



The Effect of Testosterone and Estradiol on Renal Function Markers in Two Protocols of Cisplatin Induced Nephrotoxicity Models in Surgically Orchiectomized and Ovariectomized Rats

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Received 2019 January 12; Revised 2019 March 06; Accepted 2019 March 16.

Abstract

Background: The major side effect of cisplatin (CP) therapy in patients with cancer is nephrotoxicity, which limits the treatment. In two protocols of CP treatments, we tested 3 weeks of hormone therapy against CP induced nephrotoxicity in bilateral orchiectomized (OR) and ovariectomized (OV) rats.

Methods: A total of 101 OR and OV rats were subjected to receive 3 weeks of testosterone (Ts, 10 mg/kg/week) and estradiol (Es, 250 µg/kg/week), respectively, followed by two protocols of CP therapies; continuous (divided) doses (3 mg/kg/day for period of 5 days) as protocol 1 and single dose (7.5 mg/kg) as protocol 2. The measurements were performed by the end of the 4th week.

Results: CP increased the serum levels of blood urea nitrogen (BUN) and creatinine (Cr) in both OR and OV rats, which were related to the protocols of CP treatments. Es (not Ts) in protocol 2 attenuated the serum levels of BUN and Cr. Ts significantly increased body weight loss in protocol 1 compared with control group ($P < 0.05$). Es did not attenuate kidney tissue damage score (KTDS) in protocol 1 treated animal, but KTDS was decreased by Es in protocol 2 treated Rats. Hormone replacement therapy did not alter Cr clearance compared with control group.

Conclusions: It seems that hormone therapy could not protect the kidney against CP induced nephrotoxicity.

Keywords: Testosterone, Estradiol, Cisplatin, Nephrotoxicity, Rats

1. Background

Cisplatin (CP); $\text{cis-(PtII(NH}_3)_2\text{Cl}_2)$ is known as an anti-cancer agent for treatment of solid tumors (1, 2). Nephrotoxicity is the most important side effect of CP (3) due to its accumulation in epithelial cells of tubules (4). The animals treated with single dose of CP showed an increase in the serum level of creatinine (Cr), blood urea nitrogen (BUN), kidney damage, and kidney weights (KW) (5-8). CP also causes inflammation, stress oxidative, and apoptosis in cells (9). Previously, we showed that CP induced nephrotoxicity is gender-related (2, 10, 11). The result of some animals' experiments indicated that estradiol (Es) can intensify and testosterone (Ts) may improve CP induced nephrotoxicity (12, 13). To consider renal function, it is reported that CP reduced glomerular filtration rate (GFR) (14) and sodium excretion was different between male and female rats treated with CP (15). In addition, the renal clearance in animals treated with CP is also age-related (16). CP induced

nephrotoxicity also related to platinum-based and its accumulation in the kidney (17, 18). The side effects of this drug also are dose-related (19). The single dose and the divided dose of CP therapy may alter the accumulation of CP in the kidney. Both single dose and the divided dose of CP therapy were considered in clinic for toxicity profile, and indicated the possible less kidney toxicity and more magnesium loss by single dose therapy (20); however, different results was found by others (21).

2. Objectives

Therefore, the high dose of CP treatment in divided manner may have different toxicity profile. Accordingly, we designed two protocols of CP therapy, and the effects of sex hormone on renal function markers after CP therapy were considered.

3. Methods

The male and female (180 - 250 g) Wistar rats (Animal center, Isfahan University of Medical Science, Isfahan, Iran) were used in this study. The animals were housed under conventional and controlled conditions (12/12 light/dark cycles; 23 - 25°C). All experiments were approved by the Ethics Committee of Isfahan University of Medical Sciences (IR.MUI.REC.1394.2.239).

3.1. Male Animals

The male rats were anesthetized with chloral hydrate injection (450 mg/kg; ip). A midline incision was made in the sub-abdominal region, and the epididymis and testis were pulled out and removed. One week later, they were randomly allocated to 6 experimental groups for 2 protocols of CP treatments; there were 3 groups in each protocol.

3.1.1. Protocol 1: Continues (Divided) Dose of CP Treatment

Group 1 (named vehicle, n = 6): The bilateral orchiectomized (OR) rats received sesame oil at the beginning of each week intramuscularly as vehicle for 3 weeks, and then sacrificed by the end of the 3rd week.

Group 2 (named CP, n = 4): The OR rats received the same regimen as group 1 except that CP (3 mg.kg/day, i.p) for 5 days was administrated during the 3rd week.

Group 3 (named Ts + CP, n = 6): The OR rats received Ts enanthate (10 mg/kg/week, IM, Aburaihan Co., Tehran, Iran) dissolved in sesame oil at the beginning of each week intramuscularly for 3 weeks in addition to CP for 5 days during the 3rd week, and then sacrificed by the end of 3rd week.

3.1.2. Protocol 2: Single doses of CP treatment

The protocol 2 included 3 groups of 4 (n = 5), 5 (n = 6), and 6 (n = 5). The only difference between these groups and the groups in protocol 1 (groups 1-3) was the dose of CP. The OR rats in protocol 2 received the same regimen as group 1, 2 and 3, but at the beginning of the 3rd week, they received a single dose of CP (7.5 mg.kg/day, i.p), and then sacrificed by the end of the 3rd week. The treatment dose of CP was based on our previous researches (5, 6).

3.2. Female Animals

The female rats were anesthetized with chloral hydrate injection (450 mg/kg; ip). After midline incision in the sub-abdominal region, the ovaries were removed. One week later and similar to male rats, they were randomly allocated to 6 experimental groups for 2 protocols of CP treatments; there were 3 groups in each protocol.

Groups 7 - 12 (n = 7, 6, 5, 6, 7, 6 respectively): The bilateral ovariectomized (OV) rats received the same regimen

as group 1 to 6, respectively, except Es (250 µg/kg/week, IM, Aburaihan Co., Tehran, Iran) instead of Ts. The selected dose of Es was based on previous work (12).

All the animals were placed in metabolic cages for 6 hours before the end of experiment to collect urine output. Then, the blood samples were obtained, and the animals were sacrificed humanly. The kidneys were excised and weighted immediately. The left kidney was used for histopathology investigations via hematoxylin and eosin (H&E) staining. The renal damage was assigned as kidney tissue damage score (KTDS), and it was scored from 1 to 4, while zero score was recorded for normal tissue (7, 8). The right kidney was homogenized and centrifuged. The serum levels of creatinine (Cr) and blood urea nitrogen (BUN) and the urine level of Cr were determined, using quantitative diagnostic kits (pars Azmoon, Iran) by auto-analyzer (Technicon, Ireland LTD). The serum level of nitrite was determined by Griess method as described before (22). Briefly, the sulfanilamide solution was added to the samples and the mixture was incubated. Then, N-(1-naphthyl) ethylenediamine dihydrochloride solution was added, and the absorbance was determined at a wavelength of 540 nm. The nitrite concentration was calculated compared to the nitrite standard curve. The level of malondialdehyde (MDA) was measured manually (23). Briefly, 1 mL of 10% trichloroacetic acid (TCA) was added into 0.5 mL of the sample, and centrifuged at 2000 g for 10 minutes. Then 0.5 mL of the supernatant was mixed with 0.5 mL of 0.67% thiobarbituric acid (TBA), and after 10 min of incubating in the boiling water, the absorbance was determined at the wavelength of 532 nm after cooling.

3.3. Statistical Analysis

The data are presented as Mean ± SEM. The quantitative data were compared by one-way ANOVA, using LSD. The Mann-Whitney test was applied to compare the KTDS among the groups. $P \leq 0.05$ was considered as statistically significant.

4. Results

CP increased the serum levels of BUN and Cr in both male and female rats. However, Ts did not attenuate the levels of these markers toward normal values (Figure 1) in male rats, but Es in protocol 2 attenuated the serum levels of BUN and Cr. The KW and KTDS were increased and the body weight was decreased by CP, but Ts significantly increased body weight loss in protocol 1 compared with CP treated group ($P < 0.05$). Es did not attenuate KTDS in protocol 1 treated animals, but KTDS was decreased by Es

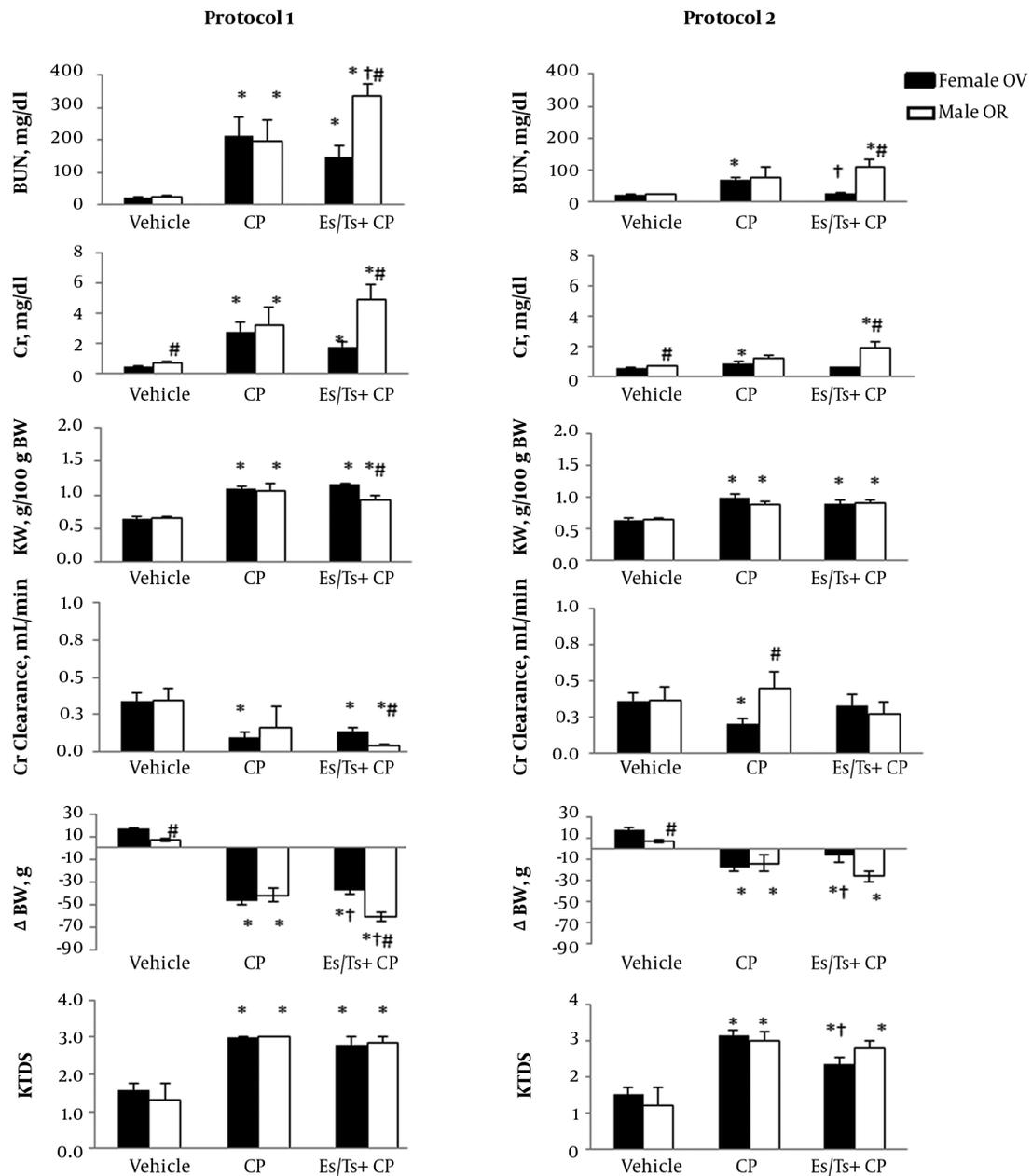


Figure 1. The serum levels of blood nitrogen urea (BUN) and creatinine (Cr), and kidney weight (KW) per body weight (BW), Cr-clearance, BW changes (Δ BW) and kidney tissue damage score (KTDS) in ovariectomized (OV) female and orchietomized (OR) male rats received two protocols of cisplatin (CP) treatments (see text for protocols detail) with and without estradiol (Es) or testosterone (Ts). * Represents significant difference from vehicle group in the same gender ($P < 0.05$). † Indicates significant difference from CP group in the same gender ($P < 0.05$). # Shows significant difference between genders that receive similar treatment ($P < 0.05$).

ment with those studies, the current finding did not indicate a nephro-protective role on renal function against CP induced nephrotoxicity. It seems that Es is a trigger to promote proximal tubules' toxicity (30, 31), and even the

antioxidant effect of Es (32, 33) did not attenuate the renal function disturbance induced by CP therapy.

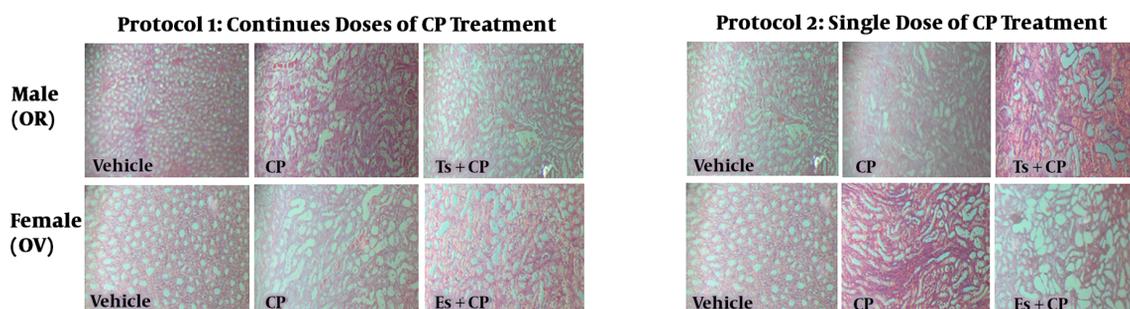


Figure 2. The samples images ($\times 100$) of kidney tissues in all experimental groups. See text for groups' detail

5.1. Conclusions

It is concluded that 3 weeks of hormone therapy in animal model may not protect the kidney against CP induced nephrotoxicity. However, the renal toxicity intensity is related to the protocol of CP treatments, and single dose treatment may prefer, because its accumulation in the kidney is the main cause of renal toxicity (18).

Acknowledgments

This research was supported by Isfahan University of Medical Sciences (grant #294239).

Footnotes

Authors' Contribution: Bahar Mazaheri, Alireza Samimiati and Mohammad-Sedigh Khosravi conducted the experimental procedure and helped in data analysis. Ardeshir Talebi conducted the pathology procedure and analysis. Mehdi Nematbakhsh conducted the study design, data analysis and preparing the article

Conflict of Interests: The authors have declared that no conflict of interest exists.

Ethical Approval: All experiments were approved by the Isfahan University of Medical Sciences Ethics Committee (IR.MUI.REC.1394.2.239).

Financial Disclosure: None declared.

Funding/Support: This research was supported by Isfahan University of Medical Sciences (grant #294239).

References

1. Florea AM, Busselberg D. Cisplatin as an anti-tumor drug: Cellular mechanisms of activity, drug resistance and induced side effects. *Cancers (Basel)*. 2011;**3**(1):1351-71. doi: [10.3390/cancers3011351](https://doi.org/10.3390/cancers3011351). [PubMed: [24212665](https://pubmed.ncbi.nlm.nih.gov/24212665/)]. [PubMed Central: [PMC3756417](https://pubmed.ncbi.nlm.nih.gov/PMC3756417/)].
2. Nematbakhsh M, Pezeshki Z, Eshraghi Jazi F, Mazaheri B, Moeini M, Safari T, et al. Cisplatin-induced nephrotoxicity; protective supplements and gender differences. *Asian Pac J Cancer Prev*. 2017;**18**(2):295-314. doi: [10.22034/APJCP.2017.18.2.295](https://doi.org/10.22034/APJCP.2017.18.2.295). [PubMed: [28345324](https://pubmed.ncbi.nlm.nih.gov/28345324/)]. [PubMed Central: [PMC5454720](https://pubmed.ncbi.nlm.nih.gov/PMC5454720/)].
3. Miller RP, Tadagavadi RK, Ramesh G, Reeves WB. Mechanisms of Cisplatin nephrotoxicity. *Toxins (Basel)*. 2010;**2**(11):2490-518. doi: [10.3390/toxins2112490](https://doi.org/10.3390/toxins2112490). [PubMed: [22069563](https://pubmed.ncbi.nlm.nih.gov/22069563/)]. [PubMed Central: [PMC3153174](https://pubmed.ncbi.nlm.nih.gov/PMC3153174/)].
4. Bagnis C, Beaufils H, Jacquiaud C, Adabra Y, Jouanneau C, Le Nahour G, et al. Erythropoietin enhances recovery after cisplatin-induced acute renal failure in the rat. *Nephrol Dial Transplant*. 2001;**16**(5):932-8. doi: [10.1093/ndt/16.5.932](https://doi.org/10.1093/ndt/16.5.932). [PubMed: [11328897](https://pubmed.ncbi.nlm.nih.gov/11328897/)].
5. Eshraghi-Jazi F, Nematbakhsh M, Nasri H, Talebi A, Haghighi M, Pezeshki Z, et al. The protective role of endogenous nitric oxide donor (L-arginine) in cisplatin-induced nephrotoxicity: Gender related differences in rat model. *J Res Med Sci*. 2011;**16**(11):1389-96. [PubMed: [22973338](https://pubmed.ncbi.nlm.nih.gov/22973338/)]. [PubMed Central: [PMC3430054](https://pubmed.ncbi.nlm.nih.gov/PMC3430054/)].
6. Nematbakhsh M, Ashrafi F, Safari T, Talebi A, Nasri H, Mortazavi M, et al. Administration of vitamin E and losartan as prophylaxes in cisplatin-induced nephrotoxicity model in rats. *J Nephrol*. 2012;**25**(3):410-7. doi: [10.5301/jn.5000018](https://doi.org/10.5301/jn.5000018). [PubMed: [21928232](https://pubmed.ncbi.nlm.nih.gov/21928232/)].
7. Nematbakhsh M, Ashrafi F, Pezeshki Z, Fatahi Z, Kianpoor F, Sanei MH, et al. A histopathological study of nephrotoxicity, hepatotoxicity or testicular toxicity: Which one is the first observation as side effect of Cisplatin-induced toxicity in animal model? *J Nephropathol*. 2012;**1**(3):190-3. doi: [10.5812/nephropathol.8122](https://doi.org/10.5812/nephropathol.8122). [PubMed: [24475415](https://pubmed.ncbi.nlm.nih.gov/24475415/)]. [PubMed Central: [PMC3886150](https://pubmed.ncbi.nlm.nih.gov/PMC3886150/)].
8. Nematbakhsh M, Ashrafi F, Nasri H, Talebi A, Pezeshki Z, Eshraghi F, et al. A model for prediction of cisplatin induced nephrotoxicity by kidney weight in experimental rats. *J Res Med Sci*. 2013;**18**(5):370-3. [PubMed: [24174938](https://pubmed.ncbi.nlm.nih.gov/24174938/)]. [PubMed Central: [PMC3810567](https://pubmed.ncbi.nlm.nih.gov/PMC3810567/)].
9. Jo SK, Cho WY, Sung SA, Kim HK, Won NH. MEK inhibitor, U0126, attenuates cisplatin-induced renal injury by decreasing inflammation and apoptosis. *Kidney Int*. 2005;**67**(2):458-66. doi: [10.1111/j.1523-1755.2005.67102.x](https://doi.org/10.1111/j.1523-1755.2005.67102.x). [PubMed: [15673293](https://pubmed.ncbi.nlm.nih.gov/15673293/)].
10. Nematbakhsh M, Ebrahimian S, Toyserkani M, Eshraghi-Jazi F, Talebi A, Ashrafi F. Gender difference in Cisplatin-induced nephrotoxicity in a rat model: Greater intensity of damage in male than female. *Nephrourol Mon*. 2013;**5**(3):818-21. doi: [10.5812/numonthly.10128](https://doi.org/10.5812/numonthly.10128). [PubMed: [24282792](https://pubmed.ncbi.nlm.nih.gov/24282792/)]. [PubMed Central: [PMC3830908](https://pubmed.ncbi.nlm.nih.gov/PMC3830908/)].
11. Nematbakhsh M, Talebi A, Nasri H, Safari T. Some evidence for sex-based differences in cisplatin-induced nephrotoxicity in rats. *Clin Experiment Med Lett*. 2012;**53**:29-32.

12. Pezeshki Z, Nematbakhsh M, Nasri H, Talebi A, Pilehvarian AA, Safari T, et al. Evidence against protective role of sex hormone estrogen in Cisplatin-induced nephrotoxicity in ovariectomized rat model. *Toxicol Int.* 2013;**20**(1):43-7. doi: [10.4103/0971-6580.111568](https://doi.org/10.4103/0971-6580.111568). [PubMed: [23833437](https://pubmed.ncbi.nlm.nih.gov/23833437/)]. [PubMed Central: [PMC3702126](https://pubmed.ncbi.nlm.nih.gov/PMC3702126/)].
13. Rostami B, Nematbakhsh M, Pezeshki Z, Talebi A, Sharifi MR, Moslemi F, et al. Effect of testosterone on Cisplatin-induced nephrotoxicity in surgically castrated rats. *Nephrourol Mon.* 2014;**6**(5): e21546. doi: [10.5812/numonthly.21546](https://doi.org/10.5812/numonthly.21546). [PubMed: [25695037](https://pubmed.ncbi.nlm.nih.gov/25695037/)]. [PubMed Central: [PMC4318011](https://pubmed.ncbi.nlm.nih.gov/PMC4318011/)].
14. Daniel G, Hahn K, Bravo L, Legendre A. The effect of a single therapeutic dose of cisplatin on GFR in dogs. *Oncol Rep.* 1997;**4**(1):153-6. doi: [10.3892/or.4.1.153](https://doi.org/10.3892/or.4.1.153). [PubMed: [21590032](https://pubmed.ncbi.nlm.nih.gov/21590032/)].
15. Stakisaitis D, Dudeniene G, Jankunas RJ, Grazeliene G, Didziapetriene J, Pundziene B. Cisplatin increases urinary sodium excretion in rats: Gender-related differences. *Medicina (Kaunas).* 2010;**46**(1):45-50. doi: [10.3390/medicina46010008](https://doi.org/10.3390/medicina46010008). [PubMed: [20234163](https://pubmed.ncbi.nlm.nih.gov/20234163/)].
16. Pezeshki Z, Maleki M, Talebi A, Nematbakhsh M. Age and gender related renal side effects of cisplatin in animal model. *Asian Pac J Cancer Prev.* 2017;**18**(6):1703-5. doi: [10.22034/APJCP.2017.18.6.1703](https://doi.org/10.22034/APJCP.2017.18.6.1703). [PubMed: [28670892](https://pubmed.ncbi.nlm.nih.gov/28670892/)].
17. Wong E, Giandomenico CM. Current status of platinum-based anti-tumor drugs. *Chem Rev.* 1999;**99**(9):2451-66. doi: [10.1021/cr980420v](https://doi.org/10.1021/cr980420v). [PubMed: [11749486](https://pubmed.ncbi.nlm.nih.gov/11749486/)].
18. Esteban-Fernandez D, Verdaguer JM, Ramirez-Camacho R, Palacios MA, Gomez-Gomez MM. Accumulation, fractionation, and analysis of platinum in toxicologically affected tissues after cisplatin, oxaliplatin, and carboplatin administration. *J Anal Toxicol.* 2008;**32**(2):140-6. doi: [10.1093/jat/32.2.140](https://doi.org/10.1093/jat/32.2.140). [PubMed: [18334097](https://pubmed.ncbi.nlm.nih.gov/18334097/)].
19. Soni KK, Kim HK, Choi BR, Karna KK, You JH, Cha JS, et al. Dose-dependent effects of cisplatin on the severity of testicular injury in Sprague Dawley rats: Reactive oxygen species and endoplasmic reticulum stress. *Drug Des Devel Ther.* 2016;**10**:3959-68. doi: [10.2147/DDDT.S120014](https://doi.org/10.2147/DDDT.S120014). [PubMed: [28003740](https://pubmed.ncbi.nlm.nih.gov/28003740/)]. [PubMed Central: [PMC5161341](https://pubmed.ncbi.nlm.nih.gov/PMC5161341/)].
20. Zhang IF, Li T, Hu XC. Toxicity profile of cisplatin given at one full dose versus three divided doses for three consecutive days. *J Clin Oncol.* 2016;**34**(15_suppl).
21. Ahmadzadeh A, Shahbazian H, Safapour N, Tulabi M, Zandifar S. Comparison between the effects of one-day treatment regimen with cisplatin on renal function and various biochemical parameters in patients with gastric and lung cancer compared with two-days divided cisplatin treatment regimen. *J Renal Inj Prev.* 2015;**4**(3):87-91. doi: [10.12861/jrip.2015.17](https://doi.org/10.12861/jrip.2015.17). [PubMed: [26468480](https://pubmed.ncbi.nlm.nih.gov/26468480/)]. [PubMed Central: [PMC4594219](https://pubmed.ncbi.nlm.nih.gov/PMC4594219/)].
22. Bryan NS, Grisham MB. Methods to detect nitric oxide and its metabolites in biological samples. *Free Radic Biol Med.* 2007;**43**(5):645-57. doi: [10.1016/j.freeradbiomed.2007.04.026](https://doi.org/10.1016/j.freeradbiomed.2007.04.026). [PubMed: [17664129](https://pubmed.ncbi.nlm.nih.gov/17664129/)]. [PubMed Central: [PMC2041919](https://pubmed.ncbi.nlm.nih.gov/PMC2041919/)].
23. Mazaheri S, Nematbakhsh M, Bahadorani M, Pezeshki Z, Talebi A, Ghannadi AR, et al. Effects of fennel essential oil on cisplatin-induced nephrotoxicity in ovariectomized rats. *Toxicol Int.* 2013;**20**(2):138-45. doi: [10.4103/0971-6580.117256](https://doi.org/10.4103/0971-6580.117256). [PubMed: [24082507](https://pubmed.ncbi.nlm.nih.gov/24082507/)]. [PubMed Central: [PMC3783680](https://pubmed.ncbi.nlm.nih.gov/PMC3783680/)].
24. Garcia MM, Acquier A, Suarez G, Gomez NV, Gorostizaga A, Mendez CF, et al. Cisplatin inhibits testosterone synthesis by a mechanism that includes the action of reactive oxygen species (ROS) at the level of P450scc. *Chem Biol Interact.* 2012;**199**(3):185-91. doi: [10.1016/j.cbi.2012.08.012](https://doi.org/10.1016/j.cbi.2012.08.012). [PubMed: [22940207](https://pubmed.ncbi.nlm.nih.gov/22940207/)].
25. Park KM, Kim JI, Ahn Y, Bonventre AJ, Bonventre JV. Testosterone is responsible for enhanced susceptibility of males to ischemic renal injury. *J Biol Chem.* 2004;**279**(50):52282-92. doi: [10.1074/jbc.M407629200](https://doi.org/10.1074/jbc.M407629200). [PubMed: [15358759](https://pubmed.ncbi.nlm.nih.gov/15358759/)].
26. Verzola D, Gandolfo MT, Salvatore F, Villaggio B, Gianiorio F, Traverso P, et al. Testosterone promotes apoptotic damage in human renal tubular cells. *Kidney Int.* 2004;**65**(4):1252-61. doi: [10.1111/j.1523-1755.2004.00497.x](https://doi.org/10.1111/j.1523-1755.2004.00497.x). [PubMed: [15086464](https://pubmed.ncbi.nlm.nih.gov/15086464/)].
27. Mooradian AD. Antioxidant properties of steroids. *J Steroid Biochem Mol Biol.* 1993;**45**(6):509-11. doi: [10.1016/0960-0760\(93\)90166-T](https://doi.org/10.1016/0960-0760(93)90166-T). [PubMed: [8518206](https://pubmed.ncbi.nlm.nih.gov/8518206/)].
28. Beleh MA, Lin YC, Brueggemeier RW. Estrogen metabolism in microsomal, cell, and tissue preparations of kidney and liver from Syrian hamsters. *J Steroid Biochem Mol Biol.* 1995;**52**(5):479-89. doi: [10.1016/0960-0760\(95\)00003-I](https://doi.org/10.1016/0960-0760(95)00003-I). [PubMed: [7748813](https://pubmed.ncbi.nlm.nih.gov/7748813/)].
29. Nematbakhsh M, Pezeshki Z, Eshraghi-Jazi F, Ashrafi F, Nasri H, Talebi A, et al. Vitamin E, vitamin C, or losartan is not nephroprotectant against cisplatin-induced nephrotoxicity in presence of estrogen in ovariectomized rat model. *Int J Nephrol.* 2012;**2012**:284896. doi: [10.1155/2012/284896](https://doi.org/10.1155/2012/284896). [PubMed: [23056943](https://pubmed.ncbi.nlm.nih.gov/23056943/)]. [PubMed Central: [PMC3463913](https://pubmed.ncbi.nlm.nih.gov/PMC3463913/)].
30. Roy D, Liehr JG. Target organ-specific inactivation of drug metabolizing enzymes in kidney of hamsters treated with estradiol. *Mol Cell Biochem.* 1992;**110**(1):31-9. doi: [10.1007/BF02385003](https://doi.org/10.1007/BF02385003). [PubMed: [1315925](https://pubmed.ncbi.nlm.nih.gov/1315925/)].
31. Butterworth M, Lau SS, Monks TJ. 2-Hydroxy-4-glutathion-S-yl-17beta-estradiol and 2-hydroxy-1-glutathion-S-yl-17beta-estradiol produce oxidative stress and renal toxicity in an animal model of 17beta-estradiol-mediated nephrocarcinogenicity. *Carcinogenesis.* 1998;**19**(1):133-9. doi: [10.1093/carcin/19.1.133](https://doi.org/10.1093/carcin/19.1.133). [PubMed: [9472704](https://pubmed.ncbi.nlm.nih.gov/9472704/)].
32. Song JY, Kim MJ, Jo HH, Hwang SJ, Chae B, Chung JE, et al. Antioxidant effect of estrogen on bovine aortic endothelial cells. *J Steroid Biochem Mol Biol.* 2009;**117**(1-3):74-80. doi: [10.1016/j.jsbmb.2009.07.006](https://doi.org/10.1016/j.jsbmb.2009.07.006). [PubMed: [19635556](https://pubmed.ncbi.nlm.nih.gov/19635556/)].
33. Mann V, Huber C, Kogianni G, Collins F, Noble B. The antioxidant effect of estrogen and selective estrogen receptor modulators in the inhibition of osteocyte apoptosis in vitro. *Bone.* 2007;**40**(3):674-84. doi: [10.1016/j.bone.2006.10.014](https://doi.org/10.1016/j.bone.2006.10.014). [PubMed: [17174166](https://pubmed.ncbi.nlm.nih.gov/17174166/)].