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Brief Report

Profile of *Streptococcus pneumoniae* Serotypes of Children with Invasive Disease in Tehran, Iran. An Implication for Vaccine Coverage

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Abstract

Objectives: This study aims to define *S. pneumoniae* serotypes in children hospitalized with an invasive pneumococcal disease from March 20, 2012 to March 10, 2013 by polymerase chain reaction (PCR) method.

Methods: Specimens from cerebrospinal fluid and blood were collected from children aged one month-18 years with suspected invasive pneumococcal infection admitted to Mofid Children's Hospital and other regional hospitals. Multiplex PCR with 13 groups of primers were used to detect 33 serotypes of *S. pneumoniae* in positive blood and cerebrospinal fluid cultures. Out of 563 samples, 83 *S. pneumoniae* isolates were identified. Sixty-seven samples were typeable.

Results: The results showed that serotypes 3 (21.7%), 23F (13.2%), and 19F (10.8%) were the most prevalent serotypes. Sixteen samples (19.3%) were non-typeable by Multiplex PCR method. The 13-valent pneumococcal vaccine provides the highest coverage (66.23%), followed by the 10-valent vaccine (34.9%) and, lastly, the 7-valent vaccine (33.71%).

Conclusions: We found that serotypes 3, 23F, and 19F accounted for almost 46% of invasive pneumococcal isolates. As per relatively high coverage of prevalent serotypes, PCV13 should be considered for routine childhood vaccination programs.

Keywords: Streptococcus pneumoniae, Polymerase Chain Reaction, Serotype, Children

1. Background

Streptococcus pneumoniae is a significant cause of mortality and morbidity worldwide; the estimated global mortality rate for *S. pneumoniae* in 2015 was reported as 45 deaths (29 - 56) per 100000 children younger than five years (1). Pneumococcal nasopharyngeal colonization, reported in 27-65% of children, is a potential cause of severe pneumococcal infections (e.g., pneumonia, meningitis, and sepsis) and a source of transmission of the pathogen (2).

In 2000, the first pneumococcal conjugate vaccine (PCV7), comprising the serotypes 4, 6B, 9V, 14, 18C, 19F, and 23F, was licensed by the American Food and Drug Administration (FDA). Since then, PCV10 and PCV13 have also been introduced, each covering additional serotypes to those covered by PCV7 (3). Following the introduction of pneumococcal vaccines, a significant reduction in incidence, hospitalization, and mortality rates related to pneumococcal diseases was observed in high-income and non-high-income countries (4). However, the available vaccines

only include the serotypes responsible for severe pneumococcal disease in developed countries, and the serotypespecific immunity induced by vaccination has led to a relative increase in non-vaccine serotypes in other geographic regions (5, 6). Local and regional detection of invasive pneumococcal serotypes and monitoring the changes over time is essential to investigate the most common invasive serotypes and their coverage by current pneumococcal vaccines.

2. Objectives

Considering the novelty of study on pneumococcal serotypes in Iran and the necessity of its performance, this study aimed to detect *S. pneumoniae* serotypes in children with pneumonia, sepsis, and meningitis by PCR method and estimate their coverage with different pneumococcal vaccines.

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3. Methods

A cross-sectional study was conducted on 563 blood, and cerebrospinal fluid (CSF) specimens collected from children aged one month -18 years and suspected sepsis, pneumonia, and meningitis hospitalized at Mofid Children's Hospital and other regional hospitals from March 20, 2012 to March 10, 2013. Patients with confirmed immunodeficiency, diagnosed underlying disease, nosocomial infection, and children with chronic cough (more than two weeks) were excluded.

Clinical specimens, including 1.5 - 2 ml blood and 1 ml CSF, were obtained by a sterile procedure according to the recommendations for collecting specimens for laboratory diagnosis of S. pneumoniae and Haemophilus influenza (7). Blood specimens were injected in culture broth and mixed gently, and CSF specimens were collected in a sterile screwcapped tube. The samples were transported to the laboratory within 4 - 6 hrs. The culture was done in BACTEC 9120 BD system. Confirmatory tests identified blood and CSF positive culture specimens. DNA extraction was performed by Roche (High pure PCR template preparation kit, Cat No: 11796828001) kit, and S. pneumoniae was confirmed by a specific primer of the cps gene and PCR technique. Confirmed serotyping was done on confirmed Pneumococcus specimens by Multiplex PCR. Specific cps gene primers and PCR solution for each Multiplex PCR group are shown in Tables 1 and 2.

Data regarding patients' characteristics and PCR results were recorded in separate checklists and analyzed using SPSS ver 22. Results were reported as numbers (percent).

4. Results and Discussion

The PCR results indicated that 83 out of 563 (14.7%) specimens were positive for *S. pneumoniae*. Multiplex PCR typed 67 of 83 (80.7%) isolated pneumococci, however 16 samples (19.3%) were non-typeable (Table 3). Overall, 16 serotypes were responsible for the confirmed *S. pneumoniae* infections, the most prevalent serotypes being serotype 3 (21.7%), 23F (13.25%), and 19F (10.84%).

Compared with other studies performed in Iran, our findings showed some differences regarding prevalent pneumococcal serotypes. According to a study in Northeast Iran, serotypes 23F, 19F, 19A, 1, 14, and serogroup 6A/B were the most common types among children with suspected meningitis (8). Serotypes 23F, 14, 3, 19, and 19A were reported as the most prevalent in children with invasive and non-invasive pneumococcal infections admitted to Shariati Hospital, Tehran (9). Moreover, in a study on clinical samples obtained from children with an invasive

pneumococcal disease from several hospitals in Tehran, 23F, 19F, 19A, and 9V as the most common serotypes (10).

Studies from other countries have reported a wide variety of prevalent serotypes. In a study conducted in 60 hospitals from 11 Asian countries, the most common typeable serotypes were 19F (23.5%), 23F (10%), and 19A (8.2%); and non-typeable serotypes accounted for (5.4%) of isolates (11). On the other hand, a Turkish study performed on 167 children with invasive pneumococcal disease reported 19F, 1, 3, and 19A as the most common invasive pneumococcal serotypes (12). The differences in pneumococcal serotype distribution can be attributed to several factors such as geographic regions, type of pneumococcal disease (invasive or non-invasive), study population characteristics, source of clinical specimens, and PCR method.

Iranian national immunization program does not include routine pneumococcal vaccination. Accordingly, there is lower PCV7, PCV10, and PCV13 coverage than some other countries (11-13). Interestingly, pneumococcal vaccine coverage reported by Abdoli et al and Houri et al were higher than our findings (8, 10). Differences in pneumococcal serotypes distribution might be explained by geographical and temporal changes and PCR techniques. Based on our results, PCV-13 is the most influential vaccine against common invasive pneumococcal serotypes, covering 66.2% of serotypes detected by PCR. As most samples were collected from patients hospitalized in Mofid children's hospital, one of the primary referral pediatric centers in Iran, PCV-13 might be considered for inclusion in the national immunization program. However, further studies in different parts of Iran are recommended, as serotype distribution may change over time.

The study's limitation was difficulties in at time specimen collection before the start of antibiotics We organized our educated physicians to obtain standard specimens for serotyping at the time of admission. Our study's advantages were using clinical isolates that represent invasive pneumococcal serotypes in our society.

5. Conclusions

We found that serotypes 3, 23F, and 19F accounted for almost 46% of invasive pneumococcal isolates. As per relatively high representation of prevalent serotypes in PCV-13, this vaccine is the preferred choice in Tehran, Iran. More research on serotypes in different geographical regions allows the design of a nationwide vaccination program.

The Ethics Committees of Shahid Beheshti University of Medical Sciences and Tehran University of Medical Sciences, Tehran, Iran approved the study (Ethics code: 1391-1-87-9954-10064).

No	Primer Name	Sequence Primer	Size Primer (bp)
1	CDSA F	GCAGTACAGCAGTTTGTTGGACTGACC	27
2	CpsA R	GAATATTTTCATTATCAGTCCCAGTC	-,
2	1 E		20
3	1 R	CCAAACAAAATACTAACATTATCACAATATTGGC	34
5	4 F	CTGTTACTTGTTCTGGACTCTCGATAATTGG	31
6	4R	GCCCACTCCTGTTAAAATCCTACCCGCATTG	31
7	5 F	ATACCTACAAACTTCTGATTATGCCTTTGTG	32
8	5 R	GCTCGATAAACATAATCAATATTTGAAAAAGTATG	35
9	6A/6B/6C F	AATTTGTATTTATTCATGCCTATATCTGG	30
10	6A/6B/6C R	TTAGCCGAGATAATTTAAAATGATGACTA	29
11	7f/7a F	CCTACGGGAGGATATAAAATTATTTTTGAG	30
12	7f/7a P		30
12	707a K		30
13	7C P	GAACATAGATGTTCAGACATCTTTTCTAATTC	30
15	8F	GATGCCATGAATCAAGCAGTGGCTATAAATC	31
16	8R	ATCCTCGTGTATAATTTCAGGTATGCCACC	30
17	9V F	CTTCGTTAGTTAAAATTCTAAATTTTTCTAAG	32
18	9V R	GTCCAATACCAGTCCTTGCAACACAAG	27
19	10A F	GGTGTAGATTTACCATTAGTGTCGGCAGAC	30
20	10A R	GAATTTCTTCTTTAAGATTCGGATATTTCTC	31
21	11A F	GGACATGTTCAGGTGATTTCCCAATATAGTG	31
22	11A R	GATTATGAGTGTAATTTATTCCAACTTCTCCC	32
23	12f F	GCAACAAACGGCGTGAAAGTAGTTG	25
24	12f R	CAAGATGAATATCACTACCAATAACAAAAC	30
25	14 F	GAAATGTTACTTGGCGCAGGTGTCAGAATT	30
26	14 R	GCCAATACTTCTTAGTCTCTCAGATGAAT	29
27	15A F	ATTAGTACAGCTGGAATATCTCTTC	25
28	15A R	GATCTAGTGAACGTACTATTCCAAAC	26
29	15B/C F	TTGGAATTTTTTAATTAGTGGCTTACCTA	29
30	15B/C R	CATCCGCTTATTAATTGAAGTAATCTGAACC	31
31	16 F	CTGTTCAGATAGGCCATTTACAGCTTTAAATC	32
32	16 R	CATTCCTTTTGTTATTAGTGCTAGTTCATCC	31
33	17 F	TTCGTGATGATAATTCCAATGATCAAACAAGAG	33
34	17 R	GATGTAACAAATTTGTAGCGACTAAGGTCTGC	32
35	18c F	CTTAATAGCTCTCATTATTCTTTTTTAAGCC	32
36	18c R	TTATCTGTAAACCATATCAGCATCTGAAAC	30
37	19 F F		31
38	19 F K		32
39	19/1 F		28
40	20 F	GAGCAACAGTTTTTCACCTGACAGCGAGAAG	27
42	20 R		33
43	20 R 22f F	GAGTATAGCCAGATTATGGCAGTTTTATTGTC	32
44	22f R	CTCCAGCACTTGCGCTGGAAACAACAGACAAC	32
45	23f F	GTAACAGTIGCTGTAGAGGGAATTGGCTTTTC	32
46	23f R	CACAACACCTAACACCGATGGCTATATGATTC	33
47	31 F	GGAAGTTTTCAAGGATATGATAGTGGTGGTGG	32
48	31 R	CCGAATAATATATTCAATATATTCCTACTC	30
49	33f F	GAAGGCAATCAATGTGATTGTGTCGCG	27
50	33f R	CTTCAAAATGAAGATTATAGTACCCTTCTAC	31
51	34 F	GCTTTTGTAAGAGGAGATTATTTTCACCCAAC	32
52	34 R	CAATCCGACTAAGTCTTCAGTAAAAAACTTTAC	33

Table 2. PCR Solution for Each Multiplex PCR Group						
Primers	Addition Primers (µL)	DNA Sample (µL)	Distilled Water (µL)	The Final Volume (µL)		
19A	2	_	7	20		
19F	2	_				
6A/B	2	3				
1	2					
CPS	2					
5	2					
14	2	3 9	20			
7F/A CPS	2					
5	2					
23F	3			20		
7F	4					
11A	2	3	3 4			
33F	2					
cps	2					
16F	4			20		
Sg18	2.5		6 5			
35B	2	- 3	3 6.5			
CPS	2					
8	3					
15B/C	3					
38	3	- 3	3 4	20		
31	4					
1	3					
10A	3		_	20		
35F	3	- 3	3 5			
34	3					
20	2					
7C	2	-				
17F	2	3	7	20		
15B/C	2					
cps	2					
4	2					
14	2	1	9	20		
12	2	3				
9V	2	1				
CPS	2	2	16	20		
15A	2	2	16	20		
22F	2	2	16	20		
		1		1		

Serotype	No. (%)	
3	18 (21/7)	
23F	11 (13/25)	
19F	9 (10/84)	
19A	4 (4/81)	
6A	4 (4/81)	
9V	4 (4/81)	
4	3 (3/61)	
11A	3 (3/61)	
15A	3 (3/61)	
6C	2 (2/4)	
35B	1 (1/2)	
34	1 (1/2)	
14	1 (1/2)	
7A	1 (1/2)	
1	1 (1/2)	
23A	1 (1/2)	
Nontyeable	16 (19.3)	
Total	83 (100)	

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Footnotes

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