Published online 2016 June 13.

Research Article

Soil Contamination With Free-Living Amoeba in North of Iran

Mohammad Ali Mohaghegh,¹ Mojtaba Azimi Resketi,¹ Reza Mohammadimanesh,¹ Mehdi Azami,²

Farzaneh Mirzaie,^{1,3} Mohammad Falahati,¹ Somayeh Jahani,⁴ and Mohsen Ghomashlooyan^{1,*}

¹Department of Parasitology and Mycology, School of Medicine, Isfahan University of Medical Sciences, Isfahan, IR Iran ²Skin Diseases and Leishmaniasis Research Center, Isfahan University of Medical Sciences, Isfahan, IR Iran

³School of Paramedicine, Shahid Sadoughi University of Medical Sciences, Yazd, IR Iran

⁴ Infectious Diseases and Tropical Medicine Research Center, Zahedan University of Medical Sciences, Zahedan, IR Iran

^{*} *Corresponding author*: Mohsen Ghomashlooyan, Department of Parasitology and Mycology, School of Medicine, Isfahan University of Medical Sciences, Isfahan, IR Iran. Tel: +98-9355530151, E-mail: ghomashlooyan@med.mui.ac.ir

Received 2016 March 17; Revised 2016 April 12; Accepted 2016 April 15.

Abstract

Background: Free-living amoebas are very abundant in nature, especially in water and soil. Some species of amoebas cause serious and sometimes fatal infections such a keratitis in people with normal and suppressed immune systems. Based on the prevalence of amoebic keratitis in Iran, it is important to consider the free-living amoeba.

Objectives: The current study aimed to determine the presence of *Acanthamoeba* and *Hartmannella* spp. in soil samples of Sari, North of Iran.

Patients and Methods: A total of 96 soil samples from three areas of Sari were collected to be examined for soil contamination with *Acanthamoeba* and *Hartmannella* spp. Soil samples were investigated for the presence of these parasites by Sheather's flotation technique and Gimsa staining method. The identification of *Acanthamoeba* and *Hartmannella* spp. at the genus level in this study was based on distinctive features of double walled cysts.

Results: Of the 96 studied samples, from different environmental locations in Sari, 38 (39.6%) *Acanthamoeba* and 5 (5.2%) *Hartmannella* species were identified.

Conclusions: The results of the present study revealed that soil resources of these areas were contaminated with opportunistic amoebas such as *Acanthamoeba* and *Hartmannella* spp. and this may lead to severe diseases in high-risk people, such as immuno-compromised patients.

Keywords: Soil Contamination, Acanthamoeba spp, Hartmannella spp

1. Background

With the exception of Entamoeba histolytica which may possibly cause brain, lung and liver abscess from a primary focus on the large intestine, pathogenic amoeba species such as Acanthamoeba spp., Naegleria folweri and Balamunthia mandrillaris can cause central nervous system disability called primary amoebic meningoencephalitis (PAM) and granulomatous amoebic encephalitis (GAE) (1, 2). According to some studies, Acanthamoeba species can be classified based on their structure (3). The current study identified 20 species so far (4). Cystic form of these amoebas can survive for many years in the environment, including water, soil, sewage and dust (5-8). Because of the wide distribution, humans can be easily exposed to these amoebas; therefore, they should be separated from skin and nose of healthy people (9). Acanthamoeba cysts are the main cause of amoeba transmission. Cysts can enter the body tissues through water, soil and dust from the outside

or from a primary focus in the lung, nose and skin ulcers and cause their virulence. *Acanthamoeba* species have relatively slow tissue invasion; therefore, granuloma formations are observed in their tissue infection. Chronic granulomatous lesions of pathogenic species of *Acanthamoeba* are reported in the tissues of brain, kidney, liver, spleen, skin, uterus and prostate (10).

Ocular lesions mainly caused by the invasion of *Acanthamoeba* spp. occur with corneal ulcers and keratitis (11). If amoebic keratitis not treated, the ulcers lead to stromal puncture, loss of vision and eventually cause blindness. In rare cases, *Acanthamoeba* spp. could be permeating to retina and cause chorioretinitis (12). Molecular studies showed that most of the amoebic keratitis cases in Iran were *A. castellani*, *A. palestiniensis* and *A. griffini* (13).

In some people, especially in HIV positive patients, *Acanthamoeba* spp. can include chronic skin ulcers, abscesses or erythematous nodules, especially in the lung. Skin ulcers caused by *Acanthamoeba* spp. are common

Copyright © 2016, Infectious Diseases and Tropical Medicine Research Center. This is an open-access article distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 International License (http://creativecommons.org/licenses/by-nc/4.0/) which permits copy and redistribute the material just in noncommercial usages, provided the original work is properly cited.

in HIV positive patients and may occur alone or in combination with central nervous system (CNS) involvement. Skin ulcers are also reported in patients with amoebic encephalitis and the ones receiving immunosuppressive drugs (7).

Besides, human pathogenic bacteria such as *Legionella* spp., *Cholera* spp., Mycobacterium tuberculosis and *Helicobacter* spp. within Acanthamoeba species are repeatedly reported. Some species of *Acanthamoeba* were proposed as a reservoir of *Legionella* spp. in nature (14, 15).

2. Objectives

The current study aimed to determine the soil contamination with *Acanthamoeba* and *Hartmannella* spp. in Sari, North of Iran.

3. Patients and Methods

3.1. Study Area

Sari is the provincial capital of Mazandaran, located in North of Iran, between the Northern slopes of the Alborz mountains and the Southern coast of the Caspian sea. Sari is the largest and most populous city of Mazandaran. Human population of Sari is around 196'000. The city is located between the parallels 35° 58 and 36° 50 of the Northern latitude and between 52° 56 and 53° 59 of the Eastern longitude (16).

3.2. Soil Examination

A total of 96 samples were collected from three areas of the city (32 samples per each area). Each specimen contained approximately 200 g of soil from a depth of 2 - 5 cm in an area not exposed to direct sunlight. The soil samples were dried overnight at room temperature. The samples were filtered through 150 μ m mesh sieves, yielding approximately 2 g of powdered soil. The powdered sand was moved to a 15 mL test tube, suspended in approximately 10 mL of Tween-80 solution with the concentration of 0.05 and centrifuged at 1500 rpm for 10 minutes.

After discarding the supernatant, the test tube containing the sediments was filled to approximately 1 cm from the top with sucrose solution (specific gravity of 1.200), vortexed, and again centrifuged for 15 minutes at 1500 rpm (17). Finally the tubes were filled to the top with sucrose solution and a cover slip placed on the tube for 30 minutes and then microscopic slides were prepared from flotation materials. The fixed smear was immersed in Giemsa (Merck, Darmstadt, Germany) solution (1: 45 mL of dH₂O) in a staining container for 60 minutes. The slide was then rinsed with dH_2O (dried at 37°C for 60 minutes) and studied under the light microscope using 100X oil immersion objective.

4. Results

The identification of *Acanthamoeba* at the genus level in the study was based on distinctive features of double walled cysts (Figure 1). Of the 96 studied samples, from different environmental locations in Sari, 38 (39.6%) *Acanthamoeba* spp. and 5 (5.2%) *Hartmannella* spp. were identified. The most contaminated soil contamination was from region 1 (Table 1).

Table 1. Frequency of Samples According to Their Localities and Number of Positive Samples^a

Region	Sample No.	Acanhtamoeba Spp.	Hartmannella Spp.
1	32	15 (15.6)	4 (4.2)
2	32	11 (11.5)	1(1)
3	32	12 (12.5)	0
Total	96	38 (39.6)	5 (5.2)

^aValues are expressed as No. (%).



Figure 1. Cyst of Acanthamoeba asteronyxis in the Gimsa stained smear (100X)

5. Discussion

It was the first study to investigate soil contamination with free-living amoeba in North of Iran. The study isolated *Acanthamoeba* and *Hartmannella* spp. from a variety of ecological habitats using flotation methods. *Acanthamoeba* spp. is isolated from a variety of habitats such as water, seashores, pools, soil, dust, food and hospitals (18). The high rate of incidence in different environmental resources represent a serious warning to the public health, especially in immunocompromised patients (19). To date, northern cities of Iran, especially Sari, attract many tourists annually. The results of the present study revealed that soil resources of this area were contaminated with opportunistic amoebas such as *Acanthamoeba* spp. and *Hartmannella* spp. that may lead to severe diseases in high-risk people such as immunocompromised patients. The ability of these protozoans to survive and proliferate in nature, especially water, shows that they can be potentially pathogenic for humans and animals (7). According to the wide dispersion of protozoan, it is expected that many people are exposed to protozoan (20-26); therefore, it seems that 80% of the people have antibodies against protozoa (27).

The 96 processed samples, from three regions in Sari, represent an enormous difference in the incidence between *Acanthamoeba* spp. and *Hartmannella* spp. In a study in Spain, fifteen of the 24 samples (62.5 %) were positive for *Acanthamoeba* spp. based on morphological analysis and PCR results (28). Studies on free-living amoebae and *Acanthamoeba* spp. in Canary Islands showed that about 40 % of samples from soil and water sources were positive for *Acanthamoeba* spp. (27). The results of these studies showed that soil contamination with free-living amoebas were similar with those of the present study.

As mentioned, *Acanthamoeba* species can serve as a vector for many pathogenic microorganisms such as viruses, fungi, protozoa and bacteria (29). Also, studies showed that primary classification of amoebas species were on the basis of morphology and based on the shape, size and amoeba cysts and the three major established groups (3). However, more studies revealed that the morphology of cysts was dependent on the ionic strength of their surrounding environment (30). Therefore, only molecular methods allow reliable differentiation of the *Acanthamoeba* species (8). To date, 18 *Acanthamoeba* genotypes are identified (31). A number of these genotypes such as T3, T11 and even T5 are noted to result in human clinical manifestations (32).

Ultimately, in order to reduce the health risk and avoid infections especially in immunocompromised patients from pathogenic strains of free-living amoebae, active surveillance to identify and control the presence of potentially hazardous *Acanthamoeba* species in other resources in these areas seem necessary. Therefore, the study can serve as a preliminary study in this area for further research to determine the genotypes of strains providing *Acanthamoeba* spp. and *Hartmannella* spp.

Acknowledgments

Authors wish to thank the parasitology and mycology department of Isfahan University of Medical Sciences, Isfahan, Iran.

Footnotes

Authors' Contribution: Mohammad Ali Mohaghegh, Mohsen Ghomashlooyan and Mojtaba Azimi Resketi, experimental design of the study; Mohammad Ali Mohaghegh, Mohsen Ghomashlooyan, Mohammad Falahati and Reza Mohammadimanesh, execution techniques and parasitological examination of soils; Farzaneh Mirzaie, statistical analysis; Mojtaba Azimi Resketi, collection of samples. All authors reviewed and contributed to the writing of this manuscript.

Conflict of Interest: The authors declared no financial or personal interest toward the results of the current study.

References

- Wickert H, Sukthana Y. Im Truben fischen. 2008 :107-11. doi: 10.1007/978-3-540-69847-0_21.
- Deol I, Robledo L, Meza A, Visvesvara GS, Andrews RJ. Encephalitis due to a free-living amoeba (Balamuthia mandrillaris): Case report with literature review. Surg Neurol. 2000;53(6):611–6. doi: 10.1016/s0090-3019(00)00232-9.
- Pussard M, Pons R. Morphology of cystic wall and taxonomy of genus Acanthamoeba (Protozoa, Amoebida). Protistologica. 1977;13(4):557– 98.
- Howe DK, Vodkin MH, Novak RJ, Visvesvara G, McLaughlin GL. Identification of two genetic markers that distinguish pathogenic and nonpathogenic strains of Acanthamoeba spp. *Parasitol Res.* 1997;83(4):345–8. [PubMed: 9134555].
- Khan NA. Acanthamoeba: biology and increasing importance in human health. FEMS Microbiol Rev. 2006;30(4):564–95. doi: 10.1111/j.1574-6976.2006.00023.x. [PubMed: 16774587].
- Ahmed Khan N. Pathogenesis of Acanthamoeba infections. Microbial Pathogenesis. 2003;34(6):277–85. doi: 10.1016/s0882-4010(03)00061-5.
- Marciano-Cabral F, Cabral G. Acanthamoeba spp. as agents of disease in humans. *Clin Microbiol Rev.* 2003;16(2):273–307. [PubMed: 12692099].
- Schuster FL, Visvesvara GS. Free-living amoebae as opportunistic and non-opportunistic pathogens of humans and animals. *Int J Parasitol.* 2004;**34**(9):1001–27. doi: 10.1016/j.ijpara.2004.06.004. [PubMed: 15313128].
- Visvesvara GS, Moura H, Schuster FL. Pathogenic and opportunistic free-living amoebae: Acanthamoeba spp., Balamuthia mandrillaris, Naegleria fowleri, and Sappinia diploidea. *FEMS Immunol Med Microbiol.* 2007;**50**(1):1–26. doi: 10.1111/j.1574-695X.2007.00232.x. [PubMed: 17428307].
- Shin HJ, Im KI. Pathogenic free-living amoebae in Korea. Korean J Parasitol. 2004;42(3):93-119. [PubMed: 15381859].
- Clarke DW, Niederkorn JY. The pathophysiology of Acanthamoeba keratitis. *Trends Parasitol.* 2006;22(4):175-80. doi: 10.1016/j.pt.2006.02.004. [PubMed: 16500148].
- 12. John DT, Petri WA, Markell EK, Voge M. Markell and Voge's medical parasitology. Elsevier Health Sciences; 2006.

- Maghsood A, Rezaian M, Rahimi F, Ghiasian SA, Farnia S. Contact lens-associated Acanthamoeba keratitis in Iran. *Iranian J Pub Health*. 2005;2(2):40–7.
- Winiecka-Krusnell J, Linder E. Bacterial infections of free-living amoebae. Res Microbiol. 2001;152(7):613-9. [PubMed: 11605981].
- Alizadeh H, Neelam S, Hurt M, Niederkorn JY. Role of contact lens wear, bacterial flora, and mannose-induced pathogenic protease in the pathogenesis of amoebic keratitis. *Infect Immun.* 2005;73(2):1061– 8. doi: 10.1128/IAI.73.2.1061-1068.2005. [PubMed: 15664950].
- Sharif M, Daryani A, Nasrolahei M, Ziapour SP. Prevalence of Toxoplasma gondii antibodies in stray cats in Sari, northern Iran. *Trop Anim Health Prod.* 2009;41(2):183–7. doi: 10.1007/s11250-008-9173-y. [PubMed: 18473184].
- Horiuchi S, Paller VG, Uga S. Soil contamination by parasite eggs in rural village in the Philippines. *Trop Biomed.* 2013;30(3):495–503. [PubMed: 24189679].
- Walochnik J, Obwaller A, Aspock H. Correlations between morphological, molecular biological, and physiological characteristics in clinical and nonclinical isolates of Acanthamoeba spp. *Appl Environ Microbiol.* 2000;**66**(10):4408–13. [PubMed: 11010891].
- Lorenzo-Morales J, Lopez-Darias M, Martinez-Carretero E, Valladares B. Isolation of potentially pathogenic strains of Acanthamoeba in wild squirrels from the Canary Islands and Morocco. *Exp Parasitol.* 2007;**117**(1):74–9. doi: 10.1016/j.exppara.2007.03.014. [PubMed: 17459380].
- Niyyati M, Lorenzo-Morales J, Rezaie S, Rahimi F, Mohebali M, Maghsood AH, et al. Genotyping of Acanthamoeba isolates from clinical and environmental specimens in Iran. *Exp Parasitol.* 2009;**121**(3):242– 5. doi: 10.1016/j.exppara.2008.11.003. [PubMed: 19059239].
- Rezaeian M, Niyyati M, Farnia SH, Haghi AM. Isolation of Acanthamoeba spp. from different environmental sources. *IJP*. 2008;3(1):44–7.
- De Jonckheere JF. Pathogenic free-living amoebae in swimming pools: survey in Belgium. *Ann Microbiol (Paris)*. 1979;**130B**(2):205–12. [PubMed: 43689].
- Jeong HJ, Yu HS. The role of domestic tap water in Acanthamoeba contamination in contact lens storage cases in Korea. *Korean J Parasitol.* 2005;43(2):47–50. [PubMed: 15951638].

- Kilvington S, Gray T, Dart J, Morlet N, Beeching JR, Frazer DG, et al. Acanthamoeba keratitis: the role of domestic tap water contamination in the United Kingdom. *Invest Ophthalmol Vis Sci.* 2004;45(1):165– 9. [PubMed: 14691169].
- Lorenzo-Morales J, Lindo JF, Martinez E, Calder D, Figueruelo E, Valladares B, et al. Pathogenic Acanthamoeba strains from water sources in Jamaica, West Indies. *Ann Trop Med Parasitol*. 2005;**99**(8):751–8. doi: 10.1179/136485905X65215. [PubMed: 16297288].
- Rivera F, Ramirez E, Bonilla P, Calderon A, Gallegos E, Rodriguez S, et al. Pathogenic and free-living amoebae isolated from swimming pools and physiotherapy tubs in Mexico. *Environ Res.* 1993;62(1):43–52. doi: 10.1006/enrs.1993.1087. [PubMed: 8325265].
- Lorenzo-Morales J, Ortega-Rivas A, Foronda P, Martinez E, Valladares B. Isolation and identification of pathogenic Acanthamoeba strains in Tenerife, Canary Islands, Spain from water sources. *Parasitol Res.* 2005;**95**(4):273–7. doi: 10.1007/s00436-005-1301-2. [PubMed: 15678350].
- Reyes-Batlle M, Todd CD, Martin-Navarro CM, Lopez-Arencibia A, Cabello-Vilchez AM, Gonzalez AC, et al. Isolation and characterization of Acanthamoeba strains from soil samples in Gran Canaria, Canary Islands, Spain. *Parasitol Res.* 2014;113(4):1383–8. doi: 10.1007/s00436-014-3778-z. [PubMed: 24449449].
- Scheid P, Schwarzenberger R. Acanthamoeba spp. as vehicle and reservoir of adenoviruses. *Parasitol Res.* 2012;111(1):479–85. doi: 10.1007/s00436-012-2828-7. [PubMed: 22290448].
- Sawyer TK, Griffin JL. A proposed new family, Acanthamoebidae n. fam.(order Amoebida), for certain cyst-forming filose amoebae. *Trans Am Micro Soci.* 1975:93–8.
- Qvarnstrom Y, Nerad TA, Visvesvara GS. Characterization of a new pathogenic Acanthamoeba Species, A. byersi n. sp., isolated from a human with fatal amoebic encephalitis. *J Eukaryot Microbiol.* 2013;60(6):626–33. doi: 10.1111/jeu.12069. [PubMed: 23879685].
- Maciver SK, Asif M, Simmen MW, Lorenzo-Morales J. A systematic analysis of Acanthamoeba genotype frequency correlated with source and pathogenicity: T4 is confirmed as a pathogen-rich genotype. *Eur J Protistol.* 2013;49(2):217–21. doi: 10.1016/j.ejop.2012.11.004. [PubMed: 23290304].