




# Evaluating and Comparing Expression Levels of miR-199a and miR-126 in Peripheral Blood of Patients with Lone Atrial Fibrillation and Healthy Controls

Khatereh Falamarzi<sup>1</sup>, Mehdi Mokhtarian<sup>2</sup>, Isa Khareshi <sup>2</sup>, Mersedeh Karvandi<sup>1,\*</sup>

<sup>1</sup> Department of Cardiovascular Disease, School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran

<sup>2</sup> Cardiovascular Research Center, Shahid Beheshti University of Medical Sciences, Tehran, Iran

\*Corresponding author: Department of Cardiovascular Disease, School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran. Email: dr.karvandi@gmail.com

Received 2024 April 6; Accepted 2024 April 16.

## Abstract

**Background:** Patients with lone atrial fibrillation (AF) are ideal subjects for investigating the relationship between AF and genetic factors. MicroRNAs are regulatory genetic factors closely related to the pathways leading to heart diseases, including AF.

**Objectives:** This investigation aimed to examine the expression levels of miRNAs involved in the pathogenesis of this disease as non-invasive diagnostic and prognostic biomarkers.

**Methods:** The participants in this case-control study included 35 individuals with lone AF and 35 healthy controls. After collecting blood samples from all participants, the expression levels of miR-126 and miR-199a were evaluated using Real-Time PCR on the RNA extracted from these samples. The expression levels of these two miRNAs were then compared between the two groups.

**Results:** Although the expression level of miR-126 in lone AF patients was significantly lower compared to healthy controls, the expression level of miR-199a did not show a significant difference between cases and controls. Additionally, the expression level of miR-126, with an AUC of 0.96, demonstrated very good prognostic power for the occurrence of lone AF.

**Conclusions:** A decreased expression level of miR-126 was associated with lone AF and could be used as a biomarker with excellent diagnostic power. This change in expression could be proposed as a mediator that accelerates the complications of lone AF.

**Keywords:** Lone Atrial Fibrillation, miRNA, Gene Expression, Diagnostic Power

## 1. Background

Atrial fibrillation (AF), the most prevalent supraventricular arrhythmia, affects up to 1% of the general population, with the prevalence increasing to 10% in the elderly (1). It is estimated that more than 33 million people worldwide suffer from AF, with the incidence being three times higher in men compared to women (2). The incidence and outcomes of AF are influenced by many genetic and epigenetic factors and vary significantly by geography and ethnicity (3). Electrocardiography (ECG) is the most common method used for AF diagnosis. However, due to its short recording duration, detecting AF in asymptomatic

patients is limited. Therefore, useful biomarkers are needed for the early identification and treatment of asymptomatic AF cases (4).

Several general risk factors, such as diabetes, heart failure, hyperthyroidism, arterial hypertension, obesity, gender, and ischemic and structural heart disease, are known to be associated with AF. In more than 20% of AF subjects, there is no evidence of any underlying risk factors, which are known as lone AF (5). It has been proven that individuals with at least one parent with a history of AF are more susceptible to developing AF, implying the role of genetic factors in the pathogenesis of AF (6). However, a large part of the heritability of AF remains unexplained (7).

Micro-RNAs (miRNAs) are single-stranded, non-protein-coding RNAs containing around 22 nucleotides that regulate the expression of target genes post-transcriptionally by binding to the 3'UTR of mRNA molecules (8). Several processes, including cell growth, differentiation, proliferation, and metabolism, are regulated by miRNAs (9). miRNAs have been commonly detected in circulating blood and cardiac muscle of humans and animal models (10). Additionally, miRNAs have been shown to be associated with the pathophysiology of several cardiovascular diseases, including AF, by regulating atrial remodeling mechanisms (11, 12). Changes in miRNA expression levels have been reported to modulate the susceptibility to AF development in humans. Hence, miRNAs have the potential to be considered targets for the diagnosis and treatment of AF (13).

Changes in miRNA expression have been reported as potential prognostic and diagnostic biomarkers (14). The high stability, sensitivity, specificity, and accessibility of circulating miRNAs in serum and plasma make them interesting targets for the early diagnosis of several cardiac diseases, including AF (15-18).

Considerable efforts have been made to identify plasma markers for AF diagnosis. Several studies have identified circulating microRNA levels and aberrant expression of microRNAs in AF tissue, paving the way for further analysis of circulating microRNAs as biomarkers for AF prediction (19-21).

## 2. Objectives

Therefore, the aim of this investigation was to explore the expression levels of miR-199a and miR-126 in lone AF patients and to evaluate their diagnostic power for the discrimination of AF.

## 3. Methods

In the present case-control investigation, 70 participants were included, of which 35 cases had lone AF and 35 were healthy controls. Individuals with lone AF were selected from those referred to Modarres Hospital in Tehran (2022 - 2023) and were subjected to ECG and 24-hour Holter monitoring due to suspected clinical symptoms of AF. Control subjects were selected from those who underwent ECG due to discomfort in the chest, feeling of tachycardia, or heart palpitations,

but had a normal sinus rhythm and absence of any arrhythmia.

Patients with inflammatory, infectious, or autoimmune diseases, a history of hematological disease, malignancy, kidney, liver, or thyroid disease, hypertension, diabetes, valvular disease, or a history of previous MI were excluded from the study. This study was performed in line with the principles of the Declaration of Helsinki and according to the accepted regulations of the Shahid Beheshti University of Medical Sciences ethical committee (IR.SBMU.MSP.REC.1402.565). Informed consent was obtained from all participants.

### 3.1. Genes Selection

The prioritization and selection of the most appropriate genes in the molecular pathways involved in the pathogenesis of lone AF were conducted by collecting and downloading data on the inflammatory pathways of lone AF pathogenesis from KEGG, Reactome, BioCarta, WikiPathways, and previous studies.

### 3.2. Expression Analysis

Five milliliters of whole blood were taken from all subjects, and the total RNA was isolated from the blood specimens using the RiboEx™ RNA purification kit (GeneAll Biotechnology Co. Ltd, Seoul, Korea). The integrity and concentration of RNA samples were assessed using the Nanodrop instrument (Thermo Fisher Scientific). Complementary DNA (cDNA) synthesis was accomplished using the Applied Biosystems High-Capacity cDNA Reverse Transcription Kit (Takara, Tokyo, Japan). The expression levels of the miR-199a and miR-126 genes in the RNA samples were assessed using the RealQ Plus 2x Master Mix Green, High ROXTM (AmpliQon, Denmark) on the LightCycler 96 Real-Time PCR System (Roche, Germany). RNU6 was used to normalize the expression results via the  $2^{-\Delta\Delta Ct}$  method. Sequences of specific primers for investigating the expression levels of miR-199a, miR-126, and RNU6 were designed using Allele ID software and are provided in Table 1.

**Table 1.** Sequence of Primers to Evaluate the Expression Level of miR-126, miR-199a, and RNU6

Gene of Interest	Primer Sequence
RNU6	
Forward	5'-GCTTCGGCAGCACATATACTAAAAT-3'

Gene of Interest	Primer Sequence
Reverse	5'-CGCTTACGAATTTGCGTGCAT-3'
<b>miR-126</b>	
Forward	5'-GGCATTATTACTTTGG-3'
Reverse	5'-TGCGTGTCTGGGAGTC-3'
<b>miR-199a</b>	
Forward	5'-ACACTCCAGCTGGGCCAGTTCAGACTAC-3'
Reverse	5'-CTCAACTGGTGTCTGGAGTCGGCAA-3'

### 3.3. Statistical Analyses

All statistical analyses were performed using PRISM version 8 and SPSS version 24 statistical software. Continuous variables were described as mean  $\pm$  standard deviation (SD), and categorical variables were presented as absolute frequency and frequency percentage. The Fisher's Exact Test was used to compare categorical variables between patients and control subjects. The t-test was applied for continuous variables after verifying the normality of the distribution using the Shapiro-Wilk test. ROC curve analysis of miR-126 and miR-199a expression levels for distinguishing lone AF patients from control subjects was performed using MedCalc software. A p-value of less than 0.05 was considered significant.

## 4. Results

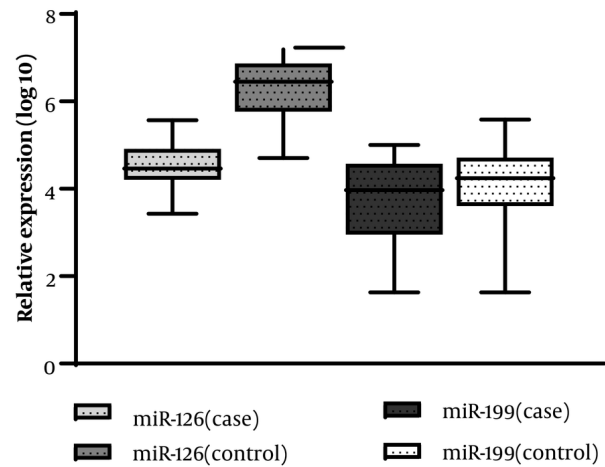
A total of 70 individuals with an average age of  $58.31 \pm 9.2$  years were included in the present study, of whom 58.6% were male. The comparison between lone AF patients and control subjects revealed significant differences only in FBS levels, with lone AF patients having significantly higher FBS levels. The basic and demographic features of the study participants are provided in Table 2.

**Table 2.** Basic Features of the Participants<sup>a</sup>

Parameters	Total (N=70)	Lone-AF Patients (N=35)	Control Subjects (N=35)	P-Value
Age (y)	58.31 $\pm$ 9.2	58.08 $\pm$ 7.58	58.54 $\pm$ 10.69	0.83
FBS (mg/dL)	97.9 $\pm$ 15.13	102.34 $\pm$ 14.95	93.45 $\pm$ 14.16	0.01
BMI (Kg/m <sup>2</sup> )	24.64 $\pm$ 2.93	25.21 $\pm$ 3.4	24.08 $\pm$ 2.28	0.1
EF	55.57 $\pm$ 3.13	55.71 $\pm$ 3.24	55.42 $\pm$ 3.06	0.7
Male	41 (58.6%)	21 (60%)	20 (57.1%)	0.99

<sup>a</sup> Values are expressed as No. (%) or mean  $\pm$  SD.

As shown in Figure 1 and Table 3, the relative expression levels of the two investigated miRNAs were compared between lone AF patients and control subjects. The expression analysis revealed that the expression level of miR-126 in patients with lone AF was significantly lower than in healthy controls ( $P = 0.0001$ ).



**Figure 1.** Comparison of patients with lone AF and controls regarding the relative expression of miR-126 and miR-199a

**Table 3.** Comparison of two Groups of Patients in Terms of Relative Expression of miR-126, miR-199a Genes

Group	Mean $\pm$ SD	P-Value <sup>a</sup>	Fold Change <sup>b</sup>	95% CI
<b>miR-126</b>		0.0001	0.013 (down-regulated)	0.08 - 0.75
Lone-AF patients	4.48 $\pm$ 0.51			
Control subjects	6.28 $\pm$ 0.71			
<b>miR-199a</b>		0.5	-	0.53 - 1.25
Lone-AF patients	3.72 $\pm$ 0.99			
Control subjects	3.89 $\pm$ 1.1			

<sup>a</sup> P-value less than or equal to 0.05 is considered significant.

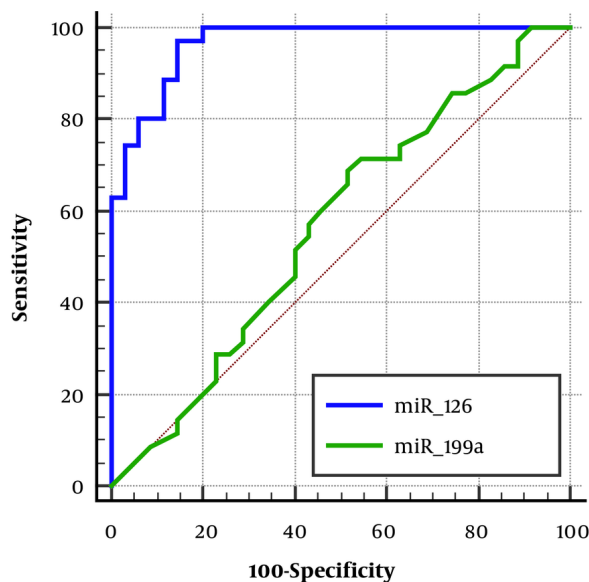
<sup>b</sup> The expression ratio (fold change) was calculated by the  $2^{-\Delta\Delta Ct}$  formula.

Finally, the diagnostic power of miR-126 and miR-199a expression levels in patients with lone AF was evaluated (Table 4). The expression level of miR-126 showed excellent diagnostic power for the occurrence of lone AF with an AUC of 0.96, while the expression level of miR-

199a did not demonstrate significant predictive power (Figure 2).

**Table 4.** ROC Curve Analysis Results of miR-126, and miR-199a Expression Levels in Identifying Lone AF Patients

Variables	AUC	95% CI	P-Value	Sensitivity	Specificity	Youden Index (Threshold)
miR-126	0.96	0.89 to 0.99	0.0001	97.14	85.71	0.82
miR-199a	0.56	0.44 to 0.68	0.33	68.57	48.57	0.17



**Figure 2.** ROC curve analysis to evaluate the diagnostic power of miR-126, and miR-199a expression levels in identifying lone AF patients

## 5. Discussion

Based on the results of this study, the expression level of miR-126 in patients with lone AF was significantly lower than in healthy controls, while the expression level of miR-199a was not significantly different between the patients and control subjects. Additionally, the expression level of miR-126, with an AUC of 0.96, showed excellent predictive power for the occurrence of lone AF, while the expression of miR-199a did not have significant predictive power in differentiating lone AF patients from control subjects.

The molecular mechanisms involved in the early progression of AF are still poorly understood. So far, only a limited number of studies have explored the role of

miRNAs as a genetic component in the pathogenesis of AF (20, 22). These investigations typically involved patients with chronic AF, where extensive atrial remodeling processes have already been established. Therefore, changes in the miRNA profile of patients with chronic AF may result from such remodeling processes. In contrast, this study focused on lone AF and revealed a specific miR-protein interaction that may be involved in the early pathogenesis of AF without causing confounding structural changes (23, 24).

These results showed that miR-126 was significantly downregulated in blood samples from patients with lone AF. miR-126, also known as angiomiR-126, acts as an important regulator in maintaining endothelial homeostasis and vascular integrity. Circulating miR-126 has the potential to be used as a biomarker for vascular injury and cardiac damage in cardiovascular diseases (CVDs) including coronary artery disease (CAD), myocardial infarction (AMI), heart failure, and AF (25-28). Decreased circulating levels of miR-126 were associated with endothelial dysfunction (29). The expression level of miR-126 was significantly decreased in persistent AF patients compared with paroxysmal AF patients (25).

Several investigations have revealed that miR-126 is highly expressed in vascular tissues, including the heart, lungs, and liver. This miRNA is also expressed in hematopoietic and endothelial cells (30, 31). It has been shown that the expression of miR-126 levels may result in the activation of the vascular endothelial growth factor signaling pathway in the endothelium (32). Additionally, endothelial cell migration was disrupted during the processes of vascular growth, development, and organization after knocking down miR-126 expression in animal models. These processes are closely related to the development of AF and HF (33). Thus, miR-126 plays an important role in regulating the activation of the vascular endothelial growth factor pathway. Abnormal expression levels of miR-126 may cause impaired angiogenesis, leading to an elevated risk of AF and HF (34).

In this study, we observed that despite matching all risk factors between the patient and control groups, fasting serum glucose levels in patients with isolated atrial fibrillation were significantly higher than in control subjects. It has been established that AF patients with Diabetes Mellitus (DM) are susceptible to cardiovascular events and mortality (35). Such patients

have a poor prognosis, decreased quality of life, and a higher risk of mortality and hospitalization compared to those without diabetes (36).

Vascular complications in DM patients are caused by endothelial dysfunction. Low levels of miR-126 have been observed in apoptotic bodies and microvesicles in diabetic individuals. Treatment of endothelial-derived EVs with high glucose and a small amount of miR-126 leads to the loss of the regenerative capacity of ECs (29). Therefore, hyperglycemia may have a role in defective angiogenesis and endothelial remodeling in AF patients with DM by contributing to endothelial dysfunction and lowering miR-126 expression in the vascular endothelium.

miR-126 suppresses inflammation in endothelial cells under hyperglycemic conditions. miR-126, expressed by endothelial cells, plays a crucial role in developmental angiogenesis in vivo. The association of miR-126 with vascular disease, inflammation, and angiogenesis has been reported (37). Recent studies have shown that miR-126 is involved in vascular inflammation (31, 38). miR-126 suppresses inflammation in endothelial cells incubated with high glucose by modulating HMGB1 expression. This suggests that miR-126 may be a useful molecular target for managing CVD, atherosclerosis, and AF in individuals with diabetes (39). These studies can explain the high levels of glucose and miR-126 in patients with lone AF in our study.

The cardioprotective protein SIRT1 is one of the predicted targets of miR-199a, and its expression level was increased in the heart tissue of individuals with AF and also in the serum of patients with chronic CAD (40, 41). Overexpression of SIRT1 is hypothesized to be a compensatory mechanism to prevent the oxidative stress process that contributes to the pathogenesis of AF and CAD (40, 42). In a recent transcriptomic study, miR-199a was designated as an up-regulated microRNA in patients with paroxysmal AF after cardiac surgery (43). It seems that miR-199a plays an important role in other types of AF. In our study, due to the absence of any risk factors such as CAD, hypertension, diabetes, or heart diseases and abnormalities, this miR did not show a significant difference between patients and controls. Therefore, it could be proposed as a marker for the differential diagnosis of AF from other types.

### 5.1. Conclusions

Our results strongly indicate that the decreased expression levels of miR-126 are associated with lone AF and that its reduced expression level could serve as an excellent diagnostic marker. This change in expression may be one of the mediators that accelerate the development of pathophysiology and complications in lone AF.

### Footnotes

**Authors' Contribution:** Study concept and design: M.K; analysis and interpretation of data: K.F, I.K., and M.M.; drafting of the manuscript: K.F; critical revision of the manuscript: All authors; statistical analysis: M.M.

**Conflict of Interests Statement:** The authors of this study declared no conflict of interests.

**Data Availability:** Data are available on request.

**Ethical Approval:** This study was performed in line with the principles of the Declaration of Helsinki and according to the accepted regulations of the Shahid Beheshti University of Medical Sciences ethical committee (IR.SBMU.MSP.REC.1402.565).

**Funding/Support:** There was no funding to be declared.

**Informed Consent:** Informed consent was obtained from all participants.

### References

- Pellman J, Sheikh F. Atrial fibrillation: mechanisms, therapeutics, and future directions. *Compr Physiol*. 2015;5(2):649-65. [PubMed ID: 25880508]. [PubMed Central ID: PMC5240842]. <https://doi.org/10.1002/cphy.c140047>.
- Ko D, Rahman F, Martins MA, Hylek EM, Ellinor PT, Schnabel RB, et al. Atrial fibrillation in women: treatment. *Nat Rev Cardiol*. 2017;14(2):113-24. [PubMed ID: 27786235]. [PubMed Central ID: PMC5520636]. <https://doi.org/10.1038/nrcardio.2016.171>.
- Vinciguerra M, Dobrev D, Nattel S. Atrial fibrillation: pathophysiology, genetic and epigenetic mechanisms. *Lancet Reg Health Eur*. 2024;37:100785. [PubMed ID: 38362554]. [PubMed Central ID: PMC10866930]. <https://doi.org/10.1016/j.lanepe.2023.100785>.
- Turagam MK, Mirza M, Werner PH, Sra J, Kress DC, Tajik AJ, et al. Circulating Biomarkers Predictive of Postoperative Atrial Fibrillation. *Cardiol Rev*. 2016;24(2):76-87. [PubMed ID: 25699982]. [PubMed Central ID: PMC4944526]. <https://doi.org/10.1097/CRD.000000000000059>.
- Lee SH, Park SJ, Byeon K, On YK, Kim JS, Shin DG, et al. Risk factors between patients with lone and non-lone atrial fibrillation. *J Korean Med Sci*. 2013;28(8):1174-80. [PubMed ID: 23960444]. [PubMed Central ID: PMC3744705]. <https://doi.org/10.3346/jkms.2013.28.8.1174>.
- Fox CS, Parise H, D'Agostino RS, Lloyd-Jones DM, Vasan RS, Wang TJ, et al. Parental atrial fibrillation as a risk factor for atrial fibrillation in



- offspring. *JAMA*. 2004;**291**(23):2851-5. [PubMed ID: [15199036](#)]. <https://doi.org/10.1001/jama.291.23.2851>.
7. Lubitz SA, Yin X, Fontes JD, Magnani JW, Rienstra M, Pai M, et al. Association between familial atrial fibrillation and risk of new-onset atrial fibrillation. *JAMA*. 2010;**304**(20):2263-9. [PubMed ID: [21076174](#)]. [PubMed Central ID: [PMC3073054](#)]. <https://doi.org/10.1001/jama.2010.1690>.
  8. Kozomara A, Griffiths-Jones S. miRBase: annotating high confidence microRNAs using deep sequencing data. *Nucleic Acids Res*. 2014;**42**(Database issue):D68-73. [PubMed ID: [24275495](#)]. [PubMed Central ID: [PMC3965103](#)]. <https://doi.org/10.1093/nar/gkt1181>.
  9. Macfarlane LA, Murphy PR. MicroRNA: Biogenesis, Function and Role in Cancer. *Curr Genomics*. 2010;**11**(7):537-61. [PubMed ID: [21532838](#)]. [PubMed Central ID: [PMC3048316](#)]. <https://doi.org/10.2174/138920210793175895>.
  10. van den Berg NWE, Kawasaki M, Berger WR, Neefs J, Meulendijks E, Tijssen AJ, et al. MicroRNAs in Atrial Fibrillation: from Expression Signatures to Functional Implications. *Cardiovasc Drugs Ther*. 2017;**31**(3):345-65. [PubMed ID: [28752208](#)]. [PubMed Central ID: [PMC5550535](#)]. <https://doi.org/10.1007/s10557-017-6736-z>.
  11. Small EM, Frost RJ, Olson EN. MicroRNAs add a new dimension to cardiovascular disease. *Circulation*. 2010;**121**(8):1022-32. [PubMed ID: [20194875](#)]. [PubMed Central ID: [PMC2847432](#)]. <https://doi.org/10.1161/CIRCULATIONAHA.109.889048>.
  12. Wyndham CR. Atrial fibrillation: the most common arrhythmia. *Texas Heart Institute J*. 2000;**27**(3):257.
  13. Neudecker V, Brodsky KS, Kreth S, Ginde AA, Eltzschig HK. Emerging Roles for MicroRNAs in Perioperative Medicine. *Anesthesiology*. 2016;**124**(2):489-506. [PubMed ID: [26632665](#)]. [PubMed Central ID: [PMC4820395](#)]. <https://doi.org/10.1097/ALN.0000000000000969>.
  14. Hodgkinson CP, Kang MH, Dal-Pra S, Mirotsov M, Dzau VJ. MicroRNAs and Cardiac Regeneration. *Circ Res*. 2015;**116**(10):1700-11. [PubMed ID: [25953925](#)]. [PubMed Central ID: [PMC4428675](#)]. <https://doi.org/10.1161/CIRCRESAHA.116.304377>.
  15. D'Alessandra Y, Devanna P, Limana F, Straino S, Di Carlo A, Brambilla PG, et al. Circulating microRNAs are new and sensitive biomarkers of myocardial infarction. *Eur Heart J*. 2010;**31**(22):2765-73. [PubMed ID: [20534597](#)]. [PubMed Central ID: [PMC2980809](#)]. <https://doi.org/10.1093/eurheartj/ehq167>.
  16. Natsume Y, Oaku K, Takahashi K, Nakamura W, Oono A, Hamada S, et al. Combined Analysis of Human and Experimental Murine Samples Identified Novel Circulating MicroRNAs as Biomarkers for Atrial Fibrillation. *Circ J*. 2018;**82**(4):965-73. [PubMed ID: [29398686](#)]. <https://doi.org/10.1253/circj.CJ-17-1194>.
  17. Wiedera C, Gupta SK, Lorenzen JM, Bang C, Bauersachs J, Bethmann K, et al. Diagnostic and prognostic impact of six circulating microRNAs in acute coronary syndrome. *J Mol Cell Cardiol*. 2011;**51**(5):872-5. [PubMed ID: [21806992](#)]. <https://doi.org/10.1016/j.yjmcc.2011.07.011>.
  18. Zampetaki A, Willeit P, Kiechl S, Mayr M. Circulating microRNAs: From single biomarkers to re-wired networks. *J Free Radical Biol Med*. 2012;**53**:S19.
  19. Kuwabara Y, Ono K, Horie T, Nishi H, Nagao K, Kinoshita M, et al. Increased microRNA-1 and microRNA-133a levels in serum of patients with cardiovascular disease indicate myocardial damage. *Circ Cardiovasc Genet*. 2011;**4**(4):446-54. [PubMed ID: [21642241](#)]. <https://doi.org/10.1161/CIRCGENETICS.110.958975>.
  20. Lu Y, Zhang Y, Wang N, Pan Z, Gao X, Zhang F, et al. MicroRNA-328 contributes to adverse electrical remodeling in atrial fibrillation. *Circulation*. 2010;**122**(23):2378-87. [PubMed ID: [21098446](#)]. <https://doi.org/10.1161/CIRCULATIONAHA.110.958967>.
  21. Meder B, Keller A, Vogel B, Haas J, Sedaghat-Hamedani F, Kayvanpour E, et al. MicroRNA signatures in total peripheral blood as novel biomarkers for acute myocardial infarction. *Basic Res Cardiol*. 2011;**106**(1):13-23. [PubMed ID: [20886220](#)]. <https://doi.org/10.1007/s00395-010-0123-2>.
  22. Xiao J, Liang D, Zhang Y, Liu Y, Zhang H, Liu Y, et al. MicroRNA expression signature in atrial fibrillation with mitral stenosis. *Physiol Genomics*. 2011;**43**(11):655-64. [PubMed ID: [21325063](#)]. <https://doi.org/10.1152/physiolgenomics.00139.2010>.
  23. Cooley N, Cowley MJ, Lin RC, Marasco S, Wong C, Kaye DM, et al. Influence of atrial fibrillation on microRNA expression profiles in left and right atria from patients with valvular heart disease. *Physiol Genomics*. 2012;**44**(3):211-9. [PubMed ID: [22147268](#)]. <https://doi.org/10.1152/physiolgenomics.00111.2011>.
  24. Nishi H, Sakaguchi T, Miyagawa S, Yoshikawa Y, Fukushima S, Saito S, et al. Impact of microRNA expression in human atrial tissue in patients with atrial fibrillation undergoing cardiac surgery. *PLoS One*. 2013;**8**(9): e73397. [PubMed ID: [24069193](#)]. [PubMed Central ID: [PMC3772020](#)]. <https://doi.org/10.1371/journal.pone.0073397>.
  25. Wei XJ, Han M, Yang FY, Wei GC, Liang ZG, Yao H, et al. Biological significance of miR-126 expression in atrial fibrillation and heart failure. *Braz J Med Biol Res*. 2015;**48**(11):983-9. [PubMed ID: [26313139](#)]. [PubMed Central ID: [PMC4671524](#)]. <https://doi.org/10.1590/1414-431X20154590>.
  26. Ali W, Mishra S, Rizvi A, Pradhan A, Perrone MA. Circulating microRNA-126 as an independent risk predictor of coronary artery disease: a case-control study. *J EIJFCC*. 2021;**32**(3):347.
  27. Fukushima Y, Nakanishi M, Nonogi H, Goto Y, Iwai N. Assessment of plasma miRNAs in congestive heart failure. *Circ J*. 2011;**75**(2):336-40. [PubMed ID: [21157109](#)]. <https://doi.org/10.1253/circj.cj-10-0457>.
  28. Long G, Wang F, Duan Q, Chen F, Yang S, Gong W, et al. Human circulating microRNA-1 and microRNA-126 as potential novel indicators for acute myocardial infarction. *Int J Biol Sci*. 2012;**8**(6):811-8. [PubMed ID: [22719221](#)]. [PubMed Central ID: [PMC3372885](#)]. <https://doi.org/10.7150/ijbs.4439>.
  29. Zampetaki A, Kiechl S, Drozdov I, Willeit P, Mayr U, Prokopi M, et al. Plasma microRNA profiling reveals loss of endothelial miR-126 and other microRNAs in type 2 diabetes. *Circ Res*. 2010;**107**(6):810-7. [PubMed ID: [20651284](#)]. <https://doi.org/10.1161/CIRCRESAHA.110.226357>.
  30. Fish JE, Santoro MM, Morton SU, Yu S, Yeh RF, Wythe JD, et al. miR-126 regulates angiogenic signaling and vascular integrity. *Dev Cell*. 2008;**15**(2):272-84. [PubMed ID: [18694566](#)]. [PubMed Central ID: [PMC2604134](#)]. <https://doi.org/10.1016/j.devcel.2008.07.008>.
  31. Harris TA, Yamakuchi M, Ferlito M, Mendell JT, Lowenstein CJ. MicroRNA-126 regulates endothelial expression of vascular cell adhesion molecule 1. *Proc Natl Acad Sci U S A*. 2008;**105**(5):1516-21. [PubMed ID: [18227515](#)]. [PubMed Central ID: [PMC2234176](#)]. <https://doi.org/10.1073/pnas.0707493105>.
  32. Nicoli S, Standley C, Walker P, Hurlstone A, Fogarty KE, Lawson ND. MicroRNA-mediated integration of haemodynamics and Vegf signalling during angiogenesis. *Nature*. 2010;**464**(7292):1196-200. [PubMed ID: [20364122](#)]. [PubMed Central ID: [PMC2914488](#)]. <https://doi.org/10.1038/nature08889>.
  33. Nikolic I, Plate KH, Schmidt MHH. EGFL7 meets miRNA-126: an angiogenesis alliance. *J Angiogenesis Res*. 2010;**2**(1):9. [PubMed ID: [20651284](#)].

- 20529320]. [PubMed Central ID: PMC2901201]. <https://doi.org/10.1186/2040-2384-2-9>.
34. Qiang L, Hong L, Ningfu W, Huaihong C, Jing W. Expression of miR-126 and miR-508-5p in endothelial progenitor cells is associated with the prognosis of chronic heart failure patients. *Int J Cardiol.* 2013;**168**(3):2082-8. [PubMed ID: 23465244]. <https://doi.org/10.1016/j.ijcard.2013.01.160>.
35. GHERASIM L. Association of atrial fibrillation with diabetes mellitus, high risk comorbidities. *J Maedica.* 2022;**17**(1):143.
36. Echouffo-Tcheugui JB, Shrader P, Thomas L, Gersh BJ, Kowey PR, Mahaffey KW, et al. Care Patterns and Outcomes in Atrial Fibrillation Patients With and Without Diabetes: ORBIT-AF Registry. *J Am Coll Cardiol.* 2017;**70**(11):1325-35. [PubMed ID: 28882229]. <https://doi.org/10.1016/j.jacc.2017.07.755>.
37. Urbich C, Kuehbacher A, Dimmeler S. Role of microRNAs in vascular diseases, inflammation, and angiogenesis. *Cardiovasc Res.* 2008;**79**(4):581-8. [PubMed ID: 18550634]. <https://doi.org/10.1093/cvr/cvn156>.
38. Hu J, Zeng L, Huang J, Wang G, Lu H. miR-126 promotes angiogenesis and attenuates inflammation after contusion spinal cord injury in rats. *Brain Res.* 2015;**1608**:191-202. [PubMed ID: 25724143]. <https://doi.org/10.1016/j.brainres.2015.02.036>.
39. Tang ST, Wang F, Shao M, Wang Y, Zhu HQ. MicroRNA-126 suppresses inflammation in endothelial cells under hyperglycemic condition by targeting HMGB1. *Vascul Pharmacol.* 2017;**88**:48-55. [PubMed ID: 27993686]. <https://doi.org/10.1016/j.vph.2016.12.002>.
40. Kilic U, Gok O, Bacaksiz A, Izmirli M, Elibol-Can B, Uysal O. SIRT1 gene polymorphisms affect the protein expression in cardiovascular diseases. *PLoS One.* 2014;**9**(2). e90428. [PubMed ID: 24587358]. [PubMed Central ID: PMC3938716]. <https://doi.org/10.1371/journal.pone.0090428>.
41. Rane S, He M, Sayed D, Vashistha H, Malhotra A, Sadoshima J, et al. Downregulation of miR-199a derepresses hypoxia-inducible factor-1 $\alpha$  and Sirtuin 1 and recapitulates hypoxia preconditioning in cardiac myocytes. *Circ Res.* 2009;**104**(7):879-86. [PubMed ID: 19265035]. [PubMed Central ID: PMC3332328]. <https://doi.org/10.1161/CIRCRESAHA.108.193102>.
42. Sun XL, Bu PL, Liu JN, Wang X, Wu XN, Zhao LX. Expression of SIRT1 in right auricle tissues and the relationship with oxidative stress in patients with atrial fibrillation. *Chinese J Cell Molecular Immunol.* 2012;**28**(9):972-4.
43. Chiang DY, Zhang M, Voigt N, Alsina KM, Jakob H, Martin JF, et al. Identification of microRNA-mRNA dysregulations in paroxysmal atrial fibrillation. *Int J Cardiol.* 2015;**184**:190-7. [PubMed ID: 25706326]. [PubMed Central ID: PMC4417399]. <https://doi.org/10.1016/j.ijcard.2015.01.075>.