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Research Article

Evaluation of Cytomegalovirus Infections in Liver Transplant Recipients Under Universal Prophylaxis: A Single Centre Experience

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Abstract

Background: Cytomegalovirus (CMV) is one of the leading viral agents that can pave the way for serious complications and organ damage in solid organ transplant (SOT) recipients after transplantation. Strategies have been developed to protect at-risk patients from CMV infection following transplantation. Since more than 90% of adults in Turkey were positive for CMV IgG, universal CMV prophylaxis was applied, and the results were evaluated.

Objectives: This study aimed to evaluate the results of universal CMV prophylaxis after liver transplantation in the long term. **Methods:** A total of 1,090 liver transplant patients were evaluated in terms of CMV infection in the Organ Transplantation Institute of Inonu University, Malatya, Turkey, from October 2014 to December 2019. In order to identify the CMV infections, quantitative nucleic acid amplification (QNAT) was used to detect potential CMV DNA. The cut-off value of CMV DNA was determined to be 1000 copies/mL after transplantation.

Results: According to the clinical and laboratory assessments, 33 (3%) patients were diagnosed with CMV infection, and 25 (2.3%) patients were evaluated as possibly having CMV syndrome. Also, eight of the 33 patients were assessed as having end-organ CMV disease and 25 as probable CMV syndrome. In the late period following prophylaxis, CMV infection was observed in 10 (0.9%) cases. The infection rate after prophylaxis (0.9%) was lower than the infection rate (2.1%) seen during prophylaxis.

Conclusions: Close clinical follow-up with CMV prophylaxis and strict monitoring of CMV DNA by determining a specific cut-off point are important in the follow-up of liver transplant patients.

Keywords: Cytomegalovirus Infection, CMV Prophylaxis, End-Organ CMV disease, Liver Transplantation, Probable CMV Syndrome, Quantitative Nucleic Acid Amplification (QNAT)

1. Background

Cytomegalovirus (CMV) is a double chain DNA virus that is a member of the Herpesviridae family and infects most people worldwide (1). Primary CMV infection can be asymptomatic or may occur as a self-limiting febrile disease in people with insufficient immunity. However, it may cause serious disease in susceptible individuals, such as solid organ transplant (SOT) recipients, in the form of a primary infection or as a result of the reactivation of a latent infection (1, 2).

CMV is an important cause of morbidity and mortality in SOT cases. Therefore, great efforts are made to prevent and manage CMV in SOT patients (2). In the United States, the overall CMV seroprevalence rate, which varies according to age, geography, and economic status, is reported to be 50% (3, 4). In Turkey, the overall seroprevalence of CMV-IgG is reported to be over 90% in all ages and genders (5, 6). In the literature, exposure to CMV has been reported to be 71.8% in liver transplant recipients according to preoperative tests (7).

Studies have reported that prophylaxis after transplantation significantly reduces the risk of CMV infection (8, 9). Without a prevention strategy, CMV infection and disease typically occurs within the first three months after SOT. It has been shown that if SOT patients do not receive prophylaxis until the 90th day after transplantation, 91.9% will have viremia, and 50-65% will develop symptomatic infection (8, 10).

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2. Objectives

Prophylaxis application, effectiveness, postoperative CMV infection rates, and CMV treatment results vary between centers around the world. In this study, we aimed to contribute to the literature by sharing our CMV infection data in liver transplant recipients who underwent CMV prophylaxis in the Organ Transplantation Institute of Inonu University, Malatya, Turkey.

3. Methods

In this study, 1,090 patients who underwent liver transplant between January 2014 and December 2019 were retrospectively evaluated. All patients included in the study were over the age of 18 years. In accordance with the universal CMV prophylaxis protocol, valganciclovir (900 mg oral daily) prophylaxis was given to all patients in the first 100 days after transplantation. In the follow-up of liver transplant recipients, molecular screening tests for CMV was carried out in accordance with the recommendations of international guidelines. EZ1 virus Mini kit V2.0 (Qiagen, Germany) was used for CMV DNA extraction, and application was executed with CMV Qs-RGQ Kit (Qiagen, Germany) on Rotor Gene Q 5 Plex HMR (Qiagen, Germany). CMV DNA positive patients underwent weekly DNA monitoring until they received two consecutive negative reports. The standard immunosuppressive treatment consisted of a calcineurin inhibitors (CNIs), including tacrolimus or cyclosporine, and steroids. Mycophenolate mofetil, basiliximab, and everolimus were among the other immunosuppressive agents used. CMV infection and disease were defined in accordance with previously published definitions (2).

3.1. The Diagnosis of CMV Infection

After transplantation, CMV infection was diagnosed based on the replication of CMV-DNA (copy/mL) by quantitative nucleic acid amplification test (QNAT). The CMV-DNA threshold value was accepted as 1000 copies/mL in our center, and values above this threshold were evaluated in favor of CMV infection.

In patients diagnosed with a CMV infection after transplantation, the hospital database system was used to determine the number of CMV DNA copies from the postoperative samples on the day positivity is detected, as well as the preoperative CMV IgG results of the recipient and donor. Along with the recipient's QNAT positivity, blood leukocyte, platelet, lymphocyte, and liver enzyme counts were recorded. Mortality was evaluated during the treatment period and one month after that. The results entered to the IBM SPSS statistics version 22.0 for Windows (SPSS, Inc., Chicago, IL).

3.2. The Diagnosis of CMV Disease

Patients were evaluated as probable CMV syndrome in the presence of at least two episodes of fever $\geq 38^{\circ}$ C, new or increased malaise or fatigue, leukopenia or neutropenia, high level of hepatic aminotransferases, and the detection of CMV with QNAT in blood. For diagnosis of endorgan CMV disease, the results of histopathological biopsy were evaluated.

Intravenous ganciclovir was administered for the treatment. CMV antiviral therapy was maintained until QNAT was negative for at least 14 days. The duration of therapy was also recorded.

For the statistical analysis, categorical variables were compared with the Pearson Chi-Square test or Fisher's exact test. A P < 0.05 value was considered significant.

The present study was approved by the noninterventional Ethical Committee of Medical Faculty at Inonu University, Turkey (approval no: 2020/262).

4. Results

A total of 1,090 liver transplantations were performed in our transplantation institute between January 2014 and December 2019. The number of QNAT examined by year in cases of clinical suspicion or during CMV screening was reported as 321, 319, 430, 283, 326, and 219 from 2014 to 2019, respectively. A total of 1,898 tests were performed on 1,090 patients. Figure 1 displays the number of transplant cases per year, number of QNAT performed, and the number of patients with CMV infection. The clinical features and results of 33 (3%) patients whose CMV DNA in blood and/or other body materials was detected above 1000 copies/mL with QNAT were evaluated. The mean age \pm SD of these 33 patients was 44.6 \pm 14.9 years, and 21 (63.6%) were male. The demographic data and baseline recipient characteristics are shown in Table 1. Twenty-four of the donors were living, and the remaining nine were cadaveric. Eight out of 33 patients died during the antiviral treatment period or within one month of follow-up. Eight of the patients were determined to have end-organ CMV disease (Table 2), and 25 (2.3%) had probable CMV syndrome (Table 3). There was no patient with refractory CMV infection or with CMV replication who did not display the clinical signs and symptoms of the disease.

4.1. CMV QNAT Assessment

CMV QNAT was positive in the plasma of 26 patients, plasma and bile of three, plasma and colon tissue of two,



Figure 1. Transplanted patients by year, number of QNAT performed, number of patients with	h CMV infectio
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Features	Values
Age, y	
Female	39.3 ± 13.5
Male	47.6 ± 15.1
Total	44.6 ± 14.9
Gender	
Female	12 (36.4)
Male	21 (63.6)
onor Type	
Deceased	9 (27)
Living	24 (73)
Retransplantation	
Yes	2 (6.1)
No	31 (93.9)

^aValues are expressed as No. (%) or mean \pm SD.

and plasma and bronchoalveolar lavage (BAL) fluid of two patients. CMV QNAT values were determined as 1780 copy/mL and 2043 copy/mL in BAL fluid, 4450 copy/mL, 1390 copy/mL, and 1037 copy/mL in endoscopic material (bile). In the colonoscopic materials, CMV QNAT was detected as 194 copy/mL (simultaneous plasma was CMV QNAT 13841 copy) in one case and 401 copy/mL (simultaneous plasma was CMV QNAT 1702 copy/mL) in the other case.

The median level of plasma CMV QNAT was 2208 copy/mL (min 1033, max 2443977). The median day of detecting first positive CMV QNAT value was 41 (min 7, max 179) days. The median duration of treatment was 14 (min 1, max 37) days.

In 23 (2.1%) of the transplanted patients, positivity was detected in the early period within the first 100 days after transplantation. The other 10 (0.9%) patients were found positive for CMV QNAT after the prophylaxis period. However, all patients were diagnosed within the first six months after liver transplantation. Four of the 23 patients were diagnosed with CMV infection within the first 100 days, and four patients from the other 10 (diagnosed with CMV infection after 100 days) died (P = 0.174).

4.2. CMV Serostatus and Other Laboratory Findings

At the time of diagnosis, blood AST (U/L), ALT (U/L), white blood cell 10³/uL), and lymphocyte count (/mL) values (mean \pm SD) were detected to be 272 \pm 609, 202 \pm 316, 9227 \pm 6476, and 570 \pm 460, respectively. In patients with end-organ disease, mean liver enzymes were higher than those with CMV syndrome, while platelets were lower (Table 4).

CMV serostatus of donors and recipients was not statistically different between diagnosis within the first 100 days and after after 100th day (P = 0.682) (Table 5).

Timing of Diagnosis (Days Post-Transplant)	Diagnosis/Classification	Method of Detection
108	Drokable geterintenting (CMV disease (2 petiente)	Appearance of upper and/or lower GI symptoms and
34	macroscopic mucosal lesion	
15		Appearance of upper and/or lower GI symptoms and
24	Probable gastrointestinal CMV disease (3 patients)	CMV demonstrated on endoscopic material (bile); but
147		
41	Proven or definite CMV pneumonia (1 patient)	Pneumonia new infiltrates on imaging, hypoxia, tachypnea, and/or dyspnea combined with CMV documented in lung tissue by DNA hybridization techniques
36	Probable CMV pneumonia (1 patient)	New infiltrates on imaging combined with detection of CMV by quantitation of CMV DNA in BAL
166	Proven or definite CMV hepatitis (1 patient)	Deterioration in liver test results and CMV demonstrated with histopathology and documented by DNA hybridization techniques of liver tissue.

Case	Time of Diagnosis, Days Post-Transplant	CMV QNAT PCR, Copies/mL)	AST, U/L	ALT, U/L	Lymphocite,/mL
1	20	9734	69	166	500
2	178	2500	14	27	500
3	111	1533	111	81	200
4	6	1820	6	9	1200
5	40	1650	40	24	530
6	31	13950	31	15	410
7	26	1647	62	55	100
8	115	1555	252	781	140
9	11	1331	66	85	170
10	41	7930	22	22	10
11	16	3050	60	60	104
12	14	14780	946	946	1328
13	22	2480	78	78	28
14	8	1536	58	58	79
15	175	2443977	60	60	118
16	43	2900	68	68	58
17	22	1307	92	92	50
18	131	22860	756	756	559
19	116	1452	47	47	62
20	90	97578	12	12	11
21	69	933	31	31	50
22	59	2208	11	11	11
23	7	4450	37	37	51
24	30	4360	3052	3052	496
25	7	79236	71	366	270

 Table 2. Description of End-Organ CMV Disease and Timing of Diagnosis (Days Post-Transplant)

Table 4. Some Laboratory Results in Patients with CMV Syndrome and CMV End-Organ Disease^a

CMV Infection	Numbers	AST, U/L	ALT, U/L	WBC,/mL	Lymphosyt,/mL	Platelet, 10 ³ /mL
CMV syndrom	25	222.3 ± 594.6	172 ± 294.9	9421.4 \pm 6625.4	554.6 ± 464.6	102.3 ± 138.3
End-organ CMV disease	8	553 ± 683	367.8 ± 414.1	8136 ± 6113.4	662 ± 477	72.6 ± 64.6
Total		272.4 ± 609.2	201.7 ± 316	9226.7 ± 6475.2	570.9 ± 460	97.8 ± 129.6

^aValues are expressed as mean \pm SD.

nosis (Within the First 100 Days)(n)	Diagnosis After 100 Days (n)
	5
8	2
15	8
	8 15

Abbreviation: Preop, preoperative.

5. Discussion

Prophylaxis strategies for CMV infection are controversial in the world. Universal prophylaxis is commonly preferred in countries that have a high seroprevalence rate as it is more effective. However, the rate of late period CMV infections after prophylaxis is an important issue with changing data between centers (8, 10, 11). When CMV prophylaxis is not given in the postoperative period of SOT patients, majority of them develop a CMV infection. To prevent these infections, either prophylaxis or preemptive treatment protocols are conducted. Since CMV seropositivity was widespread, CMV prophylaxis and close followup with real-time polymerase chain reaction (PCR) was the standard procedure after liver transplantation. The main advantages of prophylaxis to preemptive therapy in the literature is that this approach is highly successful in preventing early CMV DNAemia/infection, and it can be applied relatively easily. Despite prophylaxis, in some patients were found early CMV DNAemia in this study. It may be thought an acute infection coincidence in the patients. The dosage of valganciclovir was passed from prophylaxis to therapeutic amount in this group. The lack of CMV serology control of recipient and donor candidates before transplantation was thought another possible reason. On the other hand, it has been reported that late CMV infection/disease is more common with prophylaxis. Despite prophylaxis, research has reported the rate of CMV disease in the D+/R+ group of liver recipients to be 5% (11). In our study, prophylaxis was applied, and the overall rate of CMV infection was 3% (n = 33). In the late period, 10(0.9%)patients developed a CMV infection despite prophylaxis.

The use of common expressions in CMV infection and the definition of disease was discussed for the first time at the 1993 Fourth International CMV Conference in Paris (12). Later on, the importance of QNAT and histopathological tests for diagnosis and confirmation were emphasized. In that context, it has been emphasized to standardize the amount of CMV DNA (13). The World Health Organization (WHO) International Reference Standard for CMV quantification has become available to standardize values diagnostic of CMV infection (14, 15). Transplant centers are encouraged to achieve specific viral load thresholds based on the CMV QNAT test they use and the population at risk (2). In the study by Wadhawan et al., the viral load threshold value was specified as 500 copies/mL (10). Other studies reported that CMV viral loads above 1,000 copies/ml were generally associated with symptomatic CMV infections (16). In our center, CMV infection was determined at a CMV DNA cut-off value of 1000 copies/mL after transplantation.

CMV infection is characterized by the detection of CMV replication in patients regardless of symptoms. CMV replication can be detected by ONAT, antigen test, and culture. CMV disease is categorized as CMV syndrome and endorgan CMV disease (gastrointestinal disease, pneumonia, hepatitis, etc.) accompanying certain symptoms, including fever and/or weakness, and leukopenia or thrombocytopenia (17). In a single-center study on CMV syndrome and end-organ CMV disease, the number of CMV syndrome cases was 8 (2.4%), and the number of end-organ disease cases was 1 (0.3%) in 338 liver transplant patients over 5 years (10). In this study, 25 (2.3%) cases were diagnosed with CMV syndrome and 8 (0.7%) with end-organ CMV disease among 1,090 liver transplant patients over six years. CMV DNA was detected in the colon biopsy material of two cases with upper and lower gastrointestinal symptoms. It was also detected in the bile material taken by endoscopy in three cases exhibiting upper gastrointestinal symptoms. In these cases, QNAT also showed the presence of CMV DNA in blood. However, pathological confirmation could not be done in these cases, and they were characterized as having probable gastrointestinal CMV disease. The lack of histopathological documentation prevented the diagnosis of a proven gastrointestinal CMV disease.

In the literature, absolute lymphocyte count, which was shown to be associated with relapse in 33 of 170 participants (19.4%), was reported to be on average 1.08 \pm 0.69 cells/ μ L in relapse-free patients and 0.73 \pm 0.42 \times 103 cells/ μ L in relapsed patients (18). In this study, the absolute lymphocyte count was 570.9 \pm 460 cells/mL. However, no relapse was observed in the patients. On the other hand, absolute lymphocyte count remains both supportive and easily available in the diagnosis of CMV infection.

Diagnosis of CMV hepatitis requires histopathological and immunohistochemical (IHC) confirmation with elevated liver enzymes. There is no probable definition for CMV hepatitis (16). In only one patient, CMV hepatitis was diagnosed with definitive liver histopathological confirmation. The AST level of this patient was 241 (U/L), ALT was 312 (U/L), and CMV PCR was 1,310 copy/mL, and the diagnosis was confirmed on the postoperative 166th day. The duration of treatment was 29 days.

CMV QNAT in BAL fluid can be used to diagnose possible CMV pneumonia, and it is also recommended to define a viral load threshold (2). In our two patients who had CMV presence in BAL fluid, CMV QNAT values were determined to be 1,780 copy/mL and 2,043 copy/mL. One of the cases was characterized as histopathologically verified "proven pneumonia" and the other as "possible pneumonia".

After antiviral prophylaxis, late-onset CMV disease associated with poor long-term outcome can be seen. Patients may be monitored periodically using CMV QNAT for a certain time period even if they have completed antiviral prophylaxis. Virus quantification has been used as a method of direct measurement of the copied virus. Viral load assays play a significant role in patient management (19). The duration and interval of CMV monitoring following cessation of prophylaxis is not precisely established. One study reported that monitoring with two-week intervals was not clinically helpful in catching late-onset CMV disease after prophylaxis in SOT (17). In our study, a total of 1,898 tests were performed on 1,090 patients. Using QNAT, 33 (3%) patients had CMV DNA above 1000 copies/mL in blood and/or other body materials. Within the first 100 days after the operation, 23 (2.1%) of the transplanted patients were found to be positive for CMV DNA. The other 10 (0.9%) patients were found positive after the prophylaxis period was completed. Four of the 23 patients were diagnosed with CMV infection within the first 100 days, and four of the other 10 patients (diagnosed with CMV infection after 100 days) who had completed their prophylaxis died. There was no statistically difference in deaths between the two periods (P = 0.174). In this regard, we cannot conclude that they died due to CMV infection alone; hence, we need further prospective clinically controlled studies.

Another issue is duration of prophylaxis and duration of treatment. Protocols may vary according to centers. While some studies extended the treatment period up to three months, other studies proposed a longer period for prophylaxis (8, 20). We started prophylaxis in the first 10 days after surgery and it lasted for about three months (almost 100 days). The infection rate after prophylaxis (0.9%) was lower than the infection rate (2.1%) seen during prophylaxis. This suggested that it was not necessary to prolong prophylaxis time. The treatment period lasted until two CMV PCR results were negative for at least two weeks, with the mean \pm SD being 18.48 \pm 10.2 days. Since two cases died immediately after initiating treatment, their treatment could not be completed. Our longest treatment period was 37 days.

5.1. Conclusions

The main limitations of this study include the absence of a clinical control group, non-prospective nature of the study, and lack of CMV serostatus control. However, this study research a descriptive and self-assessment study. We consider close clinical follow-up as an important key point in liver transplant recipients who may face CMV infection. In addition, strictly monitoring CMV DNA by determining a certain cut-off can make the task easier.

Footnotes

Authors' Contribution: Study concept and design: SAT, YB, and AK. Acquisition of data: BO, SK, SY, SAT, and AK. Analysis and interpretation of data: SAT and YB. Drafting of the manuscript: SAT, YB, and AK. Critical revision of the manuscript for important intellectual content: SAT, YB, and AK. Statistical analysis: SAT. Administrative, technical, and material support: SK and SY. Study supervision: YB and SAT.

Conflict of Interests: The authors declared no conflict of interest.

Ethical Approval: The present study was approved by the non-interventional Ethical Committee of Medical Faculty at Inonu University, Turkey (approval no.: 2020/262).

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Informed Consent: In this study the patients were evaluated retrospectively from the hospital registry system. No invasive procedure was applied to the patients for the purpose of the study. There was no drug application for comparison purposes. There was no unfamiliar procedure. There was no inactive drug and plesabo usage or drug comparison.

References

- Clyde S. Cytomegalovirus (CMV). In: John E, Bennett RD, Martin. J. B, editors. Mandell, Douglas, and Bennett's Principles and Practice of Infectious Disease. 8th ed. Philadelphia: Saunders; 2015.
- Razonable RR, Humar A. Cytomegalovirus in solid organ transplant recipients-Guidelines of the American Society of Transplantation Infectious Diseases Community of Practice. *Clin Transplant*. 2019;**33**(9). e13512. doi: 10.1111/ctr.13512. [PubMed: 30817026].
- Bate SL, Dollard SC, Cannon MJ. Cytomegalovirus seroprevalence in the United States: the national health and nutrition examination surveys, 1988-2004. *Clin Infect Dis.* 2010;**50**(11):1439–47. doi: 10.1086/652438. [PubMed: 20426575].

- Cannon MJ, Schmid DS, Hyde TB. Review of cytomegalovirus seroprevalence and demographic characteristics associated with infection. *Rev Med Virol*. 2010;20(4):202–13. doi: 10.1002/rmv.655. [PubMed: 20564615].
- Hizel S, Parker S, Onde U. Seroprevalence of cytomegalovirus infection among children and females in Ankara, Turkey, 1995. *Pediatr Int*. 1999;41(5):506–9. doi: 10.1046/j.1442-200x.1999.01118.x. [PubMed: 10530062].
- Ataman S, Colak D, Gunseren F, Senol Y, Colak T, Aktekin MR, et al. [Investigation of cytomegalovirus seroepidemiology in Antalya with a population-based cross-sectional study and review of related data in Turkey]. *Mikrobiyol Bul.* 2007;41(4):545–55. [PubMed: 18173073].
- Varghese J, Subramanian S, Reddy MS, Shanmugam N, Balajee G, Srinivasan V, et al. Seroprevalence of cytomegalovirus in donors & opportunistic viral infections in liver transplant recipients. *Indian J Med Res.* 2017;**145**(4):558–62. doi: 10.4103/ijmr.IJMR_1024_14. [PubMed: 28862190]. [PubMed Central: PMC5663172].
- Pascual J, Royuela A, Fernandez AM, Herrero I, Delgado JF, Sole A, et al. Role of mTOR inhibitors for the control of viral infection in solid organ transplant recipients. *Transpl Infect Dis.* 2016;**18**(6):819–31. doi: 10.1111/tid.12601. [PubMed: 27600985].
- Chmelova K, Frankova S, Jirsa M, Neroldova M, Sticova E, Merta D, et al. IL28B rs12979860 T allele protects against CMV disease in liver transplant recipients in the post-prophylaxis and late period. *Transpl Infect Dis.* 2019;**21**(4). e13124. doi: 10.1111/tid.13124. [PubMed: 31165537].
- Wadhawan M, Gupta S, Goyal N, Vasudevan KR, Makki K, Dawar R, et al. Cytomegalovirus infection: its incidence and management in cytomegalovirus-seropositive living related liver transplant recipients: a single-center experience. *Liver Transpl*. 2012;18(12):1448–55. doi: 10.1002/lt.23540. [PubMed: 22903934].
- Harvala H, Stewart C, Muller K, Burns S, Marson L, MacGilchrist A, et al. High risk of cytomegalovirus infection following solid organ transplantation despite prophylactic therapy. *J Med Virol*. 2013;85(5):893–8. doi: 10.1002/jmv.23539. [PubMed: 23508914].
- 12. Ljungman P, Griffiths P. Definitions of cytomegalovirus infection and disease. *Multidisciplinary approach to understanding cytomegalovirus disease*. Paris, Amsterdam: Excerpta Medica International Congress

Series; 1993. p. 233-40.

- Ljungman P, Boeckh M, Hirsch HH, Josephson F, Lundgren J, Nichols G, et al. Definitions of Cytomegalovirus Infection and Disease in Transplant Patients for Use in Clinical Trials. *Clin Infect Dis*. 2017;**64**(1):87–91. doi: 10.1093/cid/ciw668. [PubMed: 27682069].
- Boaretti M, Sorrentino A, Zantedeschi C, Forni A, Boschiero L, Fontana R. Quantification of cytomegalovirus DNA by a fully automated realtime PCR for early diagnosis and monitoring of active viral infection in solid organ transplant recipients. *J Clin Virol*. 2013;**56**(2):124–8. doi: 10.1016/j.jcv.2012.10.015. [PubMed: 23182772].
- Fryer JF, Heath AB, Wilkinson DE, Minor PD; Group tCS. Collaborative study to evaluate the proposed 1st WHO international standard for Human Cytomegalovirus (HCMV) for nucleic acid amplification technology (NAT)based assays. 2010. Available from: http://apps.who.int/iris/handle/ 10665/70521.
- Kotton CN, Kumar D, Caliendo AM, Asberg A, Chou S, Danziger-Isakov L, et al. Updated international consensus guidelines on the management of cytomegalovirus in solid-organ transplantation. *Transplantation*. 2013;**96**(4):333–60. doi: 10.1097/TP.0b013e31829df29d. [PubMed: 23896556].
- Boillat Blanco N, Pascual M, Venetz JP, Nseir G, Meylan PR, Manuel O. Impact of a preemptive strategy after 3 months of valganciclovir cytomegalovirus prophylaxis in kidney transplant recipients. *Transplantation*. 2011;91(2):251–5. doi: 10.1097/TP.0b013e318200b9f0. [PubMed: 21099744].
- Gardiner BJ, Nierenberg NE, Chow JK, Ruthazer R, Kent DM, Snydman DR. Absolute Lymphocyte Count: A Predictor of Recurrent Cytomegalovirus Disease in Solid Organ Transplant Recipients. *Clin Infect Dis.* 2018;67(9):1395–402. doi: 10.1093/cid/ciy295. [PubMed: 29635432]. [PubMed Central: PMC6927884].
- Singh MP, Galhotra S, Saigal K, Kumar A, Ratho RK. Quantitative nucleic acid amplification methods and their implications in clinical virology. *Int J Appl Basic Med Res*. 2017;7(1):3–9. doi: 10.4103/2229-516X.198498. [PubMed: 28251100]. [PubMed Central: PMC5327603].
- Huprikar S. Update in infectious diseases in liver transplant recipients. *Clin Liver Dis.* 2007;11(2):337–54. doi: 10.1016/j.cld.2007.04.006. [PubMed: 17606211].