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Research Article

Post-Intervention Plasma IL-10 Level Predicts Early Tumor Response in Hepatocellular Carcinoma Treated with Transarterial Chemoembolization

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Abstract

Background: Cytokines play an important role in tumor progression, but studies have shown mixed results regarding the role of cytokines in predicting the early response to transarterial chemoembolization (TACE) in hepatocellular carcinoma (HCC). **Objectives:** This study aimed to explore the correlation between plasma levels of cytokines and early tumor response in HCC patients undergoing TACE.

Methods: Thirty HCC patients enrolled in this study from the department of liver disease of a general hospital from June 2020 to January 2021. Plasma samples were sampled at baseline and 7 days after TACE for cytokine detection by cytometric bead array (CBA). At 4 - 6 weeks after TACE, the objective response of HCC patients was confirmed according to response evaluation criteria in solid tumors (RECIST). Potential factors such as various cytokines and some clinical parameters were analyzed by univariate and multivariate analysis. The predictive effects of different factors in HCC patients undergoing TACE were analyzed by the receiver operating characteristic (ROC) curve.

Results: Plasma levels of post-TACE interleukin-10 (IL-10) were statistically significantly higher than baseline IL-10 levels. The level of plasma IL-10 after TACE was an independent risk factor for early tumor response. The patients with low plasma IL-10 levels after TACE had a favorable prognosis. Receiver operating characteristic curve analysis showed that post-TACE IL-10 had a high predictive value (area under the curve = 0.769, 95% confidence interval (CI): 0.598 - 0.939). A high level of plasma IL-10 after TACE was correlated with alpha-fetoprotein (AFP) level (P = 0.037) and post-TACE alanine aminotransferase (ALT) (r = 0.368, P = 0.045). Post-TACE plasma IL-10 did not correlate with age or tumor metastasis.

Conclusions: Our findings demonstrated that post-intervention plasma IL-10 levels could predict short-term outcomes independently after TACE. These findings were helpful in identifying the patients who might benefit from TACE.

Keywords: Interleukin-10, Carcinoma, Hepatocellular, Chemoembolization, Therapeutic

1. Background

Hepatocellular carcinoma (HCC) is the fourth most deadly cancer-related disease worldwide (1). Comprehensive treatments have been recommended as standard of care, including surgical resection, local ablation, liver transplantation, transarterial chemoembolization (TACE), and systemic therapy (2, 3). Surgical treatment remains the curative method for HCC. However, more than 60% of HCC patients are in advanced stages that are inoperable. In this regard, TACE has become the main treatment modality for patients with unresectable HCC (4). According to the clinical guideline, TACE has been identified as an alternative or combination therapy method for early or advanced HCC patients. Transarterial chemoembolization unavoidably leads to a hypoxic environment not only in the tumor itself but also in the adjacent liver tissue, which can trigger an inflammatory response in the liver environment as well as alterations within the immune system. As a result, the levels of cytokines can be altered in the tumor environment and blood. The investigation of the alteration of cytokines after TACE is helpful in improving the understanding of tumor progression and the treatment level of HCC.

In the liver, different kinds of cells produce and release cytokines in response to hepatic injury and inflammation. Cytokines play a vital role in the etiopathology

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of HCC. They mediate cellular communication and coordinate pathologic processes such as inflammation, cirrhosis, and cancer. Although many factors can trigger HCC, inflammation plays a crucial role. The HCC is actually considered a classic example of inflammation-related cancer, as approximately 90% of HCCs are caused by injury and inflammation (5). Monocytes/Kupffer cells and immature myeloid cells, following infiltration and activation, release many pro-inflammatory cytokines, such as interleukin-1 β (IL-1 β), IL-6, IL-18, and TNF- α . Meanwhile, anti-inflammatory responses (e.g., IL-10 and TGF- β) are formed in the liver to balance excessive inflammation (6). T cells are often classified into 2 subsets: CD4⁺ T cell and CD8⁺ T cell. CD4⁺ T cells can differentiate into 2 functionally distinct CD4⁺ T helper (Th) cell subsets. Th1 cells specifically secrete pro-inflammatory cytokines, including interferon- γ (IFN- γ) and IL-2, and Th2 cells initiate an anti-inflammatory response by secreting IL-4 and IL-10. As the heterogeneity of CD4⁺ T cell, an additional CD4⁺ T cell subset, called Th17 cells, is observed. It has been reported that Th17 cells can release IL-17, IL-21, and IL-22 and are associated with HCC progression (7). CD8⁺ T cells can also differentiate into cytotoxic 1 (Tc1) and Tc2 T cells, which release many cytokines.

In tumor progression, different levels of cytokines may reflect different inflammatory mechanisms. In addition, cytokines can also be the biomarkers for tumor prediction (8). However, there are few studies exploring the effect of cytokines in predicting the response of TACE in HCC.

2. Objectives

In this study, the levels of plasma cytokines pre- and post-TACE were compared, and the relationship between cytokines and early tumor response after TACE was analyzed in HCC.

3. Methods

3.1. Patients and Study Designs

Thirty patients with unresectable HCC who underwent TACE from the Second Hospital of Nanjing, Nanjing University of Chinese Medicine, were enrolled in this study from June 2020 to January 2021. Diagnosis of HCC is based on the guidelines of the European Association for the Study of the Liver (2). The exclusion criteria were as follows: (1) With TACE contraindications; (2) have ever taken immunological drugs; (3) with serious underlying diseases; (4) poor compliance; (5) clinical data were incomplete. All procedures in this study were in accordance with the ethical standards of World Medical Association Declaration of Helsinki "Ethical principles for medical research involving human subjects". The Ethics Committee of the Second Hospital of Nanjing approved this study. Written informed consent was obtained from all the participants.

3.2. Measurement of Plasma Interleukin-10 Levels

At baseline and 7 days after TACE treatment, the plasma samples were collected. The samples were stored at -80°C. Concentrations of cytokines, including IL-2, IL-4, IL-6, IL-17A, IL-9, IL-10, IFN- γ , TNF- α , and granzyme B (GranB) were assessed with cytometric bead array(CBA)kit(BD Pharmingen) in the clean bench according to the manufacturer's instructions.

3.3. Transarterial Chemoembolization Procedure

All the TACE operations were carried out by our experienced interventional radiologist at the Second Hospital of Nanjing. The procedures were as follows: Making routine preoperative preparation, then puncturing femoral artery for celiac trunk angiography. A superselective approach to the tumor was performed, by which chemotherapeutic emulsion could be transported to the active lesions to embolism tumor vessels. Finally, hepatography was performed to confirm whether the treated tumor had disappeared.

3.4. Evaluation of Transarterial Chemoembolization Response

All enrolled patients underwent imaging examination (Computed Tomography or Magnetic Resonance Imaging) 4 - 6 weeks after TACE therapy. Tumor responses were divided into 4 types according to response evaluation criteria in solid tumors (RECIST) criteria, including complete response (CR), partial response (PR), progressive disease (PD), and stable disease (SD) (9). Objective remission of tumor included CR and PR, and we defined tumor response after TACE as the objective of tumor response.

3.5. Statistical Analysis

Acquired data were analyzed by SPSS version 26.0 and GraphPad Prism version 8.0 software. The results were given as median and interquartile range. Continuous data were compared using Student's t-test, and categorical data were analyzed by χ^2 test. Non-parametric test was used when the date was skewed. Additionally, Spearman's correlation was used to find the relationship between interleukins and clinical parameters. Receiver operating characteristic (ROC) curves were generated by plotting sensitivity against 1-specificity. The determination of optimal cut-off values was due to the Youden Index. The predictive value of plasma cytokine levels with respect to tumor response was tested using a binary logistic regression model. Receiver operating characteristic curve analysis was performed with MedCalc v15.2. A P-value of < 0.05 was considered statistically significant.

Characteristics	Hepatocellular Carcinoma (n = 30)		
Sex			
Male	24 (80.00)		
Female	6 (20.00)		
Age (y)	61.87 ± 9.90		
Causes			
HBV	28 (93.33)		
HCV	2 (6.67)		
AFP (ng/mL)			
< 400	17 (56.67)		
\geq 400	13 (43.33)		
Pre-ALT (U/L)	26.60 (19.65, 31.03)		
Post-ALT (U/L)	21.05 (15.85, 32.98)		
Pre-AST (U/L)	34.20 (25.90, 46.45)		
Post-AST (U/L)	32.35 (23.53, 52.35)		
ALB (g/L)	39.02 ± 5.00		
Child-Pugh classification			
А	26 (86.67)		
В	4 (13.33)		
С	0(0)		
BCLC stage			
А	2 (6.67)		
В	16 (53.33)		
С	12 (40.00)		
D	0(0)		
Tumor size (cm)	8.09 ± 1.95		
No. of tumors, single	5 (16.67)		
Extrahepatic metastasis	5 (16.67)		

Abbreviations: AFP, alpha-fetoprotein; ALT, alanine aminotransferase; AST, aspartate aminotransferase; ALB, albumin; INR, international normalized ratio; BCLC, Barcelona Clinic Liver Cancer

 $^{\rm a}$ Data are shown as No. (%), mean \pm standard deviation or median (interquartile range (IQR)).

4. Results

4.1. Study Population and Baseline Characteristics

A total of 30 patients with unresectable HCC receiving TACE treatment enrolled in this study. Table 1 reported the baseline characteristics of all the participants. The mean age of patients was 61.87 ± 9.90 years, and 24 (80%) of the patients were male. Twenty-eight (93.33%) patients were HBV-associated, and 2 (6.67%) patients were HCV-associated. Moreover, 86.67% of the patients were identified as child-pugh class A, whereas 13.33% of the patients were in child-pugh class B. On the grounds of the Barcelona Clinic Liver Cancer (BCLC) classification, 2, 16, 12 of the patients were in the stages A, B, and C, respectively. The tumor size of these patients was 8.09 ± 1.95 cm. At enrollment, 5 (16.67%) patients had single nodular HCC, and 5 (16.67%) patients had extrahepatic metastasis.

4.2. Comparison of Levels of Plasma IL-2, IL-4, IL-6, IL-17A, IL-9, IL-10, IFN- γ , TNF- α , and Granzyme B Between Pre-transarterial Chemoembolization and Post-transarterial Chemoembolization in Patients with Hepatocellular Carcinoma

We first compared the concentrations of plasma IL-2, IL-4, IL-6, IL-17A, IL-9, IL-10, IFN- γ , TNF- α , and GranB between pre-TACE and post-TACE. Accordingly, IL-6 and IL-10 were significantly increased after TACE (P < 0.05) (Table 2). The other plasma cytokines, including IL-2, IL-4, IL-17A, IL-9, IFN- γ , TNF- α , and GranB, had no obvious difference between pre-TACE and post-TACE in our study.

4.3. Relationship Between Levels of Inflammatory Cytokines and Early Tumor Response After Transarterial Chemoembolization

The tumor responses were evaluated 4 - 6 weeks after TACE. Among the 30 patients, 15 (50%) achieved objective tumor response. In order to investigate the predictor of early tumor response after TACE, univariate and multivariate logistic regression analyses were conducted. In univariate analysis, all of those with P < 0.2, such as post-TACE plasma IL-10 level (P = 0.020), metastatic (P = 0.113), alphafetoprotein (AFP) (P = 0.072), and BCLC stage (P = 0.078) were included in the multivariate analysis (Table 3). Nevertheless, in multivariate analysis, only plasma IL-10 level after TACE was an independent risk factor for early tumor response (P = 0.043) (Table 4).

Then, ROC curve was used to determine the effectiveness of the post-TACE plasma IL-10 level in predicting early tumor response after TACE. The ROC curve analysis revealed that when the cut-off value was > 1.96 pg/mL, the

Table 2. Level of Plasma Cytokines Pre- and Post-transarterial Chemoembolization Treatment in 30 Patients with Hepatocellular Carcinoma Cohort $^{\rm a}$

Cytokines (pg/mL)	Pre-TACE	Post-TACE
IL-2	3.43 (3.01, 3.98)	3.46 (2.82, 3.89) ^b
IL-4	1.46 (0.74, 1.91)	1.43 (0.79, 1.78)
IL-6	6.25 (1.89, 29.72)	23.61 (6.81, 59.26) ^c
IL-17A	3.09 (1.70, 4.16)	2.99 (1.85, 4.27)
IL-9	0.34 (0.00, 0.94)	0.22 (0.00, 0.85)
IL-10	0.63 (0.16, 1.15)	$1.92(1.52, 2.50)^{d}$
IFN- γ	1.97 (1.16, 2.63)	1.84 (1.09, 2.58)
TNF- α	1.23 (0.58, 1.69)	1.21 (0.20, 1.58)
GranB	5.26 (3.60, 6.29)	4.31 (3.35, 5.71)

Abbreviations: TACE, transarterial chemoembolization; IL, interleukin; IFN, interferon; TNF- α , tumor necrosis factor-alpha; GranB, Granzyme B

^a Data are shown as median and interquartile range (IQR).

^b Wilcoxon matched-pairs, non-parametric test was used.

^c P < 0.01 ^d P < 0.001

Chama atomistics	Univariate Analysis		
	OR (95% CI)	P-Value	
Gender (male vs. female)	1.00 (0.17, 5.99)	1.000	
Child-Pugh classification (A vs. B/C)	1.61 (0.39, 6.62)	0.510	
Tumor size (\leq 5 cm vs. > 5 cm)	1.31 (0.31, 5.58)	0.713	
BCLC (early/mid/late)	3.43 (0.87, 13.55)	0.078 ^a	
Metastatic (presence vs. absence)	4.33 (0.71, 26.53)	0.113 ^a	
Baseline AFP	4.13 (0.88, 19.27)	0.072 ^a	
Pre-TACE plasma IL-6 level	1.01 (1.00, 1.02)	0.455	
Post-TACE plasma IL-6 level	1.00 (1.00, 1.02)	0.695	
Pre-TACE plasma IL-9 level	1.70 (0.45, 6.34)	0.433	
Post-TACE plasma IL-9 level	2.86 (0.59, 13.97)	0.193	
Pre-TACE plasma IL-10 level	1.33 (0.54, 3.27)	0.533	
Post-TACE plasma IL-10 level	5.46 (1.28, 23.33)	0.020 ^a	
Pre-TACE plasma IFN- γ level	0.79 (0.45, 1.38)	0.407	
Post-TACE plasma IFN- γ level	0.90 (0.41, 2.01)	0.806	
Pre-TACE plasma TNF- α level	0.67 (0.33, 1.39)	0.284	
Post-TACE plasma TNF- $lpha$ level	0.86 (0.64, 1.16)	0.326	
Pre-TACE plasma IL-2 level	0.81 (0.40, 1.61)	0.542	
Post-TACE plasma IL-2 level	1.70 (0.69, 4.17)	0.245	
Pre-TACE plasma IL-4 level	0.93 (0.73, 1.17)	0.527	
Post-TACE plasma IL-4 level	0.81 (0.36, 1.83)	0.611	
Pre-TACE plasma IL-17A level	0.86 (0.57, 1.31)	0.485	
Post-TACE plasma IL-17A level	0.94 (0.76, 1.14)	0.517	
Pre-TACE plasma GranB level	0.84 (0.60, 1.17)	0.306	
Post-TACE plasma GranB level	0.86 (0.67, 1.10)	0.229	

 Table 3. Univariate Logistic Regression Analysis of Prognostic Factors for Early Tumor Response After Transarterial Chemoembolization

Abbreviations: CI, confidence interval; AFP, alpha-fetoprotein; IL, interleukin; IFN, interferon; TNF- α , tumor necrosis factor-alpha; TACE, transarterial chemoembolization

^a included in the multivariate analysis

area under curve (AUC) was 0.769 (95% confidence interval (CI), 0.598 - 0.939), P < 0.01 (Figure 1). The sensitivity and specificity of post-TACE plasma IL-10 levels were 73.3% and 80%, respectively. The positive predictive value (PPV) was 78.6%, the negative predictive value (NPV) was 75%, the value of +LR was 3.69, and the value of -LR was 0.33.

4.4. Association of Interleukin-10 Plasma Level Posttransarterial Chemoembolizationwith Clinical Characteristics

The relationship between post-TACE plasma IL-10 levels and clinical parameters, including AFP, tumor metastasis, maximum tumor size, BCLC stage was assessed. High post-TACE plasma IL-10 level was associated with AFP level (P = 0.037) (Figure 2). The level of IL-10 has no significant correlation with maximum tumor size (P > 0.05) (Figure 2).

Then, Spearman's correlation analysis was used to identify the associations between post-TACE IL-10 plasma level and valuable clinical parameters. IL-10 level had a positive correlation with post-TACE serum alanine amino-transferase (ALT) level (r = 0.368, P = 0.045) (Figure 2). However, plasma IL-10 level after TACE had no significant correlation with aspartate aminotransferase (AST) before TACE AST (r = 0.170, P = 0.369), post-TACE AST (r = 0.140, P = 0.462), pre-TACE ALT (r = -0.104, P = 0.585) (Figure 2). Plasma post-TACE IL-6 level also had no significant correlation with sex, age, CHE, and international normalized ratio (INR) (data not shown).

5. Discussion

Hepatic inflammatory response, characterized by continuous expression of cytokines and infiltration of immune cells, is a risk factor for the occurrence and development of HCC and may induce alterations in the immune microenvironment of liver. The net effect of the immune system on the inflammatory process leads to tumor growth and progression. The balance between proinflammatory and anti-inflammatory cytokines in the immune system is often disrupted in HCC (10). It has been reported that elevated levels of certain inflammatory cytokines after TACE may lead to hepatic tissue injury (7, 11, 12). In our study, we compared the plasma IL-2, IL-4, IL-6, IL-17A, IL-9, IL-10, IFN- γ , TNF- α , and GranB levels between pre-TACE and post-TACE in patients with HCC and found post-TACE IL-6 plasma levels and post-TACE IL-10 plasma levels increased. The level of post-TACE plasma IL-10 can be used as an indicator to predict early tumor response after TACE.

The changes in plasma levels of cytokines after TACE in HCC patients have been explored in some studies. However, the results were heterogeneous. Most studies showed increased IL-6 in the early phase after TACE. In the late phase, the levels of IL-4 and IL-10 were higher than those before TACE (11). Our study suggested that the levels of IL-6 and IL-10 increased, whereas levels of plasma IL-2, IL-4, IL-17A, IL-9, IFN- γ , TNF- α , and GranB only showed minor changes in the early phase after TACE, which was consistent with some other studies (11, 13, 14).

The IL-6, an immunomodulatory cytokine, is a promalignant mediator in tumors (15). As described in the literature, IL-6 is involved in multiple stages of HCC development. It not only promotes primary hepatocyte proliferation and hepatocyte transformation into hepatocellular carcinoma progenitor cells but also promotes the development of hepatocellular carcinoma nodules and metastasis. Patients with large tumors, advanced tumors and

fable 4. Multivariate Logistic Regression Analysis of Prognostic Factors for Early Tumor Response After Transarterial Chemoembolization					
Characteristics	Multivariate Logistic Regression Analysis				
	OR (95% CI)	P-Value	β		
Post-TACE plasma IL-10 level	5.34 (1.05, 27.01)	0.043 ^a	1.674		
Metastatic	4.75 (0.36, 62.41)	0.236	1.557		
Baseline AFP	1.124 (0.15, 8.69)	0.911	0.117		
BCLC stage	3.54 (0.75, 16.84)	0.112	1.264		

Abbreviations: CI, confidence interval; IL, interleukin; TACE, transarterial chemoembolization, IFN, interferon; BCLC, Barcelona Clinic Liver Cancer ^a Statistically significant



Figure 1. The receiver operating characteristic (ROC) curve of plasma post-intervention IL-10 levels for diagnosis of early tumor response in 30 hepatocellular carcinoma (HCC) patients with transarterial chemoembolization (TACE)

recurrence or short survival after local treatment for HCC have higher levels of IL-6 before surgery (16). Some studies showed that high IL-6 levels after TACE statistically correlated with maximum tumor size, vascular invasion, AST levels, and BCLC stage after TACE (8, 17). Compared to dynamic

changes in IL-6, the level of IL-6 after TACE was a strong predictor of tumor response at 4 - 6 weeks after TACE (8, 12). Our results showed elevated IL-6 plasma levels in early phase after TACE. However, the post-TACE IL-6 levels had no relationship with the tumor response in our study, which



Figure 2. Correlations between post-transarterial chemoembolization (TACE) interleukin-10 (IL-10) levels and clinical parameters. A, Plasma level of post-TACE IL-10 in patients with alpha-fetoprotein (AFP) < 400 μ g/mL and AFP \geq 400 μ g/mL; B, Plasma level of post-TACE IL-10 in patients with tumor size \leq 5cm and tumor size > 5cm. In A and B, the lines and bars present mean and standard deviation (SD), respectively. Correlations between post-TACE IL-10 levels and C, pre-TACE alanine aminotransferase (ALT); D, post-TACE ALT, (E) pre-TACE aspartate aminotransferase (AST); and F, post-TACE AST

could be explained by the different cohorts, test methods, and the complexity of the immune networks in HCC.

IL-10, an immune suppressive cytokine, is involved in immune regulatory and angiogenesis (18). both tumor cells and some immune cells can secrete IL-10. In the tumor microenvironment, IL-10 secreted by tumor cells can directly inhibit cytotoxic T cells and NK cells through JAK1/STAT5 (19), and can also activate tumorassociated macrophages through the M2 macrophage polarization pathway (20, 21). IL-10 serves as a negative regulator in the progression of T cell activation. It can inhibit the release of inflammatory mediators secreted by mononuclear macrophages and enhance the release of anti-inflammatory factors. The report showed that IL-10 secreted by B cells suppressed the effector functions of CD4⁺ and CD8⁺ T cells by inhibiting their secretion of IFN- γ , TNF- α , and IL-17 (22). In addition, IL-10 limits CD8⁺ T cell activation and function by regulating cell-surface glycosylation (23). On the other side, IL-10 enhances immunosuppressive activity by promoting the generation and expansion of Treg cells (24). Meanwhile, the expression of some immunosuppressive molecules can promote the secretion of IL-10 and inhibit the secretion of IFN- γ by CD8⁺ T cells, promoting a suppressive tumor immune environment (25, 26). However, IL-10 could stimulate the immune system instead of suppressing it in some contexts. Kuhn et al. revealed that IL-10-deficient mice developed colitis (27). It is reported that serum IL-10 is associated with liver injury caused by cirrhosis and HCC (24, 28, 29). A meta-analysis has shown that IL-10 is associated with tumor prognosis (30). Only a few studies demonstrate that pre-TACE IL-10 is related to tumor response and OS in HCC patients (31, 32). The relationship between IL-10 and clinical indicators such as ALT and AST after TACE is rarely analyzed. We found that plasma IL-10 levels after TACE significantly positively correlated with baseline AFP and post-TACE ALT.

This study has some limitations. First of all, this study was a single center, and the sample size was small. Second, we only measured cytokine levels once after TACE treatment. We did not follow the dynamic changes of each cytokine and its influence on treatment response. Finally, due to the limited follow-up time, we did not evaluate the overall survival.

5.1. Conclusions

We reported that IL-10 plasma levels after TACE could serve as a potential candidate marker for predicting early tumor response after TACE treatment in HCC patients. Our future work is expected to increase the clinical sample size, appropriately extend the observation time, and conduct a multi-center study to analyze the relationship between the dynamic changes in IL-10 plasma levels and OS in HCC patients receiving TACE.

Footnotes

Authors' Contribution: Study concept and design: Y. W., and F. J.; analysis and interpretation of data: S. M. X., and L. N.; drafting of the manuscript: S. M. X. and W. L. L.; critical revision of the manuscript for important intellectual content: Y. W.; statistical analysis: S. M. X., and F. J.

Conflict of Interests: The authors are employees of Second Hospital of Nanjing, Nanjing University of Chinese Medicine.

Ethical Approval: The Second Hospital of Nanjing, Nanjing University of Chinese Medicine Ethics Committee approved the research protocol (2021-LY-kt058).

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Informed Consent: Written informed consent was obtained from all the participants.

References

- Kulik L, El-Serag HB. Epidemiology and Management of Hepatocellular Carcinoma. *Gastroenterology*. 2019;**156**(2):477-491
 e1. [PubMed: 30367835]. [PubMed Central: PMC6340716]. https://doi.org/10.1053/j.gastro.2018.08.065.
- European Association for the Study of the Liver. EASL Clinical Practice Guidelines: Management of hepatocellular carcinoma. J Hepatol. 2018;69(1):182–236. [PubMed: 29628281]. https://doi.org/10.1016/j.jhep.2018.03.019.
- Marrero JA, Kulik LM, Sirlin CB, Zhu AX, Finn RS, Abecassis MM, et al. Diagnosis, Staging, and Management of Hepatocellular Carcinoma: 2018 Practice Guidance by the American Association for the Study of Liver Diseases. *Hepatology*. 2018;68(2):723–50. [PubMed: 29624699]. https://doi.org/10.1002/hep.29913.
- Llovet JM, De Baere T, Kulik L, Haber PK, Greten TF, Meyer T, et al. Locoregional therapies in the era of molecular and immune treatments for hepatocellular carcinoma. *Nat Rev Gastroenterol Hepatol.* 2021;18(5):293-313. [PubMed: 33510460]. https://doi.org/10.1038/s41575-020-00395-0.
- Yu LX, Ling Y, Wang HY. Role of nonresolving inflammation in hepatocellular carcinoma development and progression. *NPJ Precis Oncol.* 2018;2(1):6. [PubMed: 29872724]. [PubMed Central: PMC5871907]. https://doi.org/10.1038/s41698-018-0048-z.
- Jayant K, Habib N, Huang KW, Warwick J, Arasaradnam R. Recent Advances: The Imbalance of Immune Cells and Cytokines in the Pathogenesis of Hepatocellular Carcinoma. *Diagnostics (Basel)*. 2020;10(5). [PubMed: 32466214]. [PubMed Central: PMC7277978]. https://doi.org/10.3390/diagnostics10050338.
- Lee HL, Jang JW, Lee SW, Yoo SH, Kwon JH, Nam SW, et al. Inflammatory cytokines and change of Th1/Th2 balance as prognostic indicators for hepatocellular carcinoma in patients treated with transarterial chemoembolization. *Sci Rep.* 2019;9(1):3260. [PubMed: 30824840].

[PubMed Central: PMC6397294]. https://doi.org/10.1038/s41598-019-40078-8.

- Wu Y, Fan W, Xue M, Zhong B, Zhang S, Wang Y, et al. Postintervention Interleukin-6 (IL-6) Level, Rather than the Pretreatment or Dynamic Changes of IL-6, as an Early Practical Marker of Tumor Response in Hepatocellular Carcinoma Treated with Transarterial Chemoembolization. *Oncologist.* 2019;**24**(12):e1489– 95. [PubMed: 31249138]. [PubMed Central: PMC6975952]. https://doi.org/10.1634/theoncologist.2018-0669.
- Eisenhauer EA, Therasse P, Bogaerts J, Schwartz LH, Sargent D, Ford R, et al. New response evaluation criteria in solid tumours: revised RE-CIST guideline (version 1.1). *Eur J Cancer*. 2009;**45**(2):228–47. [PubMed: 19097774]. https://doi.org/10.1016/j.ejca.2008.10.026.
- Ji L, Gu J, Chen L, Miao D. Changes of Th1/Th2 cytokines in patients with primary hepatocellular carcinoma after ultrasound-guided ablation. *Int J Clin Exp Pathol.* 2017;**10**(8):8715–20. [PubMed: 31966730]. [PubMed Central: PMC6965388].
- Kim MJ, Jang JW, Oh BS, Kwon JH, Chung KW, Jung HS, et al. Change in inflammatory cytokine profiles after transarterial chemotherapy in patients with hepatocellular carcinoma. *Cytokine*. 2013;64(2):516–22. [PubMed: 24035756]. https://doi.org/10.1016/j.cyto.2013.07.021.
- Loosen SH, Schulze-Hagen M, Leyh C, Benz F, Vucur M, Kuhl C, et al. IL-6 and IL-8 Serum Levels Predict Tumor Response and Overall Survival after TACE for Primary and Secondary Hepatic Malignancies. *Int J Mol Sci.* 2018;**19**(6). [PubMed: 29899223]. [PubMed Central: PMC6032291]. https://doi.org/10.3390/ijms19061766.
- Jekarl DW, Lee S, Kwon JH, Nam SW, Kim M, Kim Y, et al. Complex interaction networks of cytokines after transarterial chemotherapy in patients with hepatocellular carcinoma. *PLoS One*. 2019;14(11). e0224318. [PubMed: 31751357]. [PubMed Central: PMC6874208]. https://doi.org/10.1371/journal.pone.0224318.
- Hassan EA, Ahmed EH, Nafee AM, El-Gafary N, Hetta HF, El-Mokhtar MA. Regulatory T Cells, IL10 and IL6 in HCV Related Hepatocellular Carcinoma after Transarterial Chemoembolization (TACE). *Egypt J Immunol.* 2019;**26**(1):69–78. [PubMed: 31332997].
- Sheng T, Wang B, Wang SY, Deng B, Qu L, Qi XS, et al. The Relationship Between Serum Interleukin-6 and the Recurrence of Hepatitis B Virus Related Hepatocellular Carcinoma after Curative Resection. *Medicine* (*Baltimore*). 2015;**94**(24). e941. [PubMed: 26091457]. [PubMed Central: PMC4616529]. https://doi.org/10.1097/MD.000000000000941.
- Pang XH, Zhang JP, Zhang YJ, Yan J, Pei XQ, Zhang YQ, et al. Preoperative levels of serum interleukin-6 in patients with hepatocellular carcinoma. *Hepatogastroenterology*. 2011;**58**(110-111):1687–93. [PubMed: 21940335]. https://doi.org/10.5754/hge10799.
- Jang JW, Oh BS, Kwon JH, You CR, Chung KW, Kay CS, et al. Serum interleukin-6 and C-reactive protein as a prognostic indicator in hepatocellular carcinoma. *Cytokine*. 2012;60(3):686–93. [PubMed: 22906998]. https://doi.org/10.1016/j.cyto.2012.07.017.
- Peluzzo AM, Autieri MV. Challenging the Paradigm: Anti-Inflammatory Interleukins and Angiogenesis. *Cells.* 2022;11(3).
 [PubMed: 35159396]. [PubMed Central: PMC8834461]. https://doi.org/10.3390/cells11030587.
- Zhang S, Gan X, Qiu J, Ju Z, Gao J, Zhou J, et al. II-10 derived from Hepatocarcinoma cells improves human induced regulatory T cells function via JAK1/STAT5 pathway in tumor microenvironment. *Mol Immunol.* 2021;**133**:163-72. [PubMed: 33667986]. https://doi.org/10.1016/j.molimm.2021.02.014.
- 20. Wu RS, Lin J, Xing YM, Gao WL, Jiang YX, Chen LX, et al. OVOL2 in-

hibits macrophage M2 polarization by regulating IL-10 transcription, and thus inhibits the tumor metastasis by modulating the tumor microenvironment. *Immunol Lett.* 2022;**242**:17–26. [PubMed: 34033850]. https://doi.org/10.1016/j.imlet.2021.05.004.

- Colegio OR, Chu NQ, Szabo AL, Chu T, Rhebergen AM, Jairam V, et al. Functional polarization of tumour-associated macrophages by tumour-derived lactic acid. *Nature*. 2014;**513**(7519):559– 63. [PubMed: 25043024]. [PubMed Central: PMC4301845]. https://doi.org/10.1038/nature13490.
- Hu HT, Ai X, Lu M, Song Z, Li H. Characterization of intratumoral and circulating IL-10-producing B cells in gastric cancer. *Exp Cell Res.* 2019;**384**(2):111652. [PubMed: 31574287]. https://doi.org/10.1016/j.yexcr.2019.111652.
- Smith LK, Boukhaled GM, Condotta SA, Mazouz S, Guthmiller JJ, Vijay R, et al. Interleukin-10 Directly Inhibits CD8(+) T Cell Function by Enhancing N-Glycan Branching to Decrease Antigen Sensitivity. *Immunity.* 2018;48(2):299-312 e5. [PubMed: 29396160]. [PubMed Central: PMC5935130]. https://doi.org/10.1016/j.immuni.2018.01.006.
- 24. Mirlekar B. Tumor promoting roles of IL-10, TGF-beta, IL-4, and IL-35: Its implications in cancer immunotherapy. *SAGE Open Med.* 2022;**10**:20503121211069000. [PubMed: 35096390]. [PubMed Central: PMC8793114]. https://doi.org/10.1177/20503121211069012.
- Liu Z, Zhou Q, Wang Z, Zhang H, Zeng H, Huang Q, et al. Intratumoral TIGIT(+) CD8(+) T-cell infiltration determines poor prognosis and immune evasion in patients with muscle-invasive bladder cancer. *J Immunother Cancer*. 2020;8(2). [PubMed: 32817209]. [PubMed Central: PMC7430558]. https://doi.org/10.1136/jitc-2020-000978.
- Wei YY, Fan J, Shan MX, Yin DD, Wang LL, Ye W, et al. TIGIT marks exhausted T cells and serves as a target for immune restoration in patients with chronic HBV infection. *Am J Transl Res.* 2022;14(2):942–54. [PubMed: 35273697]. [PubMed Central: PMC8902551].
- Kuhn R, Lohler J, Rennick D, Rajewsky K, Muller W. Interleukin-10deficient mice develop chronic enterocolitis. *Cell*. 1993;75(2):263-74. [PubMed: 8402911]. https://doi.org/10.1016/0092-8674(93)80068-p.
- Chan SL, Mo FK, Wong CS, Chan CM, Leung LK, Hui EP, et al. A study of circulating interleukin 10 in prognostication of unresectable hepatocellular carcinoma. *Cancer*. 2012;**118**(16):3984–92. [PubMed: 22180222]. https://doi.org/10.1002/cncr.26726.
- Zhang X, Lu M, Xu Y, He G, Liu Q, Zhu J, et al. IL-10 promoter hypomethylation is associated with increased IL-10 expression and poor survival in hepatocellular carcinoma. *Transl Cancer Res.* 2019;8(4):1466–75. [PubMed: 35116889]. [PubMed Central: PMC8797925]. https://doi.org/10.21037/tcr.2019.07.33.
- Zhao S, Wu D, Wu P, Wang Z, Huang J. Serum IL-10 Predicts Worse Outcome in Cancer Patients: A Meta-Analysis. *PLoS One*. 2015;**10**(10). e0139598. [PubMed: 26440936]. [PubMed Central: PMC4595202]. https://doi.org/10.1371/journal.pone.0139598.
- Qi F, Wang L, Huang P, Zhao Z, Yang B, Xia J. Time-series clustering of cytokine expression after transarterial chemoembolization in patients with hepatocellular carcinoma. *Oncol Lett.* 2020;**19**(2):1175–86. [PubMed: 31966047]. [PubMed Central: PMC6955652]. https://doi.org/10.3892/ol.2019.11209.
- 32. Wang H, Zhang G, Fan W, Wu Y, Zhang J, Xue M, et al. Clinical Significance of Peripheral Blood Lymphocyte Subtypes and Cytokines in Patients with Hepatocellular Carcinoma Treated with TACE. *Cancer Manag Res.* 2022;14:451-64. [PubMed: 35153515]. [PubMed Central: PMC8827642]. https://doi.org/10.2147/CMAR.S342527.