



Comparison of PfHRP-2/pLDH RDTs with Light Microscopy in a Low Prevalence Setting in Southeastern Iran, Sistan and Baluchestan: Due to Implementation of Malaria Elimination Program

Mansour Rahmati Balaghaleh,¹ Mehdi Zarean,¹ Monnavar Afzal Aghae,² Seyed Aliakbar Shamsian,^{1,*} Hadi Mirahmadi,^{3,**} and Arslaan Arya⁴

¹Departments of Mycology and Parasitology, School of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran

²Department of Biostatistics and Epidemiology, Faculty of Health Medicine, Mashhad University of Medical Sciences, Mashhad, Iran

³Department of Parasitology and Mycology, Zahedan University of Medical Sciences, Zahedan, Iran

⁴School of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran

*Corresponding author: Seyed Aliakbar Shamsian, Departments of Mycology and Parasitology, School of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran. E-mail: shamsianaa@mums.ac.ir

**Corresponding author: Hadi Mirahmadi, Department of Parasitology and Mycology, Zahedan University of Medical Sciences, Zahedan, Iran. E-mail: hmiraahmadi59@gmail.com

Received 2017 May 17; Revised 2017 October 29; Accepted 2017 November 01.

Abstract

Background: The gold standard for diagnosing malaria in Iran is using light microscopy; however, this method requires equipment, time and skilled microscopists. In some circumstances, rapid diagnostic test (RDTs) can be a proper substitute. Assessing the diagnostic performance of RDT relative to microscopy, for the diagnosis of malaria in Southeastern Iran, Sistan and Baluchestan, was the purpose of this study.

Methods: From March to July 2016, which was a peak transmission season in Iran, 318 patients with suspected malaria were taken for a blood sample at the health center in Saravan, in Sistan and Baluchestan Province. The samples were surveyed instantly by light microscopy and RDT. Comparison analysis included: sensitivity, specificity, positive and negative predictive values, and other diagnostic screening performance measures for detecting malaria infections.

Results: Of the 318 malaria cases (6.6%), 21 were identified by a microscopy compared to 22 of 318 (6.9%) by RDT. RDT sensitivity and specificity for the diagnosis of malaria were 95.5% (77.2% - 99.9%, 95% CI) and 100% (98.8% - 100%, 95% CI), respectively, compared to standard microscopy, the sensitivity and specificity for *vivax* malaria were 100% (79.4-100%, 95% CI) and 99.7% (98.14% - 99.99%, 95% CI), and for *falciparum* malaria: 80% (28.4% - 99.5%, 95% CI) and 100% (98.8% - 100%, 95% CI). Two tests showed an amazing agreement with a kappa estimation of 0.975.

Conclusions: The study proved that the RDT test performs appropriately for the identification of infections with *P. falciparum* and *P. vivax* malaria in cross-border malaria. Moreover, the RDT can be a useful instrument for the identification of new clinical cases of malaria in this region.

Keywords: Malaria, Light Microscopy, Rapid Diagnosis Test, Elimination, Iran

1. Background

Malaria, transmitted by female Anopheles species mosquitoes, is one of the most significant parasitic diseases for individuals (1). World health organization (WHO) stated lately that 438000 cases out of 214 million malaria cases have resulted in death in 2015. Attributable to the endeavors worldwide, Malaria is particularly constantly one of the main health challenges in developing countries (2). It is considered as one of the leading bases of attention for public health in several countries including Iran, especially in south and south-east regions of the country (3). Much endeavors and progress has been made by

the WHO to control and eradicate malaria (4). Iran is a malaria endemic area and malaria is regarded as a main health medical challenge in this area (5, 6). Frequency of malaria has recently declined as a result of the endeavors of health establishments and specialists in Sistan and Baluchestan Province (5). Whereas parasite-based identification is growing, most doubted cases of malaria are not suitably diagnosed and thus disease monitoring continues to be obscure (7). This method entails equipment, time, and experienced microscopists, even if microscopic checkup of a tainted peripheral blood smear (PBS) is regarded as the gold standard for malaria diagnosis (8). It

was also revealed that in most secondary health structures, the quality and value of diagnosis by field microscopy was not satisfactory, which leads clinicians to treat patients for malaria regardless of the outcome of the microscopy (9). The PfHRP-2/pLDH RDTs are on the basis of immune chromatography, *P. falciparum*-specific histidine rich protein 2 (pfHRP-2), and solubility in water and can make out the type of *P. falciparum*. Besides, in sexual and nonsexual stages in all *Plasmodium* species *Plasmodium*, lactate dehydrogenase (PLDH) is produced (10, 11). Nevertheless, alternative rapid diagnostic tests (RDTs) have been improved for use in non-endemic regions and rural areas, where trained microscopists are less accessible, and endemic regions for controlling and elimination programs of malaria (12). However, the result should be approved by a microscopic method (8). The goal of this study was conducted to evaluate the sensitivity and specificity of RDTs in line with the conventional light microscopy (LM) in Southeastern Iran, Sistan and Baluchestan.

2. Methods

2.1. Study Area and Period

We conducted a cross-sectional study at a health center in Saravan, a town in Sistan and Baluchestan Province (27° 22' 15" N, 62° 20' 3" E, 1165 m above sea level) (Figure 1) from March to July 2016, which was a peak transmission season in Iran. The population of Saravan was about 175728 in 2011. The majority of its population depends on subsistence farming. Furthermore, the potential for transmission of malaria in these areas is due to population displacement with the eastern neighboring country (Pakistan), appropriate climatic conditions suitable for malaria vectors survival, and low socioeconomic conditions. Saravan health center is one of the health centers in the district where people indicating symptoms of malaria get free diagnostic services and treatment. Despite the presence of different control activities in the area, it has remained endemic for both *P. vivax* and *P. falciparum* malaria.

2.2. Study Subjects

During the study period, 318 patients, in Saravan health center, suspected of the malaria infection, were included and screened using light microscopy (LM).

2.3. Rapid Diagnostic Test

The PfHRP-2/pLDH RDT is a quick, qualitative immunoassay, parallel stream cassette gadget that utilizes 5 μ L entire blood for the identification of *P. falciparum*-specific histidine rich protein-2 (*P. falciparum*-HRP2), and pan-specific pLDH for all *Plasmodium* species. The RDT was

utilized by following the manufacturers directions offered in the product insert (First Response Malaria Ag. (pLDH / HRP2) Combo Rapid Diagnostic Test, INDIA). Reading and elucidation of test outcomes were conducted by 2 skilled technicians and elucidated by concord within the given 15 - 30 minutes test window. No test scores surpassed the 30 minutes limit. Any test that was unsuccessful to make a control band was regarded as unacceptable and the test was conducted again. Test interpretation criteria are presented in Figure 2.

2.4. Microscopy

Microscopy was applied as the suggesting and direction standard in this study. With each RDT, we arranged a corresponding thick and thin blood film and stained them with the Giemsa solution (1:10 dilution for approximately 20 minutes). We studied the slides utilizing a compound light microscopy under $\times 1000$ oil-immersion magnification by a skilled laboratory professional in the center. All slides, prior to being accounted for as either negative for malaria parasites and for the recognition of low density mixed species infections, were checked for at least 100 high-magnification fields. We accounted parasite numbers for 200 white blood cells (WBC) to evaluate parasite thickness per μ L of blood, supposing a standard mean WBC count of 8000/ μ L blood. Afterward, as an observer-blinded assessment, all slides were scrutinized by a self-sufficient practiced malaria microscopist.

2.5. Statistical Analysis

The diagnostic performance of RDT was evaluated using the microscopic method as the gold standard. We carried out the statistical analysis using SPSS 13.0 software (SPSS Inc, Chicago, IL, USA). We determined sensitivity, specificity, PPV, and NPV for both tests and compared them with one another. Kappa value was determined to see if the results among the diagnostic tools were consistent. Chi-square was used to test for significant differences between gender, age group, occupation, residence, and nationality with parasite positivity by the LM.

2.6. Ethical Approval

The study protocol was approved by the research ethics committee of Mashhad University of Medical Sciences, Mashhad, Iran (IR.MUMS.fm.REC.1395.440). Oral consent was obtained from the participants before sampling. Participants who were diagnosed with the malaria infection were given the appropriate treatment at the health center.



Figure 1. Geographical Locations Where This Study Was Carried Out

3. Results

We included a total of 318 malaria suspected patients in the study. The male to female ratio was 1.37:1 while the mean age (SD, range) of the participants was 29.1 years (22, 1 - 95). Most of the individuals participating in the test were from rural areas of the district (58.5%). Table 1 describes parasite positive results detected by LM among different sociodemographic factors of patients. Chi square test showed the sex and resident of the participants were not significantly associated with parasite positivity, however, nationality was associated with parasite positivity (Table 1). The overall parasite positivity using LM was 21 (6.6%): 4 (1.3%) for *P. falciparum*, 16 (5%) for *P. vivax*, 1 (0.3%) for *P. malariae*, and no detections of any mixed infections. Using the RDT, the total parasite positivity was 22 (6.9%): 5 (1.6%) for *P. falciparum*, 17 (5.3%) for all *Plasmodium* species. Differences in the detection of malaria parasites via either the LM or the RDT was insignificant ($P < 0.001$).

Using the LM as a standard test for malaria, we found the sensitivity and specificity of RDT to be 95.5% (77.2% - 99.9%, 95% CI) and 100% (98.8% - 100%, 95% CI), respectively. We found the positive predictive value (PPV) and the neg-

ative predictive value (NPV) to be 100% (80.8% - 100%, 95% CI) and 99.7% (97.8% - 99.9%, 95% CI), respectively. With a Kappa value of 0.975, we established an outstanding agreement between the LM and RDT. Accordingly, we calculated the sensitivity of the RDT for *P. falciparum* compared to LM to be 80% (28.4% - 99.5%, 95% CI) while a sensitivity of 100% (79.4% - 100%, 95% CI) was found for *P. vivax* species. We found the specificity for *P. falciparum* to be 100% (98.8% - 100%, 95% CI) while it was 99.7% (98.14% - 99.99%, 95% CI) for *P. vivax* species (Table 2). We also noticed a remarkable agreement between the RDT and LM in identifying diverse species of *Plasmodium*: Kappa value of 0.887 for *P. falciparum* and Kappa value of 0.968 for *P. vivax* species.

4. Discussion

Saravan is a border city (Having a common frontier with Pakistan) where for the ties and kinship of the residence of the 2 sides, especially in border villages, a high rate of commute and frequentation is observed and as a result, the transmission of Malaria to Iran is more likely. Moreover, due to geographical dispersion and scattering

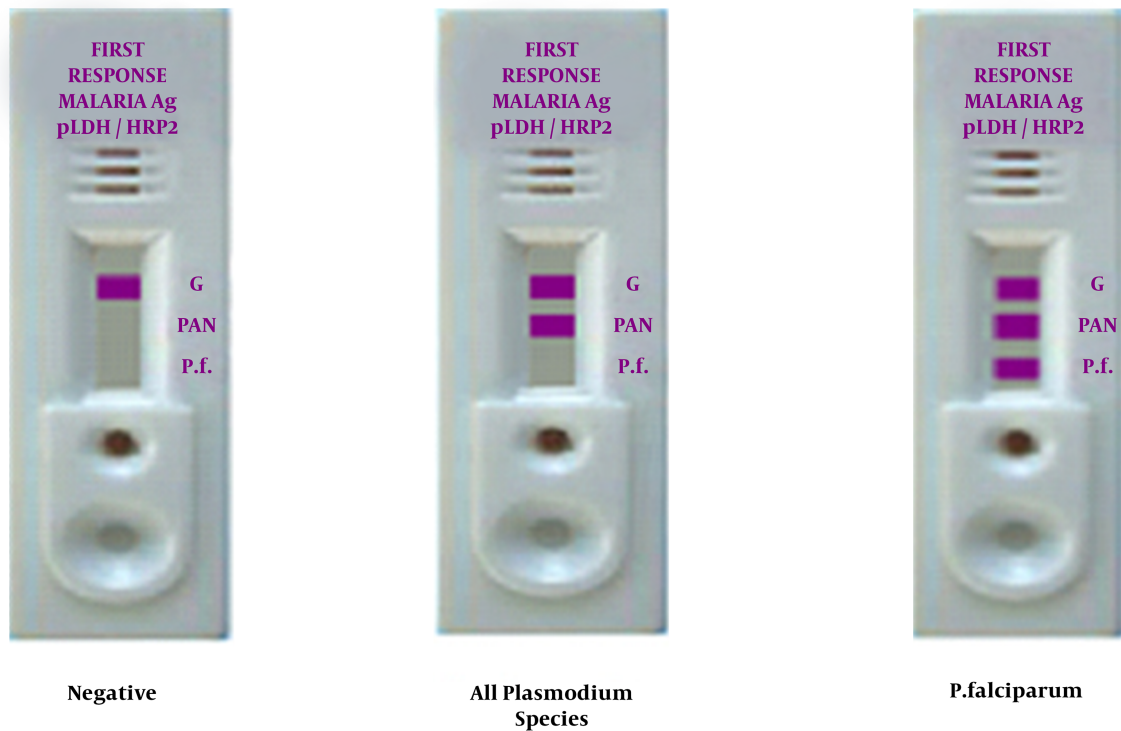


Figure 2. RDT Test for Determination of Malaria Infection to Malaria Suspected Patients in Saravan Health Center

the villages lack of laboratory facilities as well as skilled microscopists, the most significant strategy of Malaria comprehensive control and elimination program in Iran is the rapid diagnosis of Malaria in such places (13). Hence, rapid malaria diagnosis is an essential factor of the national malaria control plan in Iran, which depends on the utilization of LM and RDTs. The present study revealed a high sensitivity and specificity of the RDT (Table 2). The important finding of this study was that the sensitivity and specificity of RDT was respectively 95.5% and 100% in comparison to the gold standard microscopy method for detecting malaria. The high sensitivity of the RDT in this study were corresponding with other studies from Jimma, southwest Ethiopia (14), Madagascar (15), and northwest Ethiopia (16). The current study showed higher sensitivity and specificity than reports from China (17) and Ashton et al., from Ethiopia (16). The high specificity of the RDT in this study corresponded with another study from Jimma, southwestern Ethiopia (14). Furthermore, the present study revealed higher specificity than reports from Senegal (9), China (17) and northwest Ethiopia (16). These distinctions could be because of variant observer, differences with malaria species circulating at various regions or host elements the

number of positive cases in this study was low (18).

The great execution of the *Pf*HRP-2/pLDH RDT in the field is obviously displayed by its capacity to recognize *P. falciparum* in every single diverse level of microscopic parasite densities (19). The sensitivity of the RDT in the current study for *P. falciparum* corresponded with the reports by Bruno B Andrade et al. (20). The sensitivity of the RDT in the current study for *P. falciparum* was also higher than Maltha et al., (21) and lower than China Myanmar endemic borders (17), Indonesia (22) and Jimma, as well as southwest Ethiopia (14). The specificity of the RDT in the present study for *P. falciparum* was similar to the reports from Jimma, southwestern Ethiopia (14), and higher than the reports from southern (23) and Ethiopia and Madagascar (15, 24). WHO, in the 2000 epecificity examination of these two strategies disclosed that RDT has a specificity of 100% in diagnosing *P. falciparum* (25). In spite of the fact that *P. falciparum*, not stating *Pf*HRP2, has been accounted from some regions (26), the *pfrp2* gene was available in most of the circulating *P. falciparum* parasites in southeast Iran. Among other reasons for *P. falciparum*, in this study, we may refer to the point that Iran is in the stage of eliminating malaria.

Table 1. Parasite Positivity as Detected by Light Microscopy Among Different Socio-Demographic Factors of Patients Using Chi-Square Analysis^a

Variables	Parasite Positivity by the LM					
	No. tested%	No. positive%	No. negative%	P Value	OR	95% CI
Gender				0.19	1.9	0.72 - 5.02
Male	184 (57.9)	15 (8.2)	169 (91.8)			
Female	134 (42.1)	6 (4.5)	128 (95.5)			
Age, y						
< 5	51 (16)	1 (1.9)	50 (98.1)	0.15	0.23	0.03 - 1.75
≥ 5 and < 15	44 (13.8)	2 (4.5)	42 (95.5)	0.42	0.54	0.12 - 2.43
≥ 15 ^b	223 (70.2)	18 (8.1)	205 (91.9)			
Occupation						
Non schooled child ^b	60 (18.9)	2 (3.3)	58 (96.7)	0.54		
Students	48 (15.1)	2 (4.2)	46 (95.8)	0.26	1.13	0.15 - 8.35
Housewife	77 (24.2)	5 (6.5)	72 (93.5)	0.41	1.8	0.34 - 9.67
Farmer and worker	28 (8.8)	4 (14.3)	24 (85.7)	0.74	4.33	0.74 - 25.31
Officer and Teacher	2 (0.6)	0	2 (100)	0.29	0	0
Trader/Driver	103 (32.4)	4 (3.9)	99 (96.1)	0.99	1.05	0.19 - 5.93
Residence				0.04	0.31	0.1 - 0.95
Urban	132 (41.5)	4 (3)	128 (97)			
Rural	186 (58.5)	17 (9.1)	169 (90.9)			
Nationality				0.0001	0.01	0.003 - 0.09
Iranian	310 (97.5)	15 (4.8)	295 (95.2)			
Non-Iranian	8 (2.5)	6 (75)	2 (25)			

Abbreviations: CI, Confidence Interval; LM, Light Microscopy; OR, Odds Ratio.

^aValues are expressed as No. (%).

^bReference; 0.74.

Table 2. Performance Characteristics of RDT by Species Identified in Comparison to the Gold Standard Reference Light Microscopy

Microscopy	RDT			
	<i>P. falciparum</i>		<i>P. vivax</i>	
	Positive	Negative	Positive	Negative
Positive	4	0	16	1
Negative	1	296	0	296
Sensitivity (95% IC),%	80 (28.4 - 99.5)		100 (79.41 - 100)	
Specificity (95% IC),%	100 (98.8 - 100)		99.7(98.14 - 99.99)	
Positive predictive value (95% IC),%	100 (39.5 - 100)		94.1(69.34 - 99.12)	
Negative predictive value (95% IC),%	99.7 (98 - 99.94)		100(98 - 100)	
Kappa value	0.887		0.968	

Abbreviation: CI, Confidence Interval.

The sensitivity for the *P. vivax* in the current study was higher in China-Myanmar endemic borders (17), Indone-

sia, and the reports by Bruno B Andrade et al. (20). The specificity for the *P. vivax* in the current study was sim-

ilar to the reports of some studies (16, 22) and lower in others (14, 17, 20). For sensitivity and specificity examination of PfHRP-2 and PLDH, tests in recognizing *P. falciparum*, Iqbal J et al., outlined that the sensitivity of PfHRP-2 was more than PLDH; nonetheless, specificity of PLDH was more than PfHRP-2 (27, 28). The distinctions in the specificity of the RDT could be because of the above-mentioned grounds (18). High NPV of the RDT suggests that it was reliable on eliminating malaria. Likewise, the higher PPV suggests that patients will be detected positive more accurately for malaria and will keep away superfluous treatments. Its applicability and rapidity plays a significant role in diagnosing both *falciparum* and *vivax* malaria identification, rapid case administration, as well as cross borders malaria observing in the Iran endemic territories in the National Malaria Elimination program. By and large, the RDT proved great sensitivity and specificity when compared to the LM.

4.1. Conclusion

The study proved that the RDT test carries out appropriately well for the identification of *P. falciparum* and *P. vivax* malaria infections in cross-border malaria. Moreover, the RDT can be a helpful instrument for the identification of new clinical instances of malaria in this region. The RDT could, in this manner, be utilized instead of LM, which in poor layouts cannot be utilized regularly.

Acknowledgments

The authors would like to thank Dr. Fata from Mashhad University of Medical Sciences for kindly cooperating. This work is a part of the M.Sc thesis of Mansour Rahmati Balaghaleh. It is financially supported by a Grant (950273) from Mashhad University of Medical Sciences and is fully appreciated.

References

- White NJ, Pukrittayakamee S, Hien TT, Faiz MA, Mokuolu OA, Don-dorp AM. Malaria. *Lancet*. 2014;**383**(9918):723-35. doi: [10.1016/S0140-6736\(13\)60024-0](https://doi.org/10.1016/S0140-6736(13)60024-0). [PubMed: [23953767](https://pubmed.ncbi.nlm.nih.gov/23953767/)].
- Sheikhzadeh K, Haghdoost AA, Bahrampour A, Zolala F, Raeisi A. Assessment of the impact of the malaria elimination programme on the burden of disease morbidity in endemic areas of Iran. *Malar J*. 2016;**15**:209. doi: [10.1186/s12936-016-1267-9](https://doi.org/10.1186/s12936-016-1267-9). [PubMed: [27074734](https://pubmed.ncbi.nlm.nih.gov/27074734/)].
- Edrissian G. Malaria in Iran, past and present situation. *Iran J Parasitol*. 2006;**1**(1):14.
- Chiyaka C, Tatem AJ, Cohen JM, Gething PW, Johnston G, Gosling R, et al. Infectious disease. The stability of malaria elimination. *Science*. 2013;**339**(6122):909-10. doi: [10.1126/science.1229509](https://doi.org/10.1126/science.1229509). [PubMed: [23430640](https://pubmed.ncbi.nlm.nih.gov/23430640/)].
- Salimi Khorashad A, Salehi M, Roshanravan B. The comparison of microscopic method and rapid diagnostic test in detecting plasmodium species. *Int J Infect*. 2014;**1**(3). doi: [10.17795/iji-21441](https://doi.org/10.17795/iji-21441).
- Abdolah K, Hosein ZN, Shahabi S, Hushang K, Haghghi A, Raisii A, et al. Short communication, comparison of microscopy and RDTs techniques for laboratory detection of malaria. *Afr J Biotechnol*. 2010;**9**(10):1514-6. doi: [10.5897/ajb09.1155](https://doi.org/10.5897/ajb09.1155).
- McMorrow ML, Masanja MI, Abdulla SM, Kahigwa E, Kachur SP. Challenges in routine implementation and quality control of rapid diagnostic tests for malaria-Rufiji District, Tanzania. *Am J Trop Med Hyg*. 2008;**79**(3):385-90. [PubMed: [18784230](https://pubmed.ncbi.nlm.nih.gov/18784230/)].
- Singh N, Saxena A, Valecha N. Field evaluation of the ICT malaria Pf/Pv immunochromatographic test for diagnosis of Plasmodium falciparum and P.vivax infection in forest villages of Chhindwara, central India. *Trop Med Int Health*. 2000;**5**(11):765-70. [PubMed: [11123823](https://pubmed.ncbi.nlm.nih.gov/11123823/)].
- Faye B, Nath-Chowdhury M, Tine RC, Ndiaye JL, Sylla K, Camargo FW, et al. Accuracy of HRP2 RDT (Malaria Antigen Pf(R)) compared to microscopy and PCR for malaria diagnosis in Senegal. *Pathog Glob Health*. 2013;**107**(5):273-8. doi: [10.1179/204773213Y.0000000102](https://doi.org/10.1179/204773213Y.0000000102). [PubMed: [23916337](https://pubmed.ncbi.nlm.nih.gov/23916337/)].
- Hopkins H, Kambale W, Kamya MR, Staedke SG, Dorsey G, Rosenthal PJ. Comparison of HRP2- and pLDH-based rapid diagnostic tests for malaria with longitudinal follow-up in Kampala, Uganda. *Am J Trop Med Hyg*. 2007;**76**(6):1092-7. [PubMed: [17556616](https://pubmed.ncbi.nlm.nih.gov/17556616/)].
- Wongsrichanalai C, Barcus MJ, Muth S, Sutamihardja A, Wernsdorfer WH. A review of malaria diagnostic tools: microscopy and rapid diagnostic test (RDT). *Am J Trop Med Hyg*. 2007;**77**(6 Suppl):119-27. [PubMed: [18165483](https://pubmed.ncbi.nlm.nih.gov/18165483/)].
- Beadle C, Long GW, Weiss WR, McElroy PD, Maret SM, Oloo AJ, et al. Diagnosis of malaria by detection of Plasmodium falciparum HRP-2 antigen with a rapid dipstick antigen-capture assay. *Lancet*. 1994;**343**(8897):564-8. [PubMed: [7906328](https://pubmed.ncbi.nlm.nih.gov/7906328/)].
- Hanafi-Bojd AA, Vatandoost H, Oshaghi MA, Eshraghian MR, Haghdoost AA, Abedi F, et al. Knowledge, attitudes and practices regarding malaria control in an endemic area of southern Iran. *Southeast Asian J Trop Med Public Health*. 2011;**42**(3):491-501. [PubMed: [21706926](https://pubmed.ncbi.nlm.nih.gov/21706926/)].
- Mekonnen Z, Ali S, Belay G, Suleman S, Chatterjee S. Evaluation of the performance of CareStart Malaria Pf/Pv Combo rapid diagnostic test for the diagnosis of malaria in Jimma, southwestern Ethiopia. *Acta Trop*. 2010;**113**(3):285-8. doi: [10.1016/j.actatropica.2009.12.001](https://doi.org/10.1016/j.actatropica.2009.12.001). [PubMed: [20005196](https://pubmed.ncbi.nlm.nih.gov/20005196/)].
- Ratsimbaoa A, Randriamanantena A, Raheerinjafy R, Rasoarilalao N, Menard D. Which malaria rapid test for Madagascar? Field and laboratory evaluation of three tests and expert microscopy of samples from suspected malaria patients in Madagascar. *Am J Trop Med Hyg*. 2007;**76**(3):481-5. [PubMed: [17360871](https://pubmed.ncbi.nlm.nih.gov/17360871/)].
- Moges B, Amare B, Belyhun Y, Tekeste Z, Gizachew M, Workineh M, et al. Comparison of CareStart HRP2/pLDH COMBO rapid malaria test with light microscopy in north-west Ethiopia. *Malar J*. 2012;**11**:234. doi: [10.1186/1475-2875-11-234](https://doi.org/10.1186/1475-2875-11-234). [PubMed: [22818643](https://pubmed.ncbi.nlm.nih.gov/22818643/)].
- Xiaodong S, Tambo E, Chun W, Zhibin C, Yan D, Jian W, et al. Diagnostic performance of CareStart malaria HRP2/pLDH (Pf/pan) combo test versus standard microscopy on falciparum and vivax malaria between China-Myanmar endemic borders. *Malar J*. 2013;**12**:6. doi: [10.1186/1475-2875-12-6](https://doi.org/10.1186/1475-2875-12-6). [PubMed: [23294729](https://pubmed.ncbi.nlm.nih.gov/23294729/)].
- Central Statistical Agency, ORC Macro. *Ethiopia demographic and health survey 2005*. Addis Ababa, Ethiopia: Central Statistical Agency, (Ethiopia) and ORC Macro; 2006.
- Laban NM, Kobayashi T, Hamapumbu H, Sullivan D, Mharakurwa S, Thuma PE, et al. Comparison of a PfHRP2-based rapid diagnostic test and PCR for malaria in a low prevalence setting in rural southern Zambia: implications for elimination. *Malar J*. 2015;**14**:25. doi: [10.1186/s12936-015-0544-3](https://doi.org/10.1186/s12936-015-0544-3). [PubMed: [25888818](https://pubmed.ncbi.nlm.nih.gov/25888818/)].
- Andrade BB, Reis Filho A, Barros AM, Souza Neto SM, Nogueira LL, Fukutani KF, et al. Towards a precise test for malaria diagnosis in the Brazilian Amazon, comparison among field microscopy, a rapid diagnostic test, nested PCR, and a computational expert system based on artificial neural networks. *Malar J*. 2010;**9**(117):1-11. doi: [10.1186/1475-2875-9-117](https://doi.org/10.1186/1475-2875-9-117).

21. Maltha J, Gillet P, Bottieau E, Cnops L, van Esbroeck M, Jacobs J. Evaluation of a rapid diagnostic test (CareStart Malaria HRP-2/pLDH (Pf/pan) Combo Test) for the diagnosis of malaria in a reference setting. *Malar J.* 2010;**9**:171. doi: [10.1186/1475-2875-9-171](https://doi.org/10.1186/1475-2875-9-171). [PubMed: [20565816](https://pubmed.ncbi.nlm.nih.gov/20565816/)].
22. Fransisca L, Kusnanto JH, Satoto TB, Sebayang B, Andriyan E, et al. Supriyanto. Comparison of rapid diagnostic test Plasmodium Malaria-3, microscopy, and quantitative real-time PCR for diagnoses of Plasmodium falciparum and Plasmodium vivax infections in Mimika Regency, Papua, Indonesia. *Malar J.* 2015;**14**:103. doi: [10.1186/s12936-015-0615-5](https://doi.org/10.1186/s12936-015-0615-5). [PubMed: [25890368](https://pubmed.ncbi.nlm.nih.gov/25890368/)].
23. Sharew B, Legesse M, Anmut A, Jima D, Medhin G, Erko B. Evaluation of the performance of CareStart Malaria Pf/Pv Combo and Paracheck Pf tests for the diagnosis of malaria in Wondo Genet, southern Ethiopia. *Acta Trop.* 2009;**111**(3):321-4. doi: [10.1016/j.actatropica.2009.05.014](https://doi.org/10.1016/j.actatropica.2009.05.014). [PubMed: [19482001](https://pubmed.ncbi.nlm.nih.gov/19482001/)].
24. Ashton RA, Kefyalew T, Tesfaye G, Counihan H, Yadeta D, Cundill B, et al. Performance of three multi-species rapid diagnostic tests for diagnosis of Plasmodium falciparum and Plasmodium vivax malaria in Oromia Regional State, Ethiopia. *Malar J.* 2010;**9**:297. doi: [10.1186/1475-2875-9-297](https://doi.org/10.1186/1475-2875-9-297). [PubMed: [20979601](https://pubmed.ncbi.nlm.nih.gov/20979601/)].
25. Durrheim DN, Govere J, la Grange JJ, Mabuza A. Rapid immunochromatographic diagnosis and Rolling Back Malaria-experiences from an African control program. *Afr J Med Med Sci.* 2001;**30** Suppl:21-4. [PubMed: [14513934](https://pubmed.ncbi.nlm.nih.gov/14513934/)].
26. Cheng Q, Gatton ML, Barnwell J, Chiodini P, McCarthy J, Bell D, et al. Plasmodium falciparum parasites lacking histidine-rich protein 2 and 3: a review and recommendations for accurate reporting. *Malar J.* 2014;**13**:283. doi: [10.1186/1475-2875-13-283](https://doi.org/10.1186/1475-2875-13-283). [PubMed: [25052298](https://pubmed.ncbi.nlm.nih.gov/25052298/)].
27. Iqbal J, Hira PR, Sher A, Al-Enezi AA. Diagnosis of imported malaria by Plasmodium lactate dehydrogenase (pLDH) and histidine-rich protein 2 (PfHRP-2)-based immunocapture assays. *Am J Trop Med Hyg.* 2001;**64**(1-2):20-3. [PubMed: [11425156](https://pubmed.ncbi.nlm.nih.gov/11425156/)].
28. Jelinek T, Grobusch MP, Nothdurft HD. Use of dipstick tests for the rapid diagnosis of malaria in nonimmune travelers. *J Travel Med.* 2000;**7**(4):175-9. [PubMed: [11003728](https://pubmed.ncbi.nlm.nih.gov/11003728/)].