



Investigation of Antimicrobial Resistance Pattern Among *Escherichia coli* Strains Isolated from Patients Referred to Amir Al-Momenin Hospital, Gerash, Iran

Aliyar Pirouzi ¹, Hossain Foruozaandeh ^{1*}, Abbas Farahani², Jebreil Shamseddin ², Hamideh Mohseni³, Ahmad Abdollahi¹, Elham Rezaee¹, Zahra Forouzandeh⁴, Parisa Mohseni⁵ and Fatemeh Fariyabi⁶

¹Cellular and Molecular Research Center, Gerash University of Medical Sciences, Gerash, Iran

²Infectious and Tropical Diseases Research Center, Hormozgan Health Institute, Hormozgan University of Medical Sciences, Bandar Abbas, Iran

³Islamic Azad University of Larestan, Larestan, Iran

⁴Nursing School, Jahrom University of Medical Sciences, Jahrom, Iran

⁵Shahid Beheshti University of Medical Sciences, Tehran, Iran

⁶Blood Transfusion Research Center, High Institute for Research and Education in Transfusion Medicine, Shiraz, Iran

*Corresponding author: Blood Transfusion Research Center, High Institute for Research and Education in Transfusion Medicine, Shiraz, Iran. Email: hosainforuozaandeh@yahoo.com

Received 2019 August 28; Revised 2019 November 30; Accepted 2019 December 02.

Abstract

Background: Antibiotic resistance is now considered as a global problem. Due to the increasing use of antibiotics, and subsequent increase in antibiotic resistance, treatment of bacterial infections has faced many difficulties.

Objectives: This study aimed to investigate the pattern of antibiotic resistance in *Escherichia coli* (*E. coli*) isolated from patients referred to the hospital during the years 2016 - 2018.

Methods: In this cross-sectional study, 2250 samples were taken from outpatient and hospitalized patients (inpatients) during three years (2016 - 2018). After collection, the specimens were cultured in blood agar and EMB media. One thousand six hundred and forty-two positive samples were obtained that from these, a number of 1067 specimens were recognized as *E. coli*. Antibiotic susceptibility was determined using a disk diffusion method on Mueller Hinton agar media for eight antibiotics. The result was evaluated based on the Clinical Laboratory Standards Institute (CLSI).

Results: In this study, 1642 positive samples were diagnosed that the largest isolated strains were associated with *E. coli* (65%). About 241 samples belonged to men (22.6%) and 826 samples were isolated from women (77.4%). Based on the findings, most cases of resistance were related to co-trimoxazole with 52.6% and the highest sensitivity was related to nitrofurantoin with 81%.

Conclusions: The results of this study indicated that the pattern of antimicrobial resistance is increasing and urinary tract infection (UTI) caused by *E. coli* isolates with a high rate of resistance still remains an important healthcare problem. Therefore, continuous evaluation of the pattern of resistance in different areas is necessary for promotion of antibacterial management.

Keywords: *Escherichia coli*, Antimicrobial Resistance, Outpatients, Inpatients

1. Background

Since recognizing bacteria, humans have been always looking for effective medications against infections caused by these microorganisms. In addition, bacteria have achieved effective mechanisms to eliminate antibiotic effects. Following the development of antibiotic-resistant organisms, nowadays, we have had several reports on widespread outbreaks in different wards of hospitals (1, 2). Development of drug resistance has been considered as a common situation. The treatment of this type of illness

has caused many problems (3, 4). *E. coli* is ubiquitous and found in water, soil, plants and makes the normal flora of human and animal intestines.

This bacterium is the most important common microbial agent of urinary tract infections. It is the cause of many hospital opportunistic infections such as sepsis, gastric wounds, urethritis, and neonatal meningitis (5-7). This organism is resistant to beta-lactam antibiotics due to the acquisition of plasmids that encode broad-spectrum beta-lactamases. Based on these facts, the treatment of *E. coli* infection has been problematic (8).

Overuse and misuse of antibiotics along with factors such as long-term hospitalization of patients and the use of such devices such urinary and intravascular catheters has led to the formation of multi-drug resistant bacteria and increasing the pattern of drug resistance in the hospital (9, 10).

2. Objectives

With the increasing use of antibiotics and subsequent resistance development, we need to evaluate the patterns of resistance in all regions continuously. Therefore, this study aimed to evaluate the antibiotic resistance pattern of *E. coli* in patients referred to the hospital during the years 2016 - 2018.

3. Methods

3.1. Bacterial Isolates

In this cross-sectional study, 2250 specimens of outpatient and inpatients obtained from Gerash hospital in three years of 2016 - 2018 were studied. After collecting the specimens, they were cultured in blood agar and eosin methylene blue (EMB) agar media by standard loop and incubated in 37°C, for 24 hours.

A sample was considered positive for urinary tract infection (UTI) in the light of the number of yielded colonies (10^5 cfu per milliliter) and the cytology of the urine through microscopic detection of bacteriuria and PMNs (≥ 8 leukocytes/mm³). However, lower colony counts associated with significant pyuria or low PMN count associated with significant colony counts was considered and analyzed in the light of the clinical picture and the patient's immunological status (11). Also, the study excluded patients if they were pregnant, planned to become pregnant for woman, had used systemic antimicrobial agents within 14 days, had a chronic illness, or had a known anatomical or functional abnormality of the urinary tract. Afterwards the genus and species of isolates were determined based on standard methods and biochemical characteristics of isolates, such as oxidase test, nitrate revitalization, Simmon's citrate, TSI reaction, urease, indole, movement, MR/VP and lysine decarboxylase and finally all *E. coli* isolates were confirmed using API20E kit (bioMerieux SA, Marcy l'Etoile, France).

3.2. Antimicrobial Susceptibility Testing

Among the 1642 positive samples, 1067 nonduplicate cases were diagnosed as *E. coli* infection. In this study, patterns of antimicrobial resistance were studied using the disk diffusion method. Isolates were cultured

on the Mueller Hinton agar medium (Merck, Germany) for susceptibility test against several types of antibiotics with appropriate disks containing ceftriaxone (30 µg), gentamicin (10 µg), ciprofloxacin (5 µg), cefixime (30 µg), amikacin (30 µg), nitrofurantoin (300 µg), nalidixic acid (30 µg), and, trimethoprim-sulfamethoxazole (1.25/23.75 µg) antibiotics (MAST, Merseyside, UK), according to the recommendation of the Clinical Laboratory Standards Institute (CLSI) (12). The strain *E. coli* ATCC 25922 and *P. aeruginosa* ATCC 27853 were used as positive control.

3.3. Statistical Analysis

SPSS version 22 (IBM, Chicago, IL, USA) was used to analyze the data and a P value of < 0.05 considered to indicate statistical significance.

4. Results

In this center, during the years of 2016 - 2018, 2250 samples were submitted for cultivation, 1642 positive cultivations were obtained, based on the bacteriology results, of which 1067 (65%) were from *E. coli* strain. Of the total positive cultures for *E. coli*, 241 samples were related to men (22.6%) and 826 samples belonged to women (77.4%). The age range of patients in this study was 1 - 100 years. Based on the findings, the highest sensitivity seen in antibiotics nitrofurantoin (81%), gentamicin (73.9%), amikacin (71.9%), ceftriaxone (70.24%), ciprofloxacin (67.6%), cefixime (62.9%), nalidixic acid (48.4%), trimethoprim-sulfamethoxazole (46.5%) (in that order) (Figure 1).

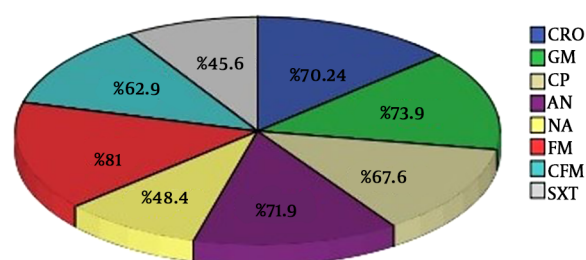


Figure 1. The percentage of antibiotic sensitivity among *Escherichia coli* isolates. CRO: ceftriaxone; GM: gentamicin; CP: ciprofloxacin; AN: amikacin; NA: nalidixic acid; FM: nitrofurantoin; CFM: cefixime; SXT: cotrimoxazole.

On the other hand, the maximum resistance rates were observed in trimethoprim-sulfamethoxazole (52.6%), nalidixic acid (49.4%), cefixime (34.3%), ciprofloxacin (29.8%), ceftriaxone (28.3%), gentamicin (17.7%), nitrofurantoin (12%) and amikacin antibiotics (9.8%) respectively (Figure 2).

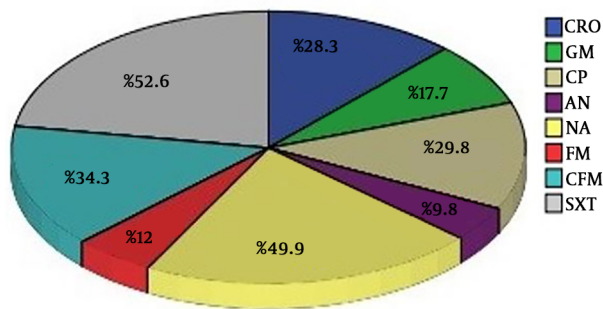


Figure 2. The percentage of antibiotic resistant among *Escherichia coli* isolates. CRO: ceftriaxone; GM: gentamicin; CP: ciprofloxacin; AN: amikacin; NA: nalidixic acid; FM: nitrofurantoin; CFM: cefixime; SXT: cotrimoxazole.

In the current study, via a chi-square test, a comparison was made between the percentage of resistance of different antibiotics among women and men. It was identified that the antibiotic resistance percentages of cefixime, ciprofloxacin, ceftriaxone, gentamicin, and amikacin had a significant difference between women and men. (P value < 0.05). Antibiotic resistance in men was higher than in women (Figure 3).

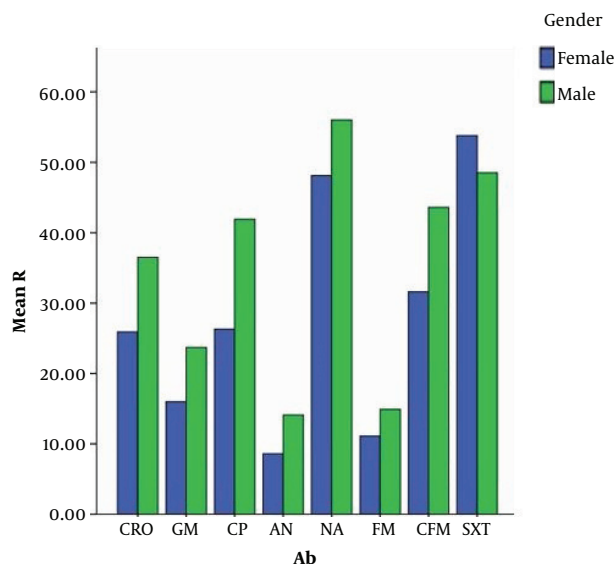


Figure 3. Antibiotic resistance according to sex

Also, through this test a significant relationship (P value < 0.05) was observed between the resistance of cefixime, ciprofloxacin, nalidixic acid, ceftriaxone, gentamicin and amikacin antibiotics and different years (2016 - 2018). Therefore, with increasing years, we observed an in-

crease in resistance (Figure 4).

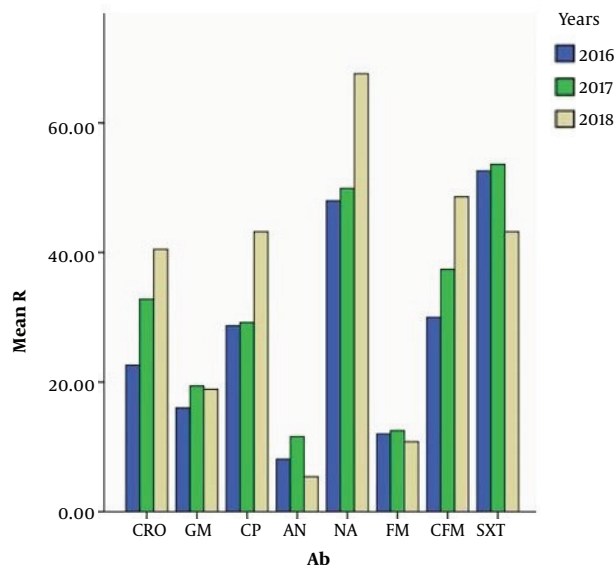


Figure 4. Antibiotic resistance according to year

In this study, the relationship between antibiotic resistance and different age groups (1 - 100) was studied. According to the statistical analyses, a significant relationship was observed between the resistance of cefixime, ciprofloxacin, ceftriaxone, gentamicin and nalidixic acid antibiotics and the age of people, such that the resistance of cefixime, ciprofloxacin, ceftriaxone, gentamicin, and antibiotics was highest in the age group of 81 - 90 and resistance to nalidixic acid in the age group of 91 - 100 has 100 percent resistant (Table 1).

5. Discussion

In this study, the most resistance is against trimethoprim-sulfamethoxazole antibiotics (52.6%) and the highest rate of sensitivity is related to nitrofurantoin (81%) which is consistent with the study of Mukherjee et al. and similar to the antimicrobial resistance of *E. coli* in the year 2018. In the Mukherjee study, the highest rate of sensitivity was related to nitrofurantoin (72.5%) (13). Resistance to antibiotics, among pathogenic bacteria, is a topic that has been considered today as a global problem. *E. coli* is one of the most important pathogens which shows resistance against common antibiotics (14, 15). In the study carried out in India by Sharma et al. (16) in order to evaluate the antimicrobial resistance of *E. coli* during 2012 - 2014, it was found that the highest rate of bacteria isolated from patients was *E. coli* with (67.66%). In this

Table 1. Antibiotic Resistance (%) by Age Group

Age	CRO	GM	CP	AN	NA	FM	CFM	SXT
1 - 10	30.3	17	13.3	12.1	44.8	15.2	36.4	60
11 - 20	23	11.5	18	16.4	42.6	6.6	23	54.1
21 - 30	22.9	14.5	21.8	8.9	44.7	11.2	30.2	48.6
31 - 40	23.6	15.5	29.7	7.4	45.3	12.2	31.8	47.3
41 - 50	21.2	18.2	22	7.6	41.7	7.6	28	53
51 - 60	23.4	10.9	40.6	9.4	58.6	10.2	25	46.1
61 - 70	42	26.1	38.6	6.8	56.8	14.8	45.5	54.5
71 - 80	33.3	25.6	47.8	14.4	57.8	18.9	43.3	55.6
81 - 90	50	30.9	58.8	10.3	67.6	8.8	57.4	57.4
91 - 100	42.9	28.6	57.1	28.6	100	28.6	57.1	71.4

Abbreviations: AN, amikacin; CFM, cefixime; CP, ciprofloxacin; CRO, ceftriaxone; FM, nitrofurantoin; GM, gentamicin; NA, nalidixic acid; SXT, cotrimoxazole

research, *E. coli* has shown the most rate of sensitivity to nitrofurantoin. Reduced sensitivity to nitrofurantoin is consonant with a recent study from 1.36 % in 2016 to 15.18 % in 2018.

The results of Barour et al. (17), showed that among the 198 *E. coli* strains, elevated resistance rates were related to ampicillin (59.09%) and tetracycline (43.43%), and low resistance rates were observed for nalidixic acid (8.08%), ciprofloxacin (7.07%), kanamycin (6.56%), cefotaxime (4.54%), chloramphenicol (4.04%), nitrofurantoin (2.52%), ceftazidime (2.02%), gentamicin (1.01%), and however did not find colistin resistance isolates. Their results were accordance with our finding.

Sharma et al. (16) indicated that the rate of resistance of 3rd generation cephalosporins had been accompanied by an increase of 15% in the years of 2012 - 2014. In our study, we have used 3rd generation cephalosporins such as cefixime and ceftriaxone, which increased the resistance of both antibiotics over the years have been reported. The rate of the resistance of *E. coli* against trimethoprim-sulfamethoxazole in the new study has risen over the three years, and it was nearly 100 percent, while in our study the rate of resistance had been 52.6% in 2016, 53.6% in 2017 and 40.3% in 2018. Similar to our results, a study from Iran showed that among 100 *E. coli* isolates collected from different clinical specimens, the maximum resistance was related to amoxicillin (96%), ceftriaxone (78%), ceftazidime (76%), co-trimoxazole (70%), cefotaxime (67%) and ceftazidime (66%) (18). The observation of drug resistance among *E. coli* strains in this study is alarming, and this could be attributed to the indiscriminate and extensive use of antibiotics, particularly beta-lactam antibiotics.

In another study performed by Sanchez et al. (19) in order to evaluate the *E. coli* antimicrobial resistance in 2002 -

2010, it turned out that ciprofloxacin resistance increased from 3.40 to 17.160 and trimethoprim-sulfamethoxazole resistance had increased from 17.9% to 24.26%. The ascending of resistance increase in the Sanchez study is consistent with the recent study. The incidence rate of resistance, an increasing trend in ceftriaxone, ciprofloxacin, nalidixic acid and cefixime and downtrend in nitrofurantoin, gentamicin, amikacin, and trimethoprim-sulfamethoxazole indicated that the resistance is contradictory with trimethoprim-sulfamethoxazole with the Sanchez study. In the study published in 2008 by Dallal et al. (20) of the 188 examined *E. coli* strains, the highest antimicrobial resistance respectively is related to cotrimoxazole (72%), nalidixic acid (69%), ciprofloxacin (99%) and gentamicin 31% which correspond with the rate of resistance in the current study that includes trimethoprim-sulfamethoxazole (52.6%), nalidixic acid (49.9%), ciprofloxacin (29.8%) and gentamicin (17.7%). But the rate of resistance in that was found in India in 2014 with the title: "*E. coli* antimicrobial resistance", the highest resistance included ceftriaxone (71.4%), cotrimoxazole (64.2%), and the highest sensitivity is for amikacin (82.6%), nitrofurantoin (78.2%). The rate of ceftriaxone resistance contradicts with the recent study. Somashekara et al. (21) during a study in 2011 found that the highest rate of *E. coli* microbial resistance is related to nalidixic acid (78.8%), trimethoprim-sulfamethoxazole (75.2%), ciprofloxacin (57.8%) and amikacin (26%) antibiotics while in our study highest rate of resistance was firstly related to trimethoprim-sulfamethoxazole (52.6%) and after that nalidixic acid with 49.4%. Also through this study, the significant relationship (P value < 0.05) was observed between the resistance of cefixime, ciprofloxacin, nalidixic acid, ceftriaxone, gentamicin and amikacin antibiotics and different years (91 - 93). Therefore with

increasing age, we have increase in the resistance. In this study, the relationship between antibiotic resistance and different age groups (1 - 100) was studied. According to the statistical analyses, a significant relationship was observed between the resistance of cefixime, ciprofloxacin, ceftriaxone, gentamicin and nalidixic acid antibiotics and the age of people, such that the resistance of cefixime, ciprofloxacin, ceftriaxone, gentamicin, and antibiotics was highest in the age group of 81 - 90 and resistance to nalidixic acid antibiotic in the age group of 91 - 100 has 100 percent resistant.

5.1. Conclusions

The results of this study showed that antibiotic resistance patterns exist in different regions, and the rate of this resistance is increasing continuously. This pattern is different from region to region due to the geographical disparity, district hygiene level, and the rate of arbitrary administration of antibiotics in different areas. The emersion of resistance to new antibiotics is a matter to be taken seriously, therefore, continuous evaluation of bacteriology, and correct and rapid diagnosis of antibiotic resistance, followed by identifying the correct line of treatment can make a significant contribution to preventing treatment failure, performing quicker treatment and saving time and cost to the patients. By recognition of resistance patterns in each area, we can prevent the resistant gene spread, and help physicians to cure patients more precisely.

Acknowledgments

The authors of this article would like to extend their gratitude to the medical staff of Gerash Amir-Almomenin Hospital and all patients who have cooperated with us in this study.

Footnotes

Authors' Contribution: Aliyar Pirouzi, Abbas Farahani, Hossein Forouzandeh, Jebreil Shamseddin, Hamideh Mohseni, and Parisa Mohseni: Substantial contributions to the conception and design of the work; final approval of the version to be published; agreement to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. Zahra Forouzandeh: Analysis, interpretation of data for the work, final approval of the version to be published. Ahmad Abdollahi, and Elham Rezaee: Agreement to be accountable for all aspects of the work in ensuring that questions related to the

accuracy or integrity of any part of the work are appropriately investigated and resolved, final approval of the version to be published. Fatemeh Fariyabi: Analysis, interpretation of data for the work.

Conflict of Interests: The author declares that he has no conflicts of interest related to the subject matter or materials discussed in this article.

Ethical Approval: This work was approved (Code: IR.GERUMS.REC. 1396.2045) by the Research Ethics Committee of the Grash University of Medical Sciences, Iran.

Funding/Support: This study funded by Grash University of Medical Sciences.

Informed Consent: All patients gave their written informed consent for participation.

References

- Kollef MH, Fraser VJ. Antibiotic resistance in the intensive care unit. *Ann Intern Med.* 2001;**134**(4):298-314. doi: [10.7326/0003-4819-134-4-200102200-00014](https://doi.org/10.7326/0003-4819-134-4-200102200-00014). [PubMed: [11182841](https://pubmed.ncbi.nlm.nih.gov/11182841/)].
- Norozi B, Farahani A, Mohajeri P, Rostami Z. Distribution of ESBL producing Uropathogenic Escherichia coli and carriage of selected β -lactamase genes in Hospital and community isolates in west of Iran. *Ann Trop Med Pub Health.* 2014;**7**(5):219. doi: [10.4103/1755-6783.154823](https://doi.org/10.4103/1755-6783.154823).
- Saeidimehr S, Geravandi S, Rahim F, Yosefi F, Salmanzadeh S, Forouzandeh H, et al. Nosocomial infection rates during one year in Naft Grand Hospital, Ahvaz, Iran. *Jundishapur J Health Sci.* 2015;**7**(4). doi: [10.17795/jjhs-30124](https://doi.org/10.17795/jjhs-30124).
- Fiore M, Maraolo AE, Gentile I, Borgia G, Leone S, Sansone P, et al. Nosocomial spontaneous bacterial peritonitis antibiotic treatment in the era of multi-drug resistance pathogens: A systematic review. *World J Gastroenterol.* 2017;**23**(25):4654-60. doi: [10.3748/wjg.v23.i25.4654](https://doi.org/10.3748/wjg.v23.i25.4654). [PubMed: [28740354](https://pubmed.ncbi.nlm.nih.gov/28740354/)]. [PubMed Central: [PMC5504381](https://pubmed.ncbi.nlm.nih.gov/PMC5504381/)].
- Wagenlehner FM, Naber KG, Weidner W. [Rational antibiotic therapy of urinary tract infections]. *Med Monatsschr Pharm.* 2008;**31**(10):385-90. quiz 391-2. German. [PubMed: [18972869](https://pubmed.ncbi.nlm.nih.gov/18972869/)].
- Sanchez UM, Bello TH, Dominguez YM, Mella MS, Zemelman ZR, Gonzalez RG. [Transference of extended-spectrum beta-lactamases from nosocomial strains of Klebsiella pneumoniae to other species of Enterobacteriaceae]. *Rev Med Chil.* 2006;**134**(4):415-20. Spanish. doi: [10.4067/s0034-98872006000400002](https://doi.org/10.4067/s0034-98872006000400002). [PubMed: [16758075](https://pubmed.ncbi.nlm.nih.gov/16758075/)].
- Mohajeri P, Darfarin G, Farahani A. Genotyping of ESBL producing uropathogenic Escherichia coli in west of Iran. *Int J Microbiol.* 2014;**2014**:276941. doi: [10.1155/2014/276941](https://doi.org/10.1155/2014/276941). [PubMed: [24839441](https://pubmed.ncbi.nlm.nih.gov/24839441/)]. [PubMed Central: [PMC4009276](https://pubmed.ncbi.nlm.nih.gov/PMC4009276/)].
- De Francesco MA, Ravizzola G, Peroni L, Negrini R, Manca N. Urinary tract infections in Brescia, Italy: Etiology of uropathogens and antimicrobial resistance of common uropathogens. *Med Sci Monit.* 2007;**13**(6):BR136-44. [PubMed: [17534228](https://pubmed.ncbi.nlm.nih.gov/17534228/)].
- Wisplinghoff H, Bischoff T, Tallent SM, Seifert H, Wenzel RP, Edmond MB. Nosocomial bloodstream infections in US hospitals: Analysis of 24,179 cases from a prospective nationwide surveillance study. *Clin Infect Dis.* 2004;**39**(3):309-17. doi: [10.1086/421946](https://doi.org/10.1086/421946). [PubMed: [15306996](https://pubmed.ncbi.nlm.nih.gov/15306996/)].
- Behzadnia S, Davoudi A, Rezai MS, Ahangarkani F. Nosocomial infections in pediatric population and antibiotic resistance of the causative organisms in north of Iran. *Iran Red Crescent Med J.* 2014;**16**(2). e14562. doi: [10.5812/ircmj.14562](https://doi.org/10.5812/ircmj.14562). [PubMed: [24719744](https://pubmed.ncbi.nlm.nih.gov/24719744/)]. [PubMed Central: [PMC3965877](https://pubmed.ncbi.nlm.nih.gov/PMC3965877/)].

11. Daoud Z, Salem Sokhn E, Masri K, Matar GM, Doron S. Escherichia coli isolated from urinary tract infections of lebanese patients between 2005 and 2012: Epidemiology and profiles of resistance. *Front Med (Lausanne)*. 2015;2:26. doi: [10.3389/fmed.2015.00026](https://doi.org/10.3389/fmed.2015.00026). [PubMed: [25984513](https://pubmed.ncbi.nlm.nih.gov/25984513/)]. [PubMed Central: [PMC4415468](https://pubmed.ncbi.nlm.nih.gov/PMC4415468/)].
12. Wayne PA. *Clinical and laboratory standards institute. Performance standards for antimicrobial susceptibility testing: Twenty-fourth informational supplement, M100-S24*. Clinical and Laboratory Standards Institute (CLSI); 2014.
13. Mukherjee M, Basu S, Mukherjee SK, Majumder M. Multidrug-resistance and extended spectrum beta-lactamase production in uropathogenic E. coli which were isolated from hospitalized patients in Kolkata, India. *J Clin Diagn Res*. 2013;7(3):449-53. doi: [10.7860/CDR/2013/4990.2796](https://doi.org/10.7860/CDR/2013/4990.2796). [PubMed: [23634394](https://pubmed.ncbi.nlm.nih.gov/23634394/)]. [PubMed Central: [PMC3616554](https://pubmed.ncbi.nlm.nih.gov/PMC3616554/)].
14. Petty NK, Ben Zakour NL, Stanton-Cook M, Skippington E, Totsika M, Forde BM, et al. Global dissemination of a multidrug resistant Escherichia coli clone. *Proc Natl Acad Sci U S A*. 2014;111(15):5694-9. doi: [10.1073/pnas.1322678111](https://doi.org/10.1073/pnas.1322678111). [PubMed: [24706808](https://pubmed.ncbi.nlm.nih.gov/24706808/)]. [PubMed Central: [PMC3992628](https://pubmed.ncbi.nlm.nih.gov/PMC3992628/)].
15. FarajzadehSheikh A, Veisi H, Shahin M, Getso M, Farahani A. Frequency of quinolone resistance genes among extended-spectrum beta-lactamase (ESBL)-producing Escherichia coli strains isolated from urinary tract infections. *Trop Med Health*. 2019;47:19. doi: [10.1186/s41182-019-0147-8](https://doi.org/10.1186/s41182-019-0147-8). [PubMed: [30872947](https://pubmed.ncbi.nlm.nih.gov/30872947/)]. [PubMed Central: [PMC6399935](https://pubmed.ncbi.nlm.nih.gov/PMC6399935/)].
16. Sharma N, Gupta A, Walia G, Bakhshi R. Pattern of antimicrobial resistance of Escherichia coli isolates from urinary tract infection patients: A three year retrospective study. *J App Pharm Sci*. 2016;6(1):62-5.
17. Barour D, Berghiche A, Boulebdia N. Antimicrobial resistance of Escherichia coli isolates from cattle in Eastern Algeria. *Vet World*. 2019;12(8):195-203. doi: [10.14202/vetworld.2019.1195-1203](https://doi.org/10.14202/vetworld.2019.1195-1203). [PubMed: [31641297](https://pubmed.ncbi.nlm.nih.gov/31641297/)]. [PubMed Central: [PMC6755386](https://pubmed.ncbi.nlm.nih.gov/PMC6755386/)].
18. Kadaei V, Rashki A. Antibiotic resistance pattern and genotype of beta-lactamase producing Escherichia coli isolates from urinary tract infections in Zabol-southeast of Iran. *J Med Microbiol Infect Dis*. 2014;2(4):153-8.
19. Sanchez GV, Master RN, Karlowsky JA, Bordon JM. In vitro antimicrobial resistance of urinary Escherichia coli isolates among U.S. outpatients from 2000 to 2010. *Antimicrob Agents Chemother*. 2012;56(4):2181-3. doi: [10.1128/AAC.06060-11](https://doi.org/10.1128/AAC.06060-11). [PubMed: [22252813](https://pubmed.ncbi.nlm.nih.gov/22252813/)]. [PubMed Central: [PMC3318377](https://pubmed.ncbi.nlm.nih.gov/PMC3318377/)].
20. Dallal MMS, Azarsa M, Shirazi MH, Lari AR, Owlia P, Mehrabadi JF, et al. The prevalence of extended-spectrum beta-lactamases and CTX-M-1 producing Escherichia coli in urine samples collected at Tabriz city Hospitals. *Tehran Univ Med J*. 2011;69(5):273-8.
21. Somashekara SC, Deepalaxmi S, Jagannath N, Ramesh B, Laveesh MR, Govindadas D. Retrospective analysis of antibiotic resistance pattern to urinary pathogens in a tertiary care hospital in South India. *J Basic Clin Pharm*. 2014;5(4):105-8. doi: [10.4103/0976-0105.141948](https://doi.org/10.4103/0976-0105.141948). [PubMed: [25316990](https://pubmed.ncbi.nlm.nih.gov/25316990/)]. [PubMed Central: [PMC4194940](https://pubmed.ncbi.nlm.nih.gov/PMC4194940/)].