



The Association of Vitamin D Receptor Gene Polymorphism with Polycystic Ovary Syndrome in a Southeast Iranian Population

Marzieh Ghasemi ^{1,2}, Marzieh Koohestani³, Anoosh Naghavi ^{4,5,*}

¹ Pregnancy Health Research Center, Zahedan University of Medical Sciences, Zahedan, Iran

² Moloud Infertility Center, Ali-ibn-Abi Taleb Hospital, Zahedan University of Medical Sciences, Zahedan, Iran

³ School of Medicine, Zahedan University of Medical Sciences, Zahedan, Iran

⁴ Cellular and Molecular Research Center, Research Institute of Cellular and Molecular Sciences in Infectious Disease, Zahedan University of Medical Sciences, Zahedan, Iran

⁵ Departments of Genetics, School of Medicine, Zahedan University of Medical Sciences, Zahedan, Iran

* Corresponding author: Cellular and Molecular Research Center, Research Institute of Cellular and Molecular Sciences in Infectious Disease, Zahedan University of Medical Sciences, Zahedan, Iran. Email: anooshnaghavi@yahoo.com

Received 2024 January 9; Revised 2024 May 12; Accepted 2024 May 20.

Abstract

Background: Polycystic ovary syndrome (PCOS) is a common endocrine disorder among women of reproductive age, characterized by hyperandrogenism, amenorrhea, and polycystic ovaries. Metabolic disorders, including insulin resistance (IR), are common comorbidities of PCOS, with vitamin D receptors playing a crucial role.

Objectives: We investigated whether the rs2228570T > C variant of the vitamin D receptor (VDR) gene is associated with the risk of PCOS.

Methods: In the current study, 112 women with PCOS and 150 healthy women participated. Genomic DNA was extracted using the standard salting-out method. Genotyping of rs2228570T > C was conducted via amplification-refractory mutation system polymerase chain reaction. Additionally, VD3 levels were measured using the ELISA system. Statistical analysis of genotyping data was performed using the SPSS V.22 package.

Results: The statistical analysis showed that the T-allele of rs2228570C/T significantly decreased the risk of PCOS. Moreover, the T-allele in the codominant model significantly eliminated the risk of PCOS in our population. Our results also showed a statistically significant difference in VD3 levels between the CC and TT genotypes.

Conclusions: Based on our result, rs2228570C/T had a protective role for PCOS risk in our population.

Keywords: Polycystic Ovary Syndrome, Vitamin D Receptor, Metabolic Disorders, Single Nucleotide Polymorphism

1. Background

Polycystic ovary syndrome (PCOS), first described in 1935 as a combination of hirsutism, amenorrhea, enlarged cystic ovaries, obesity, and infertility, affects a range of women of reproductive age (1). Polycystic ovary syndrome, a common endocrinopathy, primarily affects women of childbearing age and often begins in adolescence (2). The basic symptoms of PCOS include irregular menstruation, defects in ovulation, infertility, hirsutism, and acne (3). Additionally, there is an increased prevalence of metabolic disorders such as type 2 diabetes mellitus, impaired glucose tolerance, and insulin resistance (IR) among patients with PCOS (4, 5).

The precise etiology of PCOS is poorly understood but appears to involve familial background, genetic predisposition, and environmental factors participate in the occurrence and development of the disease (6). It is not surprising that lifestyle. Lifestyle factors, including diet, food quality, and low exercise, directly affect the risk of PCOS in susceptible individuals (7) by promoting metabolic disorders like IR and type 2 diabetes mellitus. Although genome-wide studies have identified some markers, these show low predictive values (8). Factors such as intrauterine defects, high maternal hormones, and weight gain in women are also likely to contribute to PCOS conditions (9). Thus, it is not possible to focus on a single gene as the primary candidate responsible

for the development of PCOS, leading major studies to consider the heterogeneous basis of the condition (6).

Vitamin D, converted into 1,25-dihydroxycholecalciferol in the liver and kidney, regulates calcium and phosphate homeostasis, which in turn adjusts insulin secretion from β -cells (10). The vitamin D receptor (VDR), a key functional arm of vitamin D, forms a heterodimer with the retinoid X receptor (RXR) in many organs and tissues (6). Evidence suggests that VDR may be involved in metabolic diseases (11).

Vitamin D receptor as a transcription factor regulates the expression of genes especially those that participate in the metabolism of glucose by an unrevealed mechanism, to date (10).

The VDR gene located on chromosome 12q13.11 contains 12 exons and encodes a ligand-dependent transcription factor in the steroid/thyroid hormone receptor superfamily (12).

Polymorphisms in the VDR gene may affect the conformation of the receptor as well as the interaction between vitamin D and VDR leading to a decrease in absorption of vitamin D (13). Recent findings have suggested that VDR variations were also associated with the risk of PCOS as well as the severity of the phenotype (12, 14).

2. Objectives

In this study, we aimed to investigate whether VDR gene polymorphism rs2228570T > C is associated with the risk of PCOS in a southeast population of Iran. rs2228570C/T variation occurs in the first exon of VDR mRNA and interrupts the initiator codon. In recent years, the correlation of rs2228570T > C with the risk of PCOS has been analyzed in different ethnicities (12).

3. Methods

3.1. Study Design and Subjects

In this study, 116 women clinically diagnosed with PCOS based on the Rotterdam ESHR/ASRM consensus criteria (2003) were included as the case group. The inclusion criteria consisted of clinical and/or biochemical hyperandrogenism, oligo-anovulation, and polycystic ovary, with the morphology of polycystic ovary under the guidance of Ultrasonography and patients should have at least two of these items (15). These subjects were referred to the Molood Infertility Center at Ali-Ibn-Abi Taleb Hospital, Zahedan, Iran, between August 2018 and September 2019. The control

group comprised 130 unrelated women with no history of disorders and regular menstrual cycles. These women had no complaints of hirsutism, acne, alopecia, or endocrinopathy. All subjects were informed about the study and its risks.

3.2. VD3 Levels

All subjects were referred to the biochemical laboratory of Ali-Ibn-Abi Taleb Hospital, where VD3 levels were measured using an ELISA technique (PishtazTeb company, Iran).

3.3. Genotyping

Blood samples were collected from all participants using EDTA-covered tubes. The standard salting-out method was used to extract DNA (16). Genotyping of rs2228570T > C was performed using the amplification-refractory mutation system polymerase chain reaction (ARMS PCR) method. In this technique, each tube contained three primers: Two common primers for amplifying the internal control, and one allele-specific primer for detecting the presence of either allele. The 247 bp internal control fragment was amplified using 5'-GGTCTCCACACACCCACAGATCCG-3' as the forward primer and 5'-AGGCCTGGGCCCTGGGAGAT-3' as the reverse primer. Additionally, the flanking sequence 5'-GGCCTGCTTGCTGTTCTTACAGGTAT-3' served as the T-allele-specific primer, and 5'-GGCCTGCTTGCTGTTCTTACAGGTAC-3' served as the C-allele-specific primer, producing a 112 bp fragment in combination with the forward primer (Figure 1). PCR was performed in an Applied Biosystems Veriti™ Thermal Cycler with 30 cycles of denaturation at 95°C for 30 seconds, annealing at 63°C for 30 seconds, and extension at 74°C for 30 seconds, followed by a final extension at 74°C for 5 minutes. A pre-denaturation step at 95°C for 5 minutes was applied before the ramp cycles. For each sample, a reaction tube contained 10 μ L of Taq DNA polymerase 2 \times master mix (Parstus Biotech company, Iran), 1 μ L of each primer (10 pm/ μ L) (GenFanavaran Biotech company, Iran), 0.8 μ L of extracted DNA, and up to 20 μ L of deionized water. The PCR products were subjected to 2% agarose gel electrophoresis and visualized using a UV transilluminator system.

3.4. Statistical Analysis

Statistical analysis was performed using SPSS V.22 for Windows (IBM Corporation, Armonk, NY, USA). Quantitative data were described as mean \pm standard deviation for parametric data. Qualitative data were

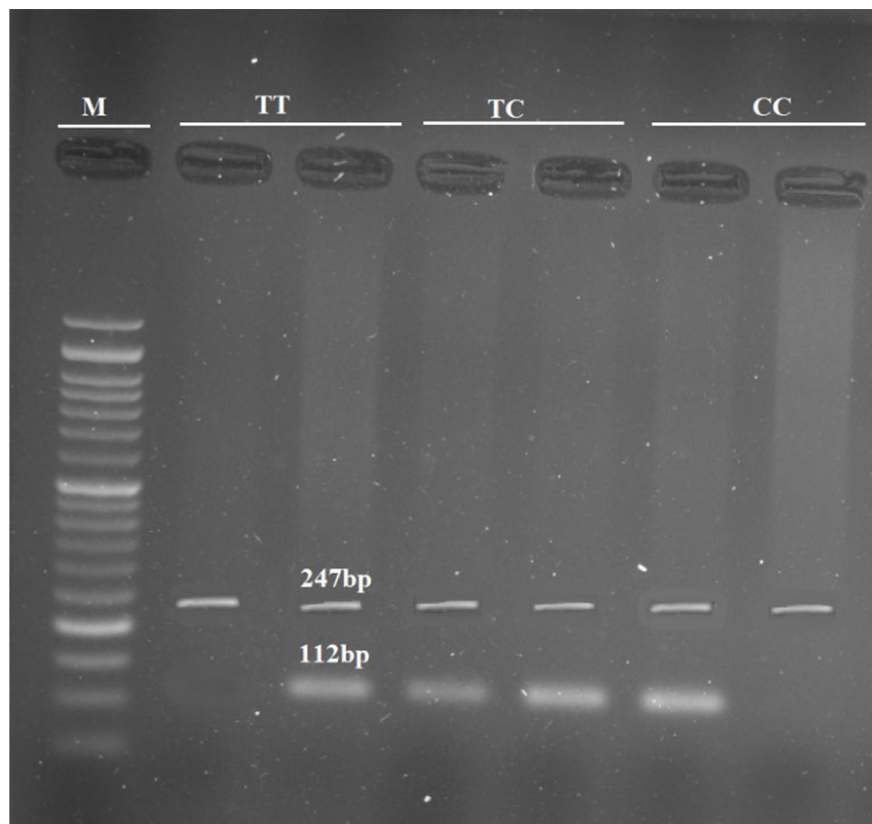


Figure 1. The gel electrophoresis of rs2228570T > C variant. M: Marker

analyzed using numbers and percentages. The Student's *t*-test and one-way ANOVA were used to compare parametric quantitative data. Genotypic and allelic differences between the two groups were compared using the chi-square (χ^2) test, odds ratios (ORs), and 95% confidence intervals.

3.5. In Silico Analysis

To determine whether rs2228570T > C affects VDR gene products, bioinformatics analyses were performed. The RNAfold server was used to explore the impact of the variant on the secondary structure of the VDR transcript (17). RNAfold introduces a thermodynamic ensemble value in which more negative ones are more stable. SpliceAid2 server was employed to detect the effect of rs2228570T > C on the splicing site patterns (18). Finally, the hypothesis implying the potential of rs2228570T > C for meaningful structural changing was

analyzed by the SNAP server which is a server for predicting the functional effects of variants (19).

4. Results

The demographic features of participants were analyzed using the Student's *t*-test. A P-value under 0.05 was considered statistically significant. There were no significant differences in age and BMI between the two groups. The mean age in the case and control groups was 26.03 ± 4.91 and 27.44 ± 5.35 , respectively ($P = 0.082$) (Table 1). The BMI in women with PCOS and healthy women was 23.73 ± 4.06 and 23.53 ± 4.02 , respectively ($P = 0.559$). A significant decrease in serum vitamin D levels was observed in PCOS cases compared to healthy women ($P = 0.001$), with levels of 17.92 ± 12.98 and 22.61 ± 13.40 , respectively (Table 1).

Table 2 shows the frequency of different genotypes in the studied population. Genotyping analysis revealed that the T-allele significantly decreased the risk of PCOS

Table 1. The Demographic Data of Women with Polycystic Ovary Syndrome and Healthy Group^a

Variables	(n ± SD)		P-Value
	PCOS (n = 112)	Controls (n = 150)	
Age	26.03 ± 4.91	27.44 ± 5.35	0.082
BMI	23.73 ± 4.06	23.53 ± 4.02	0.559
VD3	17.92 ± 12.98	22.61 ± 13.40	0.001

^z Abbreviations: BMI, Body Mass Index; VD3, cholecalciferol; PCOS, polycystic ovary syndrome; SD, standard deviation.

^a P < 0.05 was considered statistically significant.

Table 2. Genotyping and Allelic Distribution of VDR rs2228570T > C Variant in Polycystic Ovary Syndrome and Healthy Group^{a, b}

Rs2228570C > T	PCOS	Control	OR (95%CI)	P-Value
Codominant				
CC	54 (46.5)	45 (34.6)	1 [reference]	
CT	56 (48.3)	69 (53.1)	0.68 (0.40 - 1.15)	0.147
TT	6 (5.2)	16 (12.3)	0.31 (0.11 - 0.86)	0.021
Allele				
C	164 (70.7)	159 (61.2)	1 [reference]	
T	68 (29.3)	101 (38.8)	0.65 (0.49 - 0.95)	0.026
Dominant				
CC	54 (46.6)	45 (34.6)	1 [reference]	
TT + CT	62 (53.4)	85 (65.4)	0.61 (0.36 - 1.02)	0.076
Recessive				
CC + CT	110 (94.8)	114 (87.7)	1 [reference]	
TT	6 (5.2)	16 (12.3)	0.39 (0.15 - 1.03)	0.083
Over-dominant				
CC + TT	60 (51.7)	61 (46.9)	1 [reference]	
TC	56 (48.3)	69 (53.1)	0.83 (0.50 - 1.36)	0.533
Additive				
CC	54 (46.5)	45 (34.6)	1 [reference]	
CT	56 (48.3)	69 (53.1)	0.68 (0.40 - 1.15)	0.147
TT	6 (5.2)	16 (12.3)	0.46 (0.17 - 1.26)	0.193

^z Abbreviations: CI, confidence interval; OR, odds ratio; PCOS, polycystic ovary syndrome.

^a P < 0.05 was considered statistically significant.

^b Values are presented as No. (%).

Table 3. VD3 Levels of Studied Polycystic Ovary Syndrome Women According to Genotype Distribution^{a, b}

Parameter Evaluated	Case Genotypes			Test of Significance	Within Groups Significant
	CC	TC	TT		
VD3	20.05 ± 10.83	17.17 ± 14.74	5.73 ± 2.71	F: 3.62, P: 0.030	P1: 0.496, P2: 0.115, P3: 0.036

^a VD3, cholecalciferol; P1, the difference between CC & TC; P2, the difference between TC & TT; P3, the difference between CC & TT; F, One Way ANOVA test.

^b Parameters described as mean ± SD.

(OR = 0.65, 95% CI = 0.49-0.95, P = 0.026). Similarly, the TT genotype versus the CC genotype had a protective role against PCOS. The CT genotype versus the CC genotype

reduced the risk of PCOS (OR = 0.31, 95% CI = 0.11 - 0.86, P = 0.021). ANOVA showed a statistically significant difference in VD3 levels among genotypes in PCOS

Table 4. VD3 Level of Case and Control Subjects Based on Recessive Model ^{a,b}

Group and Genotype	VD3	P-Value
Control		0.821
TT	19.80 ± 7.96	
CC+TC	23.01 ± 13.97	
Case		0.001
TT	5.73 ± 2.71	
CC+TC	18.59 ± 12.99	

^a VD3, cholecalciferol.^b P < 0.05 is regarded as statistically significant.

women (P = 0.030) (Table 3). VD3 levels were significantly lower in the TT genotype compared to the CC genotype (P = 0.036). However, there was no significant difference in VD3 levels between CC vs. CT and CT vs. TT. There was a significant decrease in VD3 levels in the CC + CT genotype compared to the TT genotype in PCOS subjects (P = 0.001) (Table 4).

The related sequences of VDR genes, including transcript and amino acid sequences, were obtained from the National Center for Bioinformatics Information (NCBI) database. A 40-nt flanking region containing rs2228570T > C was selected and introduced to the RNAfold server. The RNAfold server results showed that the C-allele had more negative free energy compared to the T-allele, with thermodynamic ensemble values of -13.34 kcal/mol and -12.31 kcal/mol, respectively (Figure 2). The SpliceAid2 server revealed that the C-allele of rs2228570 creates a de novo binding sequence for the SRp30c splicing factor, while the T-allele creates three new sites for ESRP1, SC35, and hnRNPF, and disrupts the SRp30c site (Figure 3). Finally, the results of the SNAP server depicted that the T > C exchange is not likely to have a significant effect on the structure of the VDR protein. SNAP server predicted rs2228570T > C as a neutral exchange so this variant does not appear to change the structure and function of the VDR protein (Figure 4).

5. Discussion

Polycystic ovary syndrome, distinguished by prolonged lack of ovulation and hyperandrogenism, is one of the most common causes of infertility and dysmenorrhea in reproductive women (20). Due to difficulties in diagnosis, the approaches to diagnosing and managing PCOS vary among physicians and specialties, resulting in different reported prevalence rates in the literature (8). In diagnostic approaches, it is critical to exclude hormone malignancies such as

congenital adrenal hyperplasia, androgen-secreting tumors, and Cushing's syndrome due to their similar manifestations with PCOS (6).

In the current study, we found that the T-allele of rs2228570 in PCOS subjects was less frequent compared to healthy subjects and had a protective role, decreasing the risk of PCOS in the studied population. Moreover, the TT genotype versus the CC genotype significantly decreased the risk of disease. VD3 levels showed a significant decrease in PCOS subjects with the TT genotype. In silico analysis showed that the C-allele of rs2228570 is more stable compared to the T-allele. Additionally, our analysis revealed that C and T alleles create new target sites for different splicing factors such as SRp30c, ESRP1, SC35, and hnRNP. Moreover, rs2228570T > C does not appear to affect the structure of the VDR protein.

Vitamin D, resulting from sun exposure or dietary intake, is metabolized to 1,25-dihydroxyvitamin D3 (1,25(OH)₂D) in the liver and kidney. Vitamin D, along with parathyroid hormone, controls calcium homeostasis (21). Evidence suggests that vitamin D plays a role in various physiological functions in humans, including reproduction, sex hormone synthesis, and the insulin metabolic pathway (22-24). Vitamin D deficiency has been associated with some comorbidities of PCOS, including hyperandrogenism and cardiovascular disease (25, 26). Polycystic ovary syndrome patients are known to have increased levels of IR compared to healthy women (21). Insulin can increase the level of free testosterone by binding to its receptor (27). Vitamin D appears to affect the expression of insulin receptors and is essential for regulating calcium homeostasis, which is vital for insulin secretion from β-cells (28). Vitamin D regulates the transcription of genes sensitive to vitamin D levels and calcium metabolism through VDR, which acts as a transcription factor (29). Additionally, VDR is involved in estrogen metabolism, impacting ovarian function (30).

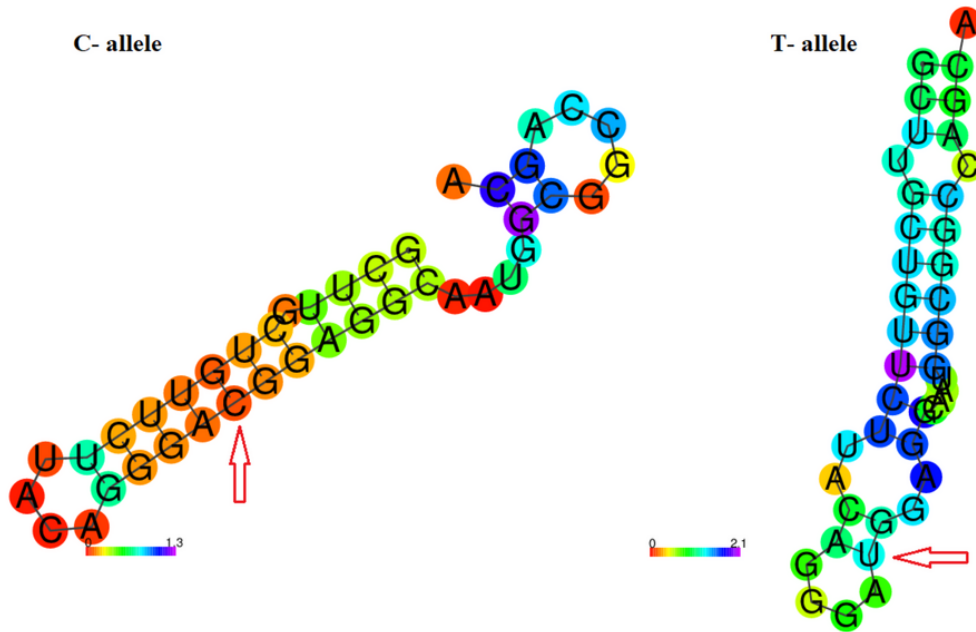


Figure 2. The result of RNAfold server. It seems that the C-allele was more stable than the T-allele regarding thermodynamic ensemble values.

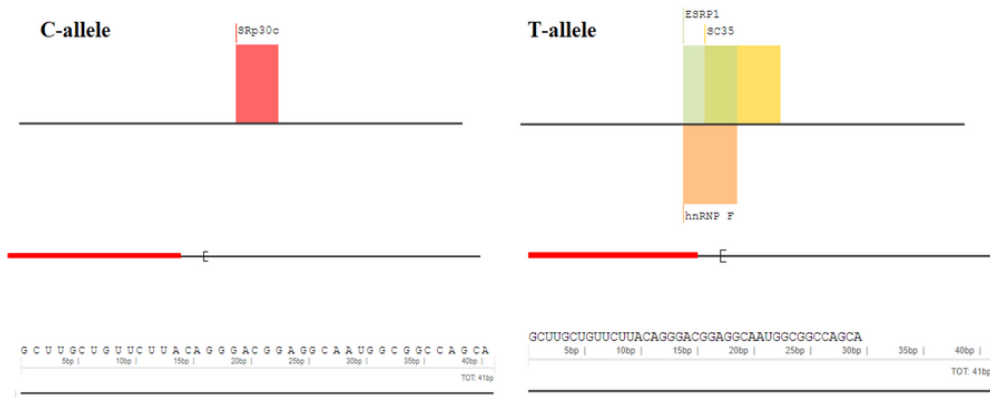


Figure 3. The SpliceAid2 server showed that some new recognition sites create in presence of both C and T-alleles.

Vitamin D receptor gene polymorphisms are likely to impact the etiology of PCOS through the insulin signaling pathway. These polymorphisms may also affect PCOS conditions via the parathyroid hormone (PTH)-vitamin D axis (10, 31), as the interaction between

vitamin D and VDR regulates PTH secretion and synthesis (12).

Many investigations have revealed the association of single nucleotide polymorphisms with the risk of PCOS. The VDR gene, involved in the insulin signaling pathway, has been identified as a crucial gene in the occurrence

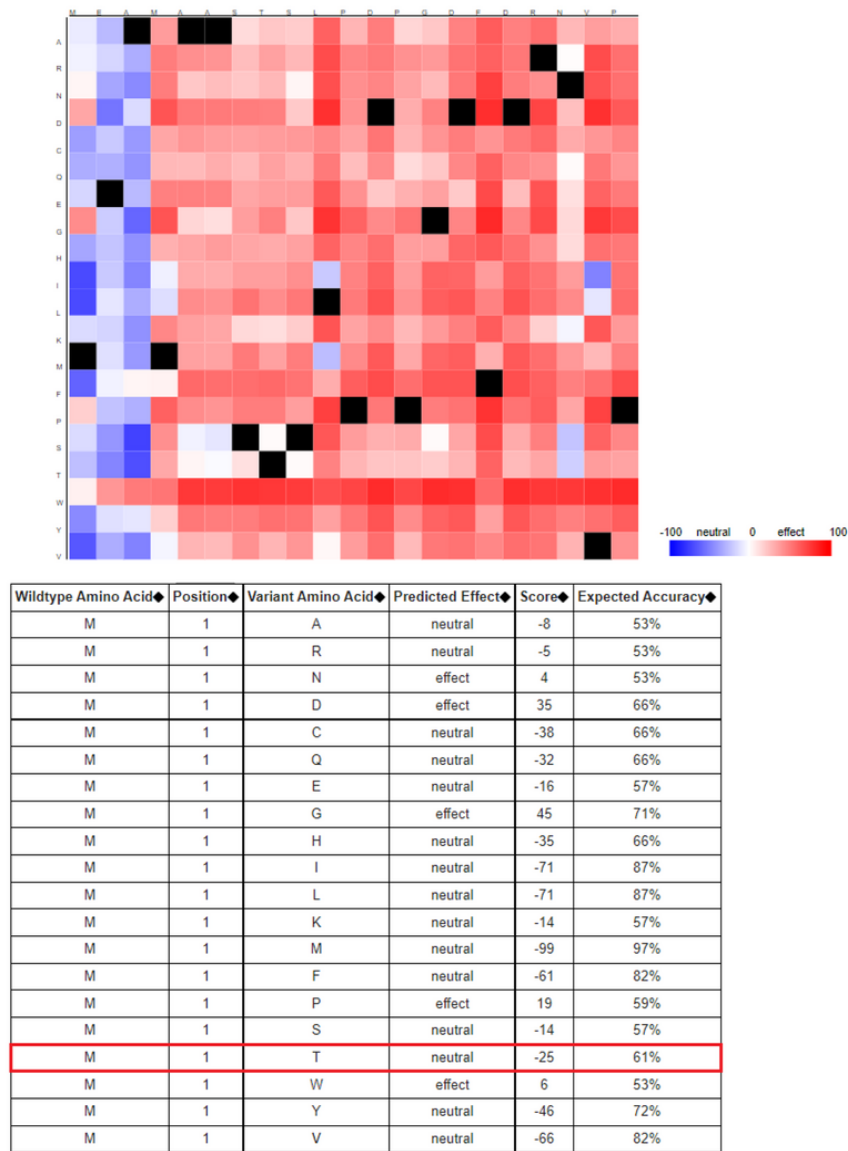


Figure 4. The results of the SNAP server imply that the rs2228570T>C variant does not seem to change the structure of the vitamin D receptor (VDR) protein.

of PCOS (32). A meta-analysis by Shi et al. showed that variations in the VDR gene, such as rs7975232 and rs1544410, are associated with the risk of PCOS, but not rs2228570T > C (12). Mahmoudi found that the CC genotype of rs2228570, compared to CT + TT, confers an increased risk for IR and higher serum insulin concentration in PCOS women (33). An association study by Tuncel et al. showed that rs2228570C > T is linked to decreased vitamin D levels in the serum of Turkish

patients compared to the common genotype (30). It is believed that the substitution of C with T leads to the formation of a longer, less active form of VDR. Therefore, carriers of the CC genotype may have higher vitamin D levels than CT and TT subjects (30). Other studies have linked rs2228570T > C with chronic kidney disease (CKD), worsened by vitamin D deficiency (34). Shaymaa and Al-Zubaidy showed that the C-allele and CC genotype of rs2228570T > C are correlated with the risk

of type 2 diabetes mellitus (35). In early 2020, Hu et al. analyzed the association of rs2228570T > C with the risk of Parkinson's disease, finding that C carriers of rs2228570T > C are prone to Parkinson's disease (36).

In the current study, we faced limitations such as sample size. We plan to extend the population size in future studies and investigate the association of more variations in the VDR gene and other related genes, overcoming technical restrictions.

5.1. Conclusions

According to the findings of the current study, we hypothesize that rs2228570C/T has a protective role against the risk of PCOS in our population.

Acknowledgements

The authors would like to thank Zahedan University of Medical Sciences (1398.8035) for its financial support of this study.

Footnotes

Authors' Contribution: Anoosh Naghavi, data collection, study supervision, and final draft; Marzieh Koohestani, genotyping, data analyzing, primary draft; Marzieh Ghasemi, study conception and primary draft.

Conflict of Interests Statement: The authors declare that there is no conflict of interest regarding the publication of this article.

Data Availability: The dataset presented in the study is available on request from the corresponding author during submission or after publication.

Ethical Approval: The ethics committee of Zahedan University of Medical Sciences approved this study under the ethics code of IR.ZAUMS.REC.1396.088.

Funding/Support: This study was financially supported by Zahedan University of Medical Sciences as a part of the MD thesis of Marzieh Koohestani.

Informed Consent: All subjects were informed about the study and its risks.

References

- Stein IF, Leventhal ML. Amenorrhea associated with bilateral polycystic ovaries. *Am J Obstetrics Gynecol.* 1935;**29**(2):181-91.
- Norman RJ, Dewailly D, Legro RS, Hickey TE. Polycystic ovary syndrome. *Lancet.* 2007;**370**(9588):685-97. [PubMed ID: 17720020]. [https://doi.org/10.1016/S0140-6736\(07\)61345-2](https://doi.org/10.1016/S0140-6736(07)61345-2).
- Guruvaiah P, Govatati S, Reddy TV, Beeram H, Deenadayal M, Shivaji S, et al. Analysis of Connexin37 gene C1019T polymorphism and PCOS susceptibility in South Indian population: case-control study. *Eur J Obstet Gynecol Reprod Biol.* 2016;**196**:17-20. [PubMed ID: 26656196]. <https://doi.org/10.1016/j.ejogrb.2015.11.002>.
- Legro RS, Gnatuk CL, Kunselman AR, Dunaif A. Changes in glucose tolerance over time in women with polycystic ovary syndrome: a controlled study. *J Clin Endocrinol Metab.* 2005;**90**(6):3236-42. [PubMed ID: 15797965]. <https://doi.org/10.1210/jc.2004.1843>.
- Maitra A, Pingle RR, Menon PS, Naik V, Gokral JS, Meherji PK. Dyslipidemia with particular regard to apolipoprotein profile in association with polycystic ovary syndrome: a study among Indian women. *Int J Fertility Women's Med.* 2001;**46**(5):271-7.
- Reis GV, Gontijo NA, Rodrigues KF, Alves MT, Ferreira CN, Gomes KB. Vitamin D receptor polymorphisms and the polycystic ovary syndrome: A systematic review. *J Obstet Gynaecol Res.* 2017;**43**(3):436-46. [PubMed ID: 28127831]. <https://doi.org/10.1111/jog.13250>.
- Diamanti-Kandaraki E, Christakou C, Marinakis E. Phenotypes and environmental factors: their influence in PCOS. *Curr Pharm Des.* 2012;**18**(3):270-82. [PubMed ID: 22229564]. <https://doi.org/10.2174/138161212799040457>.
- Norman RJ, Teede HJ. A new evidence-based guideline for assessment and management of polycystic ovary syndrome. *Med J Aust.* 2018;**209**(7):299-300. [PubMed ID: 30257632]. <https://doi.org/10.5694/mja18.00635>.
- Tata B, Mimouni NEH, Barbotin AL, Malone SA, Loyens A, Pigny P, et al. Elevated prenatal anti-Mullerian hormone reprograms the fetus and induces polycystic ovary syndrome in adulthood. *Nat Med.* 2018;**24**(6):834-46. [PubMed ID: 29760445]. [PubMed Central ID: PMC6098696]. <https://doi.org/10.1038/s41591-018-0035-5>.
- Pittas AG, Lau J, Hu FB, Dawson-Hughes B. The role of vitamin D and calcium in type 2 diabetes. A systematic review and meta-analysis. *J Clin Endocrinol Metab.* 2007;**92**(6):2017-29. [PubMed ID: 17389701]. [PubMed Central ID: PMC2085234]. <https://doi.org/10.1210/jc.2007-0298>.
- Wehr E, Moller R, Horejsi R, Giuliani A, Kopera D, Schweighofer N, et al. Subcutaneous adipose tissue topography and metabolic disturbances in polycystic ovary syndrome. *Wien Klin Wochenschr.* 2009;**121**(7-8):262-9. [PubMed ID: 19562283]. <https://doi.org/10.1007/s00508-009-1162-2>.
- Shi XY, Huang AP, Xie DW, Yu XL. Association of vitamin D receptor gene variants with polycystic ovary syndrome: a meta-analysis. *BMC Med Genet.* 2019;**20**(1):32. [PubMed ID: 30764792]. [PubMed Central ID: PMC6376757]. <https://doi.org/10.1186/s12881-019-0763-5>.
- Xavier LB, Gontijo NA, Rodrigues KF, Candido AL, Dos Reis FM, de Sousa MCR, et al. Polymorphisms in vitamin D receptor gene, but not vitamin D levels, are associated with polycystic ovary syndrome in Brazilian women. *Gynecol Endocrinol.* 2019;**35**(2):146-9. [PubMed ID: 30182771]. <https://doi.org/10.1080/09513590.2018.1512966>.
- Zadeh-Vakili A, Ramezani Tehrani F, Daneshpour MS, Zarkesh M, Saadat N, Azizi F. Genetic polymorphism of vitamin D receptor gene affects the phenotype of PCOS. *Gene.* 2013;**515**(1):193-6. [PubMed ID: 23246977]. <https://doi.org/10.1016/j.gene.2012.11.049>.
- Rotterdam EAPCWG. Revised 2003 consensus on diagnostic criteria and long-term health risks related to polycystic ovary syndrome. *Fertil Steril.* 2004;**81**(1):19-25. [PubMed ID: 14711538]. <https://doi.org/10.1016/j.fertnstert.2003.10.004>.
- Miller SA, Dykes DD, Polesky HF. A simple salting out procedure for extracting DNA from human nucleated cells. *Nucleic Acids Res.* 1988;**16**(3):1215. [PubMed ID: 3344216]. [PubMed Central ID: PMC334765]. <https://doi.org/10.1093/nar/16.3.1215>.
- Ghasemi M, Heidari Nia M, Hashemi M, Keikha N, Fazeli K, Taji O, et al. An association study of polymorphisms in the H19 imprinted gene in

- an Iranian population with the risk of polycystic ovary syndrome. *Biol Reprod.* 2020;**103**(5):978-85. [PubMed ID: 32720692]. <https://doi.org/10.1093/biolre/iaaa131>.
18. Piva F, Giulietti M, Burini AB, Principato G. SpliceAid 2: a database of human splicing factors expression data and RNA target motifs. *Hum Mutat.* 2012;**33**(1):81-5. [PubMed ID: 21922594]. <https://doi.org/10.1002/humu.21609>.
 19. Bromberg Y, Yachdav G, Rost B. SNAP predicts effect of mutations on protein function. *Bioinformatics.* 2008;**24**(20):2397-8. [PubMed ID: 18757876]. [PubMed Central ID: PMC2562009]. <https://doi.org/10.1093/bioinformatics/btn435>.
 20. Diamanti-Kandarakis E, Kouli CR, Bergiele AT, Filandra FA, Tsianateli TC, Spina GG, et al. A survey of the polycystic ovary syndrome in the Greek island of Lesbos: hormonal and metabolic profile. *J Clin Endocrinol Metab.* 1999;**84**(11):4006-11. [PubMed ID: 10566641]. <https://doi.org/10.1210/jcem.84.11.6148>.
 21. DeLuca HF. Overview of general physiologic features and functions of vitamin D. *Am J Clin Nutr.* 2004;**80**(6 Suppl):1689S-96S. [PubMed ID: 15585789]. <https://doi.org/10.1093/ajcn/80.6.1689S>.
 22. Anagnostis P, Karras S, Goulis DG. Vitamin D in human reproduction: a narrative review. *Int J Clin Pract.* 2013;**67**(3):225-35. [PubMed ID: 23294318]. <https://doi.org/10.1111/ijcp.12031>.
 23. Wojtusik J, Johnson PA. Vitamin D regulates anti-Mullerian hormone expression in granulosa cells of the hen. *Biol Reprod.* 2012;**86**(3):91. [PubMed ID: 22174023]. <https://doi.org/10.1095/biolreprod.111.094110>.
 24. Mitri J, Dawson-Hughes B, Hu FB, Pittas AG. Effects of vitamin D and calcium supplementation on pancreatic beta cell function, insulin sensitivity, and glycemia in adults at high risk of diabetes: the Calcium and Vitamin D for Diabetes Mellitus (CaDDM) randomized controlled trial. *Am J Clin Nutr.* 2011;**94**(2):486-94. [PubMed ID: 21715514]. [PubMed Central ID: PMC3142723]. <https://doi.org/10.3945/ajcn.111.011684>.
 25. de Groot PC, Dekkers OM, Romijn JA, Dieben SW, Helmerhorst FM. PCOS, coronary heart disease, stroke and the influence of obesity: a systematic review and meta-analysis. *Hum Reprod Update.* 2011;**17**(4):495-500. [PubMed ID: 21335359]. <https://doi.org/10.1093/humupd/dmr001>.
 26. Krul-Poel YH, Snackey C, Louwers Y, Lips P, Lambalk CB, Laven JS, et al. The role of vitamin D in metabolic disturbances in polycystic ovary syndrome: a systematic review. *Eur J Endocrinol.* 2013;**169**(6):853-65. [PubMed ID: 24044903]. <https://doi.org/10.1530/EJE-13-0617>.
 27. Ehrmann DA. Polycystic ovary syndrome. *N Engl J Med.* 2005;**352**(12):1223-36. [PubMed ID: 15788499]. <https://doi.org/10.1056/NEJMra041536>.
 28. Maestro B, Molero S, Bajo S, Davila N, Calle C. Transcriptional activation of the human insulin receptor gene by 1,25-dihydroxyvitamin D(3). *Cell Biochem Funct.* 2002;**20**(3):227-32. [PubMed ID: 12125099]. <https://doi.org/10.1002/cbf.951>.
 29. Fleet JC. The role of vitamin D in the endocrinology controlling calcium homeostasis. *Mol Cell Endocrinol.* 2017;**453**:36-45. [PubMed ID: 28400273]. [PubMed Central ID: PMC5529228]. <https://doi.org/10.1016/j.mce.2017.04.008>.
 30. Tuncel G, Temel SG, Ergoren MC. Strong association between VDR FokI (rs2228570) gene variant and serum vitamin D levels in Turkish Cypriots. *Mol Biol Rep.* 2019;**46**(3):3349-55. [PubMed ID: 30977086]. <https://doi.org/10.1007/s11033-019-04796-6>.
 31. Darwish H, DeLuca HF. Vitamin D-regulated gene expression. *Crit Rev Eukaryot Gene Expr.* 1993;**3**(2):89-116. [PubMed ID: 8391882].
 32. Shahrokhi SZ, Ghaffari F, Kazerouni F. Role of vitamin D in female reproduction. *Clin Chim Acta.* 2016;**455**:33-8. [PubMed ID: 26747961]. <https://doi.org/10.1016/j.cca.2015.12.040>.
 33. Mahmoudi T. Genetic variation in the vitamin D receptor and polycystic ovary syndrome risk. *Fertil Steril.* 2009;**92**(4):1381-3. [PubMed ID: 19501823]. <https://doi.org/10.1016/j.fertnstert.2009.05.002>.
 34. Williams S, Malatesta K, Norris K. Vitamin D and chronic kidney disease. *Ethn Dis.* 2009;**19**(4 Suppl 5):S5-8-11. [PubMed ID: 20077598]. [PubMed Central ID: PMC2878736].
 35. Shaymaa HM, Al-Zubaidy R. Genotyping of vitamin D receptor FOKI polymorphism as a predictor for type 2 diabetes mellitus by a tetra primer-ARMS-PCR assay. *Gene Rep.* 2019;**15**:100362.
 36. Hu W, Wang L, Chen B, Wang X. Vitamin D receptor rs2228570 polymorphism and Parkinson's disease risk in a Chinese population. *Neurosci Lett.* 2020;**717**:134722. [PubMed ID: 31877334]. <https://doi.org/10.1016/j.neulet.2019.134722>.