Published online 2018 November 12.

Review Article

Genomics Role in Cancer Immunosurveillance: Impact on Immunotherapy Response

Sanaz Tabarestani¹, Mohammad Esmaeil- Akbari ¹⁰ and Saeed Namaki^{2,*}

¹Cancer Research Center, Shahid Beheshti University of Medical Sciences, Tehran, Iran ²Department of Medical Immunology, School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran

Corresponding author: Department of Medical Immunology, School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran. Email: namaki@yahoo.com

Received 2018 October 20; Revised 2018 October 26; Accepted 2018 October 28.

Abstract

Context: One of the most important achievements in cancer research is the development of cancer immunotherapy. However, only a subset of patients respond to immunotherapy modalities, and few patients respond for a durable time. Here, we review the possible genomic mechanisms of response and resistance to these therapies, which can lead to the selection of responders, who may benefit most from immunotherapy.

Evidence Acquisition: We searched PubMed, Scopus, and Web of Science Core Collection with the following keywords: "Immunotherapy, Resistance, Response, Programmed cell death 1 receptor, CTLA-4, Cancer immunity, Tumor Genomics, and Somatic Mutations".

Results: T cells that specifically recognize cancer-associated antigens, are responsible for the immune system response against cancer. Nonsynonymous mutations, which are transcribed and translated into polypeptides, may generate new epitopes (neoepitopes), which can lead to their presentation on major histocompatibility (MHC) class I molecules and subsequently recognized by the adaptive immune system. Despite the unprecedented durable responses, the majority of patients treated with cancer immunotherapies do not respond to the therapy (primary resistance), and some patients relapse after an initial response (acquired resistance). Resistance to immunotherapy can be a result of tumor cell intrinsic or extrinsic factors. There is correlation between tumor mutation burden (TMB) and response to immunotherapy. In addition, mismatch repair deficient tumors harbor considerably more somatic mutations compared to mismatch repair proficient tumors and respond better to anti-programmed cell death protein 1 (anti-PD1) therapy. Mutations in other DNA repair genes may also affect immunotherapy response.

Conclusions: Neoantigen specific T cells constitute a major "active component" for the success of cancer immunotherapies. The genetic damage that confers tumorigenic growth, can also be targeted by the immune machinery to inhibit cancer development and progression. With further validation of experiments, genomics-based approaches can allow to select patients most likely to achieve durable responses to immunotherapy modalities.

Keywords: Cancer Genomics, Immunotherapy, Cancer Immunosurveillance, Tumor Mutation Burden

1. Context

One of the most important achievements in cancer research is the development of cancer immunotherapy (1, 2). The first cancer immunotherapy was performed by Coley, when he demonstrated that the administration of bacterial products could make tumor regression of inoperable neoplasms (3). Coley's findings resulted in the beginning of cancer immunotherapy and, consequently in the 1960s, Bacillus Calmette-Guerin (BCG) was used to treat solid malignancies, such as bladder cancer. Subsequent research efforts in the 1970s and 1980s showed that activating lymphocytes with interleukin-2 (IL-2) could kill cancer cells in vitro (4). Large clinical trials investigated the therapeutic effects of cytokines for melanoma, and renal cell carcinoma in the 1980s (5-8). During this same period, it was demonstrated that interferon- α (IFN- α) has anti-tumor activity in melanoma (9-12).

Several monoclonal antibodies targeting cancerassociated proteins (such as HER2, EGFR, VEGF, and CD20) have been approved for the treatment of solid and hematologic neoplasms. These antibodies may opsonize cancer cells and induce their death by antibody-dependent cellular cytotoxicity or phagocytosis, in addition to antagonizing oncogenic pathways (13, 14). More recently, newer approaches in cancer immunotherapy have been developed, including administration of therapeutic cancer vaccines and adoptive cell transfer by chimeric antigen

Copyright © 2018, Author(s). This is an open-access article distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 International License (http://creativecommons.org/licenses/by-nc/4.0/) which permits copy and redistribute the material just in noncommercial usages, provided the original work is properly cited.

receptor (CAR) T-cell therapy (15, 16). The finding of checkpoints in the immune machinery, such as CTL-associated protein 4 (CTLA-4) and programmed cell death protein 1 (PD-1) resulted in the development of antibodies that inhibit these checkpoints, leading to T-cell activation (17, 18). Antibodies against these checkpoints have been approved in diverse malignancies, such as non-small cell lung cancer, melanoma, Hodgkin's lymphoma, renal cell cancer, and head and neck cancer (19-24). An important characteristic of immunotherapy is durable responses, which is supposedly due to the adaptive immune system memory, resulting in increased survival in some patients.

Regrettably, merely a subset of patients respond to immunotherapy modalities, and few patients respond for a durable time. Here, we review the possible genomic mechanisms of response and resistance to these therapies, which may lead to the selection of responders, who may benefit most from immunotherapy.

2. Evidence Acquisition

We searched PubMed, Scopus, and Web of Science Core Collection with the following keywords: "Immunotherapy, Resistance, Response, Programmed cell death 1 receptor, CTLA-4, Cancer immunity, Tumor Genomics, and Somatic Mutations". We also analyzed the references mentioned by the selected articles.

3. Results

A hallmark of cancer is the accumulation of somatic genetic alterations and loss of normal cellular regulatory control (25). The number of these somatic alterations is different between and within cancer types (26). Somatic mutations in tumors may be due to intrinsic infidelity of the DNA replication system, exposure to exogenous or endogenous mutagens, or impaired DNA repair. In some cancers, most somatic alterations are a consequence of environmental exposures, such as ultraviolet light in skin and smoking in lung cancers (27). In other cancers, there are abnormalities in DNA maintenance, such as impaired DNA mismatch repair in some colorectal cancers (28).

3.1. The Catalogue of Somatic Mutations in Cancer

A cancer cell, similar to all cells of the body, is a descendent of a fertilized egg through mitotic cell division, and has a copy of diploid genome. The DNA of a tumor cell genome, similar to many normal cellular genomes, has differences from the original fertilized egg genome. These differences are known as somatic mutations, and varies from germline mutations, which are transmitted from parents to their children.

The somatic mutations in neoplastic cells include divers types of DNA change, including base substitutions, insertions, deletions, rearrangements, and copy number alterations. The tumor genome has also acquired epigenetic alterations, which change the chromatin structure and gene expression. The mutations in a malignant cell have accumulated during the lifetime of the patient with cancer. In normal cells, exogenous and endogenous mutagens continuously damage DNA. This DNA damage is mostly repaired, but a small subset becomes fixed mutations. In addition, there is a low intrinsic error rate in DNA replication machinery (29). The knowledge on somatic mutation rate in normal human cells is limited, but it is likely dependent on the specific somatic mutation class and various cell types (29).

It seems that the rate of somatic mutation is higher when a cancer cell shows phenotypic evidence of malignant change (30). This is evident at least in some cancers. For instance, there is an increased rate of single nucleotide change and small insertion/deletions at polynucleotide tracts in colorectal and endometrial cancers with impaired DNA mismatch repair (31). Other classes of "mutator phenotypes" may lead to abnormalities in chromosomal number and increased rate of chromosomal rearrangements (31). Increased mutation rate leads to increased DNA sequence diversity, on which selection acts. The catalogue of somatic mutations in a neoplastic cell is a record of all mutational processes throughout the life of the cancer patient.

Somatic mutations in a neoplastic genome may be classified as driver or passenger, based on its consequences on tumor development. Driver mutations lead to growth advantage of the cancer cell and have been selected positively during cancer development. They are located in the so called "cancer genes". The number of driver mutations and aberrant cancer genes vary between different cancers (32). The rest of mutations are passengers and do not confer growth advantage. Passenger mutations do not have functional consequences and can happen during cell division (29).

The number of driver mutations in a malignancy is an area of active research, but on the basis of age-incidence statistics, it is likely that 5 to 7 driver mutations are required in common epithelial cancers, such as breast, prostate, and colorectal cancers, whereas hematological cancers may require fewer rate-limiting events (29). Recent research have suggested that the number of positively selected cancer driver genes range from 1 in thyroid and testicular cancers to more than 10 in endometrial and colorectal cancers (33, 34). Although cancer genes frequently con-

tain driver mutations, only a fraction of the somatic mutations detected in these genes are considered true drivers and the rest are passenger mutations (35).

3.2. Cancer Genome as a Driver of Tumor Immunity

T cells that specifically recognize cancer-associated antigens are responsible for the immune system response against cancer. Nonsynonymous mutations, which are transcribed and translated into polypeptides may generate new epitopes (neoepitopes), which can lead to their presentation on major histocompatibility (MHC) class I molecules and subsequently recognized by the adaptive immune system (Figure 1) (36). However, even when T cell recognition occurs, they rarely confer protective immunity and cannot be used as a basis for therapy.

As demonstrated by mouse models, continued deletion of tumor cells expressing targets of T-cell recognition (immune editing) may help cancers evolve to avoid immune attack (37). In addition, factors in tumor microenvironment can modulate activated T cells and act as an "immunostat". Despite these results, recent research on human cancer has shown that overcoming negative regulators of T cell responses in lymphoid tissue (checkpoints) or in the tumor(immunostat function) can reverse the failure of immune protection in most patients with cancer (38).

Results from mouse models have shown that point mutations may result in the production of mutant antigens, which induce T cells and elicit endogenous immunity or immunity with the administration of a therapeutic vaccine (39, 40). However, the recognition of mutant antigens by the immune system is not efficient. A study on a mouse model showed that only 10% of nonsynonymous point mutations encoded peptides that could be recognized with high affinity by MHC class I molecules and merely a subset of generated peptides were highly immunogenic after administration as cancer vaccine (41). Immunogenic peptides contain mutations that are exposed to the T-cell receptor or mutations that make new anchor that enhance the affinity to MHC class I molecules. In addition to point mutations, frameshift mutations, insertions, or deletions may generate new peptides (neoantigens) that are supposedly more immunogenic due to their more sequence divergence (42).

Protective immunity may be conferred by mutant peptides that can bind MHC class I or class II molecules (40, 43), although it is not certain whether CD4+ T cells have a role in anti-cancer immunity. CD4 expressing T cells help the cytotoxic CD8 positive T cells and B cell antibody production. In addition, they secret IFN- γ and induce an inflammatory response that is in favor of antitumor immunity (44). A study on human melanoma detected CD4+ T cell response against tumor neoantigens, suggesting that in addition to intratumoral CD8+T cells, antigen-specific CD4+T cells also have a role in cancer immunity (45). Several studies have determined the specificities of tumor infiltrating lymphocytes (TILs) to tumor mutant epitopes by massively parallel sequencing and demonstrated that the adoptive transfer of antigen specific TILs into cancer patients is correlated with tumor regression (46-49). Together, these findings suggest that tumor mutanome is the main target of protective anti-cancer immunity and the large number of passenger mutations constitutes the most probable targets. Therefore, the mutational neoepitope load of a tumor is considered an important part of the cancer-immune set point (42).

It is important to consider the nature of the mutated genes, from which neoantigens derive and the frequency, with which they occur. It is clear that T cell responses against MHC class I-restricted and class II-restricted tumor neoantigens that are shared between subgroups of patients, sometimes occur (50, 51). At the same time, research on melanoma has shown that the majority of neoantigen-specific T cell responses are against mutated proteins unique to each tumor and likely without a key role in cellular transformation (45). This T cell preference toward patient-specific passenger mutations means that the development of personalized immunotherapies is possibly required for targeting neoantigens.

3.3. Resistance Mechanisms to Cancer Immunotherapy

With the finding of cancer immune checkpoints and the successful clinical implementation of checkpoint inhibitors, cancer treatment focus has changed from the tumor itself to the host's immune machinery. It is important to note that host immune responses and cancer genomics are closely linked, as neoantigens driving from tumor mutations may shape immune responses (52). However, the heterogeneous and evolving tumor environment may render these responses ineffective.

Despite the unprecedented durable responses, the majority of patients treated with cancer immunotherapies do not respond to the therapy (primary resistance), and some patients relapse after an initial response (acquired resistance). With regard to resistance mechanisms to cancer immunotherapies, it is necessary to consider that in each cancer patient, the immune response is dynamic and evolves either due to the patient's environmental and genetic factors or due to treatment interventions, such as surgery, radiotherapy, chemotherapy, and immunotherapy. Many of these factors may affect anti-tumor immune responses and result in the establishment of resistance (53). Furthermore, human tumors differ considerably with regard to the constituents of their microenvironment, and this variability supposedly influences the capability of the



Figure 1. Processing of tumor neoantigens and their recognition by CD8+ T cells. MHC: Major histocompatibility complex, TCR: T cell receptor, TAP: Transporter associated with antigen processing.

T cells to recognize and respond to neoantigens. Comprehensive understanding of resistance mechanisms can lead to the discovery of biomarkers of response to immunotherapies, similar to biomarkers of response to other cancer therapies (54-57).

Primary resistance to immunotherapy can be a result of tumor cell intrinsic or extrinsic factors. The most obvious reason why a tumor would not respond to immunotherapy is the absence of tumor antigens being recognized by T cells (58). Alternatively, tumor antigens may exist in tumor cells, but the neoplastic cells may develop mechanisms to escape presenting them on their surface bound to MHC molecules, such as antigen presentation alterations (including changes in proteasome subunits or transporters associated with antigen processing [TAP]), β 2microglobulin alterations (required for MHC class I folding and transport to surface of cell), or MHC alterations (59).

Various clinical trials involving immune checkpoint inhibitors have identified different response markers. A study on pembrolizumab, a PD-1 inhibitor, found that the proportion of tumor cells expressing PD-L1 affected the response rate (60). This trial showed that it is possible to use PD-L1 to select patients most likely to respond to the PD-1 inhibitor. Another study on atezolizumab, a PD-L1 inhibitor, showed that the likelihood of response was correlated with the immune cell infiltrate in the tumor, as well as PD-L1 expression on tumor cells (61). A study on pembrolizumab showed superiority of this treatment in patients with tumors that expressed PD-L1 on at least 50% of cancer cells (62).

The advent of next generation sequencing (NGS) technology has resulted in sequencing cancer genomes in an unprecedented ultra-fast and high-throughput manner. Using genomic approaches, several research groups have shown that there is a correlation between tumor mutation burden (TMB) and response to checkpoint therapy (63-67). As an example, in lung cancers mutations are often detected with a frequency of 0.1 to 100 per megabase (26). In non-small cell lung cancers with mutations in *EGFR*, *ALK*, *ROS1*, and *MEK* exon 14, TMB is usually low and these tumors usually do not respond well to check point inhibitors (68).

The association between tumor mutation (and neoantigen) load and immunotherapy response has been typically detected in tumors that have high mutational load (such as non-small cell lung cancer, melanoma, and renal carcinoma). But, some tumors (such as colorectal cancers) that generally have a significant number of mutations, usually do not respond to immunotherapy (26, 69). A likely reason is the extent of intratumoral heterogeneity, since bulk tumor sequencing might not fully show the spatial complexity of the tumor mutanome (70). As a heterogeneous population of subclones constitutes the tumor bulk, immune response against one neoantigen would target a subclonal population, thus leaving the remainder of the tumor intact.

Defects in mismatch repair generate a high mutation load. Research has shown that mismatch repair deficient tumors harbor considerably more somatic mutations compared to mismatch repair proficient tumors and respond better to anti-PD1 therapy (69, 71). Mutations in DNA repair genes may also affect immunotherapy response. A study showed that the loss of function mutations in *BRCA2* gene were enriched in melanoma patients responsive to anti-PD1 blockade (66). Another study also detected mutations in DNA repair genes in patients with non-small cell lung cancer responsive to PD1 blockade (67). Therefore, even in cancers that generally have few mutations, such genomic analysis may detect a group of patients, who may benefit from immunotherapy.

4. Conclusions

Based on data gathered over the past few years, neoantigen specific T cells constitute a major "active component" for the success of cancer immunotherapies. The genetic damage that confers tumorigenic growth can also be targeted by the immune machinery to inhibit cancer development and progression. As expression of these neoantigens is tumor restricted; cancer immunotherapies offer the promise of specificity and safety. With further validation of experiments, genomics-based approaches can allow to select patients most likely to achieve durable responses to immunotherapy modalities.

Acknowledgments

None declared.

Footnotes

Authors' Contribution: All authors designed the study. Sanaz Tabarestani performed the literature search and wrote the manuscript. All authors approved the manuscript.

Conflict of Interests: None declared.

Financial Disclosure: None declared.

References

- Mellman I, Coukos G, Dranoff G. Cancer immunotherapy comes of age. *Nature*. 2011;480(7378):480–9. doi: 10.1038/nature10673. [PubMed: 22193102]. [PubMed Central: PMC3967235].
- Topalian SL, Weiner GJ, Pardoll DM. Cancer immunotherapy comes of age. J Clin Oncol. 2011;29(36):4828–36. doi: 10.1200/JCO.2011.38.0899.
 [PubMed: 22042955]. [PubMed Central: PMC3255990].
- Nauts HC, Fowler GA, Bogatko FH. A review of the influence of bacterial infection and of bacterial products (Coley's toxins) on malignant tumors in man; a critical analysis of 30 inoperable cases treated by Coley's mixed toxins, in which diagnosis was confirmed by microscopic examination selected for special study. *Acta Med Scand Suppl.* 1953;276:1-103. [PubMed: 13039964].
- Grimm EA, Mazumder A, Zhang HZ, Rosenberg SA. Lymphokineactivated killer cell phenomenon. Lysis of natural killer-resistant fresh solid tumor cells by interleukin 2-activated autologous human peripheral blood lymphocytes. *J Exp Med.* 1982;155(6):1823–41. doi: 10.1084/jem.155.6.1823. [PubMed: 6176669]. [PubMed Central: PMC2186695].
- Dutcher JP, Creekmore S, Weiss GR, Margolin K, Markowitz AB, Roper M, et al. A phase II study of interleukin-2 and lymphokine-activated killer cells in patients with metastatic malignant melanoma. *J Clin Oncol.* 1989;7(4):477-85. doi: 10.1200/JCO.1989.7.4.477. [PubMed: 2647913].
- Atkins MB, Lotze MT, Dutcher JP, Fisher RI, Weiss G, Margolin K, et al. High-dose recombinant interleukin 2 therapy for patients with metastatic melanoma: Analysis of 270 patients treated between 1985 and 1993. J Clin Oncol. 1999;17(7):2105-16. doi: 10.1200/JCO.1999.17.7.2105. [PubMed: 10561265].
- Parkinson DR, Abrams JS, Wiernik PH, Rayner AA, Margolin KA, Van Echo DA, et al. Interleukin-2 therapy in patients with metastatic malignant melanoma: A phase II study. *J Clin Oncol*. 1990;8(10):1650–6. doi: 10.1200/JCO.1990.8.10.1650. [PubMed: 2213101].
- Rosenberg SA, Yang JC, Topalian SL, Schwartzentruber DJ, Weber JS, Parkinson DR, et al. Treatment of 283 consecutive patients with metastatic melanoma or renal cell cancer using high-dose bolus interleukin 2. *JAMA*. 1994;**271**(12):907-13. doi: 10.1001/jama.1994.03510360033032. [PubMed: 8120958].
- Krown SE, Burk MW, Kirkwood JM, Kerr D, Morton DL, Oettgen HF. Human leukocyte (alpha) interferon in metastatic malignant melanoma: The American Cancer Society phase II trial. *Cancer Treat Rep.* 1984;68(5):723-6. [PubMed: 6722830].
- Kirkwood JM, Ernstoff MS, Davis CA, Reiss M, Ferraresi R, Rudnick SA. Comparison of intramuscular and intravenous recombinant alpha-2 interferon in melanoma and other cancers. *Ann Intern Med.* 1985;103(1):32–6. doi: 10.7326/0003-4819-103-1-32. [PubMed: 4003987].
- Kirkwood JM, Strawderman MH, Ernstoff MS, Smith TJ, Borden EC, Blum RH. Interferon alfa-2b adjuvant therapy of high-risk resected cutaneous melanoma: The Eastern Cooperative Oncology Group trial EST 1684. *J Clin Oncol.* 1996;14(1):7–17. doi: 10.1200/JCO.1996.14.1.7. [PubMed: 8558223].

- Kirkwood JM, Ibrahim JG, Sosman JA, Sondak VK, Agarwala SS, Ernstoff MS, et al. High-dose interferon alfa-2b significantly prolongs relapsefree and overall survival compared with the GM2-KLH/QS-21 vaccine in patients with resected stage IIB-III melanoma: Results of intergroup trial E1694/S9512/C509801. *J Clin Oncol.* 2001;19(9):2370–80. doi: 10.1200/JCO.2001.19.9.2370. [PubMed: 11331315].
- Nahta R, Esteva FJ. Herceptin: Mechanisms of action and resistance. *Cancer Lett.* 2006;232(2):123-38. doi: 10.1016/j.canlet.2005.01.041. [PubMed: 16458110].
- Weiner GJ. Rituximab: Mechanism of action. Semin Hematol. 2010;47(2):115–23. doi: 10.1053/j.seminhematol.2010.01.011. [PubMed: 20350658]. [PubMed Central: PMC2848172].
- Gill S, June CH. Going viral: Chimeric antigen receptor T-cell therapy for hematological malignancies. *Immunol Rev.* 2015;263(1):68–89. doi: 10.1111/imr.12243. [PubMed: 25510272].
- Hacohen N, Fritsch EF, Carter TA, Lander ES, Wu CJ. Getting personal with neoantigen-based therapeutic cancer vaccines. *Cancer Immunol Res.* 2013;1(1):11–5. doi: 10.1158/2326-6066.CIR-13-0022. [PubMed: 24777245]. [PubMed Central: PMC4033902].
- Ribas A. Tumor immunotherapy directed at PD-1. N Engl J Med. 2012;366(26):2517–9. doi: 10.1056/NEJMe1205943. [PubMed: 22658126].
- Sharma P, Allison JP. The future of immune checkpoint therapy. Science. 2015;348(6230):56–61. doi: 10.1126/science.aaa8172. [PubMed: 25838373].
- Ansell SM, Lesokhin AM, Borrello I, Halwani A, Scott EC, Gutierrez M, et al. PD-1 blockade with nivolumab in relapsed or refractory Hodgkin's lymphoma. *N Engl J Med*. 2015;**372**(4):311–9. doi: 10.1056/NE-[Moa1411087. [PubMed: 25482239]. [PubMed Central: PMC4348009].
- Borghaei H, Paz-Ares L, Horn L, Spigel DR, Steins M, Ready NE, et al. Nivolumab versus docetaxel in advanced nonsquamous non-smallcell lung cancer. N Engl J Med. 2015;373(17):1627-39. doi: 10.1056/NEJ-Moa1507643. [PubMed: 26412456]. [PubMed Central: PMC5705936].
- Herbst RS, Baas P, Kim DW, Felip E, Perez-Gracia JL, Han JY, et al. Pembrolizumab versus docetaxel for previously treated, PD-L1-positive, advanced non-small-cell lung cancer (KEYNOTE-010): A randomised controlled trial. *Lancet.* 2016;**387**(10027):1540–50. doi: 10.1016/S0140-6736(15)01281-7. [PubMed: 26712084].
- Postow MA, Chesney J, Pavlick AC, Robert C, Grossmann K, McDermott D, et al. Nivolumab and ipilimumab versus ipilimumab in untreated melanoma. *N Engl J Med*. 2015;**372**(21):2006–17. doi: 10.1056/NE-JMoa1414428. [PubMed: 25891304]. [PubMed Central: PMC5744258].
- Rosenberg JE, Hoffman-Censits J, Powles T, van der Heijden MS, Balar AV, Necchi A, et al. Atezolizumab in patients with locally advanced and metastatic urothelial carcinoma who have progressed following treatment with platinum-based chemotherapy: A singlearm, multicentre, phase 2 trial. *Lancet.* 2016;**387**(10031):1909–20. doi: 10.1016/S0140-6736(16)00561-4. [PubMed: 26952546]. [PubMed Central: PMC5480242].
- 24. Seiwert TY, Burtness B, Mehra R, Weiss J, Berger R, Eder JP, et al. Safety and clinical activity of pembrolizumab for treatment of recurrent or metastatic squamous cell carcinoma of the head and neck (KEYNOTE-012): An open-label, multicentre, phase 1b trial. *Lancet Oncol.* 2016;17(7):956–65. doi: 10.1016/S1470-2045(16)30066-3. [PubMed: 27247226].
- Tian T, Olson S, Whitacre JM, Harding A. The origins of cancer robustness and evolvability. *Integr Biol (Camb)*. 2011;3(1):17-30. doi: 10.1039/c0ib00046a. [PubMed: 20944865].
- Alexandrov LB, Nik-Zainal S, Wedge DC, Aparicio SA, Behjati S, Biankin AV, et al. Signatures of mutational processes in human cancer. *Nature*. 2013;**500**(7463):415–21. doi: 10.1038/nature12477. [PubMed: 23945592]. [PubMed Central: PMC3776390].
- Pfeifer GP. Environmental exposures and mutational patterns of cancer genomes. *Genome Med.* 2010;2(8):54. doi: 10.1186/gm175. [PubMed: 20707934]. [PubMed Central: PMC2945011].
- 28. Pena-Diaz J, Bregenhorn S, Ghodgaonkar M, Follonier C, Artola-Boran M, Castor D, et al. Noncanonical mismatch repair as a source of ge-

nomic instability in human cells. *Mol Cell*. 2012;**47**(5):669–80. doi: 10.1016/j.molcel.2012.07.006. [PubMed: 22864113].

- Stratton MR, Campbell PJ, Futreal PA. The cancer genome. *Nature*. 2009;**458**(7239):719–24. doi: 10.1038/nature07943. [PubMed: 19360079]. [PubMed Central: PMC2821689].
- Loeb LA, Bielas JH, Beckman RA. Cancers exhibit a mutator phenotype: Clinical implications. *Cancer Res.* 2008;68(10):3551–7. discussion 3557. doi: 10.1158/0008-5472.CAN-07-5835. [PubMed: 18483233].
- Lengauer C, Kinzler KW, Vogelstein B. Genetic instabilities in human cancers. *Nature*. 1998;**396**(6712):643–9. doi: 10.1038/25292. [PubMed: 9872311].
- Beerenwinkel N, Antal T, Dingli D, Traulsen A, Kinzler KW, Velculescu VE, et al. Genetic progression and the waiting time to cancer. *PLoS Comput Biol.* 2007;3(11). e225. doi: 10.1371/journal.pcbi.0030225. [PubMed: 17997597]. [PubMed Central: PMC2065895].
- Iranzo J, Martincorena I, Koonin EV. Cancer-mutation network and the number and specificity of driver mutations. *Proc Natl Acad Sci* U S A. 2018;115(26):E6010–9. doi: 10.1073/pnas.1803155115. [PubMed: 29895694]. [PubMed Central: PMC6042135].
- Martincorena I, Raine KM, Gerstung M, Dawson KJ, Haase K, Van Loo P, et al. Universal patterns of selection in cancer and somatic tissues. *Cell*. 2018;**173**(7):1823. doi: 10.1016/j.cell.2018.06.001. [PubMed: 29906452]. [PubMed Central: PMC6005233].
- Vogelstein B, Papadopoulos N, Velculescu VE, Zhou S, Diaz LA Jr, Kinzler KW. Cancer genome landscapes. *Science*. 2013;**339**(6127):1546–58. doi: 10.1126/science.1235122. [PubMed: 23539594]. [PubMed Central: PMC3749880].
- Coulie PG, Van den Eynde BJ, van der Bruggen P, Boon T. Tumour antigens recognized by T lymphocytes: At the core of cancer immunotherapy. *Nat Rev Cancer*. 2014;14(2):135–46. doi: 10.1038/nrc3670. [PubMed: 24457417].
- Dunn GP, Bruce AT, Ikeda H, Old LJ, Schreiber RD. Cancer immunoediting: From immunosurveillance to tumor escape. *Nat Immunol.* 2002;3(11):991-8. doi: 10.1038/ni1102-991. [PubMed: 12407406].
- Mullard A. New checkpoint inhibitors ride the immunotherapy tsunami. Nat Rev Drug Discov. 2013;12(7):489–92. doi: 10.1038/nrd4066. [PubMed: 23812256].
- Ikeda H, Ohta N, Furukawa K, Miyazaki H, Wang L, Kuribayashi K, et al. Mutated mitogen-activated protein kinase: A tumor rejection antigen of mouse sarcoma. *Proc Natl Acad Sci U S A*. 1997;**94**(12):6375– 9. doi: 10.1073/pnas.94.12.6375. [PubMed: 9177225]. [PubMed Central: PMC21057].
- Kreiter S, Vormehr M, van de Roemer N, Diken M, Lower M, Diekmann J, et al. Mutant MHC class II epitopes drive therapeutic immune responses to cancer. *Nature*. 2015;**520**(7549):692–6. doi: 10.1038/nature14426. [PubMed: 25901682]. [PubMed Central: PMC4838069].
- Yadav M, Jhunjhunwala S, Phung QT, Lupardus P, Tanguay J, Bumbaca S, et al. Predicting immunogenic tumour mutations by combining mass spectrometry and exome sequencing. *Nature*. 2014;**515**(7528):572–6. doi:10.1038/nature14001. [PubMed: 25428506].
- Chen DS, Mellman I. Elements of cancer immunity and the cancerimmune set point. *Nature*. 2017;**541**(7637):321-30. doi: 10.1038/nature21349. [PubMed: 28102259].
- Castle JC, Kreiter S, Diekmann J, Lower M, van de Roemer N, de Graaf J, et al. Exploiting the mutanome for tumor vaccination. *Cancer Res.* 2012;**72**(5):1081–91. doi: 10.1158/0008-5472.CAN-11-3722. [PubMed: 22237626].
- 44. Tran E, Turcotte S, Gros A, Robbins PF, Lu YC, Dudley ME, et al. Cancer immunotherapy based on mutation-specific CD4+T cells in a patient with epithelial cancer. *Science*. 2014;**344**(6184):641–5. doi: 10.1126/science.1251102. [PubMed: 24812403].
- Linnemann C, van Buuren MM, Bies L, Verdegaal EM, Schotte R, Calis JJ, et al. High-throughput epitope discovery reveals frequent recognition of neo-antigens by CD4+ T cells in human melanoma. *Nat Med.* 2015;**21**(1):81–5. doi: 10.1038/nm.3773. [PubMed: 25531942].

- Robbins PF, Lu YC, El-Gamil M, Li YF, Gross C, Gartner J, et al. Mining exomic sequencing data to identify mutated antigens recognized by adoptively transferred tumor-reactive T cells. *Nat Med.* 2013;**19**(6):747-52. doi: 10.1038/nm.3161. [PubMed: 23644516]. [PubMed Central: PMC3757932].
- Rosenberg SA, Yang JC, Sherry RM, Kammula US, Hughes MS, Phan GQ, et al. Durable complete responses in heavily pretreated patients with metastatic melanoma using T-cell transfer immunotherapy. *Clin Cancer Res.* 2011;**17**(13):4550–7. doi: 10.1158/1078-0432.CCR-11-0116. [PubMed: 21498393]. [PubMed Central: PMC3131487].
- Besser MJ, Shapira-Frommer R, Treves AJ, Zippel D, Itzhaki O, Hershkovitz L, et al. Clinical responses in a phase II study using adoptive transfer of short-term cultured tumor infiltration lymphocytes in metastatic melanoma patients. *Clin Cancer Res.* 2010;**16**(9):2646–55. doi: 10.1158/1078-0432.CCR-10-0041. [PubMed: 20406835].
- Dudley ME, Wunderlich JR, Robbins PF, Yang JC, Hwu P, Schwartzentruber DJ, et al. Cancer regression and autoimmunity in patients after clonal repopulation with antitumor lymphocytes. *Science*. 2002;**298**(5594):850–4. doi: 10.1126/science.1076514. [PubMed: 12242449]. [PubMed Central: PMC1764179].
- Wolfel T, Hauer M, Schneider J, Serrano M, Wolfel C, Klehmann-Hieb E, et al. A p16INK4a-insensitive CDK4 mutant targeted by cytolytic T lymphocytes in a human melanoma. *Science*. 1995;**269**(5228):1281–4. doi: 10.1126/science.7652577. [PubMed: 7652577].
- Schumacher T, Bunse L, Pusch S, Sahm F, Wiestler B, Quandt J, et al. A vaccine targeting mutant IDH1 induces antitumour immunity. *Nature*. 2014;**512**(7514):324–7. doi: 10.1038/nature13387. [PubMed: 25043048].
- Schumacher TN, Schreiber RD. Neoantigens in cancer immunotherapy. Science. 2015;348(6230):69–74. doi: 10.1126/science.aaa4971. [PubMed: 25838375].
- Sharma P, Hu-Lieskovan S, Wargo JA, Ribas A. Primary, adaptive, and acquired resistance to cancer immunotherapy. *Cell*. 2017;**168**(4):707– 23. doi: 10.1016/j.cell.2017.01.017. [PubMed: 28187290]. [PubMed Central: PMC5391692].
- 54. Tabarestani S, Ghaderian SM, Rezvani H, Mirfakhraie R, Ebrahimi A, Attarian H, et al. Prognostic and predictive value of copy number alterations in invasive breast cancer as determined by multiplex ligation-dependent probe amplification. *Cell Oncol (Dordr)*. 2014;**37**(2):107-18. doi: 10.1007/s13402-013-0165-1. [PubMed: 24573687].
- Tabarestani S, Ghaderian SM, Rezvani H, Mirfakhraie R. Expression profiling of breast cancer patients treated with tamoxifen: Prognostic or predictive significance. *Med Oncol.* 2014;**31**(4):896. doi: 10.1007/s12032-014-0896-5. [PubMed: 24563328].
- Tabarestani S, Motallebi M, Akbari ME. Are estrogen receptor genomic aberrations predictive of hormone therapy response in breast cancer? *Iran J Cancer Prev.* 2016;9(4). e6565. doi: 10.17795/ijcp-6565. [PubMed: 27761212]. [PubMed Central: PMC5056018].
- Tabarestani S, Motallebi M, Akbari ME, Malekzadeh Moghani Mona, Shojaee Leyla. Analysis of BRCA1/2 mutations and performance of manchester scoring system in high risk Iranian breast cancer patients: A pilot study. Int J Cancer Manag. 2017;10(12). doi: 10.5812/ijcm.60392.
- Gubin MM, Zhang X, Schuster H, Caron E, Ward JP, Noguchi T, et al. Checkpoint blockade cancer immunotherapy targets tumour-specific mutant antigens. *Nature*. 2014;515(7528):577-81. doi: 10.1038/nature13988. [PubMed: 25428507]. [PubMed Central:

PMC4279952].

- Sucker A, Zhao F, Real B, Heeke C, Bielefeld N, Mabetaen S, et al. Genetic evolution of T-cell resistance in the course of melanoma progression. *Clin Cancer Res.* 2014;**20**(24):6593–604. doi: 10.1158/1078-0432.CCR-14-0567. [PubMed: 25294904].
- Garon EB, Rizvi NA, Hui R, Leighl N, Balmanoukian AS, Eder JP, et al. Pembrolizumab for the treatment of non-small-cell lung cancer. N Engl J Med. 2015;372(21):2018–28. doi: 10.1056/NEJMoa1501824. [PubMed: 25891174].
- Herbst RS, Soria JC, Kowanetz M, Fine GD, Hamid O, Gordon MS, et al. Predictive correlates of response to the anti-PD-L1 antibody MPDL3280A in cancer patients. *Nature*. 2014;**515**(7528):563– 7. doi: 10.1038/nature14011. [PubMed: 25428504]. [PubMed Central: PMC4836193].
- Reck M, Rodriguez-Abreu D, Robinson AG, Hui R, Csoszi T, Fulop A, et al. Pembrolizumab versus chemotherapy for PD-L1-positive non-small-cell lung cancer. N Engl J Med. 2016;375(19):1823-33. doi: 10.1056/NEJMoa1606774. [PubMed: 27718847].
- Yarchoan M, Hopkins A, Jaffee EM. Tumor mutational burden and response rate to PD-1 inhibition. *N Engl J Med*. 2017;**377**(25):2500–1. doi: 10.1056/NEJMc1713444. [PubMed: 29262275].
- 64. Snyder A, Makarov V, Merghoub T, Yuan J, Zaretsky JM, Desrichard A, et al. Genetic basis for clinical response to CTLA-4 blockade in melanoma. N Engl J Med. 2014;**371**(23):2189–99. doi: 10.1056/NEJ-Moa1406498. [PubMed: 25409260]. [PubMed Central: PMC4315319].
- Van Allen EM, Miao D, Schilling B, Shukla SA, Blank C, Zimmer L, et al. Genomic correlates of response to CTLA-4 blockade in metastatic melanoma. *Science*. 2015;**350**(6257):207–11. doi: 10.1126/science.aad0095. [PubMed: 26359337]. [PubMed Central: PMC5054517].
- Hugo W, Zaretsky JM, Sun L, Song C, Moreno BH, Hu-Lieskovan S, et al. Genomic and transcriptomic features of response to anti-PD-1 therapy in metastatic melanoma. *Cell.* 2016;**165**(1):35–44. doi: 10.1016/j.cell.2016.02.065. [PubMed: 26997480]. [PubMed Central: PMC4808437].
- Rizvi NA, Hellmann MD, Snyder A, Kvistborg P, Makarov V, Havel JJ, et al. Cancer immunology. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. *Science*. 2015;**348**(6230):124–8. doi: 10.1126/science.aaa1348. [PubMed: 25765070]. [PubMed Central: PMC4993154].
- Spigel DR, Schrock AB, Fabrizio D, Frampton GM, Sun J, He J, et al. Total mutation burden (TMB) in lung cancer (LC) and relationship with response to PD-1/PD-L1 targeted therapies. J Clin Oncol. 2016;34(15_suppl):9017. doi: 10.1200/JCO.2016.34.15_suppl.9017.
- Le DT, Uram JN, Wang H, Bartlett BR, Kemberling H, Eyring AD, et al. PD-1 blockade in tumors with mismatch-repair deficiency. *N Engl J Med*. 2015;**372**(26):2509–20. doi: 10.1056/NEJMoa1500596. [PubMed: 26028255]. [PubMed Central: PMC4481136].
- Gerlinger M, Rowan AJ, Horswell S, Math M, Larkin J, Endesfelder D, et al. Intratumor heterogeneity and branched evolution revealed by multiregion sequencing. *N Engl J Med*. 2012;**366**(10):883–92. doi: 10.1056/NEJM0a1113205. [PubMed: 22397650]. [PubMed Central: PMC4878653].
- Le DT, Durham JN, Smith KN, Wang H, Bartlett BR, Aulakh LK, et al. Mismatch repair deficiency predicts response of solid tumors to PD-1 blockade. *Science*. 2017;**357**(6349):409–13. doi: 10.1126/science.aan6733. [PubMed: 28596308]. [PubMed Central: PMC5576142].