Published online 2019 July 9.

# Prevalence of Methicillin-Resistant *Staphylococcus aureus* in Hospitals and Community in Duhok, Kurdistan Region of Iraq

Nawfal Hussein<sup>1,\*</sup>, Reving S Salih<sup>2</sup> and Narin A Rasheed<sup>3</sup>

<sup>1</sup>College of Medicine, University of Zakho, Kurdistan, Iraq

<sup>2</sup>Department of Internal Medicine, College of Medicine, University of Duhok, Duhok, Iraq

<sup>3</sup>Akre Technical Institute, Duhok Polytechnique University, Duhok, Kurdistan Region of Iraq

Corresponding author: College of Medicine, University of Zakho, Kurdistan, Iraq. Email: nawfal.hussein@yahoo.com

Received 2019 January 20; Revised 2019 June 20; Accepted 2019 June 30.

# Abstract

**Background:** Methicillin-resistant *Staphylococcus aureus* (MRSA) is a serious concern of the global health sector and more recently, an escalating problem in the community.

**Objectives:** This study was performed to investigate the incidence of MRSA in hospital staff and community students in Duhok, Kurdistan, Iraq, and make a molecular comparison between the strains based on the detection of *mecA* gene and Panton-Valentine Leukocidin (PVL) gene.

**Methods:** We obtained 109 and 103 samples from the nares of hospital staff and community students, respectively. Conventional laboratory tests were performed for the detection of *Staphylococcus aureus* (*S. aureus*) and antibiotic sensitivity testing to identify MRSA isolates. Besides, PCR was utilized for molecular analysis.

**Results:** All isolates from hospital staff were identified as *S. aureus*. Out of the 109 isolates, 55 (50.4%) were MRSA carrying the *mec*A gene, among which 4/55 (3.7%) were MRSA-PVL positive. Additionally, 54/109 (49.5%) isolates were methicillin-sensitive *S. aureus* (MSSA) but four isolates (3.7%) were MSSA-PVL positive. Furthermore, 23/103 (22.3%) samples from community students were identified as *S. aureus*, among which 5/23 (21.7%) and 17/23 (73.9%) isolates were MSSA-PVL positive and MSSA-PVL negative, respectively. Moreover, 1/23 (4.3%) was found as MRSA and was PVL gene-positive.

**Conclusions:** The results showed that MRSA is swarming in hospitals and community in Duhok, Iraq. The highest rate of PVL was associated with community-acquired-MSSA (CA- MRSA). With further genotypic study, immediate action is needed to control and reduce the spread of MRSA clones, determine their clonal relations, and conduct epidemiological investigations.

Keywords: Methicillin Resistance, Staphylococcus aureus, MRSA, mecA, PVL, Duhok, Hospital, Community

# 1. Background

Staphylococcus aureus is the most infectious Staphylococcus species and the most common bacterium isolated from hospital-acquired infections (1). It is also the second most prevalent pathogen among patients in outpatient clinics. *S. aureus* can cause mild to severe infections such as skin and soft tissue infections (SSTIs) and sepsis (1). Additionally, it may lead to toxin-mediated infections that are associated with high mortality. Within a decade of discovery of penicillin, about 90% of the *S. aureus* strains isolated from hospitals became resistant to penicillin by producing beta-lactamases. Hospital-acquired methicillin-resistant *Staphylococcus aureus* (HA-MRSA) includes the strains that are resistant to all b-lactams including penicillins and cephalosporins and are isolated in healthcare environments (2). MSRA rates increased steadily until the late 1990s when it experienced a sharp increase in rate (2). This was followed by the appearance of community-acquired methicillin-resistant *Staphylococcus aureus* (CA-MRSA) when community-acquired infections were identified with MRSA in healthy people with limited or no exposure to predisposing factors of MRSA (1). The *mecA* gene is responsible for methicillin resistance, which is carried by cassette chromosome *mec* (SCCmec)(3).

Life-threatening infections are predominantly associated with CA-MRSA that are more virulent due to carrying Panton-Valentine Leukocidin (PVL). PVL had been found mainly within CA-MSSA infections even before the emergence of MRSA. It is encoded by genes lukS-PV and lukF-PV that comprise two subunits that form pores in the cell wall of leukocytes by producing a substance called leucocidin (4).

In Iran, the prevalence of MRSA in healthcare workers

Copyright © 2019, International Journal of Infection. This is an open-access article distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 International License (http://creativecommons.org/licenses/by-nc/4.0/) which permits copy and redistribute the material just in noncommercial usages, provided the original work is properly cited.

was 16% - 35% (5, 6). The MRSA rate was 10.1% among healthcare workers from Jordan (7), 73% among healthcare workers from Saudi Arabia (8), and 22.5% in healthcare workers from Iraq (9). The prevalence of MRSA carriage was also studied in Iraq. The prevalence of MRSA was found to be 4.2% (10). In another study that was performed in urban and rural schools, the overall nasal carriage rates of S. aureus and MRSA isolates were 17.75% and 10%, respectively (11). MRSA infections are spreading steadily at hospitals and communities, making a public health problem. In order to control MRSA-related infections, the continuous monitoring of the prevalence of S. aureus is needed (1). Continuous study and monitoring of MRSA are critical for the prevention and treatment of MRSA-related infections (12). Such studies can provide information on the spread of the bacterium, the trends of infections and the appearance of new strains in hospitals and communities, the antibiogram and antibiotic sensitivity patterns, and the risk factors associated with infection (12). Such surveillance helps healthcare providers to plan prevention strategies and clinicians to provide accurate empiric antibiotics for infections circulating in their regions with a subsequent improvement of infection treatment outcomes (1).

# 2. Objectives

The aim of this study was to assess and compare the prevalence of CA-MRSA and HA-MRSA in Duhok city, Kurdistan region, Iraq, and detect *mecA* and PVL genes among two healthy groups of people, community students and hospital staff.

# 3. Methods

### 3.1. Sample Collection

With a confidence interval of 95% (Z = 1.96), an error margin of 10%, a design effect of 1, and an expected response rate of 0.8, the sample size of 91 was calculated. The samples were collected in the period between January 2018 and June 2018. Students who had a history of one of the following conditions were excluded: recent admission to healthcare facilities, recent surgical operations, end-stage kidney diseases on hemodialysis, using intravenous cannula at the time of swab taking, MRSA detection, and active skin infections. Both categories of samples were taken from nares by using sterile swabs, inoculated into transport media, and examined immediately after reaching the lab. All healthcare workers who agreed to participate and signed consent forms were included in the study. Healthcare workers who did not agree to participate were excluded.

## 3.2. Laboratory Investigation

#### 3.2.1. Conventional Tests

All isolates were evaluated by several conventional laboratory tests for detecting and identifying *S. aureus* and MRSA among hospital staff and community students. These tests included Gram staining and biochemical tests such as catalase and tube coagulase tests. Furthermore, all samples were cultured on Mannitol Salt Agar (Oxoid) for fermentation and Brain Heart Infusion (BHI) (Oxoid). Moreover, Mueller-Hinton Agar (Oxoid Limited, Hampshire, England) was used for antibiotic sensitivity testing to oxacillin by using Kirby-Bauer disk diffusion as recommended by the CLIS guidelines (13).

#### 3.2.2. Molecular Analyses

3.2.2.1. Detection of mecA and PVL Genes by Polymerase Chain Reaction (PCR)

PCR was applied to all S. aureus isolates for the detection of *mecA* and PVL genes according to instructions by Murakami et al. (14). The forward (GTG GAA TTG GCC AAT ACA GG) and reverse (TAG GTT CTG CAG TAC CGG AT) primers were used for the mecA gene. PCR amplification conditions for the mecA gene were 35 cycles starting with initial activation step for 2 min per each cycle at 95°C, followed by denaturation step for 30 s at 95°C. The annealing time was 1 min at 58°C and extension was 2 min at 72°C (15, 16). Luk-PV-1 (ATCATTAGGTAAAATGTCTGGACATGATCCA) and LukPV-(GCATCAAGTGTA TTGG ATAGCAAAAGC) primers were used for the PVL gene. The amplification conditions for PVL started with initial activation for 2 min at 95°C, followed by denaturation for 30 s at 95°C. Then, the annealing step was performed for 1 min at 55°C, followed by primer extension for 2 min at 72°C. The reaction was done for 35 thermal cycles (17).

#### 3.3. Ethical Statement

Consent was taken from hospital staff and community students to use their samples for research purposes. The study was approved by the Scientific Committee of Directorate of Health, Duhok, Kurdistan region of Iraq.

## 4. Results

## 4.1. Staphylococcus aureus Isolates Among Hospital Staff

In the present study, 109 samples were collected from healthcare workers at different health facilities in Duhok city, Kurdistan region of Iraq. There were 67/109 (61.4%) males in the sample. The age range of healthcare workers was 18 - 49 years. Additionally, 103 samples were collected from secondary schools in the city; 78/103 (75.7%) samples were from males with the age range of 18 - 22 years. All the isolates from hospital staff were identified as *S. aureus* according to the standard identification procedure. Based on the genotypic analysis, 55 of 109 (50.4%) isolates were MRSA carrying the *mecA* gene and 4/55 (3.7%) were MRSA-bearing PVL genes. Additionally, 54/109 (49.5%) isolates were MSSA among which, 4/54 (3.7%) isolates were MSSA-PVL gene-positive (Table 1).

Table 1. Number and Prevalence of MRSA/MSSA and PVL Gene Among Hospital Staf and Community Students in Duhok, Kurdistan, Iraq			
Sample Categories	Total Samples	MRSA, No. (%)	MSSA, No. (%)
Hospital staff	109	55 (50.4)	54 (49.5)
PVL-		51(46.8)	50 (45.9)
PVL+		4 (3.7)	4 (3.7)
Community students	103	1(4.3)	22 (95.7)
PVL+		1(4.3)	5 (21.7)
PVL-			17 (73.9)

4.2. Staphylococcus aureus Isolates Among Community Students

The results showed that 23/103 (22.3%) samples from community students were determined as *S. aureus*. There were 22 MSSA strains among which, 5/22 (21.7%) isolates were MSSA carrying the PVL gene whereas 17/22 (73.9%) isolates were MSSA PVL gene-negative. Moreover, only was 1/23 (4.3%) isolate determined as MRSA that was carrying the PVL gene (Table 1).

## 5. Discussion

In the present study, all healthcare workers were found to be *S. aureus* carriers among whom, 50.4% of the isolates were identified as MRSA that carried the *mecA* gene. A comparable rate was found in a previous study carried out in Duhok where MRSA was found in 51.7% of the isolated *S. aureus* strains in healthcare workers, implying that MRSA is prevalent at hospitals in Kurdistan (15).

Based on the genotypic analysis, 55 out of 109 (50.4%) isolates were MRSA carrying the *mec*A gene. Among healthcare worker isolates, 7% carried the PVL gene including 3.7% MRSA-PVL-positive isolates and 3.7% MSSA-PVL genepositive isolates. Noteworthy, these data were comparable with the results of a previous study that was performed in Duhok in 2016 among healthcare workers (15). A similar rate of PVL positivity was found in MRSA strains isolated from hospitals in the Netherland where about 8% of the isolates carried the PVL gene. In Iran, the prevalence of MRSA, the PVL gene, and MRSA-PVL isolates were 32%, 19%, and 10%, respectively (18). In Germany, the prevalence of MRSA isolates among healthcare workers was 11.3%, among which 9.1% of the isolates were positive for PVL (19).

On the other hand, about 22.3% of the nasal carriage isolates from community students were found to be S. aureus that was comparable to the rate found in northern Pakistan (24%) (20). In contrast, a higher rate was reported in the United State where 28.6% of the recruited samples carried S. aureus. In 2014, a study was conducted in Duhok city to evaluate the nasal carriage of CA-MRSA and the results showed that 18.4% and 2.04% carried S. aureus and MRSA, respectively (21). Additionally, a similar rate of MRSA was shown among university students with the rates of 17.5% and 4.2% for S. aureus and MRSA, respectively (10). Moreover, the MRSA rate among community students that carried the PVL gene in the current study was 4.3%, which is higher than the rate (2.5%) found among students in Palestine in 2011(20) and likewise the rate (2.4%) in Brazil (22). Interestingly, the prevalence rates of HA-MRSA and CA-MRSA carrying PVL were 3.7% and 4.3%, respectively. More studies are needed to monitor the prevalence rates of PVL and its evolution over time.

To conclude, a higher prevalence of MRSA was found in healthcare workers whereas the MRSA isolates were carrying a closely similar rate of PVL gene in both groups. Moreover, CA-MSSA was carrying the highest rate of the PVL gene. Additionally, continuous surveillance is needed to reduce the spread of isolates between hospital staff carriers and patients, as well as in the community.

# Footnotes

Authors' Contribution: All authors contributed equally.

**Conflict of Interests:** It is not declared by the authors.

**Ethical Approval:** The research proposal and consent forms were approved by the Ethics Committee of Directorate of Health, Duhok, Kurdistan region of Iraq.

**Funding/Support:** This project was funded by the College of Medicine, University of Duhok, Kurdistan region of Iraq.

**Patient Consent:** Written informed consent was obtained from the hospital staff and community students to use their samples for research purposes.

#### References

- Kong EF, Johnson JK, Jabra-Rizk MA. Community-associated methicillin-resistant staphylococcus aureus: An enemy amidst us. *PLoS Pathog.* 2016;**12**(10). e1005837. doi: 10.1371/journal.ppat.1005837. [PubMed: 27711163]. [PubMed Central: PMC5053610].
- McKinnell JA, Miller LG, Eells SJ, Cui E, Huang SS. A systematic literature review and meta-analysis of factors associated with methicillin-resistant Staphylococcus aureus colonization at time

of hospital or intensive care unit admission. *Infect Control Hosp Epidemiol*. 2013;**34**(10):1077-86. doi: 10.1086/673157. [PubMed: 24018925]. [PubMed Central: PMC3883507].

- Liu J, Chen D, Peters BM, Li L, Li B, Xu Z, et al. Staphylococcal chromosomal cassettes mec (SCCmec): A mobile genetic element in methicillinresistant Staphylococcus aureus. *Microb Pathog.* 2016;101:56–67. doi: 10.1016/j.micpath.2016.10.028. [PubMed: 27836760].
- Shallcross LJ, Fragaszy E, Johnson AM, Hayward AC. The role of the Panton-Valentine leucocidin toxin in staphylococcal disease: A systematic review and meta-analysis. *Lancet Infect Dis.* 2013;13(1):43–54. doi:10.1016/S1473-3099(12)70238-4. [PubMed: 23103172]. [PubMed Central: PMC3530297].
- Havaei SA, Ghanbari F, Rastegari AA, Azimian A, Khademi F, Hosseini N, et al. Molecular typing of hospital-acquired Staphylococcus aureus isolated from Isfahan, Iran. *Int Sch Res Notices*. 2014;2014:185272. doi: 10.1155/2014/185272. [PubMed: 27350987]. [PubMed Central: PMC4897504].
- Mirkarimi SF, Hasani A, Abdinia B, Soroush Barhaghi MH, Nikbakht M, Ahangarzadeh Rezaee M. High diversity of methicillin-resistant Staphylococcus aureus (MRSA) isolates based on hypervariable region polymorphisms. Arch Pediatr Infect Dis. 2016;4(4). doi: 10.5812/pedinfect.36433.
- Aqel AA, Alzoubi HM, Vickers A, Pichon B, Kearns AM. Molecular epidemiology of nasal isolates of methicillin-resistant Staphylococcus aureus from Jordan. J Infect Public Health. 2015;8(1):90–7. doi: 10.1016/j.jiph.2014.05.007. [PubMed: 25002017].
- Iyer A, Kumosani T, Azhar E, Barbour E, Harakeh S. High incidence rate of methicillin-resistant Staphylococcus aureus (MRSA) among healthcare workers in Saudi Arabia. J Infect Dev Ctries. 2014;8(3):372–8. doi: 10.3855/jidc.3589. [PubMed: 24619270].
- Hussein NR, Assafi MS, Ijaz T. Methicillin-resistant Staphylococcus aureus nasal colonisation amongst healthcare workers in Kurdistan region, Iraq. J Glob Antimicrob Resist. 2017;9:78–81. doi: 10.1016/j.jgar.2017.01.010. [PubMed: 28419870].
- Assafi MS, Mohammed RQ, Hussein NR. Nasal carriage rates of Staphylococcus aureus and CA-methicillin resistant Staphylococcus aureus among university students. J Microbiol Res. 2015;5(4):123–7. doi: 10.5923/j.microbiology.20150504.01.
- Hussein NR, Basharat Z, Muhammed AH, Al-Dabbagh SA. Comparative evaluation of MRSA nasal colonization epidemiology in the urban and rural secondary school community of Kurdistan, Iraq. *PLoS One*. 2015;**10**(5). e0124920. doi: 10.1371/journal.pone.0124920. [PubMed: 25932644]. [PubMed Central: PMC4416827].
- 12. Peterson LR, Schora DM. Methicillin-resistant Staphylococcus aureus control in the 21st century: Laboratory involvement affecting disease

impact and economic benefit from large population studies. *J Clin Microbiol.* 2016;**54**(11):2647-54. doi: 10.1128/JCM.00698-16. [PubMed: 27307459]. [PubMed Central: PMC5078538].

- 13. National Committee for Clinical Laboratory Standards. *Performance standards for antimicrobial susceptibility testing*. 14th ed. National Committee for Clinical Laboratory Standards; 2004.
- Murakami K, Minamide W, Wada K, Nakamura E, Teraoka H, Watanabe S. Identification of methicillin-resistant strains of staphylococci by polymerase chain reaction. *J Clin Microbiol*. 1991;**29**(10):2240–4. [PubMed: 1939577]. [PubMed Central: PMC270305].
- Hussein NR. Prevalent genotypes of Staphylococcus aureus strains isolated from healthcare workers in Duhok city, Kurdistan region, Iraq. Int J Infect. 2016;3(2). doi: 10.17795/iji-35375.
- Moussa I, Shibl AM. Molecular characterization of methicillinresistant Staphylococcus aureus recovered from outpatient clinics in Riyadh, Saudi Arabia. Saudi Med J. 2009;30(5):611–7. [PubMed: 19417957].
- McClure JA, Conly JM, Lau V, Elsayed S, Louie T, Hutchins W, et al. Novel multiplex PCR assay for detection of the staphylococcal virulence marker Panton-Valentine leukocidin genes and simultaneous discrimination of methicillin-susceptible from -resistant staphylococci. J Clin Microbiol. 2006;44(3):1141–4. doi: 10.1128/JCM.44.3.1141-1144.2006. [PubMed: 16517915]. [PubMed Central: PMC1393128].
- Lari AR, Pourmand MR, Ohadian Moghadam S, Abdossamadi Z, Ebrahimzadeh Namvar A, Asghari B. Prevalence of PVL-containing MRSA isolates among hospital staff nasal carriers. *Lab Med.* 2011;42(5):283–6. doi: 10.1309/lman7hr6vjea3nmr.
- Maltezou HC, Vourli S, Katerelos P, Maragos A, Kotsalidou S, Remoudaki E, et al. Panton-Valentine leukocidin-positive methicillinresistant Staphylococcus aureus outbreak among healthcare workers in a long-term care facility. *Int J Infect Dis.* 2009;**13**(6):e401-6. doi: 10.1016/j.ijid.2009.02.004. [PubMed: 19395300].
- Adwan K, Jarrar N, Abu-Hijleh A, Adwan G, Awwad E, Salameh Y. Molecular analysis and susceptibility patterns of methicillin-resistant Staphylococcus aureus strains causing community- and health careassociated infections in the northern region of Palestine. *Am J Infect Control.* 2013;**41**(3):195-8. doi: 10.1016/j.ajic.2012.03.040. [PubMed: 22998783].
- 21. Habeeb A. Methicillin resistant Staphylococcus aureus nasal colonization among secondary school students at Duhok city-Iraq. J Microbiol Infect Dis. 2014;4(2):59–63. doi: 10.5799/ahinjs.02.2014.02.0128.
- Prates KA, Torres AM, Garcia LB, Ogatta SF, Cardoso CL, Tognim MC. Nasal carriage of methicillin-resistant Staphylococcus aureus in university students. *Braz J Infect Dis.* 2010;14(3):316–8. [PubMed: 20835520].