



Evaluation of Potential miR-302, miR-132, miR-205, and miR-126 as Biomarkers for Laryngeal Squamous Cell Carcinoma

Amir Oliyaiezezaie¹, Habib Zarredar¹, Milad Asadi², Venus Zafari², Dariush Shanebandi³, Zahra Soleimani¹, Mohammad-Reza Firoozi¹ and Shahram Ghasembaglou^{1,*}

¹Tuberculosis and Lung Disease Research Center, Tabriz University of Medical Sciences, Tabriz, Iran

²Department of Basic Oncology, Ege University, Institute of Health Sciences, Izmir, Turkey

³Immunology Research Center, Tabriz University of Medical Sciences, Tabriz, Iran

*Corresponding author: Tuberculosis and Lung Disease Research Center, Tabriz University of Medical Sciences, Tabriz, Iran. Email: sh.gasembeglo@yahoo.com

Received 2023 March 26; Revised 2023 November 14; Accepted 2023 November 18.

Abstract

Background: Laryngeal tumor is the most commonly identified malignancy in head and neck cancers, containing 2.4% of all cancers, and is the 11th most prevalent and fatal cancer in the world. Previous research has revealed that microRNAs (miRNAs) have a significant role in the progression of laryngeal tumors. Additionally, research focuses on the association with the clinical function of miRNA in malignancy.

Objectives: Based on previous research, miR-132, miR-302, miR-126, and miR-205 have potential roles in the tumor development of the LSCC. Then, in the current study, we pointed to study if there are any significant alters in the expression of microRNAs and whether they have any significant possibility as a diagnostic or prognostic factor for laryngeal squamous cell carcinoma.

Methods: From 30 LSCC patients, we have collected tumor and marginal healthy tissues through laryngectomy. Total RNA extraction from the marginal healthy and malignancy tissues was done. Next, we control the quality of the RNAs and, then, synthesize cDNA. Lastly, the expression of the miRNAs was evaluated by qPCR. The expression level of genes and their association with the patient's clinicopathological features were studied, using related statistical tests.

Results: miR-132 ($P = 0.0001$, fold change: 1.28) and miR-302 ($P = 0.0001$, fold change: 1.05) are significantly up-regulated in tumor tissues compared to normal peripheral tissues. Furthermore, we revealed that miR-205 ($P < 0.0001$, fold change: 0.241) and miR-126 ($P = 0.0001$, fold change: 0.251) are low expressed in LSCC cancer tissue. Between these miRNAs, miR-205 had no relationship with clinical-pathological characteristics in malignancy tissue.

Conclusions: The results of the current study showed the possibility of miR-302, miR-132, miR-126, and miR-205 as diagnostic or prognostic factors in LSCC.

Keywords: Squamous Cell Carcinoma, MicroRNA, Real-time PCR

1. Background

Head and neck malignancy has been ranked as the sixth most frequently diagnosed cancer and laryngeal cancer (LC) is the most frequently observed member of this family and represents the 11th most prevalent and deadliest cancer in the world. The incidence of this disease in respiratory neoplasms is in the second place (95%) (1). LC occurs at any mucosal surface of the larynx and is the deadliest type of cancer. (2). According to the evidence, the 5-year survival rate of LC is 60%. More than half of the patients (54%) are diagnosed and cured before metastasis of the tumor and expansion of malignant cells outside the larynx, which increases the 5-year survival rate to 77%. In

contrast, the 5-year survival rate will be less than 45% if the tumor has extended to nearby tissues or lymph nodes. The 5-year survival rate in patients with metastasis is 33%. However, the site of the tumor (subglottis, supraglottis, glottis) is also another factor that influences the 5-year survival (3). The most prevalent pathological type of laryngeal malignancy is squamous cell carcinoma (LSCC). According to the results of recent studies, the rate of LC is increasing. Even though considerable advances have occurred in surgery and radiation therapy during the past decades, the 5-year survival rate of LSCC has not improved significantly to the high local recurrence rate. The main treatments for LC are surgery, radiotherapy,

chemotherapy, and combination therapy (4). Therefore, for early diagnosis of LC at all stages, even at primary stages whose clinical symptoms are concealed, recognition of greatly sensitive biomarkers seems to be essential for the amelioration of LC patients' outcomes.

MicroRNAs are a group of molecules that has recently become an interesting field of research that may be followed by the recognition of new biomarkers and new therapeutic targets. Consequently, highly specific and sensitive biomarkers are required for the prognosis prediction and early diagnosis of LC even in the early stages without clinical symptoms, and new therapeutic agents are considerably needed to be more effective for controlling and targeting LC cells. This gene family transcribes as a small non-encoding RNA of 22 nucleotides and, then, binds to the complementary mRNA leading to translational suppression or transcriptional degradation. miRNAs play a clear role in different kinds of human malignancies (5). In many diseases, the miRNA expression profile shows marked changes, which are particularly evident in tumors (6). miRNA expression analysis is helpful in the early detection and evaluation of tumor prognosis, metastasis, recurrence, and diagnosis in different malignancies (7). Early and accurate diagnosis in the different stages of the malignancy is important to reach a high survival rate in LSCC; however, early diagnosis is often inefficient due to the lack of specific symptoms (8). For this reason, we reviewed the recent literature and selected these microRNAs, which are studied as prognostic and diagnostic markers in LSCC and other cancers.

2. Objectives

In this research, we aimed at investigating the expression profile of 4 human miRNAs in 30 LC tissues in comparison to adjacent healthy tissues, hoping to detect appropriate and potent markers.

3. Methods

3.1. Study Population

SCC tissues and marginal normal tissues were collected through the laryngectomy from 30 patients between February 2018 and 2019 at Tabriz University of Medical Sciences (Imam Reza Hospital). None of the patients had any experience with radiotherapy or chemotherapy. All samples were transferred directly to the ribonuclease inhibitor solution (QIA Gene Cat NO: 76104) and stored at -80°C until the next step. Current research has been permitted by the Ethics Committee of Tabriz University of Medical Sciences

and all participants read and signed written consent (Ethical code: [IR.TBZMED.REC.1398.999](#)). The general characteristics of the participants are shown in [Table 1](#).

3.2. Total RNA Isolation, Reverse Transcription, and Quantitative PCR

Total RNA extraction from tissues was done by the manufacturer's protocol, using TRIzol reagent (Roche Cat NO: 11667165001). Afterward, the Nanodrop tool was used to check the quantity and quality of RNA (Thermo Fisher Scientific, USA). After that, the samples were stored at -80°C until the next step. For cDNA synthesis, the stem-loop method and 2x RT-PCR (Taq) pre-mixture kit (BioFACT, Seoul) were used. The gene expression level was determined by a Step-one Real-time PCR device (Applied Biosystems, USA) and SYBR Green Master mix (Takara, Korea). In addition, the housekeeping gene for the normalization of miRNA expression was U6. Formula $2^{-\Delta\text{CT}}$ was used to calculate the relative miRNA expression ([Table 2](#)).

3.3. Statistical Analysis

Statistical analysis was carried out, using Graph Pad Prism v.6.00 software to examine the gene expression panel of microRNAs in cancer and marginal samples based on the unpaired *t*-test program. ROC curve analysis was used to assess the potentiality of each of the microRNAs as a biomarker. The value of $P < 0.05$ was considered significant and all values have been defined as mean \pm standard deviation.

4. Results

4.1. miR-302 and mirR-132 are Significantly Up-regulated in Laryngeal Squamous Cell Carcinoma Cancer

Our data show significant overexpression for miR-132 (fold change: 1.28, $P < 0.0001$; [Figure 1](#)) and miR-302 (fold change: 1.05, $P < 0.0001$; [Figure 1](#)) in tumor cells compared to marginal normal cells. Analysis of the association between miR-302 and miR-132 expression and clinical-pathological features of patients had a significant connection among miR-302 and miR-132 expression and different tumor stages in the samples ($P < 0.0001$) ([Table 3](#)).

4.2. miR-126 and miR-205 are Significantly Low Expressed in Laryngeal Tumor

Based on the result of this study, miR-205 (fold change = 0.241, $P < 0.0001$, [Figure 1](#)) and miR-126 (fold change = 0.251, $P < 0.0001$, [Figure 1](#)) are low expressed in malignancy cells compared to marginal normal samples. Also, there

Table 1. Demographical Data and Clinicopathological Features of Patients

Variables	Number of Cases
Age (y)	
< 55	16
≥55	14
Gender	
Male	19
Female	11
Tobaccoexposure	
Smoker	18
Nonsmoker	12
Differentiation	
Well	19
Moderately/poorly	11
Clinicalstage	
I/II	17
III/IV	13
Lymph node metastasis	
Negative	17
Positive	13
Distant metastasis	
Negative	20
Positive	10

Table 2. Primer Sequencing

Micro-RNA	Stemloop	F Primer	R Primer
miR-126-5P	GTCGTATCCAGTGCAGGGTCCGAGGTATTGCGACTGGATACGACGCCTA	CGTGCTCATTACTTT	CCAGTGCAGGGTCCGAGGTA
miR-132-5P	GTCGTATCCAGTGCAGGGTCCGAGGTATTGCGACTGGATACGAAGTTTC	CGTGCTACCGTGGCTTTC	CCAGTGCAGGGTCCGAGGTA
miR-205-5P	GTCGTATCCAGTGCAGGGTCCGAGGTATTGCGACTGGATACGACAGACT	CGTGCTTCCTTCATTCC	CCAGTGCAGGGTCCGAGGTA
miR-302-5P	GTCGTATCCAGTGCAGGGTCCGAGGTATTGCGACTGGATACGAAGCAAG	CGTGCTACTTAACGTGG	CCAGTGCAGGGTCCGAGGTA
U6	GTCGTATCCAGTGCAGGGTCCGAGGTATTGCGACTGGATACGACAAAAATAT GCTTCGGCAGCACATATACTAAAAT	GCTTCGGCAGCACATATACTAAAAT	CGCTTCACGAATTTGCGGTGCAT

Table 3. Correlation Between micro-RNAs Expression with Demographics and Pathologic Features of Patients ($P < 0.05$)

Micro-RNAs	Age	Sex	Tobacco Exposure	Differentiation	Clinical Stage	Lymph Node Metastasis	Distant Metastases
P-Value							
miR-126-5P	0.27	0.21	0.41	0.067	< 0.0001	< 0.0001	< 0.0001
miR-132-5P	0.098	0.15	0.23	< 0.0001	0.071	0.067	0.057
miR-205-5P	0.086	0.45	0.121	0.079	0.073	0.053	0.063
miR-302-5P	miR-302-5P	0.31	0.097	0.053	< 0.0001	< 0.0001	< 0.0001

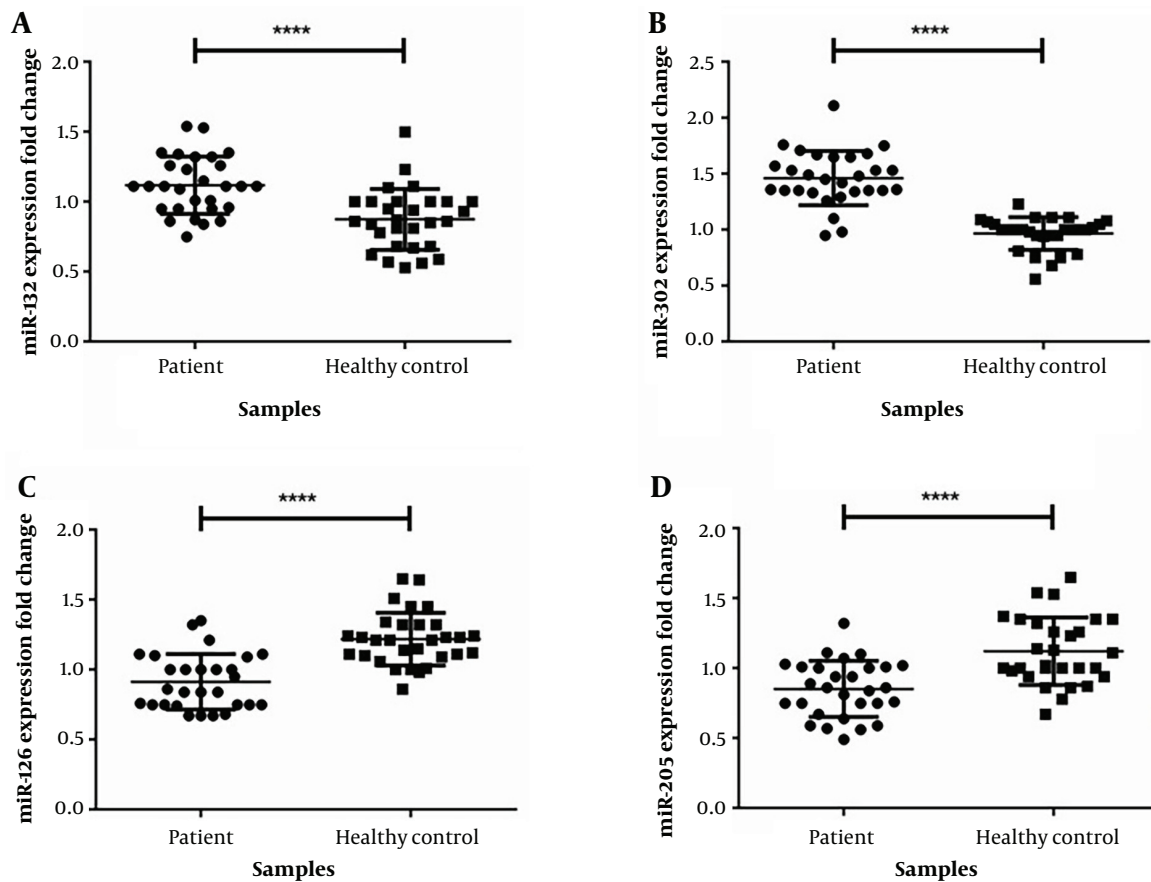


Figure 1. Dysregulation of selected 4 miRNAs in LSCC clinical specimens in comparison to the normal marginal tissue. miR-132 and miR-302 are significantly up-regulated and miR-126 and miR-205 are down-regulated in tumor tissues ($P < 0.0001$)

was a significant correlation between miR-126 expression level and tumor stages of the patients (Table 3). However, we did not find any interaction between the expression of miR-205 level and clinicopathological features of the patients.

4.3. miR-302 as a Diagnosis Marker in LSCC

We used the ROC curve to evaluate miR-205, miR-132, miR-126, and miR-302 specificity and sensitivity as new biomarkers in LSCC. The results determined a ROC region biomarker index of 0.7872, 0.8086, 0.8690, and 0.9489 in LSCC patients, respectively (Figure 2). Among these miRNAs, miR-302 has more specificity and sensitivity to discriminate cancer tissues from healthy marginal (Area: 0.9486) (Figure 2).

5. Discussion

LC is the most frequently diagnosed type of head and neck tumor and LSCC is the most prevalent type of LC (85%). In early-stage diagnosis, the rate of survival is about 90%, but in late-stage diagnosis, the survival rate is about 50%. So, it is critical to find biomarkers for early detection in LC (3, 9). miRNAs could be a precise and helpful biomarker for early diagnosis in tumors including LC (10). The miRNAs, selected for our study, were dysregulated in different types of malignancies and are involved in the pathogenesis of different malignancies including LSCC. The expression level was investigated hoping to identify appropriate biomarkers for LSCC. In this study, miR-302 and miR-132 were considerably overexpressed, and miR-205 and miR-126 were considerably low expressed in LSCC tissues. As far as we know, there is no such research that has been specifically performed to evaluate the expression level of miR-302a in LSCC tumors, but some previous

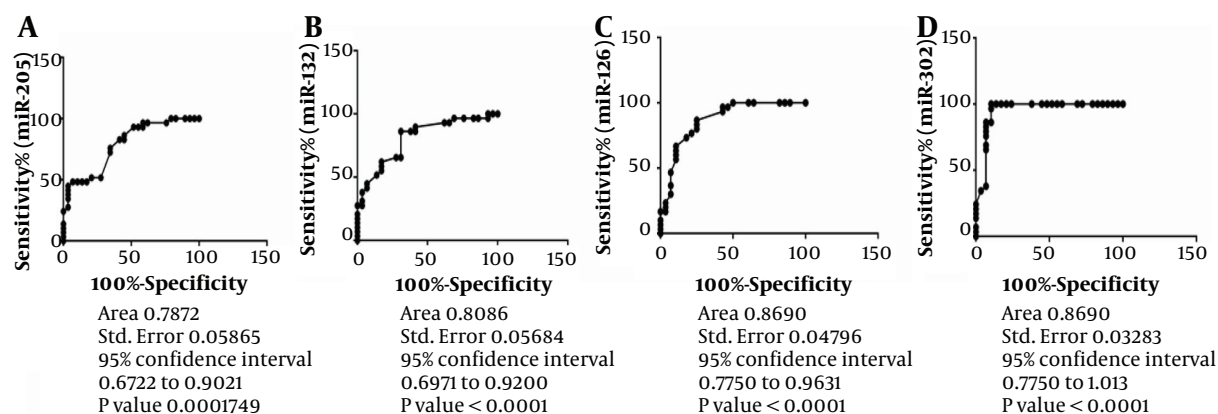


Figure 2. Schematic illustration of receiver operating characteristics (ROC) curve to evaluate the diagnostic potential of area under the curve (AUC) for miRNAs

research reported the aberrant expression of miR-302a in a variety of malignancy types. For instance, a reduction in miR-302a expression level in breast malignancy affects metastasis and invasion controlling of breast tumor cells (11). Previous research has shown that the miR302/367 cluster participates in almost all the stages of the tumor development process in several malignancies (12). Based on previous studies, the miR-302a expression correlates with necrosis and a high level of WRAP53 (13). Guo et al. revealed that miR-302a was considerably low expressed in human ovarian tumor cells, compared with the normal cells. The results suggested that changes in miR-302a expression could be involved in ovarian tumor progression. The cell growth and progression in the miR-302a transfected cells (ovarian cancer) was considerably reduced compared to the control group (14). Zhao et al. have demonstrated that the miR-302a, miR-302d, miR-302c, and miR-302b were considerably low expressed in P-glycoprotein (P-gp)-overexpressing MCF-7/ADR cells. They also revealed that miR-302a/d/b/c ectopic combination expression through targeting the P-gp gene intensifies the sensitivity of breast tumor cells to the anticancer drug PAC, VP-16 (15). Yang CM et al. have shown that miR-302/367 cluster expression in glioblastoma cells prevents the expression of the carcinogenic gene (16). Lower expression of miR-302a has a relation with the VEGF-A expression restrains cell growth and invasiveness and promotes apoptosis in HCC. On the other hand, interruption of cell growth, cell cycle suspension (in vitro), and tumor development (in vivo) in prostate cancer may be the result of miR-302 up-regulation (17). Up-regulation of the miR-302a in the colorectal tumor may prevent cancer cell progression and invasion by preventing the expression of associated

proteins via controlling the MAPK and PI3K/AKT signaling pathways (18). Our findings confirm that miR-302 is more expressed in LSCC patient tissue than in normal tissue. To the best of our knowledge, this is the first time that miR-302 expression level has been studied in LC.

Currently, there is little data on the expression of miR-132 in LSCC. Lian et al. revealed that miR-132 is highly expressed in LC cells directly targets FOXO1 (tumor suppressor), and acts as an important inhibitor of PI3K/Akt signaling. This research confirmed the miR-132 tumorigenic role in LC by controlling the PI3K/AKT/FOXO1 pathway (19).

On the other hand, an in vitro study by Chen et al. on LSCC revealed that miR-132 functions as a tumor suppressor and performs a considerable role in preventing growth, and migration, enhances chemo-sensitivity, and invasion via controlling TGF- β 1/Smad2/3 signals (20). In this research, we detected that miR-132 was highly expressed in LSCC cells and this overexpression was related to the differentiation in LSCC tissues. Most previous studies in various tumors have shown that miR-132 was down-regulated, but in our research, miR-132 was overexpressed (21).

Concerning miR-126, we found that the miR-126 plasma level was reduced in patients with LSCC. Also, we found that miR-126 low expression in patients with LSCC was involved in lymph node metastasis and differentiation. Xin Sun et al. confirmed that miR-126 plasma levels were decreased and could be used as a prognostic marker in patients with LSCC. In addition, they showed that miR-126 plasma levels were negatively correlated with Camsap1 expression. According to these findings, they hypothesize that Camsap1 may be a new target gene for miR-126 (22).

In another study, Sassahira et al. revealed that low

expression of miR-126 is involved in tumor development through induction of angiogenesis and lymphatic angiogenesis via VEGF-A activation in the oral LSCC cell line (23). Also, Yu et al. reported that low expression of miR-126 promotes oral tumors in animal models (24). Liu et al. confirmed that miR-126 is low expressed in lung tumor tissues compared to normal lung tissue and low expression of mir-126 leads to VEGF-A up-regulation in lung malignancy cells (25). They also indicated that the miR-126 expression level was low expressed in esophageal cancer (26). Guo et al. confirmed that miR-126 has a tumor suppressor role, and there is a relation between the miR-126 down-regulation and disrupted signaling via PI3K due to the low expression of its controlling subunit p85 in colon malignancy (27). The expression of mir-205 has shown a bilateral effect in various tumor types, which depends on the stage of malignancy and cell of origin. In certain cell types, miR-205 promotes the generation and progression of tumors as an oncogenic factor. In other types, it prevents cell invasion, growth, and EMT and acts as a tumor inhibitor (28).

In the current research, the expression level of the miR-205 was lower in LSCC tissues in comparison with marginal normal tissues. Tian et al. indicated the tumor suppressor outcome of the miR-205, which may inhibit cell proliferation by regulating Bcl-2 and induce LSCC apoptosis. They also proposed miR-205 as a high-value and potential novel target for therapeutic methods in LSCC (29). Boll et al. found a lower expression level of miR-205 in prostate malignancy. They also indicated that miR-205 up-regulation suppresses major carcinogenic pathways in prostate cancer (30). Lee et al. confirmed that miR-205 low expression causes to elevation of the expression levels of ZEB1 and ZEB2 and a reduction in the expression level of the E-cadherin transcriptional, thereby inducing growth and invasion in breast tumor cells (MCF-7) (31). Also, in 2011, Matsushima found that the CDK2AP1 (tumor suppressor) gene was down-regulated by miR-205, demonstrating that miR-205 controls CDK2AP1 in esophageal squamous cell carcinoma (ESCC). Thus, miR-205 affects the malignant cells as an oncogene via suppressing CDK2AP1 and promotes cell growth and motility by boosting the expression levels of Cyclin D1, C-Myc, MMP-9, and MMP-2 in ESCC cells (32). In this study, we found a positive relationship among miR-302, miR132, and miR-126 expression levels, metastasis, and lymph node involvement. In summary, our research showed that these 4 miRNAs have the potential as a diagnostic or prognostic biomarker in case of future studies confirmation.

5.1. Conclusions

In the present research, we found that miR-302 and miR-132 overexpressed and miR-126 and 205 low expressed in LSCC tissues compared with normal marginal tissues and may be used as a prognosis or diagnosis factor in LSCC.

Acknowledgments

The authors are thankful to patients and their families for their contribution to the study. This study was financially supported by a grant from the Department of Tuberculosis and Lung Diseases Research Center, University Tabriz University of Medical Sciences, Tabriz, Iran.

Footnotes

Authors' Contribution: Concept: A. O., H. Z., SH. GH., design: H. Z., M. A., V. Z., data collection or processing: M. R. F., V. Z., D. SH., analysis or Interpretation: H. Z., SH. GH., Z. S., A. O., literature search: M. A., Z. S., M. R. F., approval: SH. GH., A. O., and H. Z.

Clinical Trial Registration Code: This study is not an interventional study.

Conflict of Interests: The authors declare no conflict of interest.

Data Reproducibility: The data presented in this study are openly available in one of the repositories or will be available on request from the corresponding author by this journal representative at any time during submission or after publication.

Ethical Approval: The study was approved by the Ethics Committee of Tabriz University of Medical Sciences (Ethical code: [IR.TBZMED.REC.1398.999](https://doi.org/10.1398.999)).

Funding/Support: This study was supported by a grant from the Tuberculosis and Lung Disease Research Center, Tabriz University of Medical Science, Tabriz, Iran.

Informed Consent: All participants signed the informed consent. All the procedures were performed as a part of the routine care.

References

- Miszczuk L, Maciejewski B, Tukiendorf A, Wozniak G, Jochymek B, Gawryszuk A, et al. Split-course accelerated hyperfractionated irradiation (CHA-CHA) as a sole treatment for advanced head and neck cancer patients-final results of a randomized clinical trial. *Br J Radiol.* 2014;**87**(1041):20140212. [PubMed ID: [25027170](https://pubmed.ncbi.nlm.nih.gov/25027170/)]. [PubMed Central ID: [PMC4453153](https://pubmed.ncbi.nlm.nih.gov/PMC4453153/)]. <https://doi.org/10.1259/bjr.20140212>.

2. Saito K, Inagaki K, Kamimoto T, Ito Y, Sugita T, Nakajo S, et al. MicroRNA-196a is a putative diagnostic biomarker and therapeutic target for laryngeal cancer. *PLoS One*. 2013;**8**(8):e71480. [PubMed ID: 23967217]. [PubMed Central ID: PMC3743786]. <https://doi.org/10.1371/journal.pone.0071480>.
3. American Society of Clinical Oncology. Laryngeal and hypopharyngeal cancer. *Statistics*. 2016.
4. Song FC, Yang Y, Liu JX. Expression and significances of MiRNA Let-7 and HMGA2 in laryngeal carcinoma. *Eur Rev Med Pharmacol Sci*. 2016;**20**(21):4452-8. [PubMed ID: 27874955].
5. Zarredar H, Farajnia S, Ansarin K, Baradaran B, Aria M, Asadi M. Synergistic Effect of Novel EGFR Inhibitor AZD8931 and p38alpha siRNA in Lung Adenocarcinoma Cancer Cells. *Anticancer Agents Med Chem*. 2019;**19**(5):638-44. [PubMed ID: 30827261]. <https://doi.org/10.2174/1871520619666190301125203>.
6. Yu X, Li Z. The role of microRNAs expression in laryngeal cancer. *Oncotarget*. 2015;**6**(27):23297-305. [PubMed ID: 26079642]. [PubMed Central ID: PMC469519]. <https://doi.org/10.18632/oncotarget.4195>.
7. Li JZ, Gao W, Lei WB, Zhao J, Chan JY, Wei WI, et al. MicroRNA 744-3p promotes MMP-9-mediated metastasis by simultaneously suppressing PDCD4 and PTEN in laryngeal squamous cell carcinoma. *Oncotarget*. 2016;**7**(36):58218-33. [PubMed ID: 27533461]. [PubMed Central ID: PMC5295426]. <https://doi.org/10.18632/oncotarget.11280>.
8. Bartel DP. MicroRNAs: Genomics, biogenesis, mechanism, and function. *Cell*. 2004;**116**(2):281-97. [PubMed ID: 14744438]. [https://doi.org/10.1016/S0092-8674\(04\)00045-5](https://doi.org/10.1016/S0092-8674(04)00045-5).
9. Wang J, Wu Y, Gao W, Li F, Bo Y, Zhu M, et al. Identification and characterization of CD133(+)/CD44(+) cancer stem cells from human laryngeal squamous cell carcinoma cell lines. *J Cancer*. 2017;**8**(3):497-506. [PubMed ID: 28261352]. [PubMed Central ID: PMC5332902]. <https://doi.org/10.7150/jca.17444>.
10. Zarredar H, Ansarin K, Baradaran B, Ahdi Khosroshahi S, Farajnia S. Potential Molecular Targets in the Treatment of Lung Cancer Using siRNA Technology. *Cancer Invest*. 2018;**36**(1):37-58. [PubMed ID: 29336624]. <https://doi.org/10.1080/07357907.2017.1416393>.
11. Liang Z, Bian X, Shim H. Inhibition of breast cancer metastasis with microRNA-302a by downregulation of CXCR4 expression. *Breast Cancer Res Treat*. 2014;**146**(3):535-42. [PubMed ID: 25030358]. <https://doi.org/10.1007/s10549-014-3053-0>.
12. Chen L, Heikkinen L, Emily Knott K, Liang Y, Wong G. Evolutionary conservation and function of the human embryonic stem cell specific miR-302/367 cluster. *Comp Biochem Physiol Part D Genomics Proteomics*. 2015;**16**:83-98. [PubMed ID: 26363379]. <https://doi.org/10.1016/j.cbd.2015.08.002>.
13. Meng WJ, Pathak S, Zhang X, Adell G, Jarlsfelt I, Holmlund B, et al. Expressions of miR-302a, miR-105, and miR-888 play critical roles in pathogenesis, radiotherapy, and prognosis on rectal cancer patients: A study from rectal cancer patients in a Swedish rectal cancer trial of preoperative radiotherapy to big database analyses. *Front Oncol*. 2020;**10**:567042. [PubMed ID: 33123477]. [PubMed Central ID: PMC7573294]. <https://doi.org/10.3389/fonc.2020.567042>.
14. Guo T, Yu W, Lv S, Zhang C, Tian Y. MiR-302a inhibits the tumorigenicity of ovarian cancer cells by suppression of SDC1. *Int J Clin Exp Pathol*. 2015;**8**(5):4869-80. [PubMed ID: 2619180]. [PubMed Central ID: PMC4503052].
15. Zhao L, Wang Y, Jiang L, He M, Bai X, Yu L, et al. MiR-302a/b/c/d cooperatively sensitizes breast cancer cells to adriamycin via suppressing P-glycoprotein(P-gp) by targeting MAP/ERK kinase kinase 1 (MEKK1). *J Exp Clin Cancer Res*. 2016;**35**:25. [PubMed ID: 26842910]. [PubMed Central ID: PMC4738800]. <https://doi.org/10.1186/s13046-016-0300-8>.
16. Yang CM, Chiba T, Brill B, Delis N, von Manstein V, Vafaizadeh V, et al. Expression of the miR-302/367 cluster in glioblastoma cells suppresses tumorigenic gene expression patterns and abolishes transformation related phenotypes. *Int J Cancer*. 2015;**137**(10):2296-309. [PubMed ID: 25991553]. [PubMed Central ID: PMC4744715]. <https://doi.org/10.1002/ijc.29606>.
17. Plos One Editors. Correction: MicroRNA-302a Suppresses Tumor Cell Proliferation by Inhibiting AKT in Prostate Cancer. *PLoS One*. 2020;**15**(10):e0241462. [PubMed ID: 33091075]. [PubMed Central ID: PMC7580994]. <https://doi.org/10.1371/journal.pone.0241462>.
18. Wei ZJ, Tao ML, Zhang W, Han GD, Zhu ZC, Miao ZG, et al. Up-regulation of microRNA-302a inhibited the proliferation and invasion of colorectal cancer cells by regulation of the MAPK and PI3K/Akt signaling pathways. *Int J Clin Exp Pathol*. 2015;**8**(5):4481-91. [PubMed ID: 26191138]. [PubMed Central ID: PMC4503010].
19. Lian R, Lu B, Jiao L, Li S, Wang H, Miao W, et al. MiR-132 plays an oncogenic role in laryngeal squamous cell carcinoma by targeting FOXO1 and activating the PI3K/AKT pathway. *Eur J Pharmacol*. 2016;**792**:1-6. [PubMed ID: 27751825]. <https://doi.org/10.1016/j.ejphar.2016.10.015>.
20. Chen L, Zhu Q, Lu L, Liu Y. MiR-132 inhibits migration and invasion and increases chemosensitivity of cisplatin-resistant oral squamous cell carcinoma cells via targeting TGF-beta1. *Bioengineered*. 2020;**11**(1):91-102. [PubMed ID: 31906769]. [PubMed Central ID: PMC6961592]. <https://doi.org/10.1080/21655979.2019.1710925>.
21. Chen X, Li M, Zhou H, Zhang L. miR-132 Targets FOXA1 and Exerts Tumor-Suppressing Functions in Thyroid Cancer. *Oncol Res*. 2019;**27**(4):431-7. [PubMed ID: 29523221]. [PubMed Central ID: PMC7848280]. <https://doi.org/10.3727/096504018X15201058168730>.
22. Sun X, Wang ZM, Song Y, Tai XH, Ji WY, Gu H. MicroRNA-126 modulates the tumor microenvironment by targeting calmodulin-regulated spectrin-associated protein 1 (Camsap1). *Int J Oncol*. 2014;**44**(5):1678-84. [PubMed ID: 24603804]. <https://doi.org/10.3892/ijo.2014.2321>.
23. Sasahira T, Kurihara M, Bhawal UK, Ueda N, Shimomoto T, Yamamoto K, et al. Downregulation of miR-126 induces angiogenesis and lymphangiogenesis by activation of VEGF-A in oral cancer. *Br J Cancer*. 2012;**107**(4):700-6. [PubMed ID: 22836510]. [PubMed Central ID: PMC3419968]. <https://doi.org/10.1038/bjc.2012.330>.
24. Yu T, Wang XY, Gong RG, Li A, Yang S, Cao YT, et al. The expression profile of microRNAs in a model of 7,12-dimethyl-benz[a]anthracene-induced oral carcinogenesis in Syrian hamster. *J Exp Clin Cancer Res*. 2009;**28**(1):64. [PubMed ID: 19435529]. [PubMed Central ID: PMC2687417]. <https://doi.org/10.1186/1756-9966-28-64>.
25. Liu B, Peng XC, Zheng XL, Wang J, Qin YW. MiR-126 restoration down-regulate VEGF and inhibit the growth of lung cancer cell lines in vitro and in vivo. *Lung Cancer*. 2009;**66**(2):169-75. [PubMed ID: 19223090]. <https://doi.org/10.1016/j.lungcan.2009.01.010>.
26. Liu SG, Qin XG, Zhao BS, Qi B, Yao WJ, Wang TY, et al. Differential expression of miRNAs in esophageal cancer tissue. *Oncol Lett*. 2013;**5**(5):1639-42. [PubMed ID: 23761828]. [PubMed Central ID: PMC3678876]. <https://doi.org/10.3892/ol.2013.1251>.
27. Guo C, Sah JF, Beard L, Willson JK, Markowitz SD, Guda K. The noncoding RNA, miR-126, suppresses the growth of neoplastic cells by targeting phosphatidylinositol 3-kinase signaling and is frequently lost in colon cancers. *Genes Chromosomes Cancer*. 2008;**47**(11):939-46. [PubMed ID: 18663744]. [PubMed Central ID: PMC2739997]. <https://doi.org/10.1002/gcc.20596>.
28. Gregory PA, Bert AG, Paterson EL, Barry SC, Tsykin A, Farshid G, et al. The miR-200 family and miR-205 regulate epithelial to mesenchymal transition by targeting ZEB1 and SIP1. *Nat Cell Biol*. 2008;**10**(5):593-601. [PubMed ID: 18376396]. <https://doi.org/10.1038/ncb1722>.
29. Tian L, Zhang J, Ge J, Xiao H, Lu J, Fu S, et al. MicroRNA-205 suppresses proliferation and promotes apoptosis in laryngeal squamous cell carcinoma. *Med Oncol*. 2014;**31**(1):785. [PubMed ID: 24297308]. <https://doi.org/10.1007/s12032-013-0785-3>.
30. Boll K, Reiche K, Kasack K, Morbt N, Kretzschmar AK, Tomm JM,

- et al. MiR-130a, miR-203 and miR-205 jointly repress key oncogenic pathways and are downregulated in prostate carcinoma. *Oncogene*. 2013;**32**(3):277-85. [PubMed ID: 22391564]. <https://doi.org/10.1038/onc.2012.55>.
31. Lee JY, Park MK, Park JH, Lee HJ, Shin DH, Kang Y, et al. Loss of the polycomb protein Mel-18 enhances the epithelial-mesenchymal transition by ZEB1 and ZEB2 expression through the downregulation of miR-205 in breast cancer. *Oncogene*. 2014;**33**(10):1325-35. [PubMed ID: 23474752]. <https://doi.org/10.1038/onc.2013.53>.
32. Zhong G, Xiong X. miR-205 promotes proliferation and invasion of laryngeal squamous cell carcinoma by suppressing CDK2AP1 expression. *Biol Res*. 2015;**48**:60. [PubMed ID: 26515287]. [PubMed Central ID: PMC4625464]. <https://doi.org/10.1186/s40659-015-0052-5>.