



Antisense Non-Coding RNA in the *INK4* Locus (*ANRIL*) in Human Cancers

Mohammad Taheri¹ and Soudeh Ghafouri-Fard^{2,*}

¹Urogenital Stem Cell Research Center, Shahid Beheshti University of Medical Sciences, Tehran, Iran

²Department of Medical Genetics, Shahid Beheshti University of Medical Sciences, Tehran, Iran

*Corresponding author: Soudeh Ghafouri-Fard, Department of Medical Genetics, Shahid Beheshti University of Medical Sciences, Tehran, Iran. Tel/Fax: +98-2123872572, Email: s.ghafourifard@sbm.ac.ir

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Abstract

Context: *ANRIL* (antisense non-coding RNA in the *INK4* locus) is one of the oncogenic long non-coding RNAs (lncRNAs), which is located at the 9p21.3. It has a regulatory epigenetic function on its neighboring gene cluster *p15/CDKN2B-p16/CDKN2A-p14/ARF*. Such regulatory role has led to contribution of *ANRIL* in the pathogenesis of many human cancers.

Evidence Acquisition: We searched the PubMed, Google Scholar, EMBASE, and Web of Science databases till February 2018 with the key words "ANRIL" OR "antisense non-coding RNA in the *INK4* locus" AND "cancer" OR "tumor" OR "malignancy".

Results: *ANRIL* over-expression is documented in several cancers. The underlying mechanism for regulatory effect of *ANRIL* on the expression of its neighboring tumor suppressors has been clarified in some malignancies. Moreover, *ANRIL* over-expression has been associated with cisplatin and 5-fluorouracil (5-FU)-resistance in some cancers.

Conclusions: *ANRIL* is regarded as a potential biomarker in diverse cancer types and a therapeutic target in certain malignancies.

Keywords: lncRNA, *ANRIL*, Cancer

1. Context

Long non-coding RNAs (lncRNAs) are a group of transcripts with sizes greater than 200 nucleotides that instead of being translated into protein, themselves exert crucial functional roles (1). Almost any fundamental aspect of cell physiology is a subject of regulation by lncRNAs (2-5). Their participation in a wide range of cancers has also been evaluated (6-11). *ANRIL* (antisense non-coding RNA in the *INK4* locus) is one of the oncogenic lncRNAs, which is located at the 9p21.3 locus (12). This locus has been recognized as a common genetic susceptibility locus for coronary disease, intracranial aneurysm, type 2 diabetes, and several malignancies through genome-wide association studies (GWAS) (13). The 9p21.3 locus encompasses *p15/CDKN2B-p16/CDKN2A-p14/ARF* gene cluster as well. *ANRIL* has been shown to regulate the expression of this gene cluster through recruitment of polycomb repressive complexes PRC2 and PRC1 to this locus (14) (Figure 1). *ANRIL*-mediated recruitment of these complexes alters the epigenetic chromatin shape and, hence, suppresses the expression of this gene cluster in a cis-acting manner (15). PRC1 and PRC2 participate in chromatin remodeling, suppress transcription of several genes and interact with key signaling path-

ways. Their regulatory role on stem cell pluripotency provides further evidences for their participation in cancer pathogenesis. PRC2 and PRC1 suppressive regulatory function is exerted through two diverse histone modifications, trimethylation of lysine 27 on histone H3 (H3K27me3) and monoubiquitination of histone H2A (H2AK119ub) (16). *ANRIL*-mediated silencing of the *CDKN2B/CDKN2A/ARF* locus is exerted through *ANRIL* interaction with EZH2 (PRC2) and CBX7 (PRC1) (17). The observed up-regulation of *ANRIL* and CBX7 in prostate cancer tissues and the ability of *ANRIL* to bind with CBX7 provided further evidences for contribution of *ANRIL* in suppression of the *CDKN2A/B* locus (14). Moreover, EZH2 is a histone methyltransferase, which participates in silencing of *p15/CDKN2B-p16/CDKN2A-p14/ARF* cluster by tri-methylating H3K27 in the corresponding promoters (18). The *p15/CDKN2B-p16/CDKN2A-p14/ARF-ANRIL* genes contribute in the regulation of senescence and apoptosis. Alterations in tumor suppressor genes encoded by *p15/CDKN2B-p16/CDKN2A-p14/ARF* locus have been associated with a wide range of malignancies (19-21), implying the parallel role of *ANRIL* in the pathogenesis of these malignancies. Consequently, several studies have focused on the evaluation of *ANRIL* expression or genomic variations in human cancers.

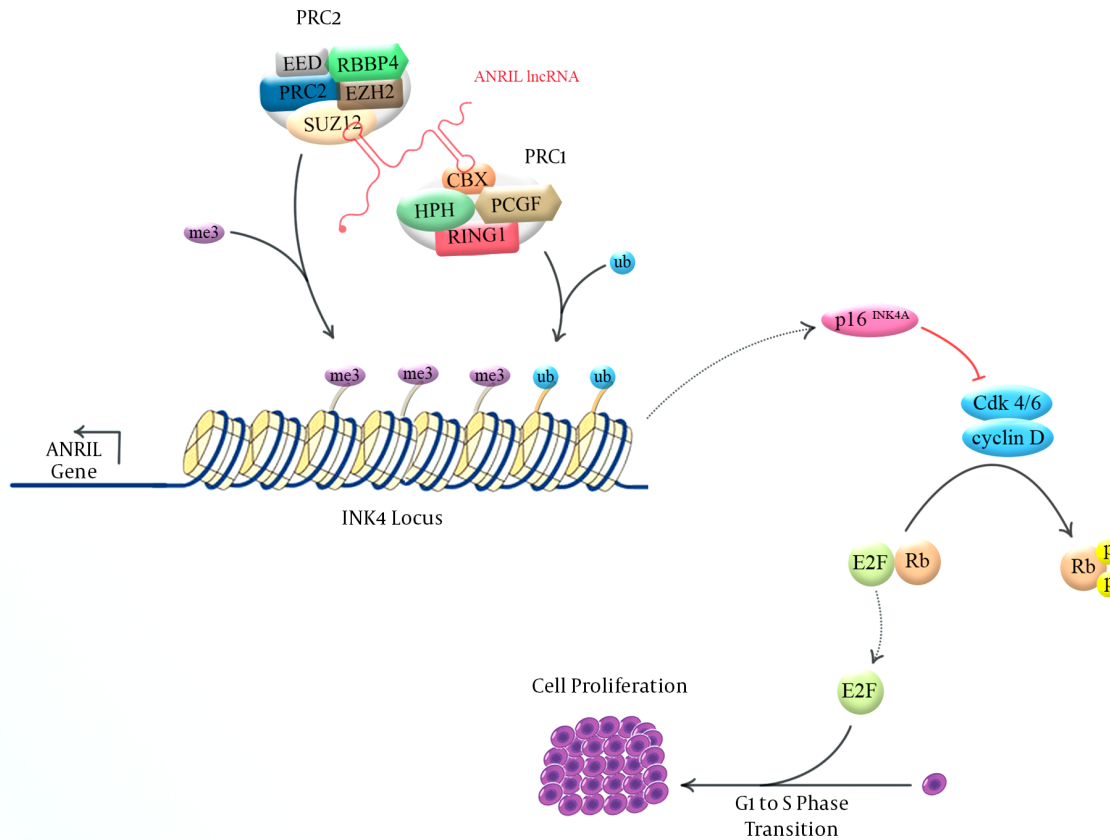


Figure 1. The 9p21.3 locus encompasses *p15/CDKN2B-p16/CDKN2A-p14/ARF* gene cluster as well. *ANRIL* regulates expression of this gene cluster through recruitment of polycomb repressive complexes PRC2 and PRC1 to this locus. *ANRIL* suppresses *p16/INK4A* thereby reverses its effects on growth suppression.

2. Evidence Acquisition

We searched the PubMed, Google Scholar, EMBASE, and Web of Science databases till February 2018 with the key words "*ANRIL*" OR "antisense non-coding RNA in the *INK4* locus" AND "cancer" OR "tumor" OR "malignancy". We only included original articles written in English, described the mechanism of *ANRIL* participation in malignancies and provided enough patients' data. We excluded other types of papers from the study.

3. Results

Several studies have pointed the significance of *ANRIL* in diverse human malignancies. Table 1 shows the summary of these studies.

3.1. *ANRIL* in Breast Cancer

In a large cohort of patients with breast cancer ($n = 456$), *ANRIL* over-expression has been detected in 19.7% of invasive breast carcinomas, particularly in the aggressive triple negative breast cancer (TNBC) subtype (16). Moreover, its expression level was correlated with estrogen receptor (ER) and progesterone receptor (PR) status but no patients' survival. A strong positive correlation has been detected between mRNA expression levels of the *ANRIL*, *ARF*, *p15*, and *p16* pointing to the presence of a complicated network between these genes (16). In spite of the proposed role for *ANRIL* in epithelial-mesenchymal transition (EMT), no significant associations were found between *ANRIL* and EMT-associated markers such as *TWIST1*, *ZEB1*, *ZEB2*, *SNAI1/Snail*, *SNAI2/Slug*, *VIM/Vimentin*, *CDH2/N-cadherin*, *CDH1/E-cadherin*, and *ZO-1* (16).

More recently, *ANRIL* has been shown to be over-expressed in TNBC tumor tissue and cell lines compared

Table 1. Expression, Genomic Variants, and Function of *ANRIL* in Human Cancers

Cancer Type	Expression	Role	Target Genes	Variant Associated with Cancer Risk	Variant Associated with Therapeutic Response	References
Lung cancer	Up	Associated with TNM stage, cisplatin resistance and poor prognosis	c-Myc, <i>KLF2</i> , P21	-	rs1333049 (associated with severe overall toxicity of cisplatin), rs10120688 and rs1333049 (associated with response to platinum)	(22-25)
Hepatocellular carcinoma	Up	Promotes cell proliferation	miR-122-5p, <i>KLF2</i>	-	-	(26, 27)
Prostate cancer	Up	Promotes cell proliferation and migration	<i>CBX7</i>	rs4977574, rs1333048, rs10757278	-	(14, 28, 29)
Breast cancer	Up	Promotes cell proliferation	<i>p15/CDKN2B-p16/CDKN2A-p14/ARF</i> gene cluster	An rs1333045, rs4977574, rs1333048 and rs10757278 haplotype, rs11515	-	(12, 53, 30)
Gastric cancer	Up	Promotes cell proliferation, Associated with cisplatin- and 5-fluorouracil (5-FU)-resistance	MDR1, MRP1, miR-449a, <i>p15^{Ink4B}</i> , <i>p16^{Ink4A}</i>	-	-	(31, 32)
Colorectal cancer	Up	Promotes cell proliferation, migration and invasion	-	-	-	(33)
Cervical cancer	Up	Promotes cell proliferation	<i>miR-186</i> , PI3K/Akt pathway, <i>p15</i> , <i>p16</i>	-	-	(34)
Ovarian cancer	Up	Promotes cell proliferation	<i>P15^{INK4}</i> , Bcl-2	-	-	(35)
Oral squamous cell carcinoma	Up	Promotes cell proliferation, inhibits metastasis, associated with cisplatin-resistance	-	-	-	
Esophageal squamous cell carcinoma (ESCC)	Up	Promotes cell proliferation	<i>p^{15INK4b}</i> , transforming growth factor β 1 (TGF β 1)	-	-	(36)
Nasopharyngeal carcinoma (NPC)	Up	Promotes cell proliferation, decreases apoptosis, increases radioresistance, reprograms cell glucose metabolism and induces side-population stem-like cancer cells	miR-125a, miR-let-	-	-	(37, 38)
Acute lymphoblastic leukemia (ALL)	-	-	-	-	rs564398 correlated with ALL phenotype	(39)
Multiple myeloma (MM)	-	Associated with melphalan-mediated apoptosis	<i>p14^{ARF}</i> , p53	-	rs2151280 associated with worse progression-free survival	(40)

to noncancerous tissue and non-TNBC cells. Such elevated expression was accompanied by poor prognosis. *In vivo* functional studies confirmed the role of *ANRIL* as molecular 'sponge' for miR-199a in TNBC (30).

The rs11515 SNP located in the 5' region of *ANRIL* has been shown to be associated with breast cancer risk in a sample of 200 patients and 200 healthy subjects from New Zealand. Notably, the CG genotype of this SNP has been

correlated with increased *ANRIL* and decreased *p16^{INK4a}* expression levels suggestive of the decreased expression of *p16^{INK4a}* as a putative mechanism for contribution of rs11515 CG genotype in cancer risk (41). Recently, we genotyped rs1333045, rs4977574, rs1333048, and rs10757278 SNPs in 122 patients with breast cancer and 200 normal age-matched subjects and found TCGA haplotype to be associated with breast cancer risk in Iranian population (12).

3.2. *ANRIL* in Prostate Cancer

The elevated expression of *ANRIL* as well as chromobox 7 (*CBX7*) within the PRC1, which interacts with *ANRIL*, has been demonstrated in prostate cancer tissues. In prostate tissues, *ANRIL* binding with *CBX7* has been shown to be crucial for suppression of the *INK4b/ARF/INK4a* locus (14). Moreover, the rs4977574, rs1333048, and rs10757278 within *ANRIL* have been shown to be associated with prostate cancer risk in Iranian population (29).

3.3. *ANRIL* in Hepatocellular Carcinoma (HCC)

The assessment of *ANRIL* expression in 77 HCC tissues and corresponding normal tissues have shown its up-regulation in HCC tissues and correlation of its transcript levels with tumor size and Barcelona Clinic Liver Cancer (BCLC) stage. Its silencing could suppress cell proliferation and invasion and trigger cell apoptosis both *in vitro* and *in vivo*. Moreover, functional studies showed the role of *ANRIL* in suppression of Kruppel-like factor 2 (*KLF2*) transcription in HCC cells by recruitment of PRC2 to the *KLF2* promoter region (42). More recently, *in vitro* studies have shown a negative correlation between *ANRIL* and miR-122-5p expressions in HCC tissues, which was confirmed through the detection of directly interaction between *ANRIL* and miR-122-5p. More importantly, *in vivo* studies showed the effect of *ANRIL* knockdown in suppression of tumor growth (26).

3.4. *ANRIL* in Lung Cancer

ANRIL is over-expressed in H1299 cancer cells compared to normal fibroblasts. Its silencing increased *p15* expression with no effect on *p16* or *ARF* expression, led to cell-cycle arrest at the G2/M phase and suppressed proliferation of these cells (43). Besides, its knock-down in A549, SPC-A1, NCI-H1650 lung cancer cells suppressed cell proliferation, migration, and invasion *in vitro* (44). In addition, its silencing in NSCLC cells could weaken cell proliferation and trigger cell apoptosis both *in vitro* and *in vivo*. Its effect on decreasing *p15* expression in PC9 cells was exerted through silencing of *KLF2* and P21 transcription (23). The expression of lncRNA *ANRIL* has been up-regulated by heparin-binding growth factor secreted from cancer-associated fibroblasts (CAF) in lung tumors. *ANRIL* is also involved in

proliferation, apoptosis, and cisplatin resistance of the tumor cells via modulation of the drug transporters MRP1 and ABCC2 (45). The physical interaction between c-Myc and *ANRIL* has also been validated *in vitro*. The effect of *ANRIL* silencing in suppression of NSCLC cell proliferation potentiate this lncRNA as a putative therapeutic target for patients with NSCLC (24). However, a single study in lung cancer H460 cells indicated *ANRIL* as an lncRNA responsible in anti-tumorigenesis caused by PLD inhibition and suggested the addition of *ANRIL* into PLD inhibition as a novel therapeutic modality for controlling lung cancer (46). The discrepancy between these two studies might be due to the difference in cultured cells or other biological variables. The results of clinical studies are in line with oncogenic role of this lncRNA. For instance, the expression level of *ANRIL* has been shown to be elevated in NSCLC tissues and lung cancer cells than in adjacent non-tumor tissues and normal human bronchial epithelial cells. Such over-expression in clinical tissues was associated with higher TNM stage, advanced lymph node metastasis as well as poor overall survival (23, 44).

Moreover, in a population of 467 Chinese patients with lung cancer treated with platinum-based chemotherapy, *ANRIL* rs1333049 was a predictor of both severe overall toxicity and severe gastrointestinal toxicity (22). Besides, *ANRIL* rs10120688 and rs1333049 were associated with response to platinum-based chemotherapy in patients with adenocarcinoma and small-cell lung cancer, respectively, in 498 Chinese patients with lung cancer (25).

3.5. *ANRIL* in Cervical Cancer

ANRIL over-expression has been detected in HeLa cancer cells compared to normal fibroblasts. Its silencing increased *p15* expression with no effect on *p16* or *ARF* expression, led to cell-cycle arrest at the G2/M phase and suppressed proliferation of these cells (43). Moreover, *ANRIL* expression has been elevated in cervical cancer tumors compared with non-cancerous tissues (34, 47). The role of this lncRNA, as a sponge for *miR-186*, has been confirmed by both bioinformatics tools and luciferase assay. Consequently, the *ANRIL/miR-186* axis has been suggested as a culprit in the pathogenesis of cervical cancer (34). Moreover, the higher expression of this lncRNA in clinical samples was associated with advanced International Federation of Gynecologists and Obstetricians stage (FIGO) stage, lymph node metastasis, and poor overall survival indicating a role for *ANRIL* expression as an independent prognostic factor in cervical cancer. *ANRIL* silencing has suppressed cell proliferation, migration, and invasion of cervical cancer via the inhibition of PI3K/Akt pathway. Consequently, this lncRNA has been suggested as a putative prognostic biomarker and therapeutic target in cervical cancer (47).

3.6. *ANRIL* in Ovarian Cancer

The expression of lncRNA *ANRIL* is increased by heparin-binding growth factor secreted from cancer-associated fibroblasts (CAF) in ovarian cancer cells (45). *ANRIL* expression was up-regulated in epithelial ovarian cancer (EOC) tissues compared with normal controls in association with advanced FIGO stage and high histological grade. Such over-expression has been demonstrated to be an independent prognostic factor for overall survival in EOC. The role of *ANRIL* in the stimulation of EOC cell proliferation has been confirmed both by *in vitro* and *in vivo*. Such role is suggested to be exerted through down-regulation of *P15INK4B* and up-regulation of *Bcl-2* (35).

3.7. *ANRIL* in Oral Squamous Cell Carcinoma (OSCC)

The expression of *ANRIL* is increased by heparin-binding growth factor secreted from cancer-associated fibroblasts (CAF) in OSCC. Moreover, its over-expression has been correlated with both high TNM stage and lymph node metastasis in OSCC. On the other hand, its silencing suppressed proliferation, induced apoptosis, and increased cisplatin cytotoxicity of the tumor cells via diminishing the drug transporters *MRP1* and *ABCC2* (45).

3.8. *ANRIL* in Esophageal Cancer

The elevated expression of *ANRIL* has been detected in human esophageal squamous cell carcinoma (ESCC) tissues compared with the corresponding adjacent non-tumor tissues. *In vitro* studies revealed the role of *ANRIL* silencing in the enhancement of the expression of *p15 (INK4b)* and transforming growth factor $\beta 1$ (*TGF β 1*) as well as the suppression of cellular proliferation (36). However, the assessment of association between *ANRIL* rs2151280 in 380 ESCC and 380 cancer-free controls from Chinese Han populations cases revealed no association between this polymorphism and ESCC (48).

3.9. *ANRIL* in Gastric Cancer

The elevated expression of *ANRIL* has been demonstrated in gastric cancer tissues of cisplatin-resistant and 5-fluorouracil (5-FU)-resistant patients as well as cisplatin-resistant cells (BGC823/DDP) and 5-FU-resistant cells (BGC823/5-FU). *ANRIL* silencing in these cells decreased their survival and invasion capability, increased their apoptosis, and down-regulated the expression of multi-drug resistance (MDR) related genes (31).

Ten-eleven translocation 2 (TET2), which converts 5-methylcytosine to 5-hydroxymethylcytosine in DNA, has been shown to bind to the promoter the region of *ANRIL* and modulate the expression of *ANRIL*, *INK4a*, *INK4b*, and *ARF* in gastric cancer cells. Besides, *ANRIL* silencing inhibits

the effects of TET2 on gastric cancer cell proliferation and colony formation (49).

Besides, in a cohort of 120 patients with gastric cancer, *ANRIL* over-expression has been correlated with a higher TNM stage and tumor size being an independent predictor for overall survival. *ANRIL* silencing decreased cell proliferation both *in vitro* and *in vivo*. E2F1 has been demonstrated to be responsible for the induction of *ANRIL* expression and *ANRIL*-mediated cell proliferation through the epigenetic suppression of miR-99a/miR-449a in gastric cancer cells (32).

3.10. *ANRIL* in Colorectal Cancer (CRC)

The expression analysis of *ANRIL* in 97 paired CRC and adjacent non neoplastic tissue samples have shown its over-expression in CRC samples compared with the adjacent non neoplastic tissues in association with decreased survival rate. Moreover, its silencing in HT29 and RKO human CRC cell lines reduced proliferation while suppressed migration and invasion (33).

3.11. *ANRIL* in Nasopharyngeal Carcinoma (NPC)

ANRIL over-expression has been demonstrated in advanced-stage cancer in a cohort of 88 NPC patients indicating a role for it an independent predictor of overall survival and disease-free survival. *In vitro* studies have shown *ANRIL* effect on the induction of side population cells (SP cells) in NPC. Moreover, *ANRIL* has a role in the enhancement of glucose uptake for glycolysis, through the regulation of the mTOR signaling pathway (50). The over-expression of *ANRIL* has been demonstrated in NPC tissues and cell lines. *ANRIL* silencing in CNE2 and HONE1 cells inhibited proliferation, triggered apoptosis, and improved radiosensitivity of cancer cells. *In vitro* studies suggested miR-125a as a target of *ANRIL* in these cells (37). Another study has shown the elevated expression of *ANRIL* while down-regulation of *let-7a* in NPC tissues and cells. The effect of *ANRIL* in down-regulation of miR-*let-7a* expression has been confirmed by luciferase assay. The role of *ANRIL* silencing in suppression of tumorigenicity and improvement of cisplatin-induced cytotoxicity has offered a hopeful treatment modality for NPC patients (38).

3.12. *ANRIL* in Hematological Malignancies

The rs564398 within *ANRIL* has been shown to be associated with acute lymphoblastic leukemia (ALL) phenotype in an association study enrolling 149 leukemia patients, including Philadelphia positive (Ph (+)) acute lymphoblastic leukemia (ALL) and acute myeloid leukemia (AML) samples. This association might show the ability of some *ANRIL* polymorphisms to alter *ANRIL* expression and, subsequently, modify *CDKN2A/B* expression signatures, which

would contribute in atypical cell proliferation and ALL susceptibility (39). Another association study in Iranian patients with acute myeloid leukemia (AML) showed no association between rs1333048, rs4977574, rs1333045, and rs10757278 polymorphisms and AML (51). *ANRIL* genotyping in 108 multiple myeloma patients treated with high-dose melphalan and subsequent hematopoietic stem cell transplantation (HSCT) have shown the association between the rs2151280 (C/T) genotypes and worse progression-free survival. The TT variant was also associated with elevated *ANRIL* expression and down-regulation of *p15*, *p14ARF*, and *p16* compared to the TC/CC variants. Consequently, *ANRIL* might contribute in melphalan-mediated apoptosis through modulation of *p14ARF* and subsequent *p53* (40).

3.13. *ANRIL* in Osteosarcoma (OS)

The transcript levels of *ANRIL* have been elevated in OS tissues compared with the adjacent normal tissues in association with tumor size and patients' outcome. siRNA mediated *ANRIL* silencing has resulted in the induction of cell apoptosis and suppression of cell proliferation and invasion both *in vitro* and *in vivo*. Nude mice injected with si*ANRIL*-transfected MNNG/HOS cells have developed smaller tumors as compared with control group (52), which might highlight the role of *ANRIL* silencing in cancer control.

4. Conclusions

ANRIL role in epigenetic regulation of expression of its adjacent tumor suppressor genes *CDKN2B*, *CDKN2A*, and *ARF* has suggested its contribution in carcinogenesis. This hypothesis has been supported by the observed over-expression of this lncRNA in a wide range of cancers. However, the associations between *ANRIL* and *CDKN2A/CDKNB/ARF* expression seem to be context-dependent. For instance, in the context of prostate cancer *CBX7* and *ANRIL* over-expressions have been correlated with *CDKN2A/CDKNB/ARF* down-regulation (14). On the contrary, in breast tumors, *CBX7* has been shown to be down-regulated, while both *ANRIL* and *CDKN2A/CDKNB/ARF* have been over-expressed (16). Such observations have led to the supposition of both tumor suppressive and oncogenic roles for *CBX7* based on type of the malignancy, microenvironment, and existence of interacting proteins (16).

In addition, certain SNPs within this gene have been shown to confer cancer risk in distinct populations. However, tissue- or disease-specific functions have been suggested for at least some SNPs (41), which might explain lack of associations with some types of cancer despite significant association with other types of malignancies.

The over-expression of *ANRIL* in cancer cells promotes cell cycle progression through the alteration of signaling

pathways such as Akt pathways or oncogenes such as Myc. In addition, *ANRIL* has a role in the inhibition of apoptosis and enhancement of cell migration and invasion possibly through the modulation of EMT pathways. Consequently, *ANRIL* has been shown to influence various aspects of carcinogenesis including the cell cycle, apoptosis, and metastasis. The results of *in vitro* and *in vivo* studies support the effectiveness of *ANRIL* silencing in the inhibition of carcinogenesis-related pathways. However, as any signaling pathway is itself affected by several lncRNAs and miRNAs, *ANRIL* silencing should be considered only as a single part of a comprehensive treatment modality for the effective eradication of cancer.

Clinical and *in vitro* studies have shown an intricate interactive network between *ANRIL*, numerous miRNAs, PRC2/PRC1 subunits, and *p15/CDKN2B-p16/CDKN2A-p14/ARF* gene cluster (16). Uncovering such interactions would facilitate the design of appropriate targeted therapies for different kinds of human malignancies. The crucial role of *ANRIL* in conferring MDR in cancer cell as being discovered in gastric cancer (31) suggests the combination of *ANRIL* targeting therapies with conventional chemotherapeutic regimens as a promising option for cancer treatment.

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Footnotes

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References

1. Soudyab M, Iranpour M, Ghafouri-Fard S. The role of long non-coding rnas in breast cancer. *Arch Iran Med*. 2016;**19**(7):508-17. [PubMed: 27362246].
2. Dianatpour A, Ghafouri-Fard S. The role of long non coding RNAs in the repair of DNA double strand breaks. *Int J Mol Cell Med*. 2017;**6**(1):1-12. [PubMed: 28868264]. [PubMed Central: PMC5568187].
3. Eftekharian MM, Ghafouri-Fard S, Soudyab M, Omrani MD, Rahimi M, Sayad A, et al. Expression analysis of long non-coding RNAs in the blood of multiple sclerosis patients. *J Mol Neurosci*. 2017;**63**(3-4):333-41. doi: 10.1007/s12031-017-0982-1. [PubMed: 28967047].
4. Dianatpour A, Ghafouri-Fard S. Long non coding RNA expression intersecting cancer and spermatogenesis: A systematic review. *Asian Pac J Cancer Prev*. 2017;**18**(10):2601-10. doi: 10.22034/APJCP.2017.18.10.2601. [PubMed: 29072050]. [PubMed Central: PMC5747377].
5. Faramarzi S, Dianatpour A, Ghafouri-Fard S. Discovering the role of long non-coding RNAs in regulation of steroid receptors signaling in cancer. *J Biol Today World*. 2017;**6**(12). doi: 10.15412/jbtw.01061202.

6. Taheri M, Omrani MD, Ghafouri-Fard S. Long non-coding RNA expression in bladder cancer. *Biophys Rev*. 2018;**10**(4):1205-13. doi: [10.1007/s12551-017-0379-y](https://doi.org/10.1007/s12551-017-0379-y). [PubMed: [29222807](https://pubmed.ncbi.nlm.nih.gov/29222807/)]. [PubMed Central: [PMC6082308](https://pubmed.ncbi.nlm.nih.gov/PMC6082308/)].
7. Nikpayam E, Soudyab M, Tasharrofi B, Sarrafzadeh S, Iranpour M, Geranpayeh L, et al. Expression analysis of long non-coding ATB and its putative target in breast cancer. *Breast Dis*. 2017;**37**(1):11-20. doi: [10.3233/BD-160264](https://doi.org/10.3233/BD-160264). [PubMed: [28598827](https://pubmed.ncbi.nlm.nih.gov/28598827/)].
8. Taheri M, Habibi M, Noroozi R, Rakhshan A, Sarrafzadeh S, Sayad A, et al. HOTAIR genetic variants are associated with prostate cancer and benign prostate hyperplasia in an Iranian population. *Gene*. 2017;**613**:20-4. doi: [10.1016/j.gene.2017.02.031](https://doi.org/10.1016/j.gene.2017.02.031). [PubMed: [28259691](https://pubmed.ncbi.nlm.nih.gov/28259691/)].
9. Nikpayam E, Tasharrofi B, Sarrafzadeh S, Ghafouri-Fard S. The role of long non-coding RNAs in ovarian cancer. *Iran Biomed J*. 2017;**21**(1):3-15. doi: [10.6091/21.1.24](https://doi.org/10.6091/21.1.24). [PubMed: [27664137](https://pubmed.ncbi.nlm.nih.gov/27664137/)]. [PubMed Central: [PMC5142151](https://pubmed.ncbi.nlm.nih.gov/PMC5142151/)].
10. Iranpour M, Soudyab M, Geranpayeh L, Mirfakhraie R, Azargashb E, Movafagh A, et al. Expression analysis of four long noncoding RNAs in breast cancer. *Tumour Biol*. 2016;**37**(3):2933-40. doi: [10.1007/s13277-015-4135-2](https://doi.org/10.1007/s13277-015-4135-2). [PubMed: [26409453](https://pubmed.ncbi.nlm.nih.gov/26409453/)].
11. Taheri M, Omrani M, Ghafouri-Fard S. Long non-coding RNAs expression in renal cell carcinoma. *J Biol Today World*. 2017;**6**(12). doi: [10.15412/j.btw.01061201](https://doi.org/10.15412/j.btw.01061201).
12. Khorshidi HR, Taheri M, Noroozi R, Sarrafzadeh S, Sayad A, Ghafouri-Fard S. ANRIL genetic variants in Iranian breast cancer patients. *Cell J*. 2017;**19**(Suppl 1):72-8. doi: [10.22074/cellj.2017.4496](https://doi.org/10.22074/cellj.2017.4496). [PubMed: [28580310](https://pubmed.ncbi.nlm.nih.gov/28580310/)]. [PubMed Central: [PMC5448323](https://pubmed.ncbi.nlm.nih.gov/PMC5448323/)].
13. Pasmant E, Sabbagh A, Vidaud M, Bieche I. ANRIL, a long, noncoding RNA, is an unexpected major hotspot in GWAS. *FASEB J*. 2011;**25**(2):444-8. doi: [10.1096/fj.10-172452](https://doi.org/10.1096/fj.10-172452). [PubMed: [20956613](https://pubmed.ncbi.nlm.nih.gov/20956613/)].
14. Yap KL, Li S, Munoz-Cabello AM, Raguz S, Zeng L, Mujtaba S, et al. Molecular interplay of the noncoding RNA ANRIL and methylated histone H3 lysine 27 by polycomb CBX7 in transcriptional silencing of INK4a. *Mol Cell*. 2010;**38**(5):662-74. doi: [10.1016/j.molcel.2010.03.021](https://doi.org/10.1016/j.molcel.2010.03.021). [PubMed: [20541999](https://pubmed.ncbi.nlm.nih.gov/20541999/)]. [PubMed Central: [PMC2886305](https://pubmed.ncbi.nlm.nih.gov/PMC2886305/)].
15. Kotake Y, Nakagawa T, Kitagawa K, Suzuki S, Liu N, Kitagawa M, et al. Long non-coding RNA ANRIL is required for the PRC2 recruitment to and silencing of p15(INK4B) tumor suppressor gene. *Oncogene*. 2011;**30**(16):1956-62. doi: [10.1038/onc.2010.568](https://doi.org/10.1038/onc.2010.568). [PubMed: [21151178](https://pubmed.ncbi.nlm.nih.gov/21151178/)]. [PubMed Central: [PMC3230933](https://pubmed.ncbi.nlm.nih.gov/PMC3230933/)].
16. Meseure D, Vacher S, Alsibai KD, Nicolas A, Chemlali W, Caly M, et al. Expression of ANRIL-polycomb complexes-CDKN2A/B/ARF genes in breast tumors: Identification of a two-gene (EZH2/CBX7) signature with independent prognostic value. *Mol Cancer Res*. 2016;**14**(7):623-33. doi: [10.1158/1541-7786.MCR-15-0418](https://doi.org/10.1158/1541-7786.MCR-15-0418). [PubMed: [27102007](https://pubmed.ncbi.nlm.nih.gov/27102007/)].
17. Popov N, Gil J. Epigenetic regulation of the INK4b-ARF-INK4a locus: In sickness and in health. *Epigenetics*. 2010;**5**(8):685-90. [PubMed: [20716961](https://pubmed.ncbi.nlm.nih.gov/20716961/)]. [PubMed Central: [PMC3052884](https://pubmed.ncbi.nlm.nih.gov/PMC3052884/)].
18. Zhao R, Choi BY, Lee MH, Bode AM, Dong Z. Implications of genetic and epigenetic alterations of CDKN2A (p16(INK4a)) in cancer. *EBioMedicine*. 2016;**8**:30-9. doi: [10.1016/j.ebiom.2016.04.017](https://doi.org/10.1016/j.ebiom.2016.04.017). [PubMed: [27428416](https://pubmed.ncbi.nlm.nih.gov/27428416/)]. [PubMed Central: [PMC4919535](https://pubmed.ncbi.nlm.nih.gov/PMC4919535/)].
19. Bostrom J, Meyer-Puttlitz B, Wolter M, Blaschke B, Weber RG, Lichter P, et al. Alterations of the tumor suppressor genes CDKN2A (p16(INK4a)), p14(ARF), CDKN2B (p15(INK4b)), and CDKN2C (p18(INK4c)) in atypical and anaplastic meningiomas. *Am J Pathol*. 2001;**159**(2):661-9. doi: [10.1016/S0002-9440\(10\)61737-3](https://doi.org/10.1016/S0002-9440(10)61737-3). [PubMed: [11485924](https://pubmed.ncbi.nlm.nih.gov/11485924/)]. [PubMed Central: [PMC1850553](https://pubmed.ncbi.nlm.nih.gov/PMC1850553/)].
20. Simon M, Koster G, Menon AG, Schramm J. Functional evidence for a role of combined CDKN2A (p16-p14(ARF))/CDKN2B (p15) gene inactivation in malignant gliomas. *Acta Neuropathol*. 1999;**98**(5):444-52. [PubMed: [10541865](https://pubmed.ncbi.nlm.nih.gov/10541865/)].
21. Rousseau E, Ruchoux MM, Scaravilli F, Chapon F, Vinchon M, De Smet C, et al. CDKN2A, CDKN2B and p14ARF are frequently and differentially methylated in ependymal tumours. *Neuropathol Appl Neurobiol*. 2003;**29**(6):574-83. [PubMed: [14631614](https://pubmed.ncbi.nlm.nih.gov/14631614/)].
22. Gong WJ, Peng JB, Yin JY, Li XP, Zheng W, Xiao L, et al. Association between well-characterized lung cancer lncRNA polymorphisms and platinum-based chemotherapy toxicity in Chinese patients with lung cancer. *Acta Pharmacol Sin*. 2017;**38**(4):581-90. doi: [10.1038/aps.2016.164](https://doi.org/10.1038/aps.2016.164). [PubMed: [28260796](https://pubmed.ncbi.nlm.nih.gov/28260796/)]. [PubMed Central: [PMC5386317](https://pubmed.ncbi.nlm.nih.gov/PMC5386317/)].
23. Nie FQ, Sun M, Yang JS, Xie M, Xu TP, Xia R, et al. Long noncoding RNA ANRIL promotes non-small cell lung cancer cell proliferation and inhibits apoptosis by silencing KLF2 and P21 expression. *Mol Cancer Ther*. 2015;**14**(1):268-77. doi: [10.1158/1535-7163.MCT-14-0492](https://doi.org/10.1158/1535-7163.MCT-14-0492). [PubMed: [25504755](https://pubmed.ncbi.nlm.nih.gov/25504755/)].
24. Lu Y, Zhou X, Xu L, Rong C, Shen C, Bian W. Long noncoding RNA ANRIL could be transactivated by c-Myc and promote tumor progression of non-small-cell lung cancer. *Onco Targets Ther*. 2016;**9**:3077-84. doi: [10.2147/OTT.S102658](https://doi.org/10.2147/OTT.S102658). [PubMed: [27307748](https://pubmed.ncbi.nlm.nih.gov/27307748/)]. [PubMed Central: [PMC4888724](https://pubmed.ncbi.nlm.nih.gov/PMC4888724/)].
25. Gong WJ, Yin JY, Li XP, Fang C, Xiao D, Zhang W, et al. Association of well-characterized lung cancer lncRNA polymorphisms with lung cancer susceptibility and platinum-based chemotherapy response. *Tumour Biol*. 2016;**37**(6):8349-58. doi: [10.1007/s13277-015-4497-5](https://doi.org/10.1007/s13277-015-4497-5). [PubMed: [26729200](https://pubmed.ncbi.nlm.nih.gov/26729200/)].
26. Ma J, Li T, Han X, Yuan H. Knockdown of lncRNA ANRIL suppresses cell proliferation, metastasis, and invasion via regulating miR-122-5p expression in hepatocellular carcinoma. *J Cancer Res Clin Oncol*. 2018;**144**(2):205-14. doi: [10.1007/s00432-017-2543-y](https://doi.org/10.1007/s00432-017-2543-y). [PubMed: [29127494](https://pubmed.ncbi.nlm.nih.gov/29127494/)].
27. Huang MD, Chen WM, Qi FZ, Xia R, Sun M, Xu TP, et al. Erratum to: Long non-coding RNA ANRIL is upregulated in hepatocellular carcinoma and regulates cell proliferation by epigenetic silencing of KLF2. *J Hematol Oncol*. 2017;**10**(1):143. doi: [10.1186/s13045-017-0513-0](https://doi.org/10.1186/s13045-017-0513-0). [PubMed: [28950881](https://pubmed.ncbi.nlm.nih.gov/28950881/)]. [PubMed Central: [PMC5530579](https://pubmed.ncbi.nlm.nih.gov/PMC5530579/)].
28. Zhao B, Lu YL, Yang Y, Hu LB, Bai Y, Li RQ, et al. Overexpression of lncRNA ANRIL promoted the proliferation and migration of prostate cancer cells via regulating let-7a/TGF-beta1/Smad signaling pathway. *Cancer Biomark*. 2018;**21**(3):613-20. doi: [10.3233/CBM-170683](https://doi.org/10.3233/CBM-170683). [PubMed: [29278879](https://pubmed.ncbi.nlm.nih.gov/29278879/)]. [PubMed Central: [PMC5859458](https://pubmed.ncbi.nlm.nih.gov/PMC5859458/)].
29. Taheri M, Poursmaeilif F, Omrani MD, Habibi M, Sarrafzadeh S, Noroozi R, et al. Association of ANRIL gene polymorphisms with prostate cancer and benign prostatic hyperplasia in an Iranian population. *Biomark Med*. 2017;**11**(5):413-22. doi: [10.2217/bmm-2016-0378](https://doi.org/10.2217/bmm-2016-0378). [PubMed: [28621612](https://pubmed.ncbi.nlm.nih.gov/28621612/)].
30. Xu ST, Xu JH, Zheng ZR, Zhao QQ, Zeng XS, Cheng SX, et al. Long non-coding RNA ANRIL promotes carcinogenesis via sponging miR-199a in triple-negative breast cancer. *Biomed Pharmacother*. 2017;**96**:14-21. doi: [10.1016/j.biopha.2017.09.107](https://doi.org/10.1016/j.biopha.2017.09.107). [PubMed: [28961506](https://pubmed.ncbi.nlm.nih.gov/28961506/)].
31. Lan WG, Xu DH, Xu C, Ding CL, Ning FL, Zhou YL, et al. Silencing of long non-coding RNA ANRIL inhibits the development of multidrug resistance in gastric cancer cells. *Oncol Rep*. 2016;**36**(1):263-70. doi: [10.3892/or.2016.4771](https://doi.org/10.3892/or.2016.4771). [PubMed: [27121324](https://pubmed.ncbi.nlm.nih.gov/27121324/)].
32. Zhang EB, Kong R, Yin DD, You LH, Sun M, Han L, et al. Long noncoding RNA ANRIL indicates a poor prognosis of gastric cancer and promotes tumor growth by epigenetically silencing of miR-99a/miR-449a. *Oncotarget*. 2014;**5**(8):2276-92. doi: [10.18632/oncotarget.1902](https://doi.org/10.18632/oncotarget.1902). [PubMed: [24810364](https://pubmed.ncbi.nlm.nih.gov/24810364/)]. [PubMed Central: [PMC4039162](https://pubmed.ncbi.nlm.nih.gov/PMC4039162/)].
33. Sun Y, Zheng ZP, Li H, Zhang HQ, Ma FQ. ANRIL is associated with the survival rate of patients with colorectal cancer, and affects cell migration and invasion in vitro. *Mol Med Rep*. 2016;**14**(2):1714-20. doi: [10.3892/mmr.2016.5409](https://doi.org/10.3892/mmr.2016.5409). [PubMed: [27314206](https://pubmed.ncbi.nlm.nih.gov/27314206/)].
34. Zhang JJ, Wang DD, Du CX, Wang Y. Long noncoding RNA ANRIL promotes cervical cancer development by acting as a sponge of miR-186. *Oncol Res*. 2017. doi: [10.3727/096504017X14953948675449](https://doi.org/10.3727/096504017X14953948675449). [PubMed: [28550682](https://pubmed.ncbi.nlm.nih.gov/28550682/)].
35. Qiu JJ, Wang Y, Liu YL, Zhang Y, Ding JX, Hua KQ. The long non-coding RNA ANRIL promotes proliferation and cell cycle progression and inhibits apoptosis and senescence in epithelial ovarian cancer. *Oncotarget*. 2016;**7**(22):32478-92. doi: [10.18632/oncotarget.8744](https://doi.org/10.18632/oncotarget.8744). [PubMed: [27095571](https://pubmed.ncbi.nlm.nih.gov/27095571/)]. [PubMed Central: [PMC5078027](https://pubmed.ncbi.nlm.nih.gov/PMC5078027/)].

36. Chen D, Zhang Z, Mao C, Zhou Y, Yu L, Yin Y, et al. ANRIL inhibits p15(INK4b) through the TGFbeta1 signaling pathway in human esophageal squamous cell carcinoma. *Cell Immunol.* 2014;**289**(1-2):91-6. doi: [10.1016/j.cellimm.2014.03.015](https://doi.org/10.1016/j.cellimm.2014.03.015). [PubMed: [24747824](https://pubmed.ncbi.nlm.nih.gov/24747824/)].
37. Hu X, Jiang H, Jiang X. Downregulation of lncRNA ANRIL inhibits proliferation, induces apoptosis, and enhances radiosensitivity in nasopharyngeal carcinoma cells through regulating miR-125a. *Cancer Biol Ther.* 2017;**18**(5):331-8. doi: [10.1080/15384047.2017.1310348](https://doi.org/10.1080/15384047.2017.1310348). [PubMed: [28402230](https://pubmed.ncbi.nlm.nih.gov/28402230/)]. [PubMed Central: [PMC5499759](https://pubmed.ncbi.nlm.nih.gov/PMC5499759/)].
38. Wang Y, Cheng N, Luo J. Downregulation of lncRNA ANRIL represses tumorigenicity and enhances cisplatin-induced cytotoxicity via regulating microRNA let-7a in nasopharyngeal carcinoma. *J Biochem Mol Toxicol.* 2017;**31**(7). doi: [10.1002/jbt.21904](https://doi.org/10.1002/jbt.21904). [PubMed: [28117929](https://pubmed.ncbi.nlm.nih.gov/28117929/)].
39. Iacobucci I, Sazzini M, Garagnani P, Ferrari A, Boattini A, Lonetti A, et al. A polymorphism in the chromosome 9p21 ANRIL locus is associated to Philadelphia positive acute lymphoblastic leukemia. *Leuk Res.* 2011;**35**(8):1052-9. doi: [10.1016/j.leukres.2011.02.020](https://doi.org/10.1016/j.leukres.2011.02.020). [PubMed: [21414664](https://pubmed.ncbi.nlm.nih.gov/21414664/)].
40. Poi M, Li J, Sborov DW, VanGundy Z, Cho YK, Lamprecht M, et al. Polymorphism in ANRIL is associated with relapse in patients with multiple myeloma after autologous stem cell transplant. *Mol Carcinog.* 2017;**56**(7):1722-32. doi: [10.1002/mc.22626](https://doi.org/10.1002/mc.22626). [PubMed: [28150872](https://pubmed.ncbi.nlm.nih.gov/28150872/)].
41. Royds JA, Pilbrow AP, Ahn A, Morrin HR, Frampton C, Russell IA, et al. The rs11515 polymorphism is more frequent and associated with aggressive breast tumors with increased ANRIL and decreased p16 (INK4a) expression. *Front Oncol.* 2015;**5**:306. doi: [10.3389/fonc.2015.00306](https://doi.org/10.3389/fonc.2015.00306). [PubMed: [26835415](https://pubmed.ncbi.nlm.nih.gov/26835415/)]. [PubMed Central: [PMC4720739](https://pubmed.ncbi.nlm.nih.gov/PMC4720739/)].
42. Huang MD, Chen WM, Qi FZ, Xia R, Sun M, Xu TP, et al. Long non-coding RNA ANRIL is upregulated in hepatocellular carcinoma and regulates cell apoptosis by epigenetic silencing of KLF2. *J Hematol Oncol.* 2015;**8**:50. doi: [10.1186/s13045-015-0146-0](https://doi.org/10.1186/s13045-015-0146-0). [PubMed: [25966845](https://pubmed.ncbi.nlm.nih.gov/25966845/)]. [PubMed Central: [PMC4434820](https://pubmed.ncbi.nlm.nih.gov/PMC4434820/)].
43. Naemura M, Murasaki C, Inoue Y, Okamoto H, Kotake Y. Long noncoding RNA ANRIL regulates proliferation of non-small cell lung cancer and cervical cancer cells. *Anticancer Res.* 2015;**35**(10):5377-82. [PubMed: [26408699](https://pubmed.ncbi.nlm.nih.gov/26408699/)].
44. Lin L, Gu ZT, Chen WH, Cao KJ. Increased expression of the long non-coding RNA ANRIL promotes lung cancer cell metastasis and correlates with poor prognosis. *Diagn Pathol.* 2015;**10**:14. doi: [10.1186/s13000-015-0247-7](https://doi.org/10.1186/s13000-015-0247-7). [PubMed: [25889788](https://pubmed.ncbi.nlm.nih.gov/25889788/)]. [PubMed Central: [PMC4599723](https://pubmed.ncbi.nlm.nih.gov/PMC4599723/)].
45. Zhang D, Ding L, Li Y, Ren J, Shi G, Wang Y, et al. Midkine derived from cancer-associated fibroblasts promotes cisplatin-resistance via up-regulation of the expression of lncRNA ANRIL in tumour cells. *Sci Rep.* 2017;**7**(1):16231. doi: [10.1038/s41598-017-13431-y](https://doi.org/10.1038/s41598-017-13431-y). [PubMed: [29176691](https://pubmed.ncbi.nlm.nih.gov/29176691/)]. [PubMed Central: [PMC5701200](https://pubmed.ncbi.nlm.nih.gov/PMC5701200/)].
46. Kang YH, Kim D, Jin EJ. Down-regulation of phospholipase d stimulates death of lung cancer cells involving up-regulation of the long ncRNA ANRIL. *Anticancer Res.* 2015;**35**(5):2795-803. [PubMed: [25964559](https://pubmed.ncbi.nlm.nih.gov/25964559/)].
47. Zhang D, Sun G, Zhang H, Tian J, Li Y. Long non-coding RNA ANRIL indicates a poor prognosis of cervical cancer and promotes carcinogenesis via PI3K/Akt pathways. *Biomed Pharmacother.* 2017;**85**:511-6. doi: [10.1016/j.biopha.2016.11.058](https://doi.org/10.1016/j.biopha.2016.11.058). [PubMed: [27899255](https://pubmed.ncbi.nlm.nih.gov/27899255/)].
48. Kang M, Sang Y, Gu H, Zheng L, Wang L, Liu C, et al. Long noncoding RNAs POLR2E rs3787016 C/T and HULC rs7763881 A/C polymorphisms are associated with decreased risk of esophageal cancer. *Tumour Biol.* 2015;**36**(8):6401-8. doi: [10.1007/s13277-015-3328-z](https://doi.org/10.1007/s13277-015-3328-z). [PubMed: [25874495](https://pubmed.ncbi.nlm.nih.gov/25874495/)].
49. Deng W, Wang J, Zhang J, Cai J, Bai Z, Zhang Z. TET2 regulates lncRNA-ANRIL expression and inhibits the growth of human gastric cancer cells. *IUBMB Life.* 2016;**68**(5):355-64. doi: [10.1002/iub.1490](https://doi.org/10.1002/iub.1490). [PubMed: [27027260](https://pubmed.ncbi.nlm.nih.gov/27027260/)].
50. Zou ZW, Ma C, Medoro L, Chen L, Wang B, Gupta R, et al. lncRNA ANRIL is up-regulated in nasopharyngeal carcinoma and promotes the cancer progression via increasing proliferation, reprogramming cell glucose metabolism and inducing side-population stem-like cancer cells. *Oncotarget.* 2016;**7**(38):61741-54. doi: [10.18632/oncotarget.11437](https://doi.org/10.18632/oncotarget.11437). [PubMed: [27557514](https://pubmed.ncbi.nlm.nih.gov/27557514/)]. [PubMed Central: [PMC5308687](https://pubmed.ncbi.nlm.nih.gov/PMC5308687/)].
51. Sayad A, Hajifathali A, Taheri M. Association of ANRIL gene polymorphisms with acute myeloid leukemia in an Iranian population. *Int J Cancer Manag.* 2017;**10**(10). doi: [10.5812/ijcm.11176](https://doi.org/10.5812/ijcm.11176).
52. Yu G, Liu G, Yuan D, Dai J, Cui Y, Tang X. Long non-coding RNA ANRIL is associated with a poor prognosis of osteosarcoma and promotes tumorigenesis via PI3K/Akt pathway. *J Bone Oncol.* 2018;**11**:51-5. doi: [10.1016/j.jbo.2018.02.002](https://doi.org/10.1016/j.jbo.2018.02.002). [PubMed: [29520337](https://pubmed.ncbi.nlm.nih.gov/29520337/)]. [PubMed Central: [PMC5842283](https://pubmed.ncbi.nlm.nih.gov/PMC5842283/)].
53. Meseure D, Vacher S, Alsibai KD, Nicolas A, Chemlali W, Pasmant E. Expression of ANRIL "Polycomb Complexes" CDKN2A/B/ARF genes in breast tumours: Identification of a two-gene (EZH2/CBX7) signature with independent prognostic value. *29th European Congress of Pathology.* RAI Amsterdam, The Netherlands. 2017.