Published online 2017 March 26.

Research Article

Clinical Value of Human Leucocyte Antigen G (HLA-G) Expression in the Prognosis of Colorectal Cancer

Roghaieh Samadi,¹ Ehsan Nazemalhosseini Mojarad,² Mahsa Molaei,² Faranak Kazerouni,¹ Hamid

Asadzadeh Aghdaei,² Masoumeh Navidinia,¹ and Ali Rahimipour^{1,*}

¹Department of Medical Laboratory Technology, School of Allied Medical Sciences, Shahid Beheshti University of Medical Sciences, Tehran, IR Iran ²Gastroenterology and Liver Diseases Research Center, Research Institute of Gastroenterology and Liver Diseases, Shahid Beheshti University of Medical Sciences, Tehran, IR Iran

Corresponding author: Ali Rahimipour, School of Allied Medical Sciences, Shahid Beheshti University of Medical Sciences, Tehran, IR Iran. Tel/Fax: +98-2126850560; +98-9126904482, E-mail: rahimipour.ali95@gmail.com

Received 2016 October 14; Revised 2016 December 03; Accepted 2017 April 03.

Abstract

Objectives: Overexpression of human leukocyte antigen G (HLA-G) in several malignant tumors has been reported. The aim of our study was to investigate HLA-G expression in colorectal cancer tumors and determine HLA-G expression relation between clinico-pathological characteristics and survival time.

Methods: HLA-G expression was evaluated by immunohistochemistry (IHC) using anti-HLA-G antibody in 100 primary tumors of colorectal cancer with different stages.

Results: Our results showed that 25% of the colorectal cancer tissues had positive HLA-G expression and 75% no stained with anti-HLA-G antibody. The HLA-G expression in advanced stages (III and IV) was more prevalent than those in earlier clinical stages (I and II) (P = 0.0001). Results showed that HLA-G expression can serve as an independent factor for overall survival (OS). In this study, patients with HLA-G expression had significantly shorter survival time than those with negative expressions (P = 0.023).

Conclusions: HLA-G expression can serve as an independent factor for OS and its expression may be directly related to aggressive tumor behavior via escape from the host antitumor immune defense. Protein expression of HLA-G correlates with poor prognosis in colorectal cancer.

Keywords: HLA-G expression, Prognostic Value, Colorectal Cancer

1. Introduction

Colorectal cancer is the fourth common cancer in men after stomach, bladder, and prostate cancer and second one among women after breast cancer (1, 2).

During the process of tumor development in colorectal cancer, evasion of immune surveillance and suppression of the immune system are two important traits of cancer cells (3, 4). These two pathways include a: down regulation of HLA class I expression, and b: ability of tumor cells to regulate the expression of non-classical HLA class I molecules (HLA-E and HLA-G) (5).

HLA-G is a non-classical major histocompatibility complex (MHC) class Ib antigen (6) whose physiologic expression is limited to extra villous trophoblastic cells, thymic epithelial cells, pancreatic islets, and erythroblasts (7-9). However, tumor cells can express HLA-G on the cell surface (10, 11).

HLA-G induces tolerance by binding to inhibitory receptors, such as the immunoglobulin-like transcript (ILT) receptor 2 present on lymphoid and myelomonocytic cells (12) and ILT-4 expresses by dendritic cell, macrophages, and monocytes (13, 14). In addition, natural killer (NK) cells represent an HLA-G-specific receptor named killer cell immunoglobulin-like receptor (KIR)(15). HLA-G directly interacts with different immune cell subpopulations and induces the maintenance of tolerance at different stages of the immune response. Therefore, further tumor cell escape from immune surveillance will occur (6).

Several studies have examined HLA-G expression in colorectal cancer with conflicting results in researches. It has been reported that expression of HLA-G is correlated with escape from immune surveillance during colon cancer development (16). Patients with positive HLA-G tumors had a significantly shorter survival time than patients with negative HLA-G tumors (17). Another study has reported there was no significant relation between clinicopathological characteristics and tumor HLA-G expression (18). Kawin et al. examined HLA-G by immunohistochemistry and reverse transcriptase polymerase chain reaction (RT-PCR), but they did not detect HLA-G expression in either mRNA or protein levels (19). Nevertheless, more studies are definitely needed to demonstrate the role of HLA-G in suppression of the immune system and maybe these findings con-

Copyright © 2017, International Journal of Cancer Management. This is an open-access article distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 International License (http://creativecommons.org/licenses/by-nc/4.0/) which permits copy and redistribute the material just in noncommercial usages, provided the original work is properly cited. tribute to the development of new cancer immunotherapy. The aim of this study is to evaluate the association of HLA-G expression with clinicpathological parameters and survival time in Iranian colorectal cancer populations.

2. Methods

2.1. Patients and Tissue Samples

Our historical cohort study was approved by the ethics committee of the Research institute for gastroenterology and liver diseases (RIGLD), Shahid Beheshti University of Medical Sciences, Tehran, Iran, and all participating patients provided written informed consent for their information to be stored in the databases of RIGLD and to be used for research. The study population consisted of 100 colorectal cancer patients, of which paraffin-embedded tumor tissue was available. All patients had been diagnosed and treated with surgery between 1995 and 2016 in the RIGLD. The clinicopathological factors were determined according to the classification of malignant tumors as set out by the international union against tumor node- metastasis (TNM).

Tumor and clinicopathological data were retrieved from the patients' medical files and pathology reports; in addition, the survival time of these patients was confirmed until May 2016 using telephone inquiries.

2.2. Immunohistochemical Staining

We detected the expression level of HLA-G by immunohistochemistry (IHC) in examining tumor. A mouse monoclonal anti-HLA G antibody (ab52455 clone 4h84: AbCam, UK); 4H84 mAb (specific for a peptide located in the $\alpha 1$ extracellular domain common to all HLA-G isoforms) was used in IHC staining. IHC for HLA-G tumor expression was performed on 4 μ m-thick sections, which were cut from each receiver block and mounted on polylysine coated slides. The slides were deparaffinized 3 times in 100% xylene, 5 minutes each, and rehydrated in a graded series of ethanol from 100% to 70%. Endogenous peroxidase activity was blocked for 20 minutes in 0.3% hydrogen peroxide solution containing methanol at room temperature. For antigen retrieval, slides were boiled in Tris-EDTA Buffer (pH 9.0) for 15 minutes at maximum power (100°C) in a microwave oven. Next, anti-HLA-G monoclonal antibody (1:100) was incubated for 1.5 hours, after which a thorough washing in a 0.01M phosphate-buffered saline (PBS) solution was performed. Binding sites of the primary antibody were visualized using a Dako Envision anti-mouse (K4001; DAKO Cytomation, Glostrup, Denmark) that was incubated for 1h at room temperature. Subsequently, sections were

visualized using 3, 3-diaminobenzidine solution (DAB). Tissue sections were counterstained with haematoxylin, dehydrated and finally mounted with glycerol gelatin.

The pathological features of all specimens and HLA-G expression results were confirmed by two pathologists from the department of pathology, gastroen-terology and liver diseases research center. IHC staining resulted in a visualization of the HLA-G as a brown-stained product. As shown in Figure 1, representative images of HLA-G immunohistochemical for cytotrophoblast from first-trimester human placenta served as a positive control (Figure 1A). For negative control, placenta which underwent the whole IHC staining without primary antibody was served (Figure 1B). Normal colorectal tissue was not stained with HLA-G but colorectal cancer tissue was stained with anti-HLA-G antibody (Figure 1C and 1D, respectively).

2.3. Statistical Analysis Result

Statistical analysis was performed using the SPSS software, version 16.0.0 (SPSS Inc., Chicago, IL). Depending upon the nature of the data, Pearson's chi-square test, Fisher's exact test, or the Mann- Whitney U test were used to compare variables.

Associations between tumor expressions of HLA-G and various clinicopathological variables, such as age, gender, tumor stage, and tumor differentiation (well, moderate and poor), were analyzed using the chi-square test. To evaluate the effect of the above-mentioned variables on survival univariate and multivariate, regression analyses were done using the Cox proportional hazard regression model, and Kaplan-Meier (log-rank test) curves were plotted. The significance of all statistics were recorded if P < 0.05.

3. Results

The analysis of HLA-G expression was performed on 100 colorectal cancer patients with stages I-IV. Clinicopathological features of patients are shown in Table 1. Of 100 samples analyzed, 59 were from male and 41 were from female subjects 49% of whom were under 62 years old. The mean age at the time of diagnosis was 50.52 years (SEM = 1.543 years). 51 patients had a primary tumor of the colon and 49 patients had primary tumor of the rectosigmoid. Of the 100 colorectal rectal cancer patients, 20% were diagnosed with metastases to other organs.

3.1. HLA-G Expression in Primary Colorectal Cancer

To find out the role of HLA-G expression in colorectal cancer, we investigated the correlation of HLA-G expression with the demographic and clinicopathological parameters such as age, gender, TNM stage, tumor differenFigure 1. Representative images of immunohistochemical HLA-G expression in human placenta and colorectal tissues



A, positive HLA-G staining in human placenta tissues; B, negative HLA-G staining in human placenta tissues; C, negative HLA-G staining in normal colorectal; D, positive HLA-G staining in colorectal cancer tissue.

tiation, tumor location, family history, vital status, and microsatellite instability (MSI) status (Table 2).

HLA-G expression was seen in 25 patients out of 100 patients (25%) and the majority of the cases (75%) were not stained with HLA-G antibody. We also found that positive staining for HLA-G in the primary tumor in men was higher than women (28.8% vs 19.5%) that was not statically significant. Tumors with positive HLA-G expression in the colon were comparable with those of the rectosigmoid. Considering tumor stage and HLA-G expression, Pearson chisquare test showed that HLA-G expression in stages III and VI (72%) was significantly higher than those in the stages I and II (28%) (P < 0.0001). We found no significant differences between positive and negative HLA-G expression regarding gender, age, differentiation, metastasis, and MSI status.

3.2. Correlation of HLA-G Expression with Overall Survival

All the characteristics with prognostic value in overall survival (age, gender, tumor stage, MSI status, tumor differentiation, family history and HLA-G expression) were inserted in Cox model (univariate and multivariate analyses). We did not find any significant correlation between overall survival and these prognostic factors, except for HLA-G expression.

Univariate and multivariate analyses showed that tumors with HLA-G expression have poor prognosis compared with tumors without HLA-G expression (Table 3).

Overall survival curves relative to HLA-G expression presented in Figure 2. Patients whose tumors showed loss of HLA-G had significantly a better overall survival (OS) (P = 0.023) compared to patients with tumors with positive HLA-G expression.

4. Discussion

HLA-G plays an important role in prevention of miscarriage because of its fetoprotective effects by suppression of the maternal immune system (7). However, it has been reported that HLA-G expression in cancer cells lead to escape from the host's immune system (20). As shown in Table 4, the published data on HLA-G expression in different types of cancers had conflicts as follows: the positive rate of 90.9% positivity in esophageal squamous cell carcinoma (21) and 62.8% in cervical cancer (22), 60% and 38.88% in

	No. (%)
ge, y	
< 60	49 (49)
> 60	51 (51)
Gender	
Male	59 (59)
Female	41 (41)
Tumor stage	
Ι	14 (14)
Ш	45 (45)
III	37 (37)
IV	4(4)
Differentiation	
Well	56 (56)
Moderate- Poor	44 (44)
Location	
Colon	51 (51)
Rectusigmoid	49 (49)
Metastasis	
Yes	20 (20)
No	80 (80)
Family history	
Yes	43 (43)
No	57 (57)
MSI status	
MSH	10 (10)
MSL	10 (10)
MSS	80 (80)
Vital status	
Live	74 (74)
Dead	26 (26)

breast cancer (23, 24) and different frequency of HLA-G expression in colorectal cancer.

Based on our results, HLA-G determined by immunohistochemistry was not expressed in normal colorectal tissues. However, 25% of the 100 colorectal cancer patients had positive HLA-G expression, which is consistent with Zeestraten et al. data in which they reported 20.3% HLA-G expression (18). In line with results of Ye et al. we detected that expression of HLA-G significantly increased as the clinical stage advanced and patients in III-IV stages had sig-



Figure 2. Overall Survival Curves Relative to HLA-G Status for All Colorectal Cancer Patients (N = 100)

nificantly higher HLA-G positivity than patients with early stages (17), but Zeestraten et al. have reported that HLA-G expression in colorectal cancer patients was not correlated with clinicopathological characteristics (18). However, we did not find a statically significant correlation between HLA-G expression, and age, gender, tumor differentiation, familial history, metastasis, location of tumor and MSI status (Table 2).

Furthermore, in our study univariate and multivariate analyses showed that tumors with HLA-G expression had a poor prognosis compared to tumors with negative expression. These results correlate with Ye and Guo reports (17, 25). Zeestraten et al. have reported that HLA-G expression was not related to OS and DFS (18) contrary to this study. We found that patients with positive HLA-G expression displayed relevance to shorter overall survival despite low frequency of expression (25%) compared with negative HLA-G expression using Kaplan-Meier analysis that is consistent with other studies (17, 25).

4.1. Conclusions

Our results demonstrate that patients with advanced stage had higher HLA-G expression and shorter survival than those with negative expression and HLA-G can be used as an independent prognostic factor. All these findings support the important role of HLA-G in immune surveillance of colorectal tumor cell that leads to escaping from the immune system. Considering Table 4, it can be gathered that HLA-G expression in colorectal cancer had controversial results. Therefore, to HLA-G antibodies such as a potentially useful prognostic indicator. Moreover, in cancer with overexpression of HLA-G, more studies are needed to design strategies for inhibiting HLA-G expression in cancer

	HLA-G				
Variables	Total	No expression	Expression	χ^{2} -P Value	
Age, y				0.419	
\leq 60	49 (49)	35 (71.4)	14 (28.6)		
> 60	51 (51)	40 (78.4)	11 (21.6)		
Gender				0.291	
Male	59 (59)	42 (71.2)	17 (28.8)		
Female	41 (41)	33 (80.5)	8 (19.5)		
INM stage				0.0001	
I, II	59 (59)	52 (88.1)	7 (11.9)		
III, IV	41 (41)	23 (56.1)	18 (43.9)		
Differentiation				0.352	
Well	56 (56)	44 (78.6)	12 (21.4)		
Moderate-Poor	44 (44)	31 (70.5)	13 (29.5)		
Location				0.083	
Colon	51 (51)	42 (82.4)	9 (17.6)		
Rectusigmoid	49 (49)	33 (67.3)	16 (32.7)		
Metastasis				0.564	
Yes	20 (20)	14 (70)	6 (30)		
No	80 (80)	61 (76.2)	19 (23.8)		
Family history				0.56	
Yes	43 (43)	31 (72.1)	12 (27.9)		
No	57 (57)	44 (77.2)	13 (22.8)		
MSI status				0.70	
MSH	10 (10)	8 (80)	2(20)		
MSS/MSL	90 (90)	67 (74.4)	23 (25.6)		
Vital status				0.018	
Live	74 (74)	60 (81.1)	14 (18.9)		
Dead	26 (26)	15 (57.7)	11 (42.3)		

Table 2. Correlation Between the HLA-G Expression and Well-Established Prognosis Factors in Colorectal Cancer Patients^a

Abbreviations: MSH, MSI-High; MSI, microsatellite instability; MSL, MSI-Low; MSS, microsatellite stable; TNM stage, tumor node- metastasis. ^aValues are expressed as No. (%).

immunotherapies that can be combined or not with other treatments.

Acknowledgments

The proposal for this study was approved by the ethics committee of Shahid Beheshti University of Medical Sciences, Tehran, Iran. This study was supported by grant number 400/117 from the Shahid Beheshti University of Medical Sciences.

Footnotes

Authors' Contribution: Non declared.

Funding/Support: Non declared.

Conflict of Interest: The authors declare no conflict of interest.

References

1. Mahmodlou R, Mohammadi P, Sepehrvand N. Colorectal cancer in northwestern Iran. *ISRN gastroenterol*. 2012;**2012**.

- Hajmanoochehri F, Asefzadeh S, Kazemifar AM, Ebtehaj M. Clinicopathological features of colon adenocarcinoma in Qazvin, Iran: a 16 year study. Asian Pac J Cancer Prev. 2014;15(2):951–5. [PubMed: 24568524].
- Parcesepe P, Giordano G, Laudanna C, Febbraro A, Pancione M. Cancer-Associated Immune Resistance and Evasion of Immune Surveillance in Colorectal Cancer. *Gastroenterol Res Pract.* 2016;2016;6261721. doi: 10.1155/2016/6261721. [PubMed: 27006653].
- Rouas-Freiss N, Moreau P, Menier C, Carosella ED. HLA-G in cancer: a way to turn off the immune system. *Semin Cancer Biol.* 2003;13(5):325– 36. [PubMed: 14708712].
- Cavallo F, De Giovanni C, Nanni P, Forni G, Lollini PL. 2011: the immune hallmarks of cancer. *Cancer Immunol Immunother*. 2011;**60**(3):319–26. doi: 10.1007/s00262-010-0968-0. [PubMed: 21267721].
- Amiot L, Ferrone S, Grosse-Wilde H, Seliger B. Biology of HLA-G in cancer: a candidate molecule for therapeutic intervention?. *Cell Mol Life Sci.* 2011;68(3):417-31. doi: 10.1007/s00018-010-0583-4. [PubMed: 21063893].
- Papamitsou T, Toskas A, Papadopoulou K, Sioga A, Lakis S, Chatzistamatiou M, et al. Immunohistochemical study of immunological markers: HLAG, CD16, CD25, CD56 and CD68 in placenta tissues in recurrent pregnancy loss. *Histol Histopathol.* 2014;29(8):1047-55. doi: 10.14670/HH-29.1047. [PubMed: 24557735].
- Lefebvre S, Adrian F, Moreau P, Gourand L, Dausset J, Berrih-Aknin S, et al. Modulation of HLA-G expression in human thymic and amniotic epithelial cells. *Hum Immunol.* 2000;61(11):1095-101. [PubMed: 11137212].
- Cirulli V, Zalatan J, McMaster M, Prinsen R, Salomon DR, Ricordi C, et al. The class I HLA repertoire of pancreatic islets comprises the nonclassical class Ib antigen HLA-G. *Diabetes*. 2006;55(5):1214–22. [PubMed: 16644675].
- Ibrahim EC, Guerra N, Lacombe MJ, Angevin E, Chouaib S, Carosella ED, et al. Tumor-specific up-regulation of the nonclassical class I HLA-G antigen expression in renal carcinoma. *Cancer Res.* 2001;61(18):6838–45. [PubMed: 11559559].
- Urosevic M, Kurrer MO, Kamarashev J, Mueller B, Weder W, Burg G, et al. Human leukocyte antigen G up-regulation in lung cancer associates with high-grade histology, human leukocyte antigen class I loss and interleukin-10 production. *Am J Pathol.* 2001;**159**(3):817–24. doi: 10.1016/S0002-9440(10)61756-7. [PubMed: 11549573].
- Zhang Y, Lu N, Xue Y, Zhang M, Li Y, Si Y, et al. Expression of immunoglobulin-like transcript (ILT)2 and ILT3 in human gastric cancer and its clinical significance. *Mol Med Rep.* 2012;5(4):910–6. doi: 10.3892/mmr.2012.744. [PubMed: 22246571].
- LeMaoult J, Zafaranloo K, Le Danff C, Carosella ED. HLA-G up-regulates ILT2, ILT3, ILT4, and KIR2DL4 in antigen presenting cells, NK cells, and T cells. FASEB J. 2005;19(6):662–4. doi: 10.1096/fj.04-1617fje. [PubMed: 15670976].
- Zilberman S, Schenowitz C, Agaugue S, Benoit F, Riteau B, Rouzier R, et al. HLA-G1 and HLA-G5 active dimers are present in malignant cells and effusions: the influence of the tumor microenvironment. *Eur J Immunol.* 2012;42(6):1599–608. doi: 10.1002/eji.201141761. [PubMed: 22678912].
- O'Callaghan CA, Bell JI. Structure and function of the human MHC class lb molecules HLA-E, HLA-F and HLA-G. *Immunol Rev.* 1998;163:129– 38. [PubMed: 9700506].
- 16. Fukushima Y, Oshika Y, Nakamura M, Tokunaga T, Hatanaka H, Abe

Y, et al. Increased expression of human histocompatibility leukocyte antigen-G in colorectal cancer cells. *Int J Mol Med.* 1998;**2**(3):349–51. [PubMed: 9855710].

- Ye SR, Yang H, Li K, Dong DD, Lin XM, Yie SM. Human leukocyte antigen G expression: as a significant prognostic indicator for patients with colorectal cancer. *Mod Pathol.* 2007;20(3):375–83. doi: 10.1038/modpathol.3800751. [PubMed: 17277760].
- Zeestraten EC, Reimers MS, Saadatmand S, Goossens-Beumer IJ, Dekker JW, Liefers GJ, et al. Combined analysis of HLA class I, HLA-E and HLA-G predicts prognosis in colon cancer patients. *Br J Cancer*. 2014;**110**(2):459–68. doi: 10.1038/bjc.2013.696. [PubMed: 24196788].
- Leelawat K, Engprasert S, Pongchai-rerk U, Tuchinda S, Suthipintawong C, Leardkamolkarn V. No expression of human leukocyte antigen G (HLA-G) in colorectal cancer cells. *J Med Assoc Thai*. 2004;87(7):816–8. [PubMed: 15521238].
- Barbara S, Simon JB. In: Resistance of Cancer Cells to CTL-Mediated Immunotherapy. Bonavida B, Chouaib S, editors. Los Angeles: Springer; 2015. Role of the Non-classical HLA Class I Antigens for Immune Escape.
- Yie SM, Yang H, Ye SR, Li K, Dong DD, Lin XM. Expression of HLA-G is associated with prognosis in esophageal squamous cell carcinoma. *Am J Clin Pathol.* 2007;**128**(6):1002–9. doi: 10.1309/JNCW1QLDFB6AM9WE. [PubMed: 18024326].
- Li XJ, Zhang X, Lin A, Ruan YY, Yan WH. Human leukocyte antigen-G (HLA-G) expression in cervical cancer lesions is associated with disease progression. *Hum Immunol.* 2012;73(9):946–9. doi: 10.1016/j.humimm.2012.07.041. [PubMed: 22820627].
- Lefebvre S, Antoine M, Uzan S, McMaster M, Dausset J, Carosella ED, et al. Specific activation of the non-classical class I histocompatibility HLA-G antigen and expression of the ILT2 inhibitory receptor in human breast cancer. J Pathol. 2002;196(3):266-74. doi: 10.1002/path.1039. [PubMed: 11857488].
- 24. de Kruijf EM, Sajet A, van Nes JG, Natanov R, Putter H, Smit VT, et al. HLA-E and HLA-G expression in classical HLA class I-negative tumors is of prognostic value for clinical outcome of early breast cancer patients. *J Immunol.* 2010;**185**(12):7452–9. doi: 10.4049/jimmunol.1002629. [PubMed: 21057081].
- Guo ZY, Lv YG, Wang L, Shi SJ, Yang F, Zheng GX, et al. Predictive value of HLA-G and HLA-E in the prognosis of colorectal cancer patients. *Cell Immunol.* 2015;**293**(1):10–6. doi: 10.1016/j.cellimm.2014.10.003. [PubMed: 25461612].
- Swets M, Konig MH, Zaalberg A, Dekker-Ensink NG, Gelderblom H, van de Velde CJ, et al. HLA-G and classical HLA class I expression in primary colorectal cancer and associated liver metastases. *Hum Immunol.* 2016;77(9):773–9. doi: 10.1016/j.humimm.2016.03.001. [PubMed: 26968946].
- Barrier BF, Kendall BS, Sharpe-Timms KL, Kost ER. Characterization of human leukocyte antigen-G (HLA-G) expression in endometrial adenocarcinoma. *Gynecol Oncol.* 2006;**103**(1):25–30. doi: 10.1016/j.ygyno.2006.01.045. [PubMed: 16530254].
- Ishigami S, Natsugoe S, Miyazono F, Nakajo A, Tokuda K, Matsumoto M, et al. HLA-G expression in gastric cancer. *Anticancer Res.* 2006;26(3B):2467-72. [PubMed: 16821634].
- 29. Yie SM, Yang H, Ye SR, Li K, Dong DD, Lin XM. Expression of human leukocyte antigen G (HLA-G) correlates with poor prognosis in gastric carcinoma. *Ann Surg Oncol.* 2007;**14**(10):2721–9. doi: 10.1245/s10434-007-9464-y. [PubMed: 17564748].

Variables		Univariate Analysis	Univariate Analysis		Multivariate Analysis	
	-	Hazard Ratio for Death	P Value	Hazard Ratio for Death	χ^{2} -P Value	
Gender			0.643		0.382	
	Female	1 ref.		1 ref.		
	Male	0.826 (0.368 - 1.854)		0.671 (0.274 - 1.642)		
Age			0.447		0.425	
	\leq 60	1 ref				
	> 60	1.349 (0.624 - 2.919)		1.425 (0.597 - 3.399)		
Location of tumor			0.173		0.432	
	Colon	1 ref.		1 ref.		
	Rectusigmoid	0.581 (0.266 - 1.269)		0.698 (0.285 - 1.711)		
Differentiation			0.121		0.078	
	Poor- moderate	1 ref.		1 ref.		
	Well	0.513 (0.22 -1.192)		0.443 (0.179 - 1.094)		
TNM stage			0.729		0.496	
		I, II	1 ref.	1 ref.		
	III, IV	0.827 (0.403 - 1.887)		2.18 (0.231 - 20.601)		
HLA-G expression			0.028		0.045	
	No	1 ref.		1 ref.		
	Yes	1.551 (1.049 - 2.295)		1.016 (1.016 - 2.574)		
MSI status			0.218		0.262	
	MSH	1 ref.		1 ref.		
	MSS/MSL	0.284 (0.038 - 2.103)		0.299 (0.036 - 2.463)		
Metastasis			0.306		0.213	
	No	1 ref.		1 ref.		
	Yes	0.635 (0.267 - 1.513)		0.533 (0.199 - 1.433)		

Table 3. Univariate and Multivariate Cox Regression Analyses of Possible Prognostic Variables and Parameters That Correlate with Overall Survival

Abbreviations: MSH, MSI-High; MSI, microsatellite instability; MSI, MSI-Low; MSS, microsatellite stable; TNM stage, tumor node-metastasis.

Table 4. HLA-G Expression in Several Solid Tumors

Type of Cancer	Method	HLA-G Protein Expression, %	Results	Reference
Colorectal cancer	IHC	64.4	1. HLA-G expression was significantly correlated with the clinicopathologic characteristics.	(17)
			2. Patients with HLA-G positive tumors had a significantly shorter survival time 3. HLA-G expression can serve as an independent factor for OS.	
Colorectal cancer/cell	RT-PCR IHC	87.17	1. The expression of HLA-G mRNA was significantly more frequent in colorectal cancer than in the extraneoplastic tissue.	(16)
			2. HLA-G protein expression on colorectal cancer cells may be correlated with escape from immunological surveillance during colon cancer development.	(10)
Colorectal cancer	ІНС	29	1. HLA-G expression in the primary tumors was not significantly correlated with liver metastasis.	(26)
			2. Regarding HLA-G expression no significant difference between synchronous or metachronous onset of liver metastasis was observed.	(20)
Colorectal cancer		20.3	1. HLA-G tumour expression was not related to OS (Overall Survival) and DFS (disease-free survival).	(18)
	IHC		2. None of the clinicopathological characteristics were significantly related to tumour expression.	(10)
Colorectal cancer	ІНС	70.6	1. Expression of HLA-G was only significantly associated with a pathological diagnosis.	(25)
			2. Patients with HLA-G expression had a significantly poorer overall survival	
Endometrial adenocarcinoma	IHC	55	1. The stage of cancer was significantly correlated with HLA-G staining	(27)
Breast cancer	IHC	60	1. Of the patients with no tumor expression of HLA-G, 56% of patients were relapse free after 10 y.	(24)
			2. Of the patients with tumor expression of HLA-G, 39% of patients were relapse free after 10 y.	()

Breast cancer	IHC	- 38.88	1. HLA-G expression significantly correlated with inflammatory grade of breast cancer lesions.	(23)
	PCR		2. There was no significant relationship between HLA-G expression and tumor grade, type, and patient age.	<-/
			1. HLA-G expression decreased as the clinical stage advanced.	
Gastric cancer	ІНС	45.21	2. Survival rate in the HLA-G-positive group was significantly higher than those with negative expression.	(28)
			3. HLA-G expression may play a role in the early clinical stages by protecting cancer cells from tumor infiltrating effectors.	
			1. HLA-G expression in the tumors was significantly correlated with clinicopathologic characteristics.	
Gastric carcinoma	IHC	71	2. Survival rate in the HLA-G-positive patients was significantly shorter than those with negative expression.	(29)
			3. HLA-G was an independent prognostic factor.	
			1.HLA-G expression was significantly correlated with clinicopathologic characteristics	
Esophageal squamous cell carcinoma	ІНС	90.9	2. Patients with positive HLA-G expression had a significantly worse prognosis.	(21)
			3. HLA-G expression was an independent prognostic factor.	
Cervical cancer	ІНС	62.8	1. HLA-G expression was associated with the disease progression in patients	(22)
			2. HIA-G expression in stages III and IV was higher than those of stages I and II.	(22)