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# Detection of Classical Enterotoxins of Staphylococcus aureus Strains Isolated From Raw Meat in Esfahan, Iran

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**Background:** *Staphylococcus aureus* is an important pathogen of humans in both community acquired as well as nosocomial infections. It is also among the four most common causes of food-borne illnesses.

**Objectives:** This study was conducted to determine the prevalence of enterotoxin producing *S. aureus* strains in different raw meat and hamburgers in Isfahan.

**Materials and Methods:** From August to December 2012, 370 samples of raw beef (n = 160; minced and carcass), lamb (n = 80), goat (n = 80), and camel (n = 50) meat were purchased from randomly selected butcheries in Isfahan, Iran, and analyzed for the presence of *S. aureus*. Isolates were also tested for their ability to produce staphylococcal enterotoxins A, B, C, D, and E by enzyme linked immunosorbent assay (ELISA) test.

**Results:** Totally, 223 (60.3%) *S. aureus* were isolated. Among the 223*S. aureus* isolates 30 (13.5%) were found to be enterotoxigenic. Twenty-six (86.7%) were positive only for one type of SEs (14 SEA, 1 SEB, 6 SEC and 5 SED) while the remaining (13.3%) were positive for more than one SEs. None of the isolates were positive for SEE.

**Conclusions:** Only 8.3% of the total meat samples examined in the current study showed this count or above. This low degree of contamination by *S. aureus* is tolerated in most food stuffs and they are not considered a risk for public health. However, we need more epidemiological investigations about enterotoxigenic *S. aureus* isolates and their toxins for better management of food products and to decrease human diseases. The results of this study showed that most *S. aureus* strains isolated from samples produced SEA and SED compared to other SEs.

Keywords: Staphylococcus; Enterotoxins; Meat

## 1. Background

Staphylococcus aureus is an important pathogen of humans in both community acquired as well as nosocomial infections (1, 2). S. aureus is also among the four most common causes of food borne illnesses. Staphylococcal food poisoning (SFP) results from consumption of food contaminated with staphylococcal enterotoxins (SEs) produced by S. aureus (3). SFP is widespread and quite frequent. The number of staphyloenterotoxicosis cases is probably underestimated significantly. There may be many reasons behind this, including: not contacting medical services by many ill people due to the short duration of the disease or mild symptoms and improper sample collection and laboratory examination. The food incriminated in SFP is variable from one country to another and mainly depends on nutritional habits. It has been proved that, the main source of staphylococcus in processed food is humans while animals are the main origin of contamination in raw food (3). The pathogenicity of S. aureus depends on a number of virulence factors

such as the heat stable enterotoxins as well as other enterotoxins (4). S. aureus produces 15 enterotoxins (5), and usually produces one or more of these toxins simultaneously (6, 7). The five classic enterotoxins (SEA, SEB, SEC, SED, and SEE) are known to be responsible for 95% of staphylococcal food poisoning cases and SEA and SED are most common enterotoxins recovered from food poisoning out-breaks (8), possibly because they can be produced in a wide range of growth conditions (aw, pH and Eh) (9). According to Balaban and Rasooly (10), 100 ng of the toxin is enough to induce symptoms when the population reaches 105 colony-forming units per gram (CFU/g). The principal symptom of SFP is vomiting within 1 - 6 hours after eating contaminated food, usually followed by diarrhoea, abdominal cramping, and exhaustion. In more severe cases, additional symptoms can include headache, muscle cramping, and changes in blood pressure and pulse rate. Death from SFP is rare, but can occur among certain high risk people such as infants, elderly; and chronically ill individuals (10). Among the foods implicated in SFP, milk, dairy products and meats, especially handled

Implication for health policy/practice/research/medical education

The content and results of the present communication has identified the risk indicators and it is useful for the policy/decision makers to reduce the food intoxication.

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foods, play an important role since enterotoxigenic strains of S. aureus have frequently been isolated from them (11-13).

## 2. Objectives

Currently, there is limited information regarding the prevalence and enterotoxin production of S. aureus in raw meat in Iran. The purpose of the present study was to determine the prevalence of enterotoxin producing *S*. aureus strains in different raw meat in Isfahan; no similar work has been previously performed in Iran.

## 3. Materials and Methods

#### 3.1. Sample Collection

Four hundred and ten samples of raw beef (n = 160;mince and carcass), lamb(n=80), goat(n=80) and camel(n = 50) were purchased and unpacked from randomly selected butcheries in Isfahan, Iran, From August to December 2012. All samples were placed in separate sterile plastic bags to prevent spilling and cross contamination and were immediately transported to the laboratory and cooled with ice packs.

## 3.2. Isolation and Identification of S. aureus

The samples were processed immediately upon arrival using aseptic techniques. Of each milk sample, 25 mL was homogenized and transferred to 225 mL of buffered peptone water (BPW). The samples were diluted with BPW, and 0.1 mL of diluted samples were streaked on Baird-Parker (BP) agar (Oxoid) supplemented with egg yolk-tellurite emulsion (Oxoid). After inoculation at 37°C for 24 h, typical colonies of S. aureus with similar morphologies, from each selective agar plate were isolated and cultured separately on slants of Brain-Heart Infusion (BHI, Oxoid). The identification was performed using standard microbiological and biochemical procedures including gram staining, production of coagulase, catalase, DNAse and oxidation and fermentation of mannitol (14).

## 3.3. Detect Staphylococcal Enterotoxins S. aureus

To detect staphylococcal enterotoxins, S. aureus isolates were cultured aerobiocally in 10 mL of nutrient broth at 37 °C, overnight. Culture supernatants of the isolated bacteria were then used for detection of S. aureus enterotoxins. Staphylococcal entrotoxins SEA, SEB, SEC, SED and SEE were detected by enzyme linked immunosorbent assay (ELISA) detection kit (RIDASCREEN® SET A, B, C, D, E; R-Biopharm AG, Darmstadt, Germany) according to the manufacture's instructions. The detection limit was 0.1 mg/mL and S. aureus and S. epidermis strains were used as positive and negative controls for each test.

## 3.4. Statistical Analysis

Data were transferred to a Microsoft Excel spreadsheet (Microsoft Corp., Redmond, WA, USA) for analysis. Using SPSS 16.0 statistical software (SPSS Inc., Chicago, IL, USA), chi-square test and fisher's exact two-tailed test analysis were performed and differences were considered significant at values of P < 0.05.

## 4. Results

Table 1 shows the prevalence of S. aureus isolated from beef (minced and carcasses), lamb, goat and camel meat samples in Isfahan, Iran. In this study, 223 of 370 meat samples (60.3%) were contaminated with S. *aureus*. The mean count for meat samples ranged from  $2.6 \times 10$  2 to  $4.8 \times 10$  3 CFU/g. The highest prevalence of S. aureus was found in minced beef (76.3%), followed by lamb meat (68.8%), beef meat (57.5%), goat meat (47.5%), and camel meat (46.0%). There were no significant differences (P > 0.05) in the level of contamination with S. aureus between different meat samples. Among the 223 S. aureus isolates, only 30 (13.5%) were found to be enterotoxigenic (> 100 ng). Twenty-six (86.7%) were positive only for one type of SEs (14 SEA, 1SEB, 6 SEC and 5 SED) while the remaining (13.3%) were positive for more than one SEs (Table 1).

Samples	Samples, No	Positive Sam- ples forS. aureus, No, (%)	Mean ofS. aureusCount, CFU/g <sup>a</sup>	<b>Positive Samples for</b> <b>Enterotoxigenic</b> S. aureus, <b>No</b>	SEs, % <sup>c</sup>					
					SEA <sup>a</sup>	SEB	SEC	SED	SEA + SEC	SEA + SED
Beef meat	80	46 (57.5)	$5.6 \times 10^{2}$	8	5	-	-	2	-	1
Minced meat <sup>b</sup>	80	61 (76.3)	$4.8 \times 10^{3}$	11	6	-	-	3	-	2
Lamb meat	80	55 (68.8)	$7.1 \times 10^{2}$	4	1	-	3	-	-	-
Goat meat	80	38 (47.5)	$2.6 \times 10^{2}$	3	1	-	2	-	-	-
Camel meat	50	23(46.0)	$4.3 \times 10^{2}$	4	1	1	1	-	1	-
Total	370	223 (60.3)	$8.3 \times 10^{2}$	30	14	1	6	5	1	3

Table 1 Provalance and Enterotoxin Production of Stanbulace Isolated From Poof Lamb Coat and Camel Most Sampler

<sup>a</sup> Abbreviations: CFU/g, colony forming units per gram; SEA, staphylococcal enterotoxins A

<sup>C</sup> None of the samples were positive for SEE

<sup>b</sup> Made from raw beef or cow meat

#### 5. Discussion

In this study, 223 of 370 meat samples (60.3%) were contaminated with S. aureus (The mean count ranged from  $2.6 \times 10^2$  to  $4.8 \times 10^3$  CFU/g). Similar results were also reported by Marthenge and Ombui (15) and Al-Tarazi et al. (16) for meat and meat products; however, lower contamination rates (10.5% to 16.4%) have also been reported (17,18). The contamination of food by S. aureus may directly occur due to skin lesions of workers containing bacterin or sneezing and coughing. Approximately 50% of the human population carries S. aureus as commensals. Other contamination sources of S. aureus are soil, water, dust and air (10,14). From 223 S. aureus isolates, 30 (13.3%) were found to be enterotoxigenic (Table 1). The relatively high percentage of classical enterotoxin-forming S. aureus strains from meat samples found in our study is confirmed by previous findings (12,16,19). However, these results were not in agreement with some other investigators including that reported by Marthenge and Ombui. They reported that 66% of the studied samples were contaminated. The examined samples contained 200 raw milk. 100 beef carcass swabs. 50 minced meat samples and 50 chicken carcasses. Variable contamination rates ranging from 21% to 66% have been recorded for food stuff including meat, milk, eggs and their derivatives in different countries (20-24). The difference between the results ofour study and other reports in the prevalence and production of enterotoxins among S. aureus isolates from meat samples may be a result of different sampling techniques employed, seasonal effects, number and kinds of examined samples, and/or laboratory methodologies employed. Recently, new super antigenic enterotoxins (SEG-SEU) have also been described. Due to technical limitations, we were not able to screen the new SEG-SEU, since commercial ELISA test kits for detecting these new enterotoxins are not available. There are no specific standards for the permissible number of S. aureus in fresh or raw meat in Iran; however, 10<sup>3</sup> CFU/g is the highest permissible count of S. aureus commonly specified by the international agencies (25). Only 8.3% of the total meat samples examined in the current study showed this count or above. This low degree of contamination by S. aureus is tolerated for most foodstuffs and is not considered as a risk for public health (3). This is expected because in fresh or chilled meat, S. aureus is not a good competitor with normal microflora (8). However, further molecular based studies are necessary to test enterotoxigenic S. aureus isolates or their toxins for improved management of food products and to decrease human diseases.

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#### **Authors' Contribution**

Study concept and design: Ebrahim Rahimi, Esmail Ataye Salehi. Analysis and interpretation of data: Fatemeh Nonahal, Ebrahim Rahimi, Manuchehr Momeni. All authors contributed to the literature review and writing of the article.

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There is no financial disclosure.

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#### References

- Al-Tarazi YH, Albetar MA, Alaboudi Biptyping AR. Enterotoxigenicity of Staphylococci isolated from fresh and frozen meat marketed in Jordan. *Food Res Int*. 2009;42:374-9.
- 2. Atanassova V, Meindl A, Ring C. Prevalence of Staphylococcus aureus and staphylococcal enterotoxins in raw pork and uncooked smoked ham-a comparison of classical culturing detection and RFLP-PCR. *Int J Food Microbiol*. 2001;**68**:105-13.
- Aitichou M, Henkens R, Sultana AM, Ulrich RG, Sofi Ibrahim M. Detection of Staphylococcus aureus enterotoxin A and B genes with PCR-EIA and a hand-held electrochemical sensor. *Mol Cell Probes.* 2004;18(6):373-7.
- Balaban N, Rasooly A. Staphylococcal enterotoxins. Int J Food Microbiol. 2000;61:1-10.
- Cenci-Goga BT, Karama M, Rossitto PV, Morgante RA, Cullor JS. Enterotoxin production by Staphylococcus aureus isolated from mastitic cows. J Food Prot. 2003;66(9):1693-6.
- Chambers HF. The changing epidemiology of Staphylococcus aureus? Emerg Infect Dis. 2001;7(2):178-82.
- Crago B, Ferrato C, Drews SJ, Svenson LW, Tyrrell G, Louie M. Prevalence of Staphylococcus aureus and methicillin-resistant S. aureus (MRSA) in food samples associated with foodborne illness in Alberta, Canada from 2007 to 2010. *Food Microbiol*. 2012;**32**(1):202-5.
- De Buyser ML, Dufour B, Maire M, Lafarge V. Implication of milk and milk products in food-borne diseases in France and in different industrialised countries. *Int J Food Microbiol*. 2001;67:1-17.
- 9. Gundogan N, Citak S, Yucel N, Devren A. A note on the incidence and antibiotic resistance of Staphylococcus aureus isolated from meat and chicken samples. *Meat Sci.* 2005;**69**(4):807-10.
- Hanson BM, Dressler AE, Harper AL, Scheibel RP, Wardyn SE, Roberts LK, et al. Prevalence of Staphylococcus aureus and methicillin-resistant Staphylococcus aureus (MRSA) on retail meat in Iowa. J Infect Public Health. 2011;4(4):169-74.
- International Commission on Microbiological Specifications for Foods (ICMSF): Microorganisms in foods 2. Sampling for microbiological analysis: Principle and specific application. 2nd ed.: UK, Blackwell Scientific Publications; 1986
- ay MJ, Lossner JM, Golden AD. Staphylococcal gastroenteritis. In: Modern food microbiology. 7th ed. New York, Springer Science; 2005. p. 545-560.
- Karlowsky JA, Jones ME, Draghi DC, Thornsberry C, Sahm DF, Volturo GA. Prevalence and antimicrobial susceptibilities of bacteria isolated from blood cultures of hospitalized patients in the United States in 2002. Ann Clin Microbiol Antimicrob. 2004;3:7.
- 14. Kitai S, Shimizu A, Kawano J, Sato E, Nakano C, Kitagawa H, et al. Prevalence and characterization of Staphylococcus aureus and enterotoxigenic Staphylococcus aureus in retail raw chicken meat throughout Japan. J Vet Med Sci. 2005;67(3):269-74.
- 15. Rahimi E, Mommtaz H, Shakerian A, Kavyani HR. The detection of classical enterotoxins of Staphylococcus aureus in raw cow milk using the ELISA method. *Turk J Vet Anim Sci.* 2012;**36**:319-22.

- Le Loir Y, Baron F, Gautier M. Staphylococcus aureus and food poisoning. *Genet Mol Res.* 2003;31:63-76.
- Marthenge JM, Ombui JN. Detection of staphylococcal enterotoxins in milk and meat in Nairobi Kenya using enzyme linked immunosorbent assay. J Trop Microbiol Biotechnol. 2007;3:23-8.
- Normanno G, Firinu A, Virgilio S, Mula G, Dambrosio A, Poggiu A, et al. Coagulase-positive Staphylococci and Staphylococcus aureus in food products marketed in Italy. Int J Food Microbiol. 2005;98(1):73-9.
- Pereira V, Lopes C, Castro A, Silva J, Gibbs P, Teixeira P. Characterization for enterotoxin production, virulence factors, and antibiotic susceptibility of Staphylococcus aureus isolates from various foods in Portugal. *Food Microbiol.* 2009;26(3):278-82.
- 20. Rahimi E, Safai HG. Detection of classical enterotoxins of Staphylococcus aureus strains isolated from bovine subclinical masti-

tis in Isfahan, Iran. Vet Microbiol. 2010;**141**(3-4):393-4.

- 21. Sally KH, Mark AS. Review of the microbiological standards for foods. *Food Control.* 2003;**14**:391-8.
- Sandel M, McKillip J. Virulence and recovery of Staphylococcus aureus relevant to the food using improvement on traditional approaches. *Food Control*. 2004;15:5-10.
- Tassew H, Abdissa A, Beyene G, Gebre-Selassie S. Microbial flora and food borne pathogens on minced meat and their susceptibility to antimicrobial agents. *Ethiop J Health Sci.* 2010;20(3):137-43.
- Tollersrud T, Kenny K, Caugant DA, Lund A. Characterisation of isolates of Staphylococcus aureus from acute, chronic and subclinical mastitis in cows in Norway. *APMIS*. 2000;**108**(9):565-72.
- 25. Zouharova M, Rysanek D. Multiplex PCR and RPLA Identification of Staphylococcus aureus enterotoxigenic strains from bulk tank milk. *Zoonoses Public Health*. 2008;**55**(6):313-9.