Review Article

Antibiotic Resistance Mechanisms Focusing on Quinolones Resistance in *Vibrio CHOLERAE*

Lawaly Maman Manzo,^{1,*} Bassira B. Issaka,¹ Issaka Seidou,¹ and Jibir Zanguina¹

¹Bacteriology and Virology Unit, Centre for Medical and Health Research (CERMES), Niamey, Niger

^{*} *Corresponding author*: Lawaly Maman Manzo, Bacteriology and Virology Unit, Centre for Medical and Health Research (CERMES), Niamey, Niger. Tel: +227-20752040, E-mail: manzolawal@yahoo.com

Received 2016 July 25; Revised 2016 August 10; Accepted 2016 August 15.

Abstract

Context: The number of quinolone-resistant *Vibrio cholerae* strains reported in various clinical studies is growing steadily since the 1990s. Important biological conditions such as alterations of DNA topoisomerases, perturbations in regard to drug entry and efflux and plasmids transferability between bacteria, are conditions that lead to the acquisition of certain power to resist quinolones during therapy. The current article aimed to review the evolution of quinolone-resistant strains of *V. cholerae*, the basis for quinolone mode of action against DNA topoisomerases and the recent advances in the understanding of the mechanisms by which *V. cholerae* strains are resistant to quinolones, including chromosomal mediated resistance involving mutations in topoisomerases, chromosomal mediated resistance resulted in response to reduced expression and/or overexpression of proteins synthesis for porins and cellular efflux pumps, respectively and plasmid mediated resistance.

Evidence Acquisition: This systematic review utilized the research results of clinical trials.gov database, the cochrane database of systematic reviews, Medline, PubMed, Google and other databases to select about 67 publications on cholera and fluoroquinolones resistance mechanisms in *V. cholerae*. Relevant publications were reviewed; findings were synthesized using a narrative method to illustrate how the treatment of cholera with quinolones as adjunct is evolved.

Results: Through the thorough literature review, it was found that despite the progress made in the development of effective fluoroquinolones against various enteric pathogens including *V. cholerae*, new concerns emerged which include the constant reports on reduced susceptibility of these drugs against *V. cholerae* strains endorsed with the emergence of various mechanisms of resistance. **Conclusions:** The current systematic review emphasized on the importance of surveying the emergence of *V. cholerae* resistance to quinolones that may occur during adjunct treatment of cholera and also the importance of determining the genetic basis of the resistance mechanisms for future development of more promising quinolones.

Keywords: Vibrio cholera, Quinolones, Quinolone Resistance Mechanisms

1. Context

Rehydration therapy by appropriate oral or intravenous solutions is strongly advised during the treatment of cholera, a gastroenteritis disease with serious public health implications. Various international organizations engaged in promoting and implementing subjects relevant to public health infectious diseases recommend antimicrobial drug treatment for severely dehydrated patients with suspected cholera. Reduced duration of diarrhea and volume of stool and reduced duration of vibrio shedding due to adjunct treatment with antibiotic were reported in a recent Cochrane meta-analysis of several clinical trials conducted on a large sample of patients (1). Amongst various groups of antimicrobials, fluoroquinolones encountered significant emergence to treat gastroenteritis diseases. Evidence for various successes of clinical experience with quinolones in cholera and other enteric infections are reported in various international journals (2). A treatment network with quinolones

elucidating various comparisons and the number of trials examining each comparison is presented in Figure 1. Ciprofloxacin, norfloxacin and ofloxacin were compared to placebo/no treatment (comparison not shown in Figure 1).

Utility of quinolone as adjunct in the treatment of cholera was first confirmed by norfloxacin as reported in studies first conducted in Calcutta, India (3), then in Lima, Peru (4), in Thailand (5), and in Salta, Argentina (6) respectively. Ciprofloxacin is another important quinolone with confirmed utility against gastroenteritis such as cholera and recorded as the most extensively studied amongst others. Excellent results with this fluoroquinolones was reported in an open, non-controlled study conducted by Doganci et al. (7), in two large randomized double-blind controlled studies conducted in Peru and Bangladesh (8), and in a large clinical trial conducted in Bangladesh (9). Results of these important studies clearly showed that quinolones were highly effective in all clinical parameters.

Copyright © 2016, Infectious Diseases and Tropical Medicine Research Center. This is an open-access article distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 International License (http://creativecommons.org/licenses/by-nc/4.0/) which permits copy and redistribute the material just in noncommercial usages, provided the original work is properly cited.

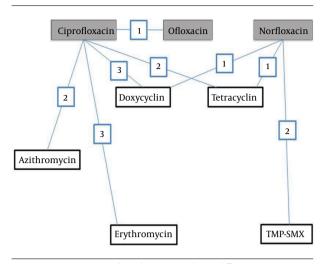


Figure 1. Treatment network implicating quinolones; different comparisons included studies which compared one quinolone vs another quinolone are described in this figure.

However, in recent years many studies from Africa, Asia and America reported the emergence of quinoloneresistant strains of V. cholerae (10, 11). Chander et al., and Saha et al., in different studies including 277 and 195 isolates of V. cholerae from Chandigarh (India) and Bangladesh respectively, both reported the emergence of ciprofloxacin-resistant strains of V. cholerae (12-14). In a study including strains of V. cholerae species isolated from 61 cases with gastroenteritis admitted to the gastroenteritis wards of Karnataka, Hubli (South India) and hospitalized for one to seven days during the cholera epidemic of 2002, Krishna et al. reported norfloxacin and ciprofloxacin resistance with the prevalence of 12.5% (15). A warning record on progressive increases in ciprofloxacin and norfloxacin resistance among V. cholerae O1 strains with the highest prevalence of 38.8% in 1999 and 25% in 2000, respectively was demonstrated by Garg et al., in a study on patients with cholera admitted to the Infectious Diseases Hospital, Calcutta, India (16). Mercy et al., in a study on V. cholerae O1 species isolated from different regions of Kenya from 2007 to 2010 reported resistance against nalidixic acid and ciprofloxacin with the prevalence of 89% and 2.3%, respectively (17). In a study aimed to determine the antibiotic susceptibility patterns of 118 strains of V. cholerae O1 isolated in 2010 at Jawaharlal Nehru institute of post graduate medical education and research, Puducherry, India, Mandal et al., reported resistance to ciprofloxacin with a modest occurrence of 4.2% (18). While a study by Eibach et al., reported that the increase in ciprofloxacin resistance reached about 98% in 2014 (19) (Table 1).

2. Evidence Acquisition

Results of the search in clinical trials.gov database and the cochrane database of systematic reviews by John Wiley & Sons, Ltd. were synthesized to illustrate and/or relate the reported clinical success and/or limits of quinolones as adjuncts to treat cholera. Medline, PubMed, Google and other databases were also used to select about 80 publications on cholera and quinolones resistant mechanisms in *V. cholera* to evaluate how the treatment of cholera with these drugs is evolved regarding the various quinoloneresistant *V. cholerae* strains.

3. Results

3.1. Quinolone Mechanisms of Activity

Understanding antibiotic resistance mechanisms requires an understanding of where antibiotics exert their effect. Cellular events such as DNA replication and mRNA transcription that take place during cell cycle or celldivision cycle require the intervention of topoisomerases such as DNA gyrase (type II topoisomerase) recognized to participate in relieving some of the stress during DNA synthesis (20). Thus, chemists and pharmacists involved in the synthesis of quinolones do exploit these conditions to target DNA-topoisomerase complexes (21, 22). Quinolones are bactericidal compounds that inhibit bacterial DNA from unwinding and duplicating through inhibition of the topoisomerase II ligase domain. This interference consequently leads to transitory DNA fragmentation and by catalyzing the passage of DNA segments within the cuts before closing them and though leading to rapid cell death (Figure 2). DNA gyrase, also referred as topoisomerase II, consists of two polypeptide subunits (GyrA and GyrB) with the function to regulate links via relaxing supercoils and by introducing both positive and negative supercoils between DNA double strands. The A subunit (encoded by gyrA gene) carries out binding of DNA, B subunit mediates the introduction of negative supercoils, and then A subunit participates in the reunion of DNA strands (23, 24). Topoisomerase IV, similar to topoisomerase II, consists of two polypeptide subunits (ParC and ParE) with the role to participate in separating new replicates of chromosomes following the replication process. Quinolones binds to the A subunit of topoisomerase II and though interfering in various steps involved in the DNA replication process.

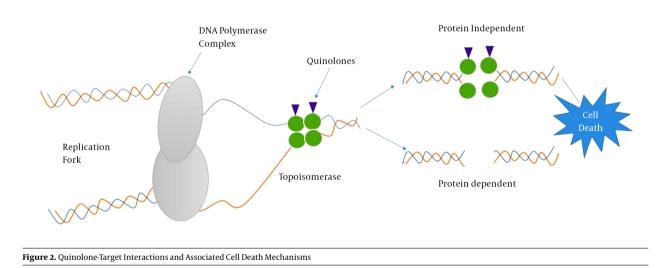
3.2. Chromosomal Mediated Resistance Involving Mutations in Topoisomerase

In many bacteria, such as *V. cholerae*, quinolones resistance is a chromosomal mediated resistance involving mutations in the quinolone resistance-determining

Year	Number of Isolates	Number of Resistant Strains (%)			
		Nalidixic Acid	Ciprofloxacin	Norfloxacin	Ref.
1989	49	3 (6.1)	ND	ND	(<mark>16</mark>)
1990	59	1 (1.7)	ND	ND	(<mark>16</mark>)
1991	30		ND	ND	(<mark>16</mark>)
1992	26	2 (7.7)			(<mark>16</mark>)
1993	20	1(5)			(<mark>16</mark>)
1994	74	73 (98.6)			(<mark>16</mark>)
1995	84	82 (97.6)	2 (2.4)	3(3.6)	(<mark>16</mark>)
1996	69	68 (98.5)	4 (5.8)		(<mark>16</mark>)
1997	53	50 (94.3)	10 (18.9)	4 (7.5)	(<mark>16</mark>)
1998	201	197 (98)	20 (10)	1(0.5)	(<mark>16</mark>)
1999	49	49 (100)	19 (38.8)	8 (16.3)	(<mark>16</mark>)
2000	16	12 (75)	3 (18.7)	4 (25)	(<mark>16</mark>)
2007-2010	44	39 (89)	1(2.3)		(17)
2010	118	ND	5(4.2)	ND	(18)
2014	92	62 (100)	61 (98.4)		(19)

Table 1. Vibrio cholerae O1 Species Isolates From Different Cholera Outbreaks and the Prevalence of Resistance to Quinolones

Abbreviation: ND, no data.



regions (QRDRs) of topoisomerases genes *gyrA* and *parC*. Quinolone resistance in *V. cholerae* species is mainly observed in strains harboring mutations in *gyrA* and *parC* genes of topoisomerase II and IV respectively (25, 26). Substitutions of single amino acid (Ser-83; Ile in gyrA and Ser-85; Leu in *parC*) were observed in genes of topoisomerase II and IV of *V. cholerae* O1 with marked reduced susceptibility to ciprofloxacin (27).

3.3. Efflux Resistance Mechanisms

The efflux system pumps out a variety of structurally and functionally distinct antimicrobials including quinolones from bacterial cells. To date, advancement in biomedicine research oriented through the understanding of the biochemistry of Gram-negative bacteria cell wall and the mechanisms involved in resisting antimicrobials such as quinolones significantly contributed to the discovery and characterization of various multidrug and drug-specific efflux pumps from bacteria.

Vibrio cholerae efflux pumps are categorized into two major families, such as the ATP-binding cassette (ABC) superfamily (28), and the proton-motive force (PMF) pump families. The PMF sub-families include the major facilitator superfamily (MFS) (29), the multidrug and toxic compound extrusion (MATE) family (30), the small multidrug resistance (SMR) family (31) and the resistance-nodulationdivision (RND) superfamily (32-34). In V. cholerae, six putative genes from MATE family of efflux pumps are reported. Involvement of genes such as VcmB, VcmD, VcmH, VcmN, VcmA and VcrM (35) with energy dependent efflux toward fluoroquinolones is demonstrated in Escherichia coli and V. cholerae background. Recently, homologue of NorM in Vibrio parahaemolyticus (36) reported to mediate resistance to hydrophilic fluoroquinolones, aminoglycosides and norfloxacin was discovered in the genome of V. cholerae O1 El Tor N16961 (37). Among the V. cholerae efflux systems, MFS transporters and the PMF-dependent efflux were reported in separate studies to confer resistance to nalidixic acid (38) and various quinolones (39) in clinical strains of V. cholerae. RND efflux systems were encoded by six operons. Four operons (VexAB, VexCD, VexGH and VexIJK) out of six were reported to confer resistance to antimicrobials in V. cholerae (40-42). VexAB was associated with resistance to several antimicrobials determined by mutational analysis (40). In a recent study conducted by Jun O et al., inherently reduced expression of gene vca0421 in V. cholerae O1 was demonstrated to confer resistance to fluoroquinolones in V. cholerae O1 (43-47).

3.4. Plasmid Mediated Quinolone Resistance

Plasmids carrying genes that encode drug resistance determinants spread very fast in mixed bacteria communities. However, the wild strains of *V. cholerae* are mostly reported as poor hosts for plasmids carrying genes. The frequency of plasmid carriage in clinical strains of *V. cholerae* O1 was less (2%) compared with clinical non-O1 strains (25%) as reported elsewhere. Most of the clinical strains of *V. cholerae* O1 isolated from different geographical regions lacked the plasmids (48, 49), and when present, the size and functions of plasmids were not uniform. A study reported some of the naturally occurring conjugative and non-conjugative plasmids harboring drug resistance determinants in environmental and clinical isolates of *V. cholerae* encoding genes inducing various antibiotics resistance.

Conjugative plasmids evolved to increase the relative fitness of bacteria had implications both for maintenance and spread of plasmids. Transfer of multiple-drugresistance from *V. cholerae* to *E. coli* (K12) was demonstrated in-vitro with conjugative plasmids. Transfer of multipledrug-resistant (MDR) plasmids of group InC through conjugation with other bacteria was reported in a study conducted by Glass et al. This plasmid conferred resistance to a number of antibiotics including quinolones (50). MDR *V. cholerae* species isolated in Kenya harbored a 100 MDa plasmid of group IncC (51). In Algeria, the IncJ plasmids (IncJ incompatibility group) containing antibiotic resistance marker was identified for the first time in *V. cholerae* El Tor (52).

An emerging clinical concern that causes reduced susceptibility to quinolones was experienced with plasmids identified to carry quinolone resistant genes (53).Plasmidquinolone resistant genes include intrinsic or *qnr* genes (22, 54, 55), reported to protect topoisomerases from quinolones in bacteria (56). Modest findings reported in a study on *V. cholerae* isolates in Brazil described *qnrVC1*, a homolog to *qnr*. Limitations regarding this study include lack of molecular mechanisms evidencing that *qnrVC1* could be linked to reduced susceptibility of ciprofloxacin in *V. cholerae* (57). However, Hong Bin Kim et al., in a recent study on *V. cholerae* species isolated from Bangladesh, described a new *qnr* homolog, *qnrVC3* and a large number of mutations in topoisomerase target enzymes that confer higher levels of transferable quinolone resistance (58, 59).

4. Conclusions

The application of antibiotics as adjunct to treat patients with cholera infection is encouraged by various international health organizations such as world health organization (WHO) and Medecin Sans Frontiere (MSF). Quinolones are one of the most important classes of antibiotics available to treat gastroenteritis including cholera. The emergence of multidrug or specific drug resistant V. cholerae strains significantly affected the clinical contribution of these drugs. To date, various studies reported comprehensive descriptions of various molecular mechanisms and mutations in topoisomerases enzymes involved in or causing drug resistance, respectively. Quinolones are in the clinics to treat cholera for several decades, but knowledge about the mechanisms of interaction between quinolone and its specific targets (i e, topoisomerases enzymes) and how mutations cause resistance is only in progress but not conclusive. Since the emergence of quinolones-resistant V. cholerae may significantly influence the strategies of controlling cholera, locally and regionally based surveillance information on antimicrobial susceptibility of V. cholerae should give importance to this aspect. Hopefully, this review can be used as a resource document to direct the development of new promising generation of quinolones to treat cholera based on better understanding of the molecular mechanisms of action of these synthetic drugs.

Acknowledgments

Authors wish to thank the center for resource management and environmental studies (CERMES) for their encouragement in submitting this manuscript.

Footnote

Authors' Contribution: Lawaly Maman Manzo: design and revision of the manuscript; Bassira B. Issaka, Issaka seidou Jibir Zanguina: revising the manuscript.

References

- Leibovici-Weissman Y, Neuberger A, Bitterman R, Sinclair D, Salam MA, Paul M. Antimicrobial drugs for treating cholera. *Cochrane Database Syst Rev.* 2014(6):CD008625. doi: 10.1002/14651858.CD008625.pub2. [PubMed: 24944120].
- 2. Seas C, Gotuzzo E. Vibrio cholerae (Cholera). USA: www.antimicrobe.org; 2014.
- Bhattacharya SK, Bhattacharya MK, Dutta P, Dutta D, De SP, Sikdar SN, et al. Double-blind, randomized, controlled clinical trial of norfloxacin for cholera. *Antimicrob Agents Chemother*. 1990;**34**(5):939–40. [PubMed: 2193628].
- Gotuzzo E, Cieza J, Estremadoyro L, Seas C. Cholera. Lessons from the epidemic in Peru. *Infect Dis Clin North Am.* 1994;8(1):183–205. [PubMed: 7980768].
- Moolasart P, Eampokalap B, Supaswadikul S. Comparison of the efficacy of tetracycline and norfloxacin in the treatment of acute severe watery diarrhea. Southeast Asian J Trop Med Public Health. 1998;29(1):108-11. [PubMed: 9740281].
- Seijo AC, Fernandez M, Campos C, Cernigoi BE, Nogueras M, San Juan J, et al. Norfloxacin monodose use in patients with cholera in Salta Argentina. *Rev Inst Med Trop Sao Paulo*. 1996;**38**(3):217–9. [PubMed: 9163987].
- Doganci L, Gun H, Baysallar M, Albay A, Cinar E, Haznedaroglu T. Shortterm quinolones for successful eradication of multiply resistant Vibrio cholerae in adult patients. *Scand J Infect Dis.* 1995;27(4):425–6. [PubMed: 8658086].
- 8. Gotuzzo E, Seas C, Echevarria J, Carrillo C, Mostorino R, Ruiz R. Ciprofloxacin for the treatment of cholera: a randomized, doubleblind, controlled clinical trial of a single daily dose in Peruvian adults. *Clin Infect Dis.* 1995;**20**(6):1485–90. [PubMed: 7548496].
- Khan W, Bennish ML, Seas C, Khan EH, Ronan A, Dhar U, et al. Randomised controlled comparison of single-dose ciprofloxacin and doxycycline for cholera caused by Vibrio cholerae O1 or O139. *The Lancet.* 1996;**348**(9023):296–300. doi: 10.1016/s0140-6736(96)01180-4.
- Jesudason MV, John TJ. Transferable trimethoprim resistance of Vibrio cholerae O1 encountered in southern India. *Trans R Soc Trop Med Hyg.* 1990;84(1):136–7. [PubMed: 2345916].
- Faruque AS, Alam K, Malek MA, Khan MG, Ahmed S, Saha D, et al. Emergence of multidrug-resistant strain of Vibrio cholerae O1 in Bangladesh and reversal of their susceptibility to tetracycline after two years. J Health Popul Nutr. 2007;25(2):241–3. [PubMed: 17985827].
- Bauer AW, Kirby WM, Sherris JC, Turck M. Antibiotic susceptibility testing by a standardized single disk method. *Am J Clin Pathol.* 1966;**45**(4):493-6. [PubMed: 5325707].

- Chander J, Kaistha N, Gupta V, Mehta M, Singla N, Deep A, et al. Epidemiology & antibiograms of Vibrio cholerae isolates from a tertiary care hospital in Chandigarh, north India. *Indian J Med Res.* 2009;**129**(5):613-7. [PubMed: 19675394].
- Saha D, Karim MM, Khan WA, Ahmed S, Salam MA, Bennish ML. Singledose azithromycin for the treatment of cholera in adults. *N Engl J Med.* 2006;**354**(23):2452–62. doi: 10.1056/NEJMoa054493. [PubMed: 16760445].
- Krishna BV, Patil AB, Chandrasekhar MR. Fluoroquinoloneresistant Vibrio cholerae isolated during a cholera outbreak in India. *Trans R Soc Trop Med Hyg.* 2006;100(3):224–6. doi: 10.1016/j.trstmh.2005.07.007. [PubMed: 16246383].
- Garg P, Sinha S, Chakraborty R, Bhattacharya SK, Nair GB, Ramamurthy T, et al. Emergence of fluoroquinolone-resistant strains of Vibrio cholerae O1 biotype El Tor among hospitalized patients with cholera in Calcutta, India. *Antimicrob Agents Chemother*. 2001;45(5):1605–6. [PubMed: 11372642].
- Mercy N, Mohamed AA, Zipporah N, Chowdhury G, Pazhani GP, Ramamurthy T, et al. Phenotypic and genetic characterization of Vibrio cholerae O1 isolated from various regions of Kenya between 2007 and 2010. *Pan Afr Med J.* 2014;**19**:8. doi: 10.11604/pamj.2014.19.8.2496. [PubMed: 25584121].
- Mandal J, Dinoop KP, Parija SC. Increasing antimicrobial resistance of Vibrio cholerae OI biotype E1 tor strains isolated in a tertiary-care centre in India. J Health Popul Nutr. 2012;30(1):12–6. [PubMed: 22524114].
- Eibach D, Herrera-Leon S, Gil H, Hogan B, Ehlkes L, Adjabeng M, et al. Molecular Epidemiology and Antibiotic Susceptibility of Vibrio cholerae Associated with a Large Cholera Outbreak in Ghana in 2014. *PLoS Negl Trop Dis.* 2016;10(5):4751. doi: 10.1371/journal.pntd.0004751. [PubMed: 27232338].
- Espeli O, Marians KJ. Untangling intracellular DNA topology. *Mol Microbiol.* 2004;**52**(4):925–31. doi: 10.1111/j.1365-2958.2004.04047.x. [PubMed: 15130115].
- Drlica K, Malik M, Kerns RJ, Zhao X. Quinolone-mediated bacterial death. *Antimicrob Agents Chemother*. 2008;**52**(2):385–92. doi: 10.1128/AAC.01617-06. [PubMed: 17724149].
- Sugino A, Peebles CL, Kreuzer KN, Cozzarelli NR. Mechanism of action of nalidixic acid: purification of Escherichia coli nalA gene product and its relationship to DNA gyrase and a novel nicking-closing enzyme. Proc Natl Acad Sci U S A. 1977;74(11):4767-71. [PubMed: 200930].
- Willmore E, de Caux S, Sunter NJ, Tilby MJ, Jackson GH, Austin CA, et al. A novel DNA-dependent protein kinase inhibitor, NU7026, potentiates the cytotoxicity of topoisomerase II poisons used in the treatment of leukemia. *Blood*. 2004;103(12):4659–65. doi: 10.1182/blood-2003-07-2527. [PubMed: 15010369].
- Chen CR, Malik M, Snyder M, Drlica K. DNA gyrase and topoisomerase IV on the bacterial chromosome: quinolone-induced DNA cleavage. *J Mol Biol.* 1996;258(4):627–37. doi: 10.1006/jmbi.1996.0274. [PubMed: 8636997].
- Everett MJ, Jin YF, Ricci V, Piddock LJ. Contributions of individual mechanisms to fluoroquinolone resistance in 36 Escherichia coli strains isolated from humans and animals. *Antimicrob Agents Chemother.* 1996;40(10):2380–6. [PubMed: 8891148].
- Vila J, Ruiz J, Marco F, Barcelo A, Goni P, Giralt E, et al. Association between double mutation in gyrA gene of ciprofloxacin-resistant clinical isolates of Escherichia coli and MICs. *Antimicrob Agents Chemother*. 1994;**38**(10):2477-9. [PubMed: 7840592].
- 27. Quilici ML, Massenet D, Gake B, Bwalki B, Olson DM. Vibrio cholerae O1 variant with reduced susceptibility to ciprofloxacin, Western Africa. *Emerg Infect Dis.* 2010;**16**(11):1804–5. doi: 10.3201/eid1611.100568. [PubMed: 21029554].
- Lubelski J, Konings WN, Driessen AJ. Distribution and physiology of ABC-type transporters contributing to multidrug resistance in bacteria. *Microbiol Mol Biol Rev.* 2007;**71**(3):463–76. doi: 10.1128/MMBR.00001-07. [PubMed: 17804667].

- Pao SS, Paulsen IT, Saier MH. Major facilitator superfamily. *Microbiol Mol Biol Rev.* 1998;62(1):1–34. [PubMed: 9529885].
- Kuroda T, Tsuchiya T. Multidrug efflux transporters in the MATE family. *Biochim Biophys Acta*. 2009;**1794**(5):763–8. doi: 10.1016/j.bbapap.2008.11.012. [PubMed: 19100867].
- Jack DL, Yang NM, Saier MH. The drug/metabolite transporter superfamily. Eur J Biochem. 2001;268(13):3620–39. [PubMed: 11432728].
- Nikaido H, Takatsuka Y. Mechanisms of RND multidrug efflux pumps. Biochim Biophys Acta. 2009;**1794**(5):769–81. doi: 10.1016/j.bbapap.2008.10.004. [PubMed: 19026770].
- Paulsen IT, Brown MH, Skurray RA. Proton-dependent multidrug efflux systems. *Microbiol Rev.* 1996;60(4):575–608. [PubMed: 8987357].
- Putman M, van Veen HW, Konings WN. Molecular properties of bacterial multidrug transporters. *Microbiol Mol Biol Rev.* 2000;64(4):672–93. [PubMed: 11104814].
- Begum A, Rahman MM, Ogawa W, Mizushima T, Kuroda T, Tsuchiya T. Gene cloning and characterization of four MATE family multidrug efflux pumps from Vibrio cholerae non-O1. *Microbiol Immunol.* 2005;49(11):949–57. [PubMed: 16301805].
- Heidelberg JF, Eisen JA, Nelson WC, Clayton RA, Gwinn ML, Dodson RJ, et al. DNA sequence of both chromosomes of the cholera pathogen Vibrio cholerae. *Nature*. 2000;406(6795):477-83. doi: 10.1038/35020000. [PubMed: 10952301].
- Morita Y, Kodama K, Shiota S, Mine T, Kataoka A, Mizushima T, et al. NorM, a putative multidrug efflux protein, of Vibrio parahaemolyticus and its homolog in Escherichia coli. *Antimicrob Agents Chemother*. 1998;**42**(7):1778–82. [PubMed: 9661020].
- Colmer JA, Fralick JA, Hamood AN. Isolation and characterization of a putative multidrug resistance pump from Vibrio cholerae. *Mol Microbiol.* 1998;27(1):63-72. [PubMed: 9466256].
- Baranwal S, Dey K, Ramamurthy T, Nair GB, Kundu M. Role of active efflux in association with target gene mutations in fluoroquinolone resistance in clinical isolates of Vibrio cholerae. *Antimicrob Agents Chemother.* 2002;46(8):2676–8. [PubMed: 12121955].
- Bina JE, Provenzano D, Wang C, Bina XR, Mekalanos JJ. Characterization of the Vibrio cholerae vexAB and vexCD efflux systems. *Arch Microbiol.* 2006;**186**(3):171–81. doi: 10.1007/s00203-006-0133-5. [PubMed: 16804679].
- Bina XR, Provenzano D, Nguyen N, Bina JE. Vibrio cholerae RND family efflux systems are required for antimicrobial resistance, optimal virulence factor production, and colonization of the infant mouse small intestine. *Infect Immun.* 2008;**76**(8):3595–605. doi: 10.1128/IAI.01620-07. [PubMed: 18490456].
- Van Bambeke F, Michot JM, Tulkens PM. Antibiotic efflux pumps in eukaryotic cells: occurrence and impact on antibiotic cellular pharmacokinetics, pharmacodynamics and toxicodynamics. *JAntimicrob Chemother*. 2003;**51**(5):1067–77. doi: 10.1093/jac/dkg225. [PubMed: 12697641].
- Okuda J, Hayashi N, Wakahara Y, Gotoh N. Reduced expression of the vca0421 gene of Vibrio cholerae O1 results in innate resistance to ciprofloxacin. *Antimicrob Agents Chemother.* 2010;**54**(11):4917–9. doi: 10.1128/AAC.01652-09. [PubMed: 20805389].
- Li XZ, Nikaido H. Efflux-mediated drug resistance in bacteria. Drugs. 2004;64(2):159–204. [PubMed: 14717618].
- Chen J, Morita Y, Huda MN, Kuroda T, Mizushima T, Tsuchiya T. VmrA, a member of a novel class of Na(+)-coupled multidrug efflux

pumps from Vibrio parahaemolyticus. *J Bacteriol*. 2002;**184**(2):572–6. [PubMed: 11751837].

- 46. Huda N, Lee EW, Chen J, Morita Y, Kuroda T, Mizushima T, et al. Molecular cloning and characterization of an ABC multidrug efflux pump, VcaM, in Non-O1 Vibrio cholerae. *Antimicrob Agents Chemother*. 2003;47(8):2413-7. [PubMed: 12878498].
- Woolley RC, Vediyappan G, Anderson M, Lackey M, Ramasubramanian B, Jiangping B, et al. Characterization of the Vibrio cholerae vceCAB multiple-drug resistance efflux operon in Escherichia coli. *J Bacteriol.* 2005;**187**(15):5500–3. doi: 10.1128/JB.187.15.5500-5503.2005. [PubMed: 16030246].
- Bag PK, Maiti S, Sharma C, Ghosh A, Basu A, Mitra R, et al. Rapid spread of the new clone of Vibrio cholerae O1 biotype El Tor in cholera endemic areas in India. *Epidemiol Infect.* 1998;**121**(2):245–51. [PubMed: 9825773].
- Clark CA, Purins L, Kaewrakon P, Manning PA. VCR repetitive sequence elements in the Vibrio cholerae chromosome constitute a mega-integron. *Mol Microbiol*. 1997;26(5):1137-8. [PubMed: 9426148].
- Glass RI, Huq MI, Lee JV, Threlfall EJ, Khan MR, Alim AR, et al. Plasmidborne multiple drug resistance in Vibrio cholerae serogroup O1, biotype El Tor: evidence for a point-source outbreak in Bangladesh. J Infect Dis. 1983;147(2):204–9. [PubMed: 6827137].
- Finch MJ, Morris JG, Kaviti J, Kagwanja W, Levine MM. Epidemiology of antimicrobial resistant cholera in Kenya and East Africa. *Am J Trop Med Hyg.* 1988;39(5):484–90. [PubMed: 3195696].
- Korichi MN, Belhocine S, Rahal K. [Inc J plasmids identified for the first time in Vibrio cholerae El Tor]. *Med Trop (Mars)*. 1997;57(3):249– 52. [PubMed: 9513150].
- Martinez-Freijo P, Fluit AC, Schmitz FJ, Grek VS, Verhoef J, Jones ME. Class I integrons in Gram-negative isolates from different European hospitals and association with decreased susceptibility to multiple antibiotic compounds. *J Antimicrob Chemother*. 1998;42(6):689–96. [PubMed: 10052890].
- Robicsek A, Jacoby GA, Hooper DC. The worldwide emergence of plasmid-mediated quinolone resistance. *Lancet Infect Dis.* 2006;6(10):629–40. doi: 10.1016/S1473-3099(06)70599-0. [PubMed: 17008172].
- Robicsek A, Strahilevitz J, Jacoby GA, Macielag M, Abbanat D, Park CH, et al. Fluoroquinolone-modifying enzyme: a new adaptation of a common aminoglycoside acetyltransferase. *Nat Med.* 2006;**12**(1):83–8. doi: 10.1038/nm1347. [PubMed: 16369542].
- Tran JH, Jacoby GA, Hooper DC. Interaction of the plasmidencoded quinolone resistance protein Qnr with Escherichia coli DNA gyrase. Antimicrob Agents Chemother. 2005;49(1):118–25. doi: 10.1128/AAC.49.1.118-125.2005. [PubMed: 15616284].
- Fonseca EL, Dos Santos Freitas F, Vieira VV, Vicente AC. New qnr gene cassettes associated with superintegron repeats in Vibrio cholerae O1. *Emerg Infect Dis.* 2008;14(7):1129–31. doi: 10.3201/eid1407.080132. [PubMed: 18598639].
- Burrus V, Marrero J, Waldor MK. The current ICE age: biology and evolution of SXT-related integrating conjugative elements. *Plasmid.* 2006;55(3):173–83. doi: 10.1016/j.plasmid.2006.01.001. [PubMed: 16530834].
- 59. Kim HB, Wang M, Ahmed S, Park CH, LaRocque RC, Faruque ASG, et al. Transferable quinolone resistance in vibrio cholerae. *Antimicrob Agents Chemother.* 2009;**54**(2):799–803. doi:10.1128/aac.01045-09.