

# TiO<sub>2</sub> Nanoparticle as a Sensitizer Drug in Radiotherapy: in Vitro Study

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## Abstract

**Background:** Radiosensitizer drugs are used to enhance the efficiency of radiotherapy. Some nanoparticles can be considered as radiosensitizers, because they enhance cytotoxicity due to oxidative stress and increase free radical yield, especially ROS, within cells resulting to cell death.

**Methods:** In this study, synergistic effect of TiO<sub>2</sub> nanoparticles was evaluated in presence of <sup>60</sup>Co gamma rays on human breast cancer (MCF-7) and gastric cancer (MKN-45) cell lines. After cell culture, cells were exposed to several doses of gamma rays and a dose of 2Gy was selected due to survival analysis. Next, several doses of nanoparticle from each type was applied and cell survival was analyzed from which a dose of 30µg/ml was selected for the remainder of study. Finally, synergistic effect of gamma rays and nanoparticles was evaluated in two time delay groups using MTT assay.

**Results:** Viability of cells in presence of gamma radiation and nanoparticles, significantly reduced compared to viability of cells exposed only to radiation or nanoparticle, alone (P-value≤0.05). The effect was dependent on nanoparticle type, time between addition of nanoparticle to cells and exposure to gamma rays and also cell dependent.

**Conclusion:** TiO<sub>2</sub> increased sensitivity of cancer cells to gamma radiation, due to an increase in ROS production and cytotoxicity. Anatase crystals have more severe effects than Rutile crystal because of having a larger surface area and creation of more free radicals. Therefore, this nanoparticle has the potential to be used as a radiosensitizer and further studies should be considered on other cell lines and in vivo.

**Keywords:** Titanium dioxide; Breast cancer; Gastric cancer; Radiation-sensitizing agents radiotherapy

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## Introduction

Radiotherapy is one of the major cancer treatment in which ionizing radiation is used to destroy cancerous cells [1]. In this treatment a high radiation dose should be delivered to tumor in which in some cases due to existence of some functional and healthy tissues, it is not possible to increase radiation dose practically [1] and to increase efficacy of this treatment, some chemicals are used to increase radiosensitivity of tumoral cells or reduce radiosensitivity of healthy cells [2]. Mechanism of induced radiosensitivity by most of these sensitizers is free radical and specifically ROS production. There exist a wide variety of literature on different materials have been used as sensitizers [3, 4]; along which, nanomaterials have been

considered markedly which include: gold nanoparticles [5, 6], carbon nanotubes [7, 8] and metallic nanoparticles [9]. It has been shown that simultaneous exposure to gamma-rays and nano-C60 causes a reduced survival in tumor cells and this agent increases gamma-ray effect by induction of damage in cell membrane which may be used in radiotherapy; but some problems such as low sensitizing effect, short half life and side effects should be studied [7]. There are several studies on mechanism of toxicity of nanomaterials which have shown that oxidative stress, lipid peroxidation and reaction of DNA with ROS has an important role on DNA damage, destruction of membrane and finally cell death [8, 10-12]. TiO<sub>2</sub> is a biocompatible material which in nano size causes some inflammatory effects and confirms the idea that

nanomaterials have different properties [8]. Some evidence show that nano-TiO<sub>2</sub> causes H<sub>2</sub>O<sub>2</sub> and hydroxyl free radical formation which result to cell toxicity in mammals [13-15]. Nanomaterials play an important role in DNA damage, membrane destruction and finally cell death via oxidative stress and lipid per-oxidation [13] which can develop effective modalities to destroy tumor with least side effects [14]. Among nanomaterials, TiO<sub>2</sub> is a biocompatible agent which causes inflammation in nano-domain [13] and leads to cell toxicity by super-oxide, H<sub>2</sub>O<sub>2</sub> and free hydroxyl radical formation in mammals [14]. Increasing free radicals due to induction of oxidative stress activates necrosis and apoptosis reactions and finally leads to cell death [12,15]. In this study, due to potential of TiO<sub>2</sub> nanoparticle in free radical formation -specially ROS- and also initial passive accumulation of nanoparticles in tumor cells because of increased angiogenesis [16], probability of increasing radiosensitivity of tumoral cells in presence of nanoparticle was assessed to reach a similar therapeutic efficiency using lower doses of radiation.

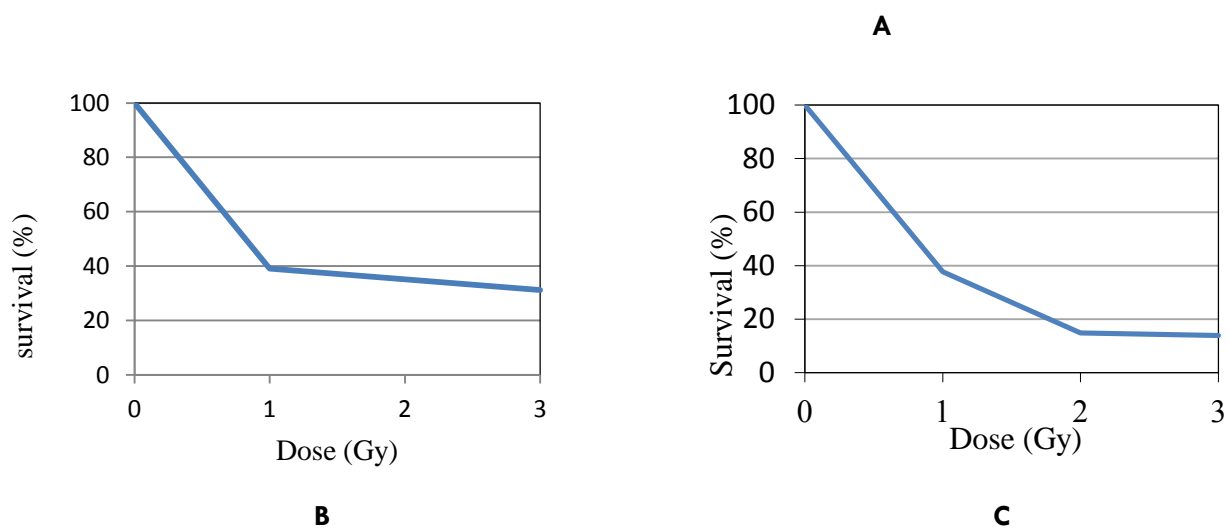
### Cell culture

In this study MCF-7 and MKN-45 cell lines were purchased from Iran Pasteur Institute. Cells were cultured in DMEM (Gibco, Invitrogen GmbH, Darmstadt, Germany) containing 10% FBS (Gibco, Invitrogen GmbH, Darmstadt, Germany), 100unit/ml Penicillin and 100 µg/ml Streptomycin and incubated at 37°C and 5% CO<sub>2</sub>.

### Irradiation condition

Cells were irradiated using a <sup>60</sup>Co therapeutic unit (AECL Theratron, Canada) at the radiotherapy department of Shohada Hospital (Tehran, Iran). Samples were placed in a 15×15cm<sup>2</sup> field of at a Source to Surface Distance (SSD) of 80cm and were irradiated. In order to find out response of selected cell lines to radiation, they were irradiated with 1, 2 and 3Gy <sup>60</sup>Co gamma rays and their survival was calculated in percents. Within three irradiation groups, 2Gy exposure was selected for the remainder of study in which survival was reduced to 20% and 37% for MKN-45 and MCF-7, respectively. Cell survival curves for these three groups are presented in Figure 1.

## Materials and Methods



**Figure 1. A)** Irradiation set up, **B)** MCF-7 survival curve under <sup>60</sup>Co irradiation, **C)** MKN-45 survival curve under <sup>60</sup>Co irradiation

### TiO<sub>2</sub> nanoparticle

To obtain effect of TiO<sub>2</sub> nanoparticles on cells, TiO<sub>2</sub> Anatase and Rutile nanoparticles (Grafen Chemical Industries, Ankara, Turkey) were added to DMEM cell culture separately. Required concentration was obtained from the pilot study. After 24 hours from initial cell culture in 96 well plates, cell culture medium was replaced with cell culture medium including nanoparticle and after 7 days, MTT assay was done. In order to find out required concentration of nanoparticles of each type, several concentrations of nanoparticle in culture media was added to two cell lines and survival percentage was obtained by MTT assay.

### Cytotoxicity assessment using MTT assay

To analyze cell survival, MTT assay was implemented. To do so, a flask of cell with concentration of 60% was trypsinized and after cell counting, cells were moved to a 96 well plate flask to embed 5000 cells in 200 µl of cell culture medium. After 24 hours, experimental groups were exposed to nanoparticle and gamma rays and were incubated at 37°C for 7 days. On the day of assessment, 20 µl of MTT solution was added to each well and after 3 hours incubation, contents of wells were replaced with 100 µl of DMSO to solvate formazon crystals. In order to run assay, 3 similar samples were obtained and their absorbance was read at 570 nm with Rayto software of Elisa reader system. Finally, survival percentage was calculated as the ratio of percentage optical density (mean light absorbance) in experimental group (O<sub>det</sub>) to

control group (O<sub>Dcont</sub>) as below:

$$\text{Survival Percentage} = \text{OD}_{\text{test}} / \text{OD}_{\text{cont}} \times 100$$

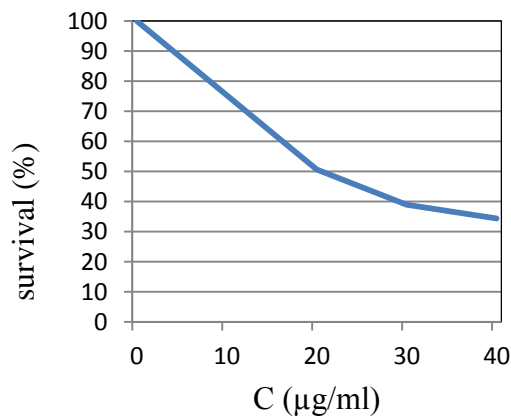
### Statistical analysis

Each experiment was repeated three times and data were presented as Mean (Standard Deviation). Statistical analyses were performed with SPSS v.17 (SPSS/PC Inc., Chicago, IL, USA) and graphs were prepared using Microsoft Excel 2007. After verifying normality and homogeneity of variables, analysis of variance (ANOVA) was performed with a 95% confidence interval (P-value ≤ 0.05); Tukey was used for multiple comparisons.

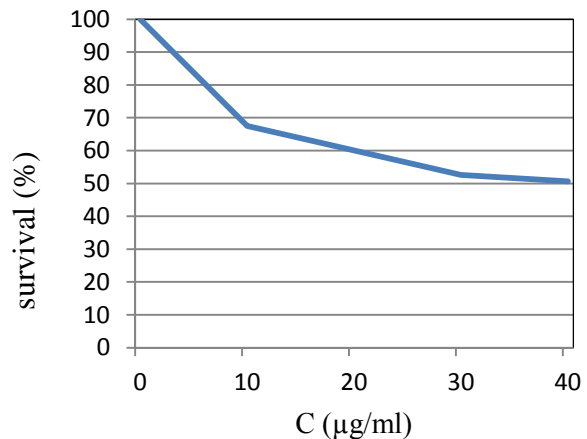
## Results

### Cytotoxicity induced by nanoparticles

Figure 2 shows the effect of nanoparticle concentration on cell survival. It was shown that in the case of Anatase, a concentration of 30 µg/ml was resulted to a 60% and 90% reduction in survival relative to non-exposed control groups for MCF-7 and MKN-45, respectively. Besides, in the case of Rutile a concentration of 30 µg/ml resulted to a 50% reduction in survival for both cell lines. Figure 3 shows microscopic images of nanoparticle exposed cell lines. As it is obvious, Rutile nanoparticle produced less cytotoxicity compared to Anatase and within two cell lines; MKN-45 was more sensitive to Anatase than MCF-7. This might be due to their different physical structure and chemical properties (Figure 2).



2A



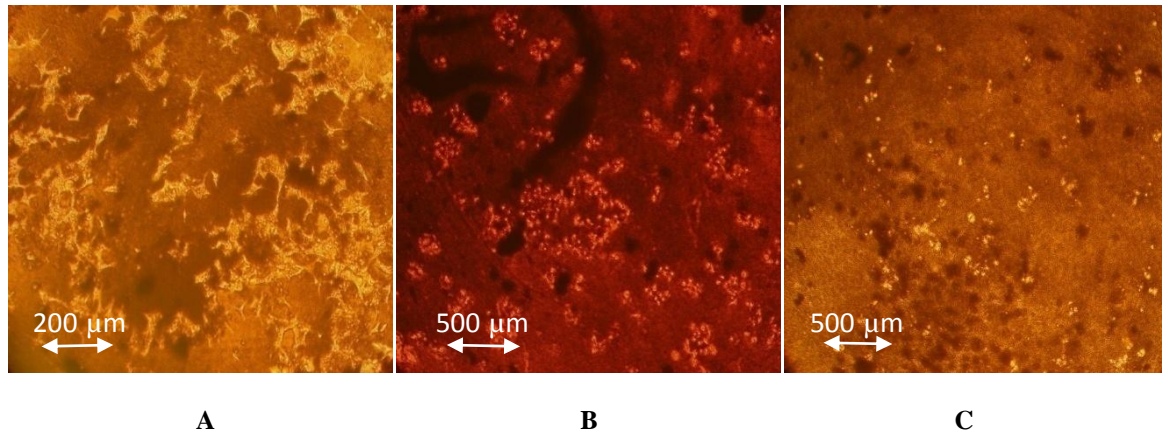
2B

**Figure 2.** A) Effect of several concentrations of Anatase on MCF-7 cell line, B) Effect of several concentrations of Rutile on MCF-7 cell line

### Microscopic study

In order to analyze cell morphology and observe effect of nanoparticle and radiation on cell structure in vitro, study was run under several groups. These groups included: absence of

nanoparticle and radiation, presence of nanoparticle with no radiation, absence of nanoparticle with radiation and presence of nanoparticle and radiation. The obtained results revealed that nanoparticle induces some morphological changes in cells (Figure 3).



**Figure 3.** A) MCF-7 cell line in the presence of Anatase with concentration of 30μg/ml, B) MKN-45 cell line in the presence of Anatase with concentration of 30μg/ml, C) MKN-45 cell line in the presence of Rutile with concentration of 30μg/ml

### Radiosensitivity induced by Anatase nanoparticle

#### MCF-7 cell line

After addition of 30μg/ml Anatase nanoparticle to MCF-7 cell line and under 2Gy of gamma-irradiation with 10 hours and 72 hours time delay, survival reduced to 12% and 14%, respectively (Figure 4). As it is obvious, presence of Anatase nanoparticle in cell culture medium alone had a lower effect on survival than radiation and simultaneous application of nanoparticle and radiation with both 10 hours and 72 hours time delays and did not reduce cell survival significantly compared to control group (P-value≤0.05).

#### MKN-45 cell line

Addition of Anatase with concentration of 30μg/ml to MKN-45 cell line and exposing cells to a dose of 2Gy from gamma rays with 10 hours and 72 hours time delay, reduced cell survival to

less than 8%. This might be due to higher sensitivity of this cell line to radiation (Figure 5).

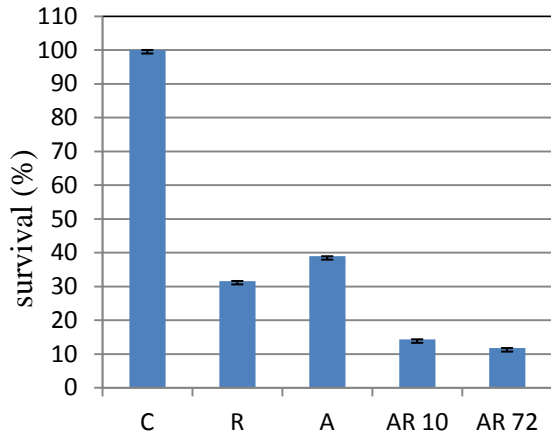
### Radiosensitivity induced by Rutile nanoparticle

#### MCF-7 cell line

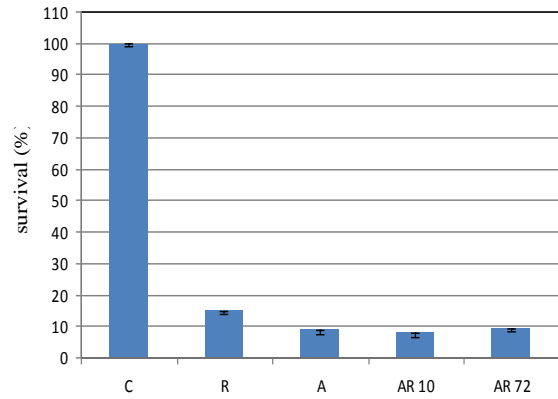
In the case of Rutile nanoparticle, after addition of nanoparticle with a dose of 30 μg/ml and irradiation with a dose of 2Gy, 10 hours and 72 hours later, survival was reduced to 35% which compared to radiation alone, had no significant difference (P-value≤0.05) (Figure 6).

#### MKN-45 cell line

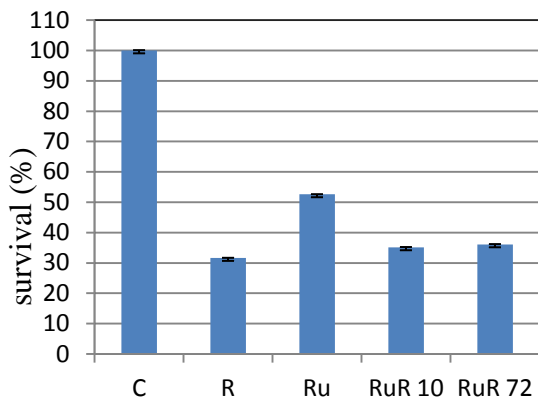
A concentration of 30μg/ml from Rutile nanoparticles was added to MKN-45 cell line and a dose of 2Gy from gamma rays was applied to them with a 10 hours and 72 hours time delay. Our findings showed that cell survival was reduced to less than 30% in both groups which effect was less than Anatase nanoparticle, and had no significant difference with gamma irradiated cells (Figure 7).



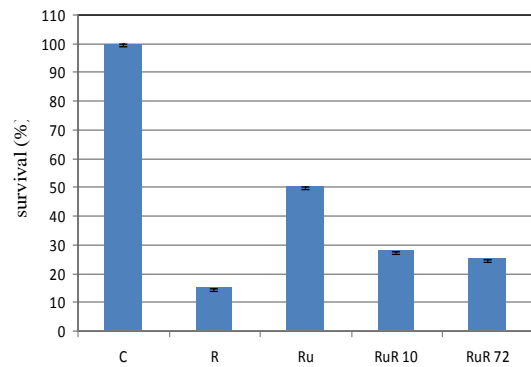
**Figure 4.** Survival percentage in MCF-7 cell line in experimental groups; **C:** Control, **R:** Radiation, **A:** Anatase nanoparticle, **AR10:** Anatase nanoparticle and irradiation with 10 hours time delay and **AR72:** Anatase nanoparticle and irradiation with 72 hours time delay



**Figure 5.** Survival percentage in MKN-45 cell line in experimental groups; **C:** Control, **R:** Radiation, **A:** Anatase nanoparticle, **AR10:** Anatase nanoparticle and irradiation with 10 hours time delay and **AR72:** Anatase nanoparticle and irradiation with 72 hours time delay



**Figure 6.** Survival percentage in MCF-7 cell line in experimental groups; **C:** Control, **R:** Radiation, **Ru:** Rutile nanoparticle, **RuR10:** Rutile nanoparticle and irradiation with 10 hours time delay and **RuR72:** Rutile nanoparticle and irradiation with 72 hours time delay



**Figure 7.** Survival percentage in MKN-45 cell line in experimental groups; **C:** Control, **R:** Radiation, **Ru:** Rutile nanoparticle, **RuR10:** Rutile nanoparticle and irradiation with 10 hours time delay and **RuR72:** Rutile nanoparticle and irradiation with 72 hours time delay

## Discussion

With increasing development of nanotechnology and using different aspects of nanomaterials, a new scope has been formed in all scientific fields. This technology has found application in several fields from industrials, home appliances and specially medicine [17]. In the field of medicine, using nanomaterial properties in both fields of diagnosis and therapy of disease has profoundly studied [18-20]. TiO<sub>2</sub> is a member of metallic nanomaterials and has been considered widely due to its specific chemical properties. This nanoparticle has different forms with different physical and chemical properties and the most important forms are Anatase and Rutile. Rutile has a higher stability and larger size than Anatase [21]. This nanoparticle has some photocatalytic characteristics which in higher concentrations, stimulates with UV radiation and produces free radicals [22-24]. This nanoparticle interacts with water molecules in cell medium and using electron capture pathway, produces free radicals specially ROS [17]. The exact mechanism of free radical production by TiO<sub>2</sub> has not determined yet, but several studies have shown that Anatase crystals aggregate in mitochondria and cause some defect in electron chain and destruct its function. This leads to more and more production of free radicals. But in the case of Rutile, the problem is different. Rutile crystals place in cell sparsely and don't enter in mitochondria [25]. Our findings support this problem when Anatase alone has a higher effect than Rutile on the breast cancer cell line. There exists a wide variety of literature on cytotoxicity induced by TiO<sub>2</sub>. These include fibroblast and epithelial cells [26, 27], kidney cells [28], neuroblast cells [27] and endothelial cells [29]. All of these studies relate this toxicity to ROS production. Some studies consider synergistic effect of UV radiation and TiO<sub>2</sub> nanoparticles on several cells such as CHO [24], glioma [22] and HeLa [23]. These studies relate the enhanced cytotoxicity to the capability of this nanoparticle to react with water molecule in cell medium and yield ROS via electron capture pathway. This capability provides the potential for this particle to be considered as a radiosensitizer. According to production of ROS by TiO<sub>2</sub> nanoparticle and their aggregation in tumoral cells via active or passive targeting because of their high angiogenesis [30], it has been implemented in photodynamic therapy as a photosensitizing agent [23, 24, 31]. UV and laser light that are used as external stimuli in this treatment has low penetration capability, therefore this treatment is only effective for surface tumors [23]. Besides, we observed more radiosensitivity induced by Anatase compared to Rutile. Our results showed that by increasing the concentration of Anatase nanoparticles, survival percentage was reduced, but the amount of this reduction was not as severe

as lower concentrations which might be due to high toxicity induced in this lower concentration. In addition, the amount of reduction in survival was more in MKN-45 compared to MCF-7; it was due to faster cell cycle of MKN-45 which caused a higher sensitization. In the case of Rutile nanoparticle, it had a lower effect on cell survival compared to Anatase nanoparticle in both cell lines. This was in a manner that simultaneous application of nanoparticle and radiation resulted to a significant reduction in Anatase groups' survival in both cell lines ( $P\text{-value} \leq 0.05$ ), but Rutile did not cause a significant reduction in cell survival when simultaneously exposed to gamma rays ( $P\text{-value} \geq 0.05$ ). The different observed effect might be due to the higher surface to volume ratio in Anatase compared to Rutile which makes it more effective than Rutile. So, it produces more ROS and because Anatase has higher aggregation in mitochondria than Rutile and causes more disorder on electron transfer chain function and mitochondria [32]. This phenomenon might be the reason for different radiosensitization induced by two nanoparticles. There were no published work on the radiosensitization of TiO<sub>2</sub>, but several studies has showed the different effect of these two types and higher ROS production in Anatase compared to Rutile [21, 24, 26, 28, 32-34].

Results of statistical analysis showed that time delay between application of nanoparticles and exposure to gamma rays doesn't induce any significant difference between groups ( $P\text{-value} \geq 0.05$ ). This might be due to fast and stable effect of nanoparticle, so any increase in time delay doesn't change their effect, but it is necessary to design more studies.

## Conclusion

Our findings showed that the toxicity of Anatase have been increased in presence of gamma irradiation therefore it is possible to consider this particle as a radiosensitizer drug; but it is necessary to assess this in vivo and examine its possible side effects. The effect of this nanoparticle is both dose and cell type dependent. In the case of Rutile, it is not a good candidate for a radiosensitizer drug but it is recommended to assess the effect of particle size on cell survival, because in nano domain, size and surface to volume ratio play an important role in observed effects. Besides, we recommend different types of radiations and energies to be considered in combination of this nanoparticle.

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## Conflict of Interest

The authors have no conflict of interest in this study.

## Authors' Contribution

Mostafa Rezaei-Tavirani, Hadi Hasanzadeh and Samaneh Sadat Seyyedi designed the study, gathered and analyzed the data and wrote the paper. Elham Dolat and Vahid Semnani contributed to study design, sample collection and indentation. Sara Sobhi helped in writing and overall correction of the manuscript.

## References

1. Khan FM. The physics of radiation therapy. 3rd ed. Philadelphia: Lippincott Williams & Wilkins; 2003; 4(4):110-20.
2. Bump EA, Hoffman SJ, Foye WO, Abraham DJ. Radiosensitizers and Radioprotective Agents. *Burger's Medicinal Chemistry and Drug Discovery: John Wiley & Sons, Inc.*; 2003; p. 151-214.
3. Geng CX, Zeng ZC, Wang JY, Xuan SY, Lin CM. Docetaxel shows radiosensitization in human hepatocellular carcinoma cells. *World J Gastroenterol.* 2005; 11(19): 2990-3.
4. Javvadi P, Segan AT, Tuttle SW, Koumenis C. The chemopreventive agent curcumin is a potent radiosensitizer of human cervical tumor cells via increased reactive oxygen species production and overactivation of the mitogen-activated protein kinase pathway. *Mol Pharmacol.* 2008; 73(5): 1491-501.
5. Jain S, Coulter JA, Hounsell AR, Butterworth KT, McMahon SJ, Hyland WB, et al. Cell-specific radiosensitization by gold nanoparticles at megavoltage radiation energies. *Int J Radiat Oncol Biol Phys.* 2011; 79(2): 531-9.
6. Lim ZZ, Li JE, Ng CT, Yung LY, Bay BH. Gold nanoparticles in cancer therapy. *Acta Pharmacol Sin.* 2011; 32(8): 983-90.
7. Ni J, Wu Q, Li Y, Guo Z, Tang G, Sun D, et al. Cytotoxic and radiosensitizing effects of nano-C60 on tumor cells in vitro. *Journal of Nanoparticle Research.* 2008; 10: 643-51.
8. Oberdorster E. Manufactured nanomaterials (fullerenes, C60) induce oxidative stress in the brain of juvenile largemouth bass. *Environ Health Perspect.* 2004; 112(10): 1058-62.
9. Le Sech C, Kobayashi K, Usami N, Furusawa Y, Porcel E, Lacombe S. Comment on 'Therapeutic application of metallic nanoparticles combined with particle-induced x-ray emission effect'. *Nanotechnology.* 2012; 23(7).
10. Hsin YH, Chen CF, Huang S, Shih TS, Lai PS, Chueh PJ. The apoptotic effect of nanosilver is mediated by a ROS- and JNK-dependent mechanism involving the mitochondrial pathway in NIH3T3 cells. *Toxicol Lett.* 2008; 179(3): 130-9.
11. Li N, Xia T, Nel AE. The role of oxidative stress in ambient particulate matter-induced lung diseases and its implications in the toxicity of engineered nanoparticles. *Free Radic Biol Med.* 2008; 44(9): 1689-99.
12. Sharma V, Shukla RK, Saxena N, Parmar D, Das M, Dhawan A. DNA damaging potential of zinc oxide nanoparticles in human epidermal cells. *Toxicol Lett.* 2009; 185(3): 211-8.
13. Gurr JR, Wang AS, Chen CH, Jan KY. Ultrafine titanium dioxide particles in the absence of photoactivation can induce oxidative damage to human bronchial epithelial cells. *Toxicology.* 2005; 213(1-2): 66-73.
14. Kang JL, Moon C, Lee HS, Lee HW, Park EM, Kim HS, et al. Comparison of the biological activity between ultrafine and fine titanium dioxide particles in RAW 264.7 cells associated with oxidative stress. *J Toxicol Environ Health A.* 2008; 71(8): 478-85.
15. Kang SJ, Kim BM, Lee YJ, Chung HW. Titanium dioxide nanoparticles trigger p53-mediated damage response in peripheral blood lymphocytes. *Environ Mol Mutagen.* 2008; 49(5): 399-405.
16. Robertson F, Ferrari M. Introduction and rationale for nanotechnology in cancer therapy. In: Amiji MM, editor. *Nanotechnology for Cancer Therapy: Taylor & Francis group*; 2006; 3-10.
17. Singh N, Manshian B, Jenkins GJS, Griffiths SM, Williams PM, Maffei TGG, et al. NanoGenotoxicology: The DNA damaging potential of engineered nanomaterials. *Biomaterials.* 2009; 30(23-24): 3891-914.
18. Shrivastava S, Dash D. Applying Nanotechnology to Human Health: Revolution in Biomedical Sciences. *Journal of Nanotechnology.* 2009; 2009: 1-14.
19. Hasanzadeh H, Mokhtari-Dizaji M, Bathaie SZ, Hassan ZM. Effect of local dual frequency sonication on drug distribution from polymeric nanomicelles. *Ultrason Sonochem.* 2011; 18(5): 1165-71.
20. Barati AH, Hejazi P, Hasanzadeh H. Hematoporphyrin encapsulated polymeric nanomicelles for photodynamically treatment of cancer. *JPS.* 2012; 3(3): 15-9.
21. Liu H, Ma L, Zhao J, Liu J, Yan J, Ruan J, et al. Biochemical toxicity of nano-anatase TiO<sub>2</sub> particles in mice. *Biol Trace Elem Res.* 2009; 129(1-3): 170-80.
22. Yamaguchi S, Kobayashi H, Narita T, Kanehira K, Sonezaki S, Kubota Y, et al. Novel photodynamic therapy using water-dispersed TiO<sub>2</sub>-polyethylene glycol compound: evaluation of antitumor effect on glioma cells and spheroids in vitro. *Photochem Photobiol.* 2010; 86(4): 964-71.
23. Matsui K, Karasaki M, Segawa M, Hwang SY, Tanaka T, Ogino C, et al. Biofunctional TiO<sub>2</sub> nanoparticle-mediated photokilling of cancer cells

using UV irradiation. *Med Chem Comm.* 2010; 1(3): 209-11.

24. Uchino T, Tokunaga H, Ando M, Utsumi H. Quantitative determination of OH radical generation and its cytotoxicity induced by TiO<sub>2</sub>-UVA treatment. *Toxicol In Vitro.* 2002; 16(5): 629-35.

25. Jin C, Tang Y, Yang FG, Li XL, Xu S, Fan XY, et al. Cellular toxicity of TiO<sub>2</sub> nanoparticles in anatase and rutile crystal phase. *Biol Trace Elem Res.* 2011; 141(1-3): 3-15.

26. Sayes CM, Wahi R, Kurian PA, Liu Y, West JL, Ausman KD, et al. Correlating nanoscale titania structure with toxicity: a cytotoxicity and inflammatory response study with human dermal fibroblasts and human lung epithelial cells. *Toxicol Sci.* 2006; 92(1): 174-85.

27. Lai JC, Lai MB, Jandhyam S, Dukhande VV, Bhushan A, Daniels CK, et al. Exposure to titanium dioxide and other metallic oxide nanoparticles induces cytotoxicity on human neural cells and fibroblasts. *Int J Nanomedicine.* 2008; 3(4): 533-45.

28. L'Azou B, Jorly J, On D, Sellier E, Moisan F, Fleury-Feith J, et al. In vitro effects of nanoparticles on renal cells. *Particle and Fibre Toxicology.* 2008; 19: 5-22.

29. Peters K, Unger RE, Kirkpatrick CJ, Gatti AM, Monari E. Effects of nano-scaled particles on endothelial cell function in vitro: studies on viability, proliferation and inflammation. *J Mater Sci Mater Med.* 2004; 15(4): 321-5.

30. Moghimi SM. Passive targeting of solid tumors: pathophysiological principles and physicochemical aspects of delivery systems. *Nanotechnology for cancer therapy.* 2007: 11-8.

31. Yamaguchi S, Kobayashi H, Narita T, Kanehira K, Sonezaki S, Kubota Y, et al. Novel Photodynamic Therapy Using Water dispersed TiO<sub>2</sub> Polyethylene Glycol Compound: Evaluation of Antitumor Effect on Glioma Cells and Spheroids In Vitro. *Photochemistry and photobiology.* 2010; 86(4): 964-71.

32. Jin C, Tang Y, Yang FG, Li XL, Xu S, Fan XY, et al. Cellular toxicity of TiO<sub>2</sub> nanoparticles in anatase and rutile crystal phase. *Biol Trace Elem Res.* 2010; 141(1-3): 3-15.

33. Zhu RR, Wang SL, Chao J, Shi DL, Zhang R, Sun XY, et al. Bio-effects of Nano-TiO<sub>2</sub> on DNA and cellular ultrastructure with different polymorph and size. *Materials Science and Engineering: C.* 2009; 29(3): 691-6.

34. Liu H, Ma L, Liu J, Zhao J, Yan J, Hong F. Toxicity of nano-anatase TiO<sub>2</sub> to mice: Liver injury, oxidative stress. *Toxicological & Environmental Chemistry.* 2010; 92: 175-186.