Published online 2016 June 13.

#### **Research Article**

# Immunohistochemistry Study of *P*53 and *C-erbB-2* Expression in Trophoblastic Tissue and Their Predictive Values in Diagnosing Malignant Progression of Simple Molar Pregnancy

Malihe Hasanzadeh,<sup>1</sup> Norrie Sharifi,<sup>2</sup> Marjaneh Farazestanian,<sup>3</sup> Seyed Saman Nazemian,<sup>4</sup> and Faezeh Madani Sani<sup>5,\*</sup>

<sup>1</sup>Department of Obstetrics and Gynecology, Women Health Research Center, Qaem Hospital, Mashhad University of Medical Sciences, Mashhad, IR Iran

<sup>2</sup>Department of Pathology, Qaem Hospital, Mashhad University of Medical Sciences, Mashhad, IR Iran

<sup>3</sup>Women Health Research Center, Qaem Hospital, Mashhad University of Medical Sciences, Mashhad, IR Iran

<sup>4</sup>Emam Reza Hospital, Mashhad University of Medical Sciences, Mashhad, IR Iran

<sup>5</sup>Qaem Hospital, Mashhad University of Medical Sciences, Mashhad, IR Iran

Corresponding author: Faezeh Madani Sani, Qaem Hospital, Mashhad University of Medical Sciences, Number 36, 9th Malek Ashtar St., Ershad Blvd., Mashhad, IR Iran. Tel: +98-9153221067, E-mail: fmadanisani@yahoo.com

Received 2015 September 22; Revised 2015 December 09; Accepted 2016 June 05.

#### Abstract

**Background:** Finding a tumor marker to predict the aggressive behavior of molar pregnancy in early stages has yet been a topic for studies.

**Objectives:** In this survey we planned to study patients with molar pregnancy to 1) assess the *p*53 and *c-erbB-2* expression in trophoblastic tissue, 2) to study the relationship between their expression intensity and progression of a molar pregnancy to gestational trophoblastic neoplasia, and 3) to determine a cut off value for the amount of *p*53 and *c-erbB-2* expression which might correlate with aggressive behavior of molar pregnancy.

**Patients and Methods:** In a prospective cross sectional study by using a high accuracy technique EnVision <sup>Tm</sup> system for immunohistochemistry staining of molar pregnancy samples, we evaluated *p*53 and *c-erbB-2* expression in cytotrophoblast and syncytiotrophoblast and the correlation of their expression with progression of molar pregnancy to gestational trophoblastic neoplasia (GTN). Normal prostatic tissue and Breast cancer tissue were used as positive controls.

**Results:** We studied 28 patients with simple molar pregnancy (SMP) and 30 with GTN. Cytotrophobalst had significantly higher expression of *p*53 and *c-erbB-2* and syncytiotrophoblast had greater expression of *p*53 in GTN group as compared to SMP group. The cut off values for percentage of *p*53 positive immunostained cytotrophoblast and syncytiotrophoblast were 5.5% and 2.5%. In *c-erbB-2* positive membranous stained cytotrophoblast the cut off was 12.5%.

**Conclusions:** Our data suggests that over expression of *p*53 and *c-erbB-2* is associated with malignant progression of molar pregnancy. We encountered that high expression of *p*53 and *c-erbB-2* in trophoblastic cells could predict gestational trophoblastic neoplasia during the early stages.

Keywords: Complete Hydatiform Mole, P53, C-erbB-2, Prognosis

# 1. Background

Gestational trophoblastic disease is a group of heterogeneous conditions; ranging from simple molar pregnancy to gestational trophoblastic neoplasia with aggressive behavior and metastasis (1-3). It has an incidence rate of 1 per 1000 pregnancies in Western countries (4) and is more common among Eastern populations (5, 6). Although the metastatic form could be potentially fatal, early diagnosis and chemotherapy makes it as one of the most curable solid tumors (7-9).

Current guidelines suggest measuring weekly serum  $\beta$ hCG (human chorionic gonadotropin) following the evacuation of pregnancy products and in case of plateau or rising pattern; the persistent gestational trophoblastic

disease would be suspected and chemotherapy should be started (10, 11) it is a great advantage if one could predict aggressive behavior of the disease before an increase in serum  $\beta$ hCG. Researchers have studied molecular pathogenesis of gestational trophoblastic neoplasia and some found *p53* and *c-erbB-2* to have a role in malignant behaviors of these tumors (12-16). However, there were controversies whether *p53* and *c-erbB-2* expression could act as tumor markers.

## 2. Objectives

In this survey we planned to study patients with molar pregnancy using immunohistochemistry staining to 1) as-

Copyright © 2016, Iranian Journal of Cancer Prevention. This is an open-access article distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 International License (http://creativecommons.org/licenses/by-nc/4.0/) which permits copy and redistribute the material just in noncommercial usages, provided the original work is properly cited.

sess *p*53 and *c-erbB-2* expression in trophoblastic tissue, 2) to study the relationship between their expression intensity and progression of a molar pregnancy to gestational trophoblastic neoplasia, and 3) to determine a cut off value for *p*53 and *c-erbB-2* expression intensity in case of correlation with aggressive behavior of molar pregnancy.

## 3. Patients and Methods

#### 3.1. Population

In a prospective cross sectional study, we included patients with primary diagnosis of molar pregnancy referring to oncology clinic of Qaem hospital, affiliated to MUMS. All patients underwent evacuation and curettage, followed by weekly  $\beta$ hCG measurements. Patients were divided into two groups: (1) gestational trophoblastic neoplasia (GTN) group if serum  $\beta$ hCG level rose or did not change during study; (2) simple molar pregnancy group whose serum  $\beta$ hCG underwent gradual decrease. Serum  $\beta$ hCG level < 5 mIU/mL was considered as normal. Patients' specimen of curettage were referred to pathology laboratory of hospital for histological and immunochemistry studies.

#### 3.2. Histological and Immunochemistry Studies

Immunohistochemistry staining was performed on multiple 4  $\mu$ m sections of paraffin blocks provided from formalin fixed trophoblastic tissues. In order to evaluate the immunoreactivity of *c-erbB-2* oncogene and *p53* tumor suppressor gene, we applied a polymer based Dako Envision <sup>Tm</sup> system technique; (Do-7, Dakocytomation, N1581, DAKO Corporation, Carpiteria, CA 93013 USA) for p53 antigen and (Clone PN2A, Dakocytomation, Denmark A/S, DK-2600 Glostrup, Denmark) for c-erbB-2. Normal prostatic tissue and breast cancer slides were used as positive controls for p53 and c-erbB-2 respectively due to company protocols. As negative controls, phosphate buffered saline (PBS) was substituted with antibodies. All slides were observed by a single pathologist under a light microscope (Olympus  $B \times 50$ ; Olympus optical Co, Ltd, Tokyo Japan). The rate of p53 expression was reported as percentage of cytotrophoblastic and syncytiotrophoblastic cells with positive nuclear immunoreactivity. The c-erbB-2 oncogene expression rate was calculated as percentage of cells with positive membranous staining. To grade p53 staining intensity semi quantitatively, we applied 0 for no stained cells, + for staining of less than 10% of cells, ++ for 10 to 50% of cells, +++ for staining in more than 50% of cells. To score cerbB-2 staining intensity we used negative as no or less than 10% of cells' membranes stained, 1+ for faint membranous staining in more than 10% of cells, 2+ for weak to moderate

complete membranous staining in more than 10% of cells and evaluate 3+ as strong for complete membranous staining in more than 30% of cells (17). All tissue preparation stages were performed based on Dako Envision Tm company protocols (18).

# 3.3. Statistical Analysis

Data were entered on SPSS for windows software version 21. Categorical data were analyzed by chi-square or exact Fischer test. Mann-Whitney test and independent sample t-test were applied to compare continuous variables. To estimate a cut off for percentage of positive immunostained cells ROC (receiver operating characteristic) curve analysis was applied to evaluate the risk of transformation of molar pregnancy to gestational trophoblastic neoplasia. The P value < 0.05 was considered statistically significant.

## 3.4. Ethics

Informed consents were signed by all patients. All diagnostic and therapeutic interventions including evacuation of pregnancy products, serial weekly measurement of serum  $\beta$ hCG, and histological evaluation for their primary diagnosis (complete or partial molar pregnancy) were performed according to indications for patients with molar pregnancy diagnosis. Immunohistochemistry expenses were covered by the research budget.

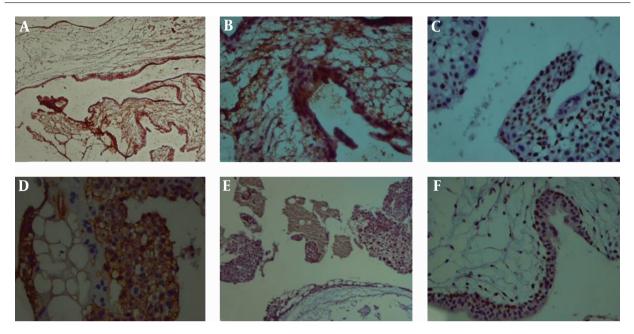
## 4. Results

We included 58 patients: 30 with final diagnosis of Gestational Trophoblastic Neoplasia (GTN) and 28 with simple molar pregnancy. Although Patients with GTN diagnosis had a higher age average, gravidity and parity number in comparison with simple molar pregnancy group, only age had a significant correlation with GTN. The primary diagnosis of 28 patients with (GTN) was complete molar pregnancy (P value < 0.05). Table 1 displays patients' demographics.

The immunohistochemistry staining results in GTN group showed a significantly higher average percentage of cytotrophobalst and syncytiotrophoblast with positive nuclear immunoreactivity for *p*53 in comparison with simple molar patients. The membranous immunostaining of cytotrophoblast for *c-erbB-2* was also significantly greater in GTN group. Patients with primary diagnosis of complete mole had significantly higher percentage of *p*53 positive cytotrophoblast and syncytiotrophoblast in comparison with partial mole group (Table 2 and Figure 1).

Patients in GTN group displayed a significantly higher immunoreactivity score for *p*53 among both cytotrophoblast and syncytiotrophoblast as compared to patients

## Figure 1. Hydropic Villi with Trophoblastic Proliferation and Their Diffuse Nuclear Immunoreactivity for P53 ( $\times$ 40)



(A), Nuclear cytotrophoblasts and syncytiotrophoblasts immunoreactivity ( $\times$  100) (B), Negative immunoreactivity of syncytiotrophoblasts and positive nuclear immunoreactivity of more than 80% of trophoblasts (C), positive cytoplasmic immunoreactivity of trophoblasts for *c-erbB-2* with fairly no staining of syncytiotrophoblasts (D), Hydropic villi with trophoblasts (vacuolization and positive immunoreactivity in about 80% of trophoblasts ( $\times$  40)(E), ( $\times$  100) (F).

Table 1.	Demographics of	Patients and	Results of	Chi-Square	and Independent	T-
Test <sup>a</sup>						

Simple Mole	GTN	P Value
28	30	
		0.00
18	28	
10	2	
$26.2\pm7.4$	$31.9 \pm 9.0$	0.01
$11.3\pm4.0$	$11\pm3.2$	0.79
$1.7\pm1.5$	$3.1\pm2.3$	
$0.5\pm1.5$	$1.9\pm2.2$	
	$ 28 18 10 26.2 \pm 7.4 11.3 \pm 4.0 1.7 \pm 1.5 $	28     30       18     28       10     2       26.2 ± 7.4     31.9 ± 9.0       11.3 ± 4.0     11 ± 3.2       1.7 ± 1.5     3.1 ± 2.3

Abbreviation: GTN, Gestational Trophoblastic Neoplasia.

<sup>a</sup>P value < 0.05 considered significant.

with simple molar pregnancy. Membranous immunoreactivity score of cytotrophoblast for *c-erbB-2* marker were also higher among GTN group. Syncytiotrophoblast showed fairly similar immunoreactivity for *c-erbB-2* marker in both GTN and simple molar pregnancy groups with no statistical meaningful difference (Table 3).

The receiver operating characteristic (ROC) curve analysis displayed the 5.5% as a cutoff percentage for cytotrophoblast with *p*53 nuclear immunostaining (93.3% sensitivity, 88% specificity). For syncytotrophoblast with sensitivity and specificity of 90% and 88% respectively, the cutoff value of 2.5% was determined. We found the cut off value of 12% for the percentage of cytotrophoblast with c-erbB-2 membranous staining (sensitivity of 90% and a specificity of 92%) which might increase the risk of progression of a molar pregnancy to GTN (Figure 2). The positive predictive values were as 90%, 88.8% 88.4% for calculated cut off of *p*53positive cytotrophoblast, *p*53 positive syncytiotrophoblast and *c-erbB-2* positive cytotrophoblast respectively (Table 4).

# 5. Discussion

In patients with simple molar pregnancy, gynecologists are always concerned about their progression toward the gestational trophoblastic neoplasia. To date the only way to evaluate patients following the evacuation and curettage, is serial measurement of serum  $\beta$ hCG (11). Regarding the invasive and metastatic behavior of malignant transformations of molar pregnancy, finding a marker with high predictive value to diagnose the malignant forms early after evacuation is of great importance. In this survey, we found that *p*53 and *c-erbB-2* genes had higher expressions in both cytotrophoblast and syncyTable 2. Percentage of Cytotrophobalsts and Syncytiotrophoblasts with Positive Immunoreactivity for *c-erbB-2* and *P*53 of Patients Regarding Their Final Diagnosis, Primary Diagnosis and Mann-Whitney Test Results<sup>a,b</sup>

	GTN	Simple Mole	P Value	Complete Mole	Partial Mole	P Value
P53 Cytotrophobalsts	$41.6\pm25.4$	$5.3\pm9.3$	0.000	$27.9\pm26.7$	$9.0\pm21.1$	0.007
P53 Syncytiotrophoblasts	$19.3 \pm 18.8$	$1.2\pm1.1$	0.000	$12.1\pm17.1$	$4.0\pm9.8$	0.027
C-erbB-2 Cytotrophobalsts	$18.4\pm26.4$	$2.5\pm8.6$	0.000	$11.7\pm21.6$	$6.2\pm20.1$	0.053
C-erbB-2 Syncytiotrophoblasts	$5.4\pm14.5$	$1.6\pm5.4$	0.508	$4.0\pm12.1$	$1.6\pm5.7$	0.437

Abbreviation: GTN, Gestational Trophoblastic Neoplasia.

<sup>a</sup>P value < 0.05 considered significant.

<sup>b</sup>Values are expressed as mean (SD).

Table 3. Semi Quantitative Immunoreactivity Score of Cytotrophoblasts and Syncytiotrophoblasts for P53 and C-erbB-2 Markers<sup>a,b,c</sup>

	P53 Cytotrophoblast		P53 Syncytio	P53 Syncytiotrophoblast		C-erbB-2 Cytotrophoblasts		C-erbB-2 Syncytiotrophoblast	
	SMP	GTN	SMP	GTN	SMP	GTN	SMP	GTN	
Negative	1(3.8)	1(3.3)	6 (23.1)	2 (6.9)	23 (88.5)	7 (23.3)	23 (88.5)	25 (83.3)	
+	22 (84.6)	3 (10)	20 (76.9)	11 (37.9)	1(3.8)	12 (40)	1(3.8)	1(3.3)	
++	3 (11.5)	17 (56.7)	0(0)	15 (51.7)	2 (7.7)	7 (23.3)	2 (7.7)	3 (10)	
+++	0(0)	9 (30)	0(0)	1(3.4)	0(0)	4 (13.3)	0(0)	1 (3.3)	
P Value	0.000		0.0	00	0.0	00	1.0	00	

Abbreviations: GTN, Gestational Trophoblastic Neoplasia; SMP, Simple Molar Pregnancy.

<sup>a</sup> For *p*53 marker, we applied 0 for no stained cells, + for staining of less than 10% of cells, ++ for 10 to 50% of cells, +++ for staining in more than 50% of cells. To score *c-erbB-2* staining intensity we used negative as of no or less than 10% of cells' membranes stained, + for faint membranous staining in more than 10% of cells, +++ for weak to moderate complete membranous staining in more than 30% of cells, results of chi-square test.

<sup>b</sup>P value < 0.05 considered significant.

<sup>c</sup>Values are expressed as No. (%).

Table 4. The Positive and Negative Predictive Values for Calculated Cut Offs for Percentage of Cells with Positive Immunostaining to Diagnose Malignant Progression of Molar Pregnancy

	P53 Cytotrophoblasts, (%)	P53 Syncytiotrophoblast, (%)	C-erbB-2 Cytotrophoblasts, (%)
Cut off for percentage of cells with positive immunostaining	5.5	2.5	12.5
Positive predictive value	90	88.8	88.4
Negative predictive value	92	82.1	76.6

tiotrophoblast of GTN patients in comparison with simple molar patients with significant difference.

*P*53 is known as a tumor suppressor gene which encodes a nuclear phosphoprotein and its mutation seems to involve in many human cancers' pathogenesis (13, 19, 20). Several studies have been performed on the role of *p*53 in gestational trophoblastic neoplasia. Petignat et al. reported the over expression of mutant *p*53 in complete moles and malignant forms (21). Although many studies reported an increase in *p*53 expression in GTN and complete mole in comparison with simple molar pregnancy and partial mole (12, 14 - 16, 22-24), some found that the in-

creased type is rather the wild type to the mutant type of *p53* (12, 23, 25, 26). Yang et al. also reported an increased expression of *p53* in GTN and complete mole although he did not find a significant predictive value for such increase to diagnose the malignant forms in early stages (16). Whether the mutant type or wild type are over expressed, in this survey we found that *p53* expression increased significantly in cytotrophoblast and syncytiotrophoblast of GTN and complete moles. We found the positive predictive values of 90% and 88.8% when 5.5% and 2.5% of cytotrophoblast and syncytiotrophoblast with positive nuclear immunoactivity were used as the cut off respectively. Using the same

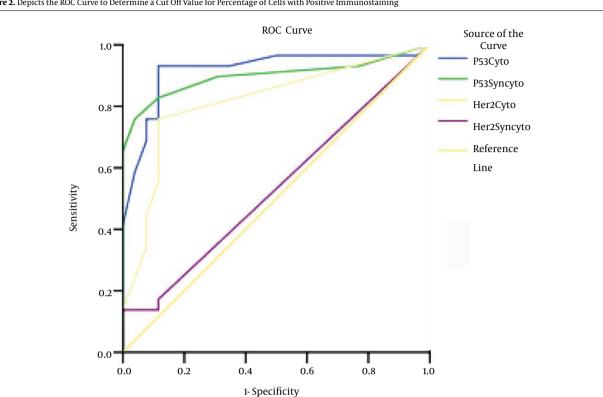


Figure 2. Depicts the ROC Curve to Determine a Cut Off Value for Percentage of Cells with Positive Immunostaining

Diagonal segments are produced by ties.

Test Result		Std. Error <sup>a</sup>	L	Asymptotic %95 Confidence Interval		
Variable (s)	Area		Asymptotic Sig. <sup>b</sup>	Lower Bound	Upper Bound	
P53Cyto	.917	.042	.000	.835	.999	
P53Syncyto	.899	.047	.000	.807	.991	
Her2Cyto	.816	.060	.000	.698	.934	
Her2Syncyto	.534	.078	.661	.381	.688	

For p53 and c-erbB-2 markers among cytotrophoblasts and syncytiotrophoblasts in molar tissue to estimate their risk to GTN transformation. The test result variable(s): p53Cyto, p53Syncyto, Her2Cyto, Her2Syncyto has at least one tie between the positive actual state group and the negative actual state group. Statistics may be biased. a, Under the nonparametric assumption. b, Null hypothesis, true area = 0.5.

method of immunostaining, Chen Y et al. results support our data and find it useful to evaluate p53 expression as adjuncts to conventional methods of diagnosis (24).

Epidermal growth factor receptors (EGFRs) are a big family of transmembrane signaling proteins which are involved in many human neoplasms pathogenesis (27-29). C-erbB-2 is a member of (EGFRs) family and is involved in pathogenesis of different malignancies including melanomas, breast cancer, and colorectal cancer (30, 31) as well as complete mole and choriocarcinoma (16, 27, 32, 33). Yang et al. reported a prognostic value of 84% for

percentage of cytorophoblasts with positive cytoplasmic immunostaining to predict the malignant progression of simple molar pregnancy (16). However, there are conflicting reports: Cameron et al. reported the expression of *c*erbB-2 in only one case out of 20 patients with persistent gestational trophoblastic disease (34); Dehaghani AS et al. found that the mean serum *c-erbB-2* does not differ significantly between GTD patients and normal pregnant controls (35). In contrast to these reports, we found an over expression of *c-erbB-2* in cytotrophoblast of patients with GTN and complete mole with significant difference to simple molar pregnancy and partial mole. We determined a cut off value of 12.5% for the percentage of cytotrophoblast with *c-erbB-2* membranous staining (sensitivity of 90% and a specificity of 92%) which might increase the risk of progression of a molar pregnancy to GTN. Many researchers have supported our data: Yang et al. has found *c-erbB-2* a strong predictor for malignant behavior of molar pregnancies; Yazaki et al. proposed to use *c-erbB-2* expression as well as  $\beta$ hCG in therapeutic protocols (16, 27, 32, 33, 36).

Our data is noteworthy because of the more accurate method of immunohistochemistry we have used; unlike the most studies on this issue using avidin-biotin methods (16, 23, 32, 33, 37, 38), our study was performed on En-Vision <sup>Tm</sup> system. Avidin-biotin methods are widely used since their introduction in 1981 but because of the background staining due to tissues endogenous biotin and decreasing the expression of biotin in formalin fixation and paraffin blocking of tissues, the new method of polymer based methods have been established (39, 40). EnVision <sup>Tm</sup> system is a new polymer based technique which has higher sensitivity than routine avidin-biotin method without its limitations (41).

Although our study was performed only by one pathologist and was not a blind study, using a more accurate technique of immunostaining and counting immunostained cells separately on each cell population (cytotrophoblast and syncytiotrophoblast) might make our results more practical to be a base for future studies. Finally, further collaboration of pathologists and gynecologists would be suggested to establish comprehensive guidelines for early diagnosis of malignant progression of molar pregnancies.

Our data suggests that over expression of *p*53 and *c*-*erbB*-2 is associated with malignant progression of molar pregnancy. We encountered that high expression of *p*53 and *c*-*erbB*-2 in trophoblastic cells could predict gestational trophoblastic neoplasia in early stages. Supposed our data could be supported with more studies on this issue, it might be useful to evaluate the immuno-expression of *p*53 and *c*-*erbB*-2 genes on primary samples of pregnancy products of patients with molar pregnancy to estimate their risk of progression toward the malignant forms.

#### Acknowledgments

None declared.

# Footnotes

**Authors' Contribution:** Malihe Hasanzadeh designed the study, performed the operations and biopsy procedures and supervised the entire project. Norrie Sharifi performed the staining step and studied the samples and supervised the pathological aspect of discussion. Marjaneh Farazestanian contributed to data collection and entry; literature review and writing process. Saman Nazemian and Faezeh Madani sani contributed to data analysis; literature review and writing up process.

**Conflict of Interest:** All authors have declared no conflict of interests and the whole project has paid from Dr. Hasanzade grant from research budget of Mashhad University of Medical Sciences.

Financial Disclosure: None declared.

**Funding/Support:** This survey was financially supported by Mashhad University of Medical Sciences (MUMS) as Malihe Hasanzade research grant.

### References

- Berkowitz RS, Goldstein DP, Bernstein MR. Evolving concepts of molar pregnancy. J Reprod Med. 1991;36(1):40–4. [PubMed: 1848899].
- Roberts DJ, Mutter GL. Advances in the molecular biology of gestational trophoblastic disease. *J Reprod Med.* 1994;**39**(3):201–8. [PubMed: 8035375].
- Szulman AE, Surti U. The syndromes of hydatidiform mole. I. Cytogenetic and morphologic correlations. *Am J Obstet Gynecol.* 1978;**131**(6):665–71. [PubMed: 686053].
- Atrash HK, Hogue CJ, Grimes DA. Epidemiology of hydatidiform mole during early gestation. *Am J Obstet Gynecol.* 1986;**154**(4):906–9. [PubMed: 3963081].
- Chun D, Ma HK. Choriocarcinoma in Hong Kong. J R Coll Surg Edinb. 1974;19(2):69–81. [PubMed: 4824316].
- Ma HK, Wong LC. Gestational trophoblastic disease in Hong Kong. Semin Oncol. 1982;9(2):224-33. [PubMed: 6289466].
- Berkowitz RS, Goldstein DP. Current management of gestational trophoblastic diseases. *Gynecol Oncol.* 2009;**112**(3):654–62. doi: 10.1016/j.ygyno.2008.09.005. [PubMed: 18851873].
- Lurain JR. Gestational trophoblastic tumors. Semin Surg Oncol. 1990;6(6):347-53. [PubMed: 2175931].
- Soper JT. Gestational trophoblastic disease. *Obstet Gynecol.* 2006;**108**(1):176-87. doi: 10.1097/01.AOG.0000224697.31138.a1. [PubMed: 16816073].
- Benedet JL, Bender H, Jones H, Ngan HY, Pecorelli S. FIGO staging classifications and clinical practice guidelines in the management of gynecologic cancers. FIGO Committee on Gynecologic Oncology. *Int J Gynaecol Obstet*. 2000;**70**(2):209–62. [PubMed: 11041682].
- Classifications FS. Clinical Practice Guidelines of Gynecological Cancers by the FIGO Committee on Gynecologic Oncology. London: Elsevier; 2000.
- Cheung AN, Srivastava G, Chung LP, Ngan HY, Man TK, Liu YT, et al. Expression of the p53 gene in trophoblastic cells in hydatidiform moles and normal human placentas. *J Reprod Med.* 1994;**39**(3):223–7. [PubMed: 8035377].
- Hollstein M, Sidransky D, Vogelstein B, Harris CC. p53 mutations in human cancers. Science. 1991;253(5015):49–53. [PubMed: 1905840].
- Lee YS. p53 expression in gestational trophoblastic disease. Int J Gynecol Pathol. 1995;14(2):119–24. [PubMed: 8601523].
- Yamauchi H, Katayama K, Ueno M, He XJ, Mikami T, Uetsuka K, et al. Essential role of p53 in trophoblastic apoptosis induced in the developing rodent placenta by treatment with a DNA-damaging agent. *Apoptosis*. 2007;**12**(10):1743–54. doi: 10.1007/s10495-007-0099-z. [PubMed: 17594519].

- Yang X, Zhang Z, Jia C, Li J, Yin L, Jiang S. The relationship between expression of c-ras, c-erbB-2, nm23, and p53 gene products and development of trophoblastic tumor and their predictive significance for the malignant transformation of complete hydatidiform mole. *Gynecol Oncol.* 2002;85(3):438–44. [PubMed: 12051871].
- Fletcher CD. Diagnostic Histopathology of Tumors. 3 ed. 2. Philadelphia: Elsevier; 2007.
- Kumar GL, Rudbeck L, DAKO A. Education Guide: Immunohistochemical Staining Methods: Pathology. Carpinteria: Dako North America; 2009.
- Greenblatt MS, Bennett WP, Hollstein M, Harris CC. Mutations in the p53 tumor suppressor gene: clues to cancer etiology and molecular pathogenesis. *Cancer Res.* 1994;**54**(18):4855-78. [PubMed: 8069852].
- Efeyan A, Serrano M. p53: guardian of the genome and policeman of the oncogenes. *Cell Cycle*. 2007;6(9):1006–10. doi: 10.4161/cc.6.9.4211. [PubMed: 17457049].
- Petignat P, Laurini R, Goffin F, Bruchim I, Bischof P. Expression of matrix metalloproteinase-2 and mutant p53 is increased in hydatidiform mole as compared with normal placenta. *Int J Gynecol Cancer.* 2006;**16**(4):1679-84. doi: 10.1111/j.1525-1438.2006.00643.x. [PubMed: 16884384].
- 22. Qiao S, Nagasaka T, Harada T, Nakashima N. p53, Bax and Bcl-2 expression, and apoptosis in gestational trophoblast of complete hydatidiform mole. *Placenta*. 1998;**19**(5-6):361-9. [PubMed: 9699956].
- 23. Shi YF, Xie X, Zhao CL, Ye DF, Lu SM, Hor JJ, et al. Lack of mutation in tumour-suppressor gene p53 in gestational trophoblastic tumours. *Br J Cancer.* 1996;**73**(10):1216–9. [PubMed: 8630281].
- 24. Chen Y, Shen D, Gu Y, Zhong P, Xie J, Song Q. The diagnostic value of Ki-67, P53 and P63 in distinguishing partial Hydatidiform mole from hydropic abortion. *Wien Klin Wochenschr.* 2012;**124**(5-6):184–7. doi:10.1007/s00508-011-0119-4. [PubMed: 22218717].
- Chen CA, Chen YH, Chen TM, Ko TM, Wu CC, Lee CN, et al. Infrequent mutation in tumor suppressor gene p53 in gestational trophoblastic neoplasia. *Carcinogenesis*. 1994;15(10):2221-3. [PubMed: 7955057].
- Fulop V, Mok SC, Genest DR, Gati I, Doszpod J, Berkowitz RS. p53, p21, Rb and mdm2 oncoproteins. Expression in normal placenta, partial and complete mole, and choriocarcinoma. *J Reprod Med.* 1998;43(2):119–27. [PubMed: 9513873].
- Fulop V, Mok SC, Genest DR, Szigetvari I, Cseh I, Berkowitz RS. c-myc, c-erbB-2, c-fms and bcl-2 oncoproteins. Expression in normal placenta, partial and complete mole, and choriocarcinoma. *J Reprod Med.* 1998;43(2):101–10. [PubMed: 9513871].
- Kew TY, Bell JA, Pinder SE, Denley H, Srinivasan R, Gullick WJ, et al. c-erbB-4 protein expression in human breast cancer. *Br J Cancer*. 2000;82(6):1163-70. doi: 10.1054/bjoc.1999.1057. [PubMed: 10735500].
- Srinivasan R, Gillett CE, Barnes DM, Gullick WJ. Nuclear expression of the c-erbB-4/HER-4 growth factor receptor in invasive breast cancers. *Cancer Res.* 2000;**60**(6):1483–7. [PubMed: 10749108].
- 30. Bodey B, Bodey BJ, Groger AM, Luck JV, Siegel SE, Taylor CR, et al.

Clinical and prognostic significance of the expression of the c-erbB-2 and c-erbB-3 oncoproteins in primary and metastatic malignant melanomas and breast carcinomas. *Anticancer Res.* 1997;**17**(2B):1319–30. [PubMed: 9137492].

- Maurer CA, Friess H, Kretschmann B, Zimmermann A, Stauffer A, Baer HU, et al. Increased expression of erbB3 in colorectal cancer is associated with concomitant increase in the level of erbB2. *Hum Pathol.* 1998;**29**(8):771–7. [PubMed: 9712416].
- Jelincic D, Hudelist G, Singer CF, Bauer M, Horn LC, Bilek K, et al. Clinicopathologic profile of gestational trophoblastic disease. *Wien Klin Wochenschr.* 2003;115(1-2):29–35. [PubMed: 12658908].
- Yazaki-Sun S, Daher S, de Souza Ishigai MM, Alves MT, Mantovani TM, Mattar R. Correlation of c-erbB-2 oncogene and p53 tumor suppressor gene with malignant transformation of hydatidiform mole. *J Obstet Gynaecol Res.* 2006;**32**(3):265-72. doi: 10.1111/j.1447-0756.2006.00397.x. [PubMed: 16764615].
- Cameron B, Gown AM, Tamimi HK. Expression of c-erb B-2 oncogene product in persistent gestational trophoblastic disease. *Am J Obstet Gynecol.* 1994;**170**(6):1616–21. [PubMed: 7911272].
- Dehaghani AS, Rad NR, Fattahi MJ, Khadang B, Kashef MA, Sarraf Z, et al. Investigation of soluble HER2 and transforming growth factor Beta-1 serum levels in gestational trophoblastic disease. *Pathol Oncol Res.* 2009;**15**(1):37-40. doi: 10.1007/s12253-008-9115-z. [PubMed: 18975137].
- Menczer J, Schreiber L, Berger E, Golan A, Levy T. Assessment of Her-2/neu expression in hydatidiform moles for prediction of subsequent gestational trophoblastic neoplasia. *Gynecol Oncol.* 2007;**104**(3):675– 9. doi: 10.1016/j.ygyno.2006.10.012. [PubMed: 17126893].
- Halperin R, Peller S, Sandbank J, Bukovsky I, Schneider D. Expression of the p53 gene and apoptosis in gestational trophoblastic disease. *Placenta*. 2000;**21**(1):58–62. doi: 10.1053/plac.1999.0442. [PubMed: 10692252].
- Hussein MR. Analysis of p53, BCL-2 and epidermal growth factor receptor protein expression in the partial and complete hydatidiform moles. *Exp Mol Pathol.* 2009;87(1):63–9. doi: 10.1016/j.yexmp.2009.03.005. [PubMed: 19348791].
- Heras A, Roach CM, Key ME, editors. Enhanced polymer detection system for immunohistochemistry. Laboratory Investigation. 1995; Williams & Wilkins 351 West Camden St, Baltimore, Md 21201-2436; p. 165.
- Hsu SM, Raine L, Fanger H. Use of avidin-biotin-peroxidase complex (ABC) in immunoperoxidase techniques: a comparison between ABC and unlabeled antibody (PAP) procedures. *J Histochem Cytochem.* 1981;29(4):577-80. [PubMed: 6166661].
- Sabattini E, Bisgaard K, Ascani S, Poggi S, Piccioli M, Ceccarelli C, et al. The EnVision++ system: a new immunohistochemical method for diagnostics and research. Critical comparison with the APAAP, Chem-Mate, CSA, LABC, and SABC techniques. *J Clin Pathol.* 1998;**51**(7):506-11. [PubMed: 9797726].