



Synthesis of Zinc Nanoparticles in *Hibiscus sabdariffa* Plant Extract and Investigation of Its Antimicrobial Properties

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Abstract

Background: The use of nanotechnology in various biomedical and pharmaceutical fields is expanding rapidly.

Objectives: The aim of this study was to synthesize zinc nanoparticles using *Hibiscus sabdariffa* plant extract and investigate their antimicrobial properties.

Methods: In this experimental study, the aqueous extract of sour tea was mixed with a 0.1 M zinc sulfate solution at a 1:1 ratio and allowed to react for 15 minutes at room temperature to produce nanoparticles. The synthesis of zinc oxide nanoparticles was confirmed using spectrophotometric methods, measuring the average diameter of nanoparticles, X-ray diffraction, and scanning electron microscopy (SEM). The minimum inhibitory concentration (MIC) of the zinc oxide nanoparticles was then measured against standard strains.

Results: The results showed that the largest inhibition zone diameter was observed at a concentration of 1024 µg/mL against *Listeria monocytogenes*, while the smallest inhibition zone was observed against *Aeromonas hydrophila*. At a concentration of 512 µg/mL, an inhibition zone of 10 mm was developed against two strains: *Vibrio cholerae* and *Escherichia coli*.

Conclusions: Based on the results of this study, zinc oxide nanoparticles, along with the sour tea aqueous extract and zinc sulfate, exhibited antibacterial properties. However, the most pronounced antimicrobial effects were observed with the zinc oxide nanoparticles.

Keywords: *Hibiscus Sabdariffa*, Zinc Nanoparticles, Biosynthesis, Antimicrobial Activity

1. Background

Using herbal compounds to treat diseases is an ancient practice, and until the 19th century, the use of natural resources, mainly plants, was one of the primary methods for treating diseases. The expansion of various scientific fields, such as phytochemical drugs and pharmacology, introduced the use of chemicals in the production of antibacterial drugs. However, due to their unnecessary and incorrect use, scientists have been compelled to revisit plant compounds for the treatment of infectious diseases (1, 2).

Plants, particularly those long used in traditional and folk medicine to combat microbial infections, can be valuable sources for developing new antimicrobial drugs (3).

In Sudan, as in many African countries, the majority of people still rely on traditional or folk medicine for disease treatment. This approach is an integral part of an informal healthcare system, with roots in Islamic and West African medicine (4).

Nanotechnology has gained significant popularity over the last decade due to its important applications and foundations in various scientific fields (5). Among metal nanoparticles, silver and gold nanoparticles, and among metal oxide nanoparticles, titanium dioxide nanoparticles are commonly used in medical devices, electronics, and the food industry due to their unique properties (6, 7).

Historically, nanoparticles have been synthesized only through physical and chemical methods, which are typically expensive and require high temperatures and pressures. Additionally, these methods can be toxic and

harmful to the environment and living organisms due to the production of toxic chemicals. Recent developments highlight the important role of biological systems (green methods) in the synthesis of metal nanoparticles, presenting a viable alternative to traditional methods. The low-cost green methods are simple, environmentally friendly, and capable of large-scale synthesis (8, 9).

Nanoparticles are known for their non-toxic and biocompatible nature, making them highly suitable for a range of biomedical applications. These include anticancer (10), anti-inflammatory (11), and antimicrobial properties, as well as targeted drug delivery (12). They also exhibit capabilities in wound healing and biological imaging (13, 14).

Nanoparticles can be synthesized using various methods, including chemical, physical, and biosynthesis techniques, each offering a wide array of properties and applications. While plant-based synthesis of ZnO-NPs has been documented, there is still limited literature on their diverse biological properties, such as antimicrobial, antilarvicidal, protein kinase, and anticancer activities.

Sour tea (*Hibiscus sabdariffa* L.), a member of the *Malvaceae* family, is a small tropical annual shrub native to Africa and also found in Southeast Asia and Central America (15, 16). Locally known as Karkede, *H. sabdariffa* is well-regarded internationally. Various parts of *H. sabdariffa* are utilized in traditional medicine across numerous countries, including those in Africa, India, Mexico, Brazil, China, and Iran (17).

The leaves, when consumed as a vegetable, possess diuretic, antiseptic, digestive, purgative, sedative, and astringent properties (18, 19). Although the seeds are less commonly mentioned in traditional medicine compared to other parts of the plant, they are smoked and consumed as food and traditionally used as a demulcent, laxative, and tonic (20).

2. Objectives

The purpose of this study is to investigate the synthesis of zinc nanoparticles using the extract of the sour tea plant and to explore its antimicrobial properties.

3. Methods

Mix 5 grams of powder prepared from the sour tea plant with 100 cc of deionized water. Heat the mixture on the stove until it reaches boiling temperature and maintain it for 15 minutes. After it cools, strain it using

Whatman No. 1 filter paper and use the resulting aqueous extract for subsequent tests. The extract should be stored at 4°C.

3.1. Synthesis of Nanoparticles

10 cc of the prepared extract was mixed with 90 cc of a 1 mM zinc salt solution, and the mixture was placed on a magnetic stirrer at laboratory temperature for 24 hours. To observe color changes, the absorbance of the solution was measured using a spectrophotometer in the range of 300 - 700 nm. The prepared nanoparticle solution was then centrifuged at 1200 rpm for 15 minutes, and the supernatant was discarded.

3.2. Vegetative Electron Microscope

The shape and size of silver oxide nanoparticles were examined using scanning electron microscopy (SEM). To do this, 15 microliters of the zinc nanoparticle solution were applied to specialized SEM grids. After drying, the antibacterial properties of the nanoparticles were assessed.

4. Results

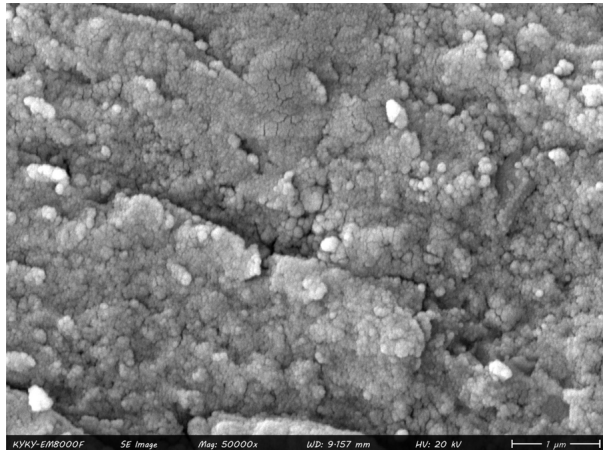
The results of this study indicated that the largest diameter of the inhibition zone was observed at a concentration of 1024 µg/mL against *Listeria monocytogenes* bacteria (Figure 1), while the smallest inhibition zone was noted for *Aeromonas hydrophila* bacteria. At a concentration of 512 µg/mL, the diameter of the inhibition zone was 10 mm against two strains of *Vibrio cholerae* and *Escherichia coli* bacteria (Figure 2 and Table 1). The image of the synthesized nanoparticles is shown in Figure 3.



Figure 1. The diameter of the inhibition zone against *L. monocytogenes* in different concentrations of nanoparticles

Table 1. Diameter of Sour Tea Nanoparticles Synthesized in Sour Tea Medicinal Plant (mm)

Variables	1024 µg/mL	512 µg/mL	256 µg/mL
<i>Vibrio cholerae</i>	14	10	5
<i>Escherichia coli</i>	15	10	5
<i>Listeria monocytogenes</i>	20	15	6
<i>Aeromonas hydrophila</i>	4	3	1

**Figure 2.** The diameter of the inhibition zone against *E.coli* in different concentrations of nanoparticles**Figure 3.** Electron microscope photo of synthesized nanoparticles

5. Discussion

In the study by Tabatabai Yazdi et al., the antibacterial effects of sour tea extracts were examined in vitro against several antibiotic-resistant pathogenic bacteria. The results revealed that *E.coli* showed resistance to penicillin (75.9%), erythromycin (58.3%), tetracycline (Tet) (56.9%), and cefixime (37%), while *Staphylococcus*

aureus exhibited resistance to penicillin (83%), cefixime (80%), erythromycin (55.6%), and Tet (26.1%). This study demonstrated the beneficial effect of the ethanolic extract of *H.sabdariffa* on antibiotic-resistant strains of *E.coli* and *S.aureus*, with the minimum inhibitory concentration (MIC) of the extract being 16 mg/mL for *E.coli* and 4 mg/mL for *S.aureus* (21).

In the study by Rashidi et al., the antioxidant and antimicrobial effects of ethanolic extracts of sour tea and tea grass were investigated. The antibacterial properties of these extracts against pathogenic bacteria (*Salmonella enterica*, *Bacillus cereus*, *E.coli*, and *S.aureus*) were assessed using the well diffusion method. Additionally, the antioxidant activity of the extracts was measured using the ABTS method, with results compared to the antioxidant capacity of Trolox. The findings indicated that *S.aureus* was the most sensitive bacteria to both tea grass and sour tea extracts. There was no significant difference between the two plants regarding their antibacterial properties against this microorganism (22).

In another study, the antibacterial test results indicated that the methanolic extract of *H. sabdariffa* calyces contained effective antibacterial agents. This extract demonstrated a significant zone of inhibition against all tested gram-negative and gram-positive bacteria, performing as well as or better than gentamicin, and substantially more effective than penicillin, which showed weak or no effect (23).

In a study comparing the antimicrobial effects of aqueous extract of red rose (RE), chlorhexidine (CH), amoxicillin-clavulanic acid (ACA), Tet, and metronidazole (Met) against *Streptococcus mutans*, *S.aureus*, and *Enterococcus faecalis*, it was found that at a dilution of 25 mg/mL, the diameters of the inhibition zones were 9.1 mm for *S. mutans*, 7.5 mm for *S. aureus*, and 8 mm for *E. faecalis* (24).

In another study, the antimicrobial activity of sour tea extract was investigated against *Streptococcus mitis* and *Streptococcus oralis* bacteria. The results demonstrated that sour tea extract inhibited 29.1% of *S. mitis* and 63.23% of *S. oralis* bacteria (25).

Marquez-Rodriguez et al. explored the in vitro antibacterial activity of the phenolic extract of sour tea and its application in extending the shelf life of beef. The study found that the minimum inhibitory concentrations against *E. coli*, *S. enterica*, *S. aureus*, *L. monocytogenes*, and *B.cereus* were 300, 300, 200, 200, and 200 mg/mL, respectively (26).

Anvarinezhad et al. conducted a study on the green synthesis of zinc oxide nanoparticles using clove extract and three different heating methods. They evaluated the properties of the nanoparticles and found that the average crystallite sizes of nanoparticles synthesized using microwave, autoclave, and stirring heater methods were 45, 50, and 52 nm, respectively. The antioxidant properties of the nanoparticles were 89%, 85%, and 80% in inhibiting free radicals, and their methylene blue color removal properties were 79%, 70%, and 66%, respectively. The synthesized nanoparticles also exhibited high antibacterial activity against both *E.coli* and *S.aureus* (27).

A study aimed at synthesizing zinc oxide nanoparticles under laboratory conditions evaluated their antimicrobial properties for inhibiting biofilm formation and eradicating *Klebsiella pneumoniae* biofilm. The synthesized zinc oxide nanoparticles had a circular structure with a size of 30 nm. Biofilm formation by *K. pneumoniae* was assessed using the microtiter plate method. The anti-biofilm activity and biofilm eradication of the zinc oxide nanoparticles were observed at concentrations of 50 and 500 micrograms/mL, respectively.

Ranjbar et al. conducted a study to synthesize green zinc nanoparticles using the extract of brown algae *Sargassum ilicifolium* and evaluate their antibacterial properties. The results showed that the formed nanoparticles were spherical and crystalline, with sizes ranging from 15.1 to 27 nm. The antibacterial activity of the biosynthesized zinc oxide nanoparticles was assessed using MIC and MBC methods against *S.aureus* and *E.coli*. The results indicated that the biosynthesized zinc nanoparticles exhibited significant antibacterial effects against the tested bacteria (28).

In the study by Soosani et al., which aimed to achieve green and extracellular synthesis of zinc oxide nanoparticles using a cell-free extract from *Rhodotorula pacifica* NSO₂, the antimicrobial properties of the nanoparticles were investigated. The results showed that zinc oxide nanoparticles with an average size of 42.6 nm were obtained using the cell-free extract solution. Due to the small size and proper distribution of the nanoparticles, a significant inhibitory effect was

observed against the tested clinical bacterial isolates (29).

In a study, zinc nanoparticles were synthesized using the aqueous extract of *Limonium pruinosum* L. Chaz. TEM images revealed that the green ZnO nanoparticles had a hexagonal/cubic shape with an average size of approximately 41 nm. The results demonstrated that both the synthesized zinc oxide nanoparticles and the plant extract exhibited the highest inhibition zones against *E.coli*, measuring 29 mm and 31 mm, respectively, followed by *C. albicans* with inhibition zones of 28 mm and 29 mm (30).

In a study, nanoparticles were synthesized using the aqueous extract of *Myristica fragrans* fruit. The findings indicated that *K.pneumoniae* was the most sensitive strain to both the nanoparticles (27 ± 1.73 mm) and the nanoparticles coated with imipenem (26 ± 1.5 mm) (31).

5.1. Conclusions

The results demonstrated that the nanoparticles synthesized from the sour tea medicinal plant exhibit a very high inhibitory effect against fish pathogenic bacteria.

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Footnotes

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Data Availability: The dataset presented in the study is available on request from the corresponding author during submission or after publication.

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References

1. Mercy R, David Udo E. Natural products as lead bases for drug discovery and development. *Res Rep Med Sci*. 2018;2(1):1-2.

2. Al-Rifai A, Aqel A, Al-Warhi T, Wabaidur SM, Al-Othman ZA, Badjah-Hadj-Ahmed AY. Antibacterial, Antioxidant Activity of Ethanolic Plant Extracts of Some Convolvulus Species and Their DART-ToF-MS Profiling. *Evid Based Complement Alternat Med*. 2017;**2017**:5694305. [PubMed ID: 29317894]. [PubMed Central ID: PMC5727833]. <https://doi.org/10.1155/2017/5694305>.
3. Abdallah EM. Plants: An alternative source for antimicrobials. *J Appl Pharmaceutical sci*. 2011;(Issue):16-20.
4. World Health Organization. *Legal Status of Traditional Medicine and Complementary Alternative Medicine: A Worldwide Review*. Geneva, Switzerland: World Health Organization; 2001. Available from: <https://www.who.int/publications/i/item/WHO-EDM-TRM-2001.2>.
5. Ahmad N, Alam MK, Shehbaz A, Khan A, Mannan A, Hakim SR, et al. Antimicrobial activity of clove oil and its potential in the treatment of vaginal candidiasis. *J Drug Target*. 2005;**13**(10):555-61. [PubMed ID: 16390816]. <https://doi.org/10.1080/10611860500422958>.
6. Vivek R, Thangam R, Muthuchelian K, Gunasekaran P, Kaveri K, Kannan S. Green biosynthesis of silver nanoparticles from *Annona squamosa* leaf extract and its in vitro cytotoxic effect on MCF-7 cells. *Process Biochem*. 2012;**47**(12):2405-10.
7. Roopan SM, Madhumitha G, Rahuman AA, Kamaraj C, Bharathi A, Surendra TV. Low-cost and eco-friendly phyto-synthesis of silver nanoparticles using *Cocos nucifera* coir extract and its larvicidal activity. *Industrial Crops and Products*. 2013;**43**:631-5.
8. Makarov VV, Love AJ, Sinitsyna OV, Makarova SS, Yaminsky IV, Taliensky ME, et al. "Green" nanotechnologies: synthesis of metal nanoparticles using plants. *Acta Naturae (англоязычная версия)*. 2014;**6**(1(20)):35-44.
9. Iravani S. Green synthesis of metal nanoparticles using plants. *Green Chem*. 2011;**13**(10):2638-50.
10. Mishra PK, Mishra H, Ekielski A, Talegaonkar S, Vaidya B. Zinc oxide nanoparticles: a promising nanomaterial for biomedical applications. *Drug Discov Today*. 2017;**22**(12):1825-34. [PubMed ID: 28847758]. <https://doi.org/10.1016/j.drudis.2017.08.006>.
11. Nagajyothi PC, Cha SJ, Yang JJ, Sreekanth TV, Kim KJ, Shin HM. Antioxidant and anti-inflammatory activities of zinc oxide nanoparticles synthesized using *Polygala tenuifolia* root extract. *J Photochem Photobiol B*. 2015;**146**:10-7. [PubMed ID: 25777265]. <https://doi.org/10.1016/j.jphotobiol.2015.02.008>.
12. Cai X, Luo Y, Zhang W, Du D, Lin Y. pH-Sensitive ZnO Quantum Dots-Doxorubicin Nanoparticles for Lung Cancer Targeted Drug Delivery. *ACS Appl Mater Interfaces*. 2016;**8**(34):22442-50. [PubMed ID: 27463610]. <https://doi.org/10.1021/acsami.6b04933>.
13. Gutha Y, Pathak JL, Zhang W, Zhang Y, Jiao X. Antibacterial and wound healing properties of chitosan/poly(vinyl alcohol)/zinc oxide beads (CS/PVA/ZnO). *Int J Biol Macromol*. 2017;**103**:234-41. [PubMed ID: 28499948]. <https://doi.org/10.1016/j.ijbiomac.2017.05.020>.
14. Lai L, Zhao C, Su M, Li X, Liu X, Jiang H, et al. In vivo target bio-imaging of Alzheimer's disease by fluorescent zinc oxide nanoclusters. *Biomater Sci*. 2016;**4**(7):1085-91. [PubMed ID: 27229662]. <https://doi.org/10.1039/c6bm00233a>.
15. Voon HC, Bhat R, Rusul G. Flower extracts and their essential oils as potential antimicrobial agents for food uses and pharmaceutical applications. *Compr Rev Food Sci Food Safety*. 2012;**11**(1):34-55.
16. Morales-Cabrera M, Hernández-Morales J, Leyva-Rúelas G, Salinas-Moreno Y, Soto-Rojas L, Castro-Rosas J. Influence of variety and extraction solvent on antibacterial activity of roselle (*Hibiscus sabdariffa* L.) calyces. *J Med Plants Res*. 2013;**7**(31):2319-22.
17. Da-Costa-Rocha I, Bonnlaender B, Sievers H, Pischel I, Heinrich M. *Hibiscus sabdariffa* L. - a phytochemical and pharmacological review. *Food Chem*. 2014;**165**:424-43. [PubMed ID: 25038696]. <https://doi.org/10.1016/j.foodchem.2014.05.002>.
18. Obouayeba AP, Djyh NB, Diabate S, Djaman AJ, N'guessan JD, Kone M, et al. Phytochemical and antioxidant activity of roselle (*Hibiscus sabdariffa* L.) petal extracts. *Res J Pharm Bio Chem Sci*. 2014;**5**.
19. Ewansiha JU. Evaluation of the antimicrobial activity of roselle (*Hibiscus sabdariffa* L.) leaf extracts and its phytochemical properties. *Peak J Med Plant Res*. 2014;**2**(1):1-5.
20. Ismail A, Ikram EHK, Nazri HSM. Roselle (*Hibiscus sabdariffa* L.) seeds nutritional composition protein quality and health benefits. *Food*. 2008;**2**(1):1-16.
21. Tabatabaei Yazdi F, Alizadeh Behbahani B, Vagadi A, Mortazavi SA, Moradi S. [Antibacterial effect of sour tea extracts on some antibiotic-resistant pathogenic bacteria in vitro]. *Iran Food Sci Industry Assoc*. 2016;**13**(55):23-31. Persian.
22. Rashidi M, Moslehi SH, Ziarati P, Qamari F. [Study of antioxidant and antibacterial effects of ethanolic extract of sour tea and tea grass against *Salmonella enterica*, *Bacillus cereus*, *Escherichia coli* and *Staphylococcus aureus*]. *Food Industry Res (Agricultural Knowledge)*. 2017;**28**(3):45-37. Persian.
23. Abdallah EM. Antibacterial efficiency of the Sudanese Roselle (*Hibiscus sabdariffa* L.), a famous beverage from Sudanese folk medicine. *J Intercult Ethnopharmacol*. 2016;**5**(2):186-90. [PubMed ID: 27104041]. [PubMed Central ID: PMC4835995]. <https://doi.org/10.5455/jice.20160320022623>.
24. Abass AA, Al-Magsoosi MJN, Kadhim WA, Mustafa R, Ibrahim SA, Aljdaimi AI, et al. Antimicrobial effect of Red Roselle (*Hibiscus Sabdariffa*) against different types of oral bacteria. *J Med Life*. 2022;**15**(1):89-97. [PubMed ID: 35186141]. [PubMed Central ID: PMC8852637]. <https://doi.org/10.25122/jml-2021-0184>.
25. Salah R, Haleem AM, Mazin H. Potential effect of Roselle (*Hibiscus sabdariffa*) ethanolic extract against *Streptococcus mitis* and *Streptococcus oralis*. *J Herbal Med*. 2023;**41**:100710.
26. Marquez-Rodriguez AS, Nevarez-Baca S, Lerma-Hernandez JC, Hernandez-Ochoa LR, Nevarez-Moorillon GV, Gutierrez-Mendez N, et al. In Vitro Antibacterial Activity of *Hibiscus sabdariffa* L. Phenolic Extract and Its In Situ Application on Shelf-Life of Beef Meat. *Food*. 2020;**9**(8). [PubMed ID: 32784385]. [PubMed Central ID: PMC7464790]. <https://doi.org/10.3390/foods9081080>.
27. Anvarinezhad M, Jafarizadeh-Malmiri H, Javadi A, Azadmard-Damirchi S. [Green Synthesis of Zinc Oxide Nanoparticles Using Clove Extract by Three Different Heating Methods and Evaluation of their Properties]. *Iran Chem Engin J*. 2021;**20**(118):78-87. Persian. <https://doi.org/10.22034/ijche.2021.276471.1094>.
28. Ranjbar MS, Jangizahi S, Yousefzadi M, Attaran Fariman G, Zarei M. [Green synthesis of zinc oxide nanoparticles using brown algae *Sargassum ilicifolium* extract and evaluation of its antibacterial properties]. *Aquatic Physiol Biotechnol*. 2023. Persian. <https://doi.org/10.22124/japb.2023.25411.1510>.
29. Soosani N, Ashengroph M, Chehri K. Extracellular green synthesis of zinc oxide nanoparticle by using the cell-free extract *Rhodotorula pacifica* NS02 and investigation of their antimicrobial activities. *Nova Biologica Reperta*. 2021;**8**(3):195-205.
30. Naiel B, Fawzy M, Halmy MWA, Mahmoud AED. Green synthesis of zinc oxide nanoparticles using Sea Lavender (*Limonium pruinosum* L. Chaz.) extract: characterization, evaluation of anti-skin cancer, antimicrobial and antioxidant potentials. *Sci Rep*. 2022;**12**(1):20370. [PubMed ID: 36437355]. [PubMed Central ID: PMC9701696]. <https://doi.org/10.1038/s41598-022-24805-2>.
31. Faisal S, Jan H, Shah SA, Shah S, Khan A, Akbar MT, et al. Green synthesis of zinc oxide (ZnO) nanoparticles using aqueous fruit extracts of *Myristica fragrans*: their characterizations and biological and environmental applications. *ACS omega*. 2021;**6**(14):9709-22.