costly measures, the development of cold-tolerant citrus cultivars using genetic engineering methods provides long-term protection against cold (16, 17).

Advances in biotechnology and identification of the genes regulating tolerance to different environmental conditions, such as cold temperature, using genetic engineering and their introduction into the commercial citrus cultivars might be a solution for this issue (18). Noting the periodical occurrence of chilling stress in the north of Iran (2007, 2013, 2016, and 2018) and the resulting economic and social challenges, efforts to alleviate this damage seem necessary. Due to the generic nature of cold stress tolerance (polygenic) in citrus species, it appears that cold patience and knowledge about the mechanisms involved in it to select suitable cultivars are among the significant priorities for citrus breeding programs in subtropical regions. The use of suitable cultivars is a reasonable and cheap way to reach sustainable production in an area, and the selection of tolerant species and implementation of good farming practices might also be effective.

There are various methods for the analysis of traits, and the researcher chooses one of them according to their aims. Simple correlation, regression models, and pathway analysis are commonly used to analyze physiological traits (19). The investigation of stress tolerance relationships and their interactions is critically important to obtain high levels of tolerance (20). In general, the correlation coefficient is mainly used to describe the relationship among traits. However, this coefficient might be misleading in some instances, as a high correlation between two traits might be due to the indirect effects of traits on each other (21). In these types of studies, selection based on simple correlation alone cannot provide optimum results. Therefore, it is essential to determine the direct and indirect effects of some traits affecting other important traits. In this regard, stepwise regression and pathway analysis are of utmost importance, and various studies have been performed to investigate the causality relationships of traits (22). In stepwise regression, all independent variables enter the model, and those independent variables with no significant ef-

fect on the dependent variable will be removed from the model. This method is a combination of forward and backward regressions. In forward regression, no variable is present in the model at the beginning, and the first variable entering the model is the one with the highest correlation with the dependent variable (23).

Stress tolerance or the median lethal dose (LT_{50}) is a quantitative trait controlled by multiple genes. This trait is highly influenced by the environment. Various characteristics affect the LT_{50} alone or in combination with each other (24). For the purpose of the determination of the role of physiological traits in improved stress tolerance and increased selection efficiency by a limited number of traits, which are important parameters to achieve breeding goals, stepwise selection of variables in a multiple linear regression might be used (25). The investigation of the traits affecting stress tolerance in citrus using stepwise regression analysis is not available in the literature.

2. Objectives

The present study was conducted to identify the physiological and biochemical traits of some citrus cultivars with the greatest influence on stress tolerance and determine the direct and indirect effects of these traits on the LT_{50} using integrated pathway analysis (i.e., stepwise regression and causality coefficient). The findings of the present study might be used to develop breeding programs to improve citrus stress tolerance.

3. Methods

3.1. Experiment Conditions and Treatments

The present study was conducted at the Horticulture Laboratory of Guilan University in 2021 to investigate the biochemical traits of four citrus species (i.e., Japanese mandarin, Thomson orange, Ruby Star grapefruit, and Lisbon lemon) at three temperature levels (i.e., -8 , -4 , and $+4^{\circ}\text{C}$) (Table 1). Two-year-old seedlings were selected and placed in 10 kg pots filled with the typical soil of the region (i.e., silty loam texture) (26). Before the application of treatments and for the adaptation of the plant material to cold, the seedlings were transferred to temperature conditions (27) and then to a storage room with a $+4^{\circ}\text{C}$ temperature and $65 \pm 5\%$ relative humidity. This relative humidity was chosen according to the moisture required for citrus growth in the North of Iran. The temperature of the test chamber was $+4^{\circ}\text{C}$, which decreased by 1.5°C every hour. The samples were kept at the aforementioned temperature treatments for 10 hours when the test chamber device reached that temperature, and the traits were measured after this period (i.e., 10 hours) (28).

Table 1. Experiment Treatments

Treatment Code	Citrus Genotype	Temperature Levels (°C)
1	Japanese mandarin	+4
2	Thomson orange	+4
3	Ruby Star grapefruit	+4
4	Lisbon lemon	+4
5	Japanese mandarin	-4
6	Thomson orange	-4
7	Ruby Star grapefruit	-4
8	Lisbon lemon	-4
9	Japanese mandarin	-8
10	Thomson orange	-8
11	Ruby Star grapefruit	-8
12	Lisbon lemon	-8

3.2. Measurement of Traits

Ion leakage, relative water content, and LT_{50} traits were immediately measured after the application of each temperature treatment. For the measurement of the traits, such as leaf chlorophyll and carotenoid, total flavonoid, peroxidation of membrane lipids, and the antioxidant activity of catalase, superoxide dismutase, and ascorbate peroxidase after the induction of stress, the samples were immediately frozen in liquid nitrogen and kept at -80°C to be measured later. The biochemical traits were measured according to [Table 2](#).

3.3. Statistical Analysis

Prior to performing the analysis of variance on the data, the Roc Univariate command was employed to check the normality of data distribution, and ensuring this issue was followed by the analysis of descriptive statistics and correlation coefficient. On the other hand, stepwise regression analysis was employed at a significance level of 1% to specify the key traits affecting stress tolerance in these cultivars. The causal analysis method (based on the traits with the most justification in stress tolerance) was used to specify the direct and indirect effects of each of the traits included in the stress tolerance model. All the aforementioned calculations (i.e., checking the normality of data distribution, determining simple correlation coefficients, stepwise regression analysis, and causal analysis) were conducted using SPSS software (version 24).

4. Results

[Table 3](#) shows descriptive statistics related to the studied traits. These results indicated a suitable variation

Table 2. Measurement of the Studied Traits

Studied Trait	Measurement Method
Total flavonoid	Campos et al. (29)
Malondialdehyde	Campos et al. (29)
Ascorbate peroxidase	Maehly and Chance (30)
Catalase	Clairborne (31)
Chlorophyll a	Barnes et al. (32)
Chlorophyll b	Barnes et al. (32)
Total chlorophyll	Barnes et al. (32)
Carotenoid	Barnes et al. (32)
Superoxide dismutase	Wu et al. (33)
RWC	Ritchie et al. (34)
Ion leakage	Sullivan and Ross (35)
LT_{50}	Lim et al. (36)
Anthocyanin	Wagner (37)
Proline	Bates et al. (38)
Glycine-betaine	Grieve and Grattan (39)
Hydrogen peroxide	Alexieva (40)

Abbreviations: RWC, relative water content; LT_{50} , median lethal dose

among the studied populations. According to the standard deviation value, relative moisture, superoxide dismutase, and ascorbate peroxidase traits had the highest variations. Relative moisture content had a range of variation equal to 179.5. This value was reported as 109 and 55 for superoxide dismutase and ascorbate peroxidase, respectively, indicating a high variation. In this study, the effect of citrus cultivar and temperature on the biochemical traits was studied for the first time. The results showed that the traits of various citrus cultivars had a wide range of biochemical properties. Descriptive statistics show general information on the studied traits and help the researchers to have a better and more precise knowledge of the studied traits.

4.1. Correlation Coefficients

The correlation coefficient was used to determine the extent of the relationship between the linear changes in the two traits. This coefficient described the degree of linear relationship and direction of changes between two traits. [Table 4](#) shows Pearson's correlation coefficients among the studied traits. The results showed that the LT_{50} with total flavonoid (0.443**), chlorophyll a with chlorophyll b (0.613**), carotenoid with chlorophyll a (0.929**), chlorophyll b (0.573**), and total chlorophyll (0.849**), relative moisture content with malondialdehyde (0.559**), glycine-betaine with catalase (0.919**), hydrogen peroxide with total flavonoid (0.405**), and catalase (0.611**) had the

Table 3. Descriptive Statistics Related to Various Traits in Citrus Cultivars

Trait	Mean \pm Standard Variation	Range	Minimum	Maximum
Total flavonoid	11.92 \pm 9.55	30.36	3.06	33.42
Ascorbate peroxidase	31.55 \pm 15.75	55	12	67
Catalase	30.42 \pm 3.01	10.5	26.6	37.1
Malondialdehyde	0.66 \pm 0.35	1.4	0.22	1.62
Chlorophyll a	3.18 \pm 3.11	10.45	0.18	10.63
Chlorophyll b	0.83 \pm 0.7	2.9	0.16	3.06
Total chlorophyll	4.02 \pm 3.52	12.5	0.73	13.22
Carotenoid	1.56 \pm 1.11	3.52	0.34	3.86
Superoxide dismutase	126.4 \pm 29.84	109	53	162
RWC	93.12 \pm 56.64	179.5	28	207.5
Ion leakage	26.7 \pm 17.1	54.93	13.89	68.82
LT ₅₀	5.54 \pm 3.13	10.73	12	1.27
Anthocyanin	0.164 \pm 0.07	0.202	0.035	0.238
Proline	13.2 \pm 3.57	15.3	7	22.3
Glycine-betaine	6.22 \pm 4.11	12.94	1.91	14.85
Hydrogen peroxide	0.24 \pm 0.14	0.47	0.035	0.51

Abbreviations: RWC, relative water content; LT₅₀, median lethal dose

highest positive and significant correlations, which indicated that improvement in each of these traits might lead to enhanced stress tolerance.

4.2. Stepwise Regression

Stepwise regression was first performed to determine the most important traits influencing the LT₅₀, total flavonoid, and proline and the elimination of negligible variables and pathway analysis. Table 5 shows the results of stepwise regression with the LT₅₀ as the independent variable and other traits as the dependent variables. The LT₅₀, total flavonoid, and proline with partial regression coefficients of 88.5, 96%, and 93.8% entered the model, respectively. In general, the first and second variables that entered the model were total flavonoid and proline, which had the highest share in the description of the LT₅₀. In addition, due to the high correlation coefficient of these traits with the LT₅₀, they are the most important traits in this study and should be noted in citrus breeding programs.

4.3. Path Analysis

The variables that entered the model were subjected to pathway analysis to further interpret the results. Table 6 shows that all the coefficients of the direct path (except for proline) are significant in the final pattern. Overall, these traits described 77.5% of changes in the LT₅₀. Table 6 shows

the direct and indirect effects of the traits on the LT₅₀, according to which total flavonoid and proline had the highest positive and significant effect on the other biochemical traits.

5. Discussion

Cold stress can be classified as chilling (0 - 15°C) and freezing (< 0°C) stresses. Generally, plants originating from temperate regions, such as spinach and arabidopsis, exhibit a variable degree of chilling tolerance and can increase their freezing tolerance during exposure to chilling and nonfreezing temperatures. This process is known as cold acclimation (41). On the other hand, plants of subtropical origins are sensitive to chilling stress and lack the cold acclimation mechanism (42). Information regarding the physiological traits of such stepwise regression will provide the knowledge to develop cold-tolerant cultivars.

Knowledge of various physiological aspects helps researchers to select breeding strategies for citrus. The present study was applied research to facilitate the selection of the best citrus genotypes. Overall, the results of Pearson's correlation, stepwise regression, and pathway analysis confirmed each other.

Pearson's correlation coefficients described the degree of linear relationship and direction of changes between

Table 4. Pearson's Correlation Coefficients of the Studied Traits in Citrus Cultivars

Traits	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
1) Total flavonoids	1															
2) AXP	0.211 ^a	1														
3) CAT	0.081 ^a	0.765 **	1													
4) Malondialdehyde	0.027 ^a	0.108 ^a	0.236 ^a	1												
5) Chlorophyll a	0.270 ^a	0.262 ^a	0.106 ^a	0.457 **	1											
6) Chlorophyll b	0.142 ^a	0.136 ^a	0.056 ^a	0.141 ^a	0.613 **	1										
7) Total chlorophyll	0.303 ^a	0.316 ^a	0.138 ^a	0.356 *	0.835 **	0.543 **	1									
8) Carotenoid	0.202 ^a	0.464 **	0.114 *	0.464 **	0.929 **	0.573 **	0.849 **	1								
9) SOD	0.134 ^a	0.179 ^a	0.343 *	0.050 ^a	0.380 *	0.184 ^a	0.429 **	0.386 *	1							
10) RWC	0.255 ^a	0.185 ^a	0.040 ^a	0.559 **	0.114 ^a	0.094 ^a	0.001 ^a	0.103 ^a	0.061 ^a	1						
11) Ion leakage	0.310 ^a	-0.328 ^a	0.481 **	0.048 ^a	0.354 *	0.237 ^a	0.178 ^a	0.246 ^a	0.289 ^a	0.639 **	1					
12) LT ₅₀	0.443 **	-0.165 ^a	0.237 ^a	0.332 *	0.101 ^a	0.162 ^a	0.187 ^a	0.095 ^a	0.264 ^a	0.204 ^a	0.115 ^a	1				
13) Anthocyanin	0.201 ^a	0.572 **	0.784 **	0.056 ^a	0.096 ^a	0.120 ^a	0.118 ^a	0.092 ^a	0.093 ^a	0.042 ^a	0.418 *	0.308 ^a	1			
14) Proline	0.013 ^a	0.157 ^a	0.464 **	0.495 **	0.113 ^a	0.217 ^a	0.227 ^a	0.087 ^a	0.252 ^a	0.138 ^a	0.227 ^a	0.264 ^a	0.347 *	1		
15) Glycine-betaine	0.284 ^a	0.646 **	0.919 **	0.239 ^a	0.130 ^a	0.109 ^a	0.167 ^a	0.059 ^a	0.304 ^a	0.13 ^a	0.411 *	0.402 *	0.804 **	0.464 **	1	
16) Hydrogen peroxide	0.405 *	0.161 ^a	0.611 **	0.133 ^a	0.562 **	0.121 ^a	0.526 **	0.376 *	0.341 *	0.181 ^a	0.429 *	0.154 ^a	0.520 **	0.156 ^a	0.561 **	1

Abbreviations: RWC, relative water content; LT₅₀, median lethal dose; AXP, ascorbate peroxidase; CAT, catalase; SOD, superoxide dismutase
^a Non-significant

Table 5. Results of Stepwise Regression for Median Lethal Dose, Total Flavonoid, and Proline as Dependent Variables and Other Traits as Independent Variables

Stage of Variable Entry to Model	Entered Variable	Partial Regression Coefficient	Standard Error	F	Coefficient of Determination (R ²)
1	Total flavonoid	0.960	0.46	17.53 ^a	0.869
2	Proline	0.938	1.64	9.744 ^a	0.880
3	LT ₅₀	0.885	1.84	6.102 ^a	0.783

Abbreviation: LT₅₀, median lethal time

^a Significant at P < 0.01

Table 6. Direct and Indirect Effects of Median Lethal Dose, Total Flavonoid, and Proline

Variable	Direct Effect	Indirect Effect			
		Total Flavonoid	Proline	LT ₅₀	Residual Effects
Total flavonoid	0.608 **	-	-0.374 ^a	1.465 **	1.091 **
Proline	0.559 **	-0.061 ^a	-	0.079 ^a	0.018 ^a
LT ₅₀	0.414 **	0.146 **	0.236 ^a	-	0.382 **

Abbreviation: LT₅₀, median lethal time

^a Non-significant

two traits. Quantification is very important in breeding (25). Table 4 shows Pearson's correlation coefficients of the studied traits. The results showed that the LT₅₀ with total flavonoid, chlorophyll a with chlorophyll b, carotenoid with chlorophyll a, chlorophyll b, and total chlorophyll, relative moisture content with malondialdehyde, glycine-betaine with catalase, hydrogen peroxide with total flavonoid, and catalase had the highest positive and significant correlations, which indicated that improvement in each of these traits might lead to enhanced

stress tolerance. Abouzari et al. (43) showed a positive and significant correlation between ion leakage and leaf water core in citrus cultivars. A positive correlation between proline and carbohydrates was reported in Page mandarin by Tadjvar et al. (26). Table 4 shows a significant positive correlation between malondialdehyde and proline content (0.99; P < 0.01); the aforementioned results are consistent with the results of a study performed by Li et al. (41).

Correlation between the traits might help breeders in indirect selection for important stress traits through other

traits that are easier to measure (44). Although the correlation coefficients of physiological traits help determine the traits related to stress, they fail to describe the relationship correctly, and it is required to determine the direct and indirect effects of these traits (44, 45). Therefore, step-wise regression is used to select valuable variables among numerous measure traits to identify traits with the highest share in the description of stress tolerance (46). This method can eliminate traits that are ineffective or have a negligible effect on tolerance in the regression model and only identify traits that significantly describe the changes (47). In this study, due to the numerousness of traits with significant and positive correlation with the LT_{50} , it is reasonable to further investigate the traits using other statistical methods to determine the important traits affecting citrus performance. Therefore, integrated pathway analysis was used for further evaluation to obtain more sound information regarding the traits and their effect on citrus LT_{50} .

5.1. Conclusions

Based on the results of this study, it was concluded that there is a clear link between the expression of the LT_{50} and other physiological traits examined under cold stress at -8°C . Total flavonoid and proline show a positive correlation with the LT_{50} under cold stress in citrus. The aforementioned results provide necessary information for understanding the mechanism of the stress response and the adaptation of citrus to cold conditions. This study will also help breeders and molecular biologists to evaluate existing germplasm for cold tolerance and/or develop new stress-resistant cultivars.

Footnotes

Authors' Contribution: A. S. S. designed the idea and carried out the experiments. M. S. and M. K. S. analyzed and wrote the article. M. G. helped with analyzing and editing the manuscript.

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References

- Malhi GS, Kaur M, Kaushik P, Alyemeni MN, Alsahli AA, Ahmad P. Arbuscular mycorrhiza in combating abiotic stresses in vegetables: An eco-friendly approach. *Saudi J Biol Sci.* 2021;**28**(2):1465–76. doi: [10.1016/j.sjbs.2020.12.001](https://doi.org/10.1016/j.sjbs.2020.12.001). [PubMed: [33613074](https://pubmed.ncbi.nlm.nih.gov/33613074/)]. [PubMed Central: [PMC7878692](https://pubmed.ncbi.nlm.nih.gov/PMC7878692/)].
- Sharma RK, Meena RP, Chhokar RS, Singh S, Khokhar J, Yadav VK, et al. Increase in wheat production through management of abiotic stresses : A review. *J Appl Nat Sci.* 2015;**7**(2):1070–80. doi: [10.31018/jans.v7i2.733](https://doi.org/10.31018/jans.v7i2.733).
- Liu L, Ji H, An J, Shi K, Ma J, Liu B, et al. Response of biomass accumulation in wheat to low-temperature stress at jointing and booting stages. *Environ Exp Bot.* 2019;**157**:46–57. doi: [10.1016/j.envexpbot.2018.09.026](https://doi.org/10.1016/j.envexpbot.2018.09.026).
- Negrao S, Schmockel SM, Tester M. Evaluating physiological responses of plants to salinity stress. *Ann Bot.* 2017;**119**(1):1–11. doi: [10.1093/aob/mcw191](https://doi.org/10.1093/aob/mcw191). [PubMed: [27707746](https://pubmed.ncbi.nlm.nih.gov/27707746/)]. [PubMed Central: [PMC5218372](https://pubmed.ncbi.nlm.nih.gov/PMC5218372/)].
- Farooq M, Wahid A, Kobayashi N, Fujita D, Basra SMA. Plant drought stress: effects, mechanisms and management. *Agron Sustain Dev.* 2009;**29**(1):185–212. doi: [10.1051/agro:2008021](https://doi.org/10.1051/agro:2008021).
- Ranger C, Reding ME, Schultz PB, Oliver JB. Influence of flood-stress on ambrosia beetle host-selection and implications for their management in a changing climate. *Agric For Entomol.* 2013;**15**(1):56–64. doi: [10.1111/j.1461-9563.2012.00591.x](https://doi.org/10.1111/j.1461-9563.2012.00591.x).
- Shahid M. Effect of soil amendments on trace element-mediated oxidative stress in plants: Meta-analysis and mechanistic interpretations. *J Hazard Mater.* 2021;**407**:124881. doi: [10.1016/j.jhazmat.2020.124881](https://doi.org/10.1016/j.jhazmat.2020.124881). [PubMed: [33360193](https://pubmed.ncbi.nlm.nih.gov/33360193/)].
- Fedele G, Bove F, González-Domínguez E, Rossi V. A Generic Model Accounting for the Interactions among Pathogens, Host Plants, Biocontrol Agents, and the Environment, with Parametrization for Botrytis cinerea on Grapevines. *Agronomy.* 2020;**10**(2). doi: [10.3390/agronomy10020222](https://doi.org/10.3390/agronomy10020222).
- Ali S, Abbas Z, Seleiman MF, Rizwan M, Yava SI, Alhammad BA, et al. Glycine Betaine Accumulation, Significance and Interests for Heavy Metal Tolerance in Plants. *Plants (Basel).* 2020;**9**(7). doi: [10.3390/plants9070896](https://doi.org/10.3390/plants9070896). [PubMed: [32679909](https://pubmed.ncbi.nlm.nih.gov/32679909/)]. [PubMed Central: [PMC7412461](https://pubmed.ncbi.nlm.nih.gov/PMC7412461/)].
- Wang N. The Citrus Huanglongbing Crisis and Potential Solutions. *Mol Plant.* 2019;**12**(5):607–9. doi: [10.1016/j.molp.2019.03.008](https://doi.org/10.1016/j.molp.2019.03.008). [PubMed: [30947021](https://pubmed.ncbi.nlm.nih.gov/30947021/)].
- Taghizadeh-Alisaraei A, Hosseini SH, Ghobadian B, Motevali A. Biofuel production from citrus wastes: A feasibility study in Iran. *Renew Sust Energ Rev.* 2017;**69**:1100–12. doi: [10.1016/j.rser.2016.09.102](https://doi.org/10.1016/j.rser.2016.09.102).
- Dahmoune F, Boulekbache L, Moussi K, Aoun O, Spigno G, Madani K. Valorization of Citrus limon residues for the recovery of antioxidants: Evaluation and optimization of microwave and ultrasound application to solvent extraction. *Ind Crops Prod.* 2013;**50**:77–87. doi: [10.1016/j.indcrop.2013.07.013](https://doi.org/10.1016/j.indcrop.2013.07.013).
- Primo-Capella A, Martínez-Cuenca MR, Forner-Giner MÁ. Cold Stress in Citrus: A Molecular, Physiological and Biochemical Perspective. *Horticulturae.* 2021;**7**(10). doi: [10.3390/horticulturae7100340](https://doi.org/10.3390/horticulturae7100340).
- Mohammadian MA, Largani Z, Sajedi RH. Quantitative and qualitative comparison of antioxidant activity in the flavedo tissue of three cultivars of citrus fruit under cold stress. *Aust J Crop Sci.* 2012;**6**(3):402–6.
- Lang P, Zhang CK, Ebel RC, Dane F, Dozier WA. Identification of cold acclimated genes in leaves of Citrus unshiu by mRNA differential display. *Gene.* 2005;**359**:111–8. doi: [10.1016/j.gene.2005.06.013](https://doi.org/10.1016/j.gene.2005.06.013). [PubMed: [16125877](https://pubmed.ncbi.nlm.nih.gov/16125877/)].
- Atta AA, Morgan KT, Hamido SA, Kadyampakeni DM, Mahmoud KA. Water and Soil Nutrient Dynamics of Huanglongbing-Affected Citrus Trees as Impacted by Ground-Applied Nutrients. *Agronomy.* 2020;**10**(10). doi: [10.3390/agronomy10101485](https://doi.org/10.3390/agronomy10101485).
- Huang Y, Si Y, Dane F. Impact of grafting on cold responsive gene expression in Satsuma mandarin (Citrus unshiu). *Euphytica.* 2010;**177**(1):25–32. doi: [10.1007/s10681-010-0243-7](https://doi.org/10.1007/s10681-010-0243-7).

18. Poles L, Licciardello C, Distefano G, Nicolosi E, Gentile A, La Malfa S. Recent Advances of In Vitro Culture for the Application of New Breeding Techniques in Citrus. *Plants (Basel)*. 2020;**9**(8). doi: [10.3390/plants9080938](https://doi.org/10.3390/plants9080938). [PubMed: [32722179](https://pubmed.ncbi.nlm.nih.gov/32722179/)]. [PubMed Central: [PMC7465985](https://pubmed.ncbi.nlm.nih.gov/PMC7465985/)].
19. Rahimi MH, Houshmand S, Khodambashi M. Determination of the most important agronomic traits affecting seed yield in lentil (*Lens culinaris Medik*) recombinant inbred lines. *Iran J Crop Sci*. 2016;**18**(2).
20. Blum A. *Plant Breeding for Water-Limited Environments*. New York, United States: Springer; 2011. doi: [10.1007/978-1-4419-7491-4](https://doi.org/10.1007/978-1-4419-7491-4).
21. Darroch BA, Baker RJ. Two Measures of Grain Filling in Spring Wheat. *Crop Sci*. 1995;**35**(1):164-8. doi: [10.2135/cropsci1995.0011183X003500010030x](https://doi.org/10.2135/cropsci1995.0011183X003500010030x).
22. Wang ML, Wang W, Du S, Li CF, He Z. Causal relationships between carbon dioxide emissions and economic factors: Evidence from China. *Sustain Dev*. 2019;**28**(1):73-82. doi: [10.1002/sd.1966](https://doi.org/10.1002/sd.1966).
23. Smith G. Step away from stepwise. *J Big Data*. 2018;**5**(1). doi: [10.1186/s40537-018-0143-6](https://doi.org/10.1186/s40537-018-0143-6).
24. Amini A, Ghanadha MR, Abdmishani S. [Genetic variation and correlation between different traits in common bean]. *Journal of Iran Agricultural Sciences*. 2002;**33**(4). Persian.
25. Berry W, Feldman S. *Multiple Regression in Practice*. California, USA: SAGE Publications, Inc; 1985. doi: [10.4135/9781412985208](https://doi.org/10.4135/9781412985208).
26. Tadjvar Y, Fotouhi Ghazvini R, Hamidoghli Y, Hassan Sajedi R. [Physiological and biochemical responses of page mandarin on citrange rootstock to low temperature stress]. *Iranian Journal of Plant Biology*. 2011;**3**(9):1-12. Persian.
27. Pietrini F, Chaudhuri D, Thapliyal AP, Massacci A. Analysis of chlorophyll fluorescence transients in mandarin leaves during a photooxidative cold shock and recovery. *Agric Ecosyst Environ*. 2005;**106**(2-3):189-98. doi: [10.1016/j.agee.2004.10.007](https://doi.org/10.1016/j.agee.2004.10.007).
28. Hashempour A, Ghasemnezhad M, Sohani MM, Ghazvini RF, Abedi A. Effects of Freezing Stress on the Expression of Fatty Acid Desaturase (FAD2, FAD6 and FAD7) and Beta-Glucosidase (BGLC) Genes in Tolerant and Sensitive Olive Cultivars. *Russ J Plant Physiol*. 2019;**66**(2):214-22. doi: [10.1134/S1021443719020079](https://doi.org/10.1134/S1021443719020079).
29. Campos PS, Quartin V, Ramalho JC, Nunes MA. Electrolyte leakage and lipid degradation account for cold sensitivity in leaves of *Coffea* sp. plants. *J Plant Physiol*. 2003;**160**(3):283-92. doi: [10.1078/0176-1617-00833](https://doi.org/10.1078/0176-1617-00833). [PubMed: [12749085](https://pubmed.ncbi.nlm.nih.gov/12749085/)].
30. Maehly AC, Chance B. The assay of catalases and peroxidases. *Methods Biochem Anal*. 1954;**1**:357-424. doi: [10.1002/9780470110171.ch14](https://doi.org/10.1002/9780470110171.ch14). [PubMed: [13193536](https://pubmed.ncbi.nlm.nih.gov/13193536/)].
31. Claiborne A. Catalase activity. In: Greenwald RA, editor. *CRC Handbook of Methods for Oxygen Radical Research*. Florida, United States: CRC-Press; 1986. p. 283-4.
32. Barnes JD, Balaguer L, Manrique E, Elvira S, Davison AW. A reappraisal of the use of DMSO for the extraction and determination of chlorophylls a and b in lichens and higher plants. *Environ Exp Bot*. 1992;**32**(2):85-100. doi: [10.1016/0098-8472\(92\)90034-y](https://doi.org/10.1016/0098-8472(92)90034-y).
33. Wu Q, Zou YN, Xia RX. Effects of water stress and arbuscular mycorrhizal fungi on reactive oxygen metabolism and antioxidant production by citrus (*Citrus tangerine*) roots. *Eur J Soil Biol*. 2006;**42**(3):166-72. doi: [10.1016/j.ejsobi.2005.12.006](https://doi.org/10.1016/j.ejsobi.2005.12.006).
34. Ritchie SW, Nguyen HT, Holaday AS. Leaf Water Content and Gas-Exchange Parameters of Two Wheat Genotypes Differing in Drought Resistance. *Crop Sci*. 1990;**30**(1):105-11. doi: [10.2135/cropsci1990.0011183X003000010025x](https://doi.org/10.2135/cropsci1990.0011183X003000010025x).
35. Sullivan CY, Ross WM. Selecting for drought and heat resistance in grain sorghum. In: Sullivan CY, Ross WM, editors. *Stress Physiology in Crop Plants*. New Jersey, United States: Wiley; 1979.
36. Lim C, Arora R, Townsend EC. Comparing Gompertz and Richards Functions to Estimate Freezing Injury in Rhododendron Using Electrolyte Leakage. *J Am Soc Hortic Sci*. 1998;**123**(2):246-52. doi: [10.21273/jashs.123.2.246](https://doi.org/10.21273/jashs.123.2.246).
37. Wagner GJ. Content and vacuole/extravacuole distribution of neutral sugars, free amino acids, and anthocyanin in protoplasts. *Plant Physiol*. 1979;**64**(1):88-93. doi: [10.1104/pp.64.1.88](https://doi.org/10.1104/pp.64.1.88). [PubMed: [16660921](https://pubmed.ncbi.nlm.nih.gov/16660921/)]. [PubMed Central: [PMC543030](https://pubmed.ncbi.nlm.nih.gov/PMC543030/)].
38. Bates LS, Waldren RP, Teare ID. Rapid determination of free proline for water-stress studies. *Plant Soil*. 1973;**39**(1):205-7. doi: [10.1007/bf00018060](https://doi.org/10.1007/bf00018060).
39. Grieve CM, Grattan SR. Rapid assay for determination of water soluble quaternary ammonium compounds. *Plant Soil*. 1983;**70**(2):303-7. doi: [10.1007/bf02374789](https://doi.org/10.1007/bf02374789).
40. Alexieva V, Sergiev I, Mapelli S, Karanov E. The effect of drought and ultraviolet radiation on growth and stress markers in pea and wheat. *Plant Cell Environ*. 2001;**24**(12):1337-44. doi: [10.1046/j.1365-3040.2001.00778.x](https://doi.org/10.1046/j.1365-3040.2001.00778.x).
41. Li J, Yang Y, Iqbal A, Qadri R, Shi P, Wang Y, et al. Correlation analysis of cold-related gene expression with physiological and biochemical indicators under cold stress in oil palm. *PLoS One*. 2019;**14**(11). e0225768. doi: [10.1371/journal.pone.0225768](https://doi.org/10.1371/journal.pone.0225768). [PubMed: [31774880](https://pubmed.ncbi.nlm.nih.gov/31774880/)]. [PubMed Central: [PMC6881061](https://pubmed.ncbi.nlm.nih.gov/PMC6881061/)].
42. Salehi Sardoei A, Fazeli-Nasab B. Non-destructive estimation of leaf area of Citrus varieties of the Kotra Germplasm Bank. *Plant Biotechnology Persa*. 2021;**3**(2):18-31. doi: [10.52547/pbp.3.2.18](https://doi.org/10.52547/pbp.3.2.18).
43. Abouzari A, Solouki M, Golein B, Fakheri BA, Sabouri A. [Frost Stress Tolerance in Citrus Genotypes under Two Subfreezing Temperatures]. *Plant Production Technology*. 2020;**12**(1):193-209. Persian. doi: [10.22084/PPT.2020.19704.1938](https://doi.org/10.22084/PPT.2020.19704.1938).
44. Zahedi F, Nabati Ahmadi D, Mohammadi M, Karimizadeh R. [Path Analysis To Study Morph-Physiological Traits, Yield And Traits Related To Yield Of Lentil Genotypes Under Rain Fed Condition]. *The Plant Production (Scientific Journal Of Agriculture)*. 2016;**39**(2). Persian.
45. Moosavi SS, Abdollahi MR, Ghanbari F, Kanouni H. [Detection and selection of effective traits on grain yield in chickpea (*Cicer arietinum* L.) under normal moisture condition]. *Journal of Plant Productions (Agronomy, Breeding and Horticulture)*. 2016;**39**(1):119-31. Persian.
46. Johnson RA, Wichern DW. *Applied Multivariate Statistical Analysis*. New Jersey, United States: Prentice Hall; 1996.
47. de Marsily G. An overview of the world's water resources problems in 2050. *Ecophysiol Hydrobiol*. 2007;**7**(2):147-55. doi: [10.1016/S1642-3593\(07\)70180-5](https://doi.org/10.1016/S1642-3593(07)70180-5).