



Glutamine Supplementation Reduced Fasting Levels of High-Sensitivity C-Reactive Protein in Hospitalized Children with Acute Respiratory Infection: A Randomized Controlled Trial

Reza Alipanah-Moghadam^{#1}, Manuchehr Barak^{#1}, Reza Mohammadi¹, Ali Nemati^{1,*}, Vahideh Aghamohammadi^{2,**}, Mahsa Mohajeri¹ and Elahe Mohammadi²

¹Ardabil University of Medical Sciences, Ardabil, Iran

²Khalkhal University of Medical Sciences, Khalkhal, Iran

*Corresponding author: Department of Clinical Biochemistry, School of Medicine, Ardabil University of Medical Sciences, Ardabil, Iran. Email: nutrition1391@gmail.com

**Corresponding author: Department of Nutrition, Khalkhal University of Medical Sciences, Khalkhal, Iran. Email: v_agamohammadi@yahoo.com

Reza Alipanah-Moghadam and Manuchehr Barak should be regarded as the first author.

Received 2022 January 19; Revised 2022 March 08; Accepted 2022 March 10.

Abstract

Background: Glutamine (Gln), as a precursor of glutathione and attenuation of pro-inflammatory cytokines, has a vital role in the antioxidant and anti-inflammatory defense of the body. Oxidative stress and inflammatory cytokines increase in respiratory diseases.

Objectives: We sought to investigate the effect of Gln supplementation on serum levels of some inflammatory and oxidative stress indices in hospitalized children with ARI.

Methods: We conducted a 5-day parallel-group, randomized controlled trial. This clinical trial was held for 5 days to assess the efficacy of the 0.5 g/kg body weight Gln, along with medical therapy, in hospitalized children with ARI.

Results: The difference in the high-sensitivity C-reactive protein (hs-CRP) between the Gln and placebo groups was significant after the intervention (analyzed by analysis of covariance [ANCOVA] after adjusting for the duration of cough and biochemical baseline values, 10.67 [7.77] vs 14.04 [6.57], respectively; $P = 0.005$). Moreover, at the end of the trial, there was no significant difference regarding the duration of hospitalization between the Gln and placebo groups (3.25 [1.37] vs 3.35 [0.8], respectively; $P = 0.70$).

Conclusions: The effect of Gln supplementation on the reduction of hs-CRP in children with ARI was demonstrated in this study. Further research is needed to determine the exact effect of Gln on inflammatory and oxidative stress biomarkers in children with ARI.

Keywords: Acute Respiratory Infection, Children, Glutamine, Cytokines, Oxidative Stress

1. Background

Acute respiratory infection (ARI) is one of the most deadly infectious diseases in children worldwide (in both developed and developing countries), causing discomfort, frequent healthcare visits, and deaths (1). It is estimated that between 1.6 and 2.2 million children die each year from ARI (2). The mortality of ARI varies in different regions of the world (3) and accounts for up to 50% of visits of children to health facilities globally (4). Several socio-cultural, demographic, and environmental risk factors, such as female sex, age, comorbid diseases, nutrition, low-income status, maternal lower age, maternal lower education, place of residence (urban or rural), and wet season, predispose children younger than 5 years to ARI (5). Approaches to control ARIs according to the pneumo-

nia severity usually include 4 basic classes: immunization against specific pathogens, early detection, and therapy of disease, enhancements in nutrition, and appropriate environments (6). Assessments of the World Health Organization (WHO) indicate that improvements in nutrition may decrease the risk of ARI incidence or mortality in children of developing countries (7). Therefore, any nutritional interventions can ameliorate child survival from infectious respiratory.

Glutamine (Gln) is the most plentiful free amino acid in plasma and tissue, synthesized in the lungs, liver, brain, skeletal muscles, and adipose tissue and secreted into the circulation (8). Most consumers of Gln are the small intestine, leukocytes, liver, and kidneys (9). Gln has important and regulatory functions in metabolism (as the lipogenic

and glucogenic precursor and oxidative energy), protein synthesis and degradation, cell survival and growth, and expression of genes associated with metabolism. Moreover, Gln, as a precursor of glutathione and attenuation of pro-inflammatory cytokines, has a vital role in the antioxidant and anti-inflammatory defense of the body (10-14). As a conditionally essential amino acid, Gln is an essential element in the proliferation and function of immune cells; hence, Gln deficiency may have an intense impact on the immune system and may elevate the risk of respiratory infections. In catabolic conditions, Gln levels fall below normal, mainly in the muscle and liver (15, 16). Endogenous Gln synthesis does not provide the human body's needs in catabolic conditions, including severe and long-term physical exercise, trauma, cancer, surgeries, sepsis, and infections. Under physiological conditions, Gln is efficiently synthesized in the liver and skeletal system. However, under catabolic situations and oxidative stress, concentrations of Gln in tissues decrease swiftly to assist the further demands of the body, resulting in energy metabolism disruptions and a weakened immune system (16). These disturbances can be lessened by supplementation with Gln; accordingly, it is currently a component of clinical nutrition supplementation practices and/or introduced for patients with immune suppression (17).

Given the increase of oxidative stress (18-20) and inflammatory cytokines in respiratory diseases (21, 22) and the high prevalence of ARI in children, we sought for the first time to investigate the effects of Gln supplementation on serum levels of tumor necrosis factor α (TNF- α), interleukin 1 beta (IL-1 β), high-sensitivity C-reactive protein (hs-CRP), malondialdehyde (MDA), and total antioxidant capacity (TAC) in hospitalized children with ARI.

2. Methods

2.1. Study Design

We conducted a 5-day parallel-group, randomized controlled trial. This clinical trial was carried out at the Bu Ali Hospital, Ardabil University of Medical Sciences, for 5 days to determine the efficacy of the 0.5 g/kg body weight Gln, along with medical therapy, in hospitalized children with ARI (Figure 1).

2.2. Ethics and Trial Registration

The study protocol was approved by the Ethics Committee of Ardabil University of Medical Sciences (code: IR.ARUMS.REC.1398.122), and it was conducted in accordance with the Helsinki Declaration. This clinical trial was registered on the Iranian Registry of Clinical Trials website (registration number: IRCT20210913052455N1).

2.3. Participants and Intervention

By referring to the Bu Ali Hospital, Ardabil, Iran, 44 children with ARI who met the inclusion criteria were recruited in this trial, and written informed consent was obtained from their parents (May 2020 to July 2020). Inclusion criteria were age between 2 to 6 years, having ARI, and not having any chronic disease such as congenital heart diseases, chronic liver or renal diseases, immune deficiency, or malignancy. Exclusion criteria were refusal to continue and consumption of less than 10% of total administered powders. The diagnosis of ARI and the general management of ARI were made by pediatric physicians. The sample size was determined based on serum MDA as the primary outcome obtained from our previous study (23). The sample size was computed using the following formula: $N = [Z_{1-\frac{\alpha}{2}} + Z_{1-\beta}]^2 \times (\delta_1^2 + \delta_2^2) / (\mu_2 - \mu_1)^2$ ($\alpha = 0.05$ and $\beta = 0.2$). Considering a withdrawal, 22 subjects were recruited. A member of the research team who was not involved in assessing the outcome of the study was responsible for generating the allocation sequence and allocating participants to the sequence. The Random Allocation Software (RAS) was used for permuted-block randomization to allocate participants to 2 groups (the Gln supplementation or intervention group and sugar supplementation or placebo group) stratified by age and gender. The Gln group (n = 22) received Gln powder (0.5g/kg body weight) + 5 g of sugar dissolved in 100 mL of water per day for 5 days (24-26). Also, each subject in the placebo group (n = 22) received 5 g of sugar dissolved in 100 mL of water. Participants in both groups were advised to dissolve the content of 1 sachet in 100 mL of water and drink it immediately every evening. Gln was purchased from the Karen Company, Iran. The company had no role in the conception, design, and procedure of the study or in the analysis and interpretation of the data. In the beginning, demographic characteristics, including economic status, duration of disease, and place of residence (rural or urban), were recorded by the researcher via a sociodemographic questionnaire. The subjects were fully informed about the study's protocol. The parents were requested to consume supplements regularly, and to ensure that participants consumed the supplement properly, participants were contacted every day by one of the authors.

2.4. Assessment of Anthropometric Parameters

A trained dietitian measured anthropometric variables, including body weight, height, and body mass index (BMI), after overnight fasting (27). Body weight was measured by a Seca scale with an accuracy of 100 g at the beginning and end of the trial. Height was assessed in a relaxed

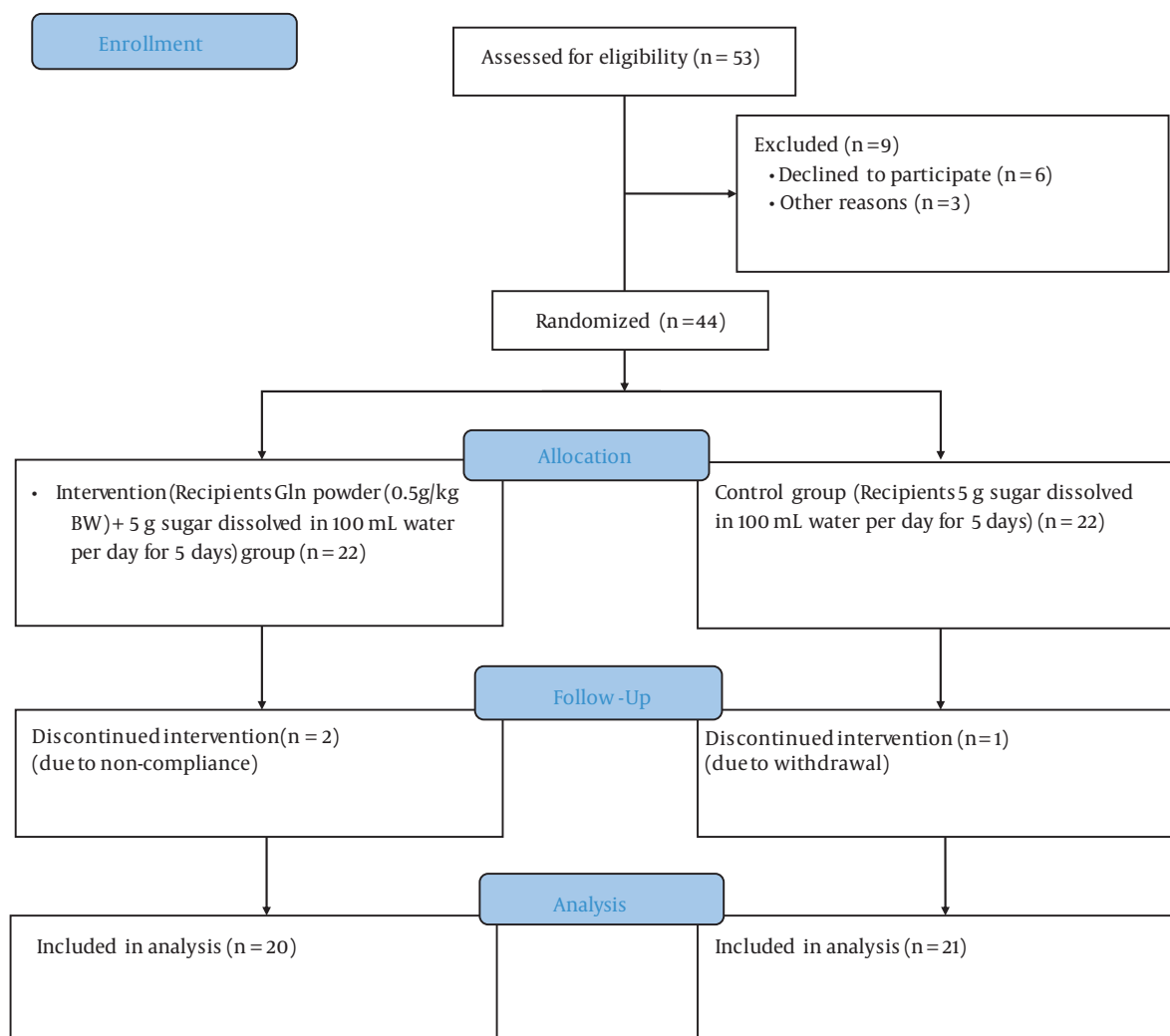


Figure 1. Consolidated Standards of Reporting Trials 2010 flow diagram

position using a Seca portable stadiometer with an accuracy of 0.5 cm. BMI was calculated as body weight in kilograms divided by the square of height in meters at baseline and the end of the study (28).

2.5. Assessment of Appetite

Appetite was evaluated by the Council on Nutrition Appetite Questionnaire (CNAQ), which was adapted by Wilson et al. (29). CNAQ contains 8 items, and the scales of each item range from 1 to 5. Thus, the range of the total score is 8 to 40 points. A score of less than 28 is a cause for concern. The validity of CNAQ has been confirmed in the Iranian population (Cronbach $\alpha = 0.77$) (30).

2.6. Assessment of Biochemistry Variables

Before and after the trial, venous blood samples (2 mL) were collected after 10-12 hours of overnight fasting. Enzyme-linked immunosorbent assay (ELISA) kits (Zell Bio GmbH) were used to determine serum levels of IL-1 β , TNF- α , MDA, and TAC. Moreover, an ELISA kit (Pars Azmoun Co, Karaj, Iran) was applied to determine serum levels of hs-CRP (31).

2.7. Statistical Analysis

The data analyst was blinded after assignment to interventions. Statistical analysis was performed using SPSS version 23 (SPSS Inc, Chicago, Ill, USA). P values less than 0.05 were considered statistically significant. The Kolmogorov-Smirnov test was used to assess the normal distribution of

Table 1. Baseline Characteristics of the Study Groups ^a

Variables	Gln Group (n = 20)	Placebo Group (n = 21)	Statistical Indicators
Economic status ^b			0.629
Equal income and expense	8 (40.0)	6 (30.0)	
Income more than expense	7 (35.0)	10 (50.0)	
Income less than expense	5 (25.0)	4 (20.0)	
Gender ^b			0.501
Girl	15 (75.0)	12 (60.0)	
Boy	5 (25.0)	8 (40.0)	
Fever at the beginning ^b			0.366
Yes	15 (75.0)	13 (65.0)	
No	5 (25.0)	7 (35.0)	
Dyspnea ^b			0.698
Yes	18 (90.0)	18 (90.0)	
No	2 (10.0)	2 (10.0)	
Age ^c	4 (2.25 - 4.75)	3.75 (2.25 - 4)	0.741
Weight (kg) ^d	15.36 (3.12)	15.15 (2.63)	0.817
Height (cm) ^c	98.2 (7.61)	98.88 (6.96)	0.796
BMI (kg/m²) ^c	15.49 (2.01)	15.23 (1.74)	0.660
Duration of cough (day) ^d	2.50 (2 - 3)	2 (1 - 2)	0.005
Appetite ^c	23.2 (9.09)	18.9 (5.95)	0.086

Abbreviations: Gln, glutamine; BMI, body mass index.

^a Independent *t* test or Mann-Whitney U test for numeric variables and Pearson chi-square test for categorical variables.

^b Values are expressed as No. (%).

^c Values are expressed as mean (SD).

^d Values are expressed as Median (percentiles)

the data. A chi-square test was applied to compare the categorical variables between the 2 groups at the baseline. The independent samples *t* test and Mann-Whitney U test were used to compare differences in parametric continuous and nonparametric data between the 2 groups, respectively. To compare within-group changes in variables, the paired-samples *t* test or Wilcoxon signed-rank test were used. To control confounding variables (TNF- α , IL-1 β , hs-CRP, MDA, and TAC baseline values), analysis of covariance (ANCOVA) was applied to identify any differences between the groups after the intervention, adjusting for baseline values and covariates.

3. Results

Two patients in the Gln group and 1 patient in the placebo group were lost to follow-up due to non-compliance and withdrawal, respectively. The Gln and placebo groups were homogenous regarding the economic status, gender, place of residence, fever at the beginning, dyspnea age, height, weight, BMI, and appetite ($P > 0.05$; Table 1). However, the study groups had a significant difference in days of cough (2.45 [0.76] vs 1.8 [0.61]; $P = 0.005$).

As presented in Table 2, there were no significant differences between the Gln and placebo groups in terms of biochemical measurements ($P > 0.05$) at the baseline, except for hs-CRP ($P = 0.004$). At the end of the intervention, the serum values of IL-1 β , TNF- α , and hs-CRP significantly decreased in the Gln group ($P < 0.05$). In the placebo group, there were no significant changes in serum levels of biochemical variables at the end of the trial compared to baseline ($P < 0.05$). The difference in hs-CRP between the Gln and placebo groups after 5 days of intervention was significant (analyzed by ANCOVA after adjusting for the duration of cough and biochemical baseline values). Moreover, at the end of the study, there was no significant difference regarding the duration of hospitalization between the Gln and placebo groups (3.25 [1.37] vs 3.35 [0.8], respectively; $P = 0.70$).

4. Discussion

According to the best of our knowledge, no study has examined the effects of Gln supplementation, along with medical therapy, in hospitalized children with ARI. It was shown that in the intervention group, hs-CRP was significantly reduced compared to the placebo group. Higher

Table 2. Biochemical Parameter Values of the Study Groups at Baseline and at the End of the Intervention

Variable	Gln Group (n = 20)	Placebo Group (n = 21)	P Value
IL-1β (mIU/mL)^a			
Before	2917.18 (2403.70)	4206.25 (2989.75)	0.183 ^b
After	2000.11 (1493.45)	2934.29 (2190.28)	0.391 ^c
MD, p ^d	-917.05, 0.028 ^e	-1271.96, 0.117	
TNF-α (ng/L)^f			
Before	433.2 (231.10 - 924.65)	358.1 (152.3 - 873.52)	0.277 ^b
After	216.8 (143.60 - 503.52)	219.20 (175.77 - 649.50)	0.798 ^c
MD, p ^d	-239.22, 0.002 ^e	-110.17, 0.167	
hs-CRP (mg/L)			
Before	18.37 (7.02)	11.53 (7.11)	0.004 ^{b,e}
After	10.67 (7.77)	14.04 (6.57)	0.005 ^{c,e}
MD, p ^d	-7.69, < 0.001 ^e	-2.51, 0.341	
MDA (nmol/mL)			
Before	2.08 (0.70)	1.99 (0.63)	0.779 ^b
After	1.84 (0.35)	1.81(0.25)	0.930 ^c
MD, p ^d	-0.24, 0.136	-0.122, 0.465	
TAC (nmol/mL)^f			
Before	0.85 (0.74 - 1.07)	0.84 (0.73 - 1.17)	0.373 ^b
After	0.86 (0.74 - 1.10)	1.1 (0.96 - 1.37)	0.100 ^c
MD, p ^d	0.06, 0.395	0.185, 0.769	

Abbreviations: IL-1 β , interleukin 1 beta; TNF- α , tumor necrosis factor α ; hs-CRP, high-sensitivity C-reactive protein; TAC, total antioxidant capacity; MD, mean difference; MDA, malondialdehyde.

^a Values are expressed as Mean (SD).

^b Independent t test or Mann-Whitney U test.

^c Analysis of covariance (adjusted for duration of cough and biochemical baseline values).

^d Paired t test or Wilcoxon signed-rank test.

^e P values of statistical significance (P < 0.05).

^f Values are expressed as median (percentiles).

hs-CRP levels were significantly associated with poor respiratory function in children (32). In ARI, periodic assessment of CRP can instruct the true recovery or deteriorating phases of infection independently from viable signs and symptoms (33). In a recent study, reduced immunoglobulin A and CD4+ CD25+ T cells percentage, as well as elevated hs-CRP, IL-10, and procalcitonin, were related to pneumonia in children with COVID-19 (34). Abdollahi et al. concluded that hs-CRP measurement could be effective in the prediction of early neonatal sepsis (35). Moreover, according to the findings of studies by Xia et al. (36) and Lu et al. (37), the detection of hs-CRP or CRP is helpful for differential diagnosis and estimating the therapeutic effect of ARI in children. Since the elevation of hs-CRP and other inflammatory biomarkers is related to pneumonia in children, a decrease of inflammatory biomarkers through safe supplements such as Gln, along with medicine, can help to cure ARI and reduce infectious morbidity rates.

In the randomized controlled trial by Cai et al., Gln supplementation and recombinant human growth hormone

for 14 days induced a significant reduction in CRP in critically ill elderly patients (38). In another study, parenteral Gln decreased CRP levels and infectious morbidity rates in patients with severe acute pancreatitis (39). Moreover, the combination of normal saline, hydroxyethyl starch, and Gln in severe acute pancreatitis resulted in a reduction in serum TNF- α , IL-8, and CRP concentrations (40). In the study by Wischmeyer et al. parenteral Gln administration in severe burn patients for 14 days reduced CRP in the Gln group, although this reduction was not statistically significant compared with the control group (41). We failed to show any significant reduction in other inflammatory indices compared to the placebo group. However, in the study by Ameho et al., Gln contributed to reducing the concentrations of the pro-inflammatory biomarkers, including IL-8 and TNF- α , in inflamed colonic tissues, leading to disease amelioration in experimental trinitrobenzene sulfuric acid-induced colitis in rats (42). Various studies have revealed that nutrition support supplemented with specific immunonutrients such as Gln may improve intestinal

integrity and modulate acute phase responses (43-45), but the precise mechanism of Gln's protection is unknown.

In the present study, the Gln administration did not change MDA and TAC levels. In line with our findings, enteral Gln administration of 45 g/d for 5 days did not change serum levels of MDA compared to the control group in patients with peritonitis or abdominal trauma (24). Seven days of Gln supplementation (0.15 g/kg) did not improve oxidative stress indices (MDA and TAC) in young, healthy men (46). Gln supplementation for 5 days increased the levels of 70-kd heat-shock protein as an oxidative stress marker and did not affect IL-10 and IL-6 levels in critically ill children (26). We presume that the differences between the conducted studies may be related to various patient groups, doses, and durations of Gln administration. The strengths of our study were the use of a placebo, regular follow-up of participants for supplement consumption, and evaluation of the effect of Gln on inflammatory and oxidative stress biomarkers levels in ARI for the first time. Limitations of this trial were lack of nutritional intake evaluation, short duration of Gln supplementation, and small sample size.

4.1. Conclusions

The effect of Gln supplementation on the reduction of hs-CRP in children with ARI was demonstrated in this study. Further studies are needed to determine the exact effects of Gln on inflammatory and oxidative stress biomarkers in children with ARI.

Acknowledgments

We hereby express our gratitude to Ardabil University of Medical Sciences for funding this research.

Footnotes

Authors' Contribution: Study concept and design: M. B. and A. N.; statistical analysis: V. A. and M. B.; analysis and interpretation of data: V. A., R. A., and M. M.; drafting of the manuscript: M. M., R. A., and M. B.; critical revision of the manuscript for important intellectual content: V. A., E. M., and A. N.

Clinical Trial Registration Code: This clinical trial was registered on the Iranian Registry of Clinical Trials website (registration number: IRCT2021091305245N1).

Conflict of Interests: The authors declare no conflict of interest.

Data Reproducibility: The dataset presented in the study is available on request from the corresponding author during submission or after its publication.

Ethical Approval: The study protocol was approved by the Ethics Committee of Ardabil University of Medical Sciences (code: IR.ARUMS.REC.1398.122). Link: ethics.research.ac.ir/EthicsProposalView.php?id=70728

Funding/Support: This study was financially supported by Ardabil University of Medical Sciences.

Informed Consent: Written informed consent was obtained from all patients' parents.

References

- Rudasingwa G. Potential risk factors contributing to acute respiratory infections among under 5 years children in Rwanda. *Int J Infect Dis.* 2020;**101**. doi: [10.1016/j.ijid.2020.09.831](https://doi.org/10.1016/j.ijid.2020.09.831).
- Williams BG, Gouws E, Boschi-Pinto C, Bryce J, Dye C. Estimates of world-wide distribution of child deaths from acute respiratory infections. *Lancet Infect Dis.* 2002;**2**(1):25-32. doi: [10.1016/S1473-3099\(01\)00170-0](https://doi.org/10.1016/S1473-3099(01)00170-0). [PubMed: [11892493](https://pubmed.ncbi.nlm.nih.gov/11892493/)].
- G. B. D. Child Mortality Collaborators. Global, regional, national, and selected subnational levels of stillbirths, neonatal, infant, and under-5 mortality, 1980-2015: a systematic analysis for the Global Burden of Disease Study 2015. *Lancet.* 2016;**388**(10053):1725-74. doi: [10.1016/S0140-6736\(16\)31575-6](https://doi.org/10.1016/S0140-6736(16)31575-6). [PubMed: [27733285](https://pubmed.ncbi.nlm.nih.gov/27733285/)]. [PubMed Central: [PMC5224696](https://pubmed.ncbi.nlm.nih.gov/PMC5224696/)].
- West TE, Goetghebuer T, Milligan P, Mulholland EK, Weber MW. Long-term morbidity and mortality following hypoxaemic lower respiratory tract infection in Gambian children. *Bull World Health Organ.* 1999;**77**(2):144-8. [PubMed: [10083713](https://pubmed.ncbi.nlm.nih.gov/10083713/)]. [PubMed Central: [PMC2557604](https://pubmed.ncbi.nlm.nih.gov/PMC2557604/)].
- Dagne H, Andualem Z, Dagnew B, Taddese AA. Acute respiratory infection and its associated factors among children under-five years attending pediatrics ward at University of Gondar Comprehensive Specialized Hospital, Northwest Ethiopia: institution-based cross-sectional study. *BMC Pediatr.* 2020;**20**(1):93. doi: [10.1186/s12887-020-1997-2](https://doi.org/10.1186/s12887-020-1997-2). [PubMed: [32111196](https://pubmed.ncbi.nlm.nih.gov/32111196/)]. [PubMed Central: [PMC7047350](https://pubmed.ncbi.nlm.nih.gov/PMC7047350/)].
- Simoes EA, Cherian T, Chow J, Shahid-Salles SA, Laxminarayan R, John TJ. Acute respiratory infections in children. In: Jamison DT, Breman JG, Measham AR, Alleyne G, Claeson M, Evans DB, et al., editors. *Disease Control Priorities in Developing Countries*. 2nd ed. New York: Oxford University Press; 2006.
- Roth DE, Caulfield LE, Ezzati M, Black RE. Acute lower respiratory infections in childhood: opportunities for reducing the global burden through nutritional interventions. *Bull World Health Organ.* 2008;**86**(5):356-64. doi: [10.2471/blt.07.049114](https://doi.org/10.2471/blt.07.049114). [PubMed: [18545738](https://pubmed.ncbi.nlm.nih.gov/18545738/)]. [PubMed Central: [PMC2647440](https://pubmed.ncbi.nlm.nih.gov/PMC2647440/)].
- Dechelotte P, Hasselmann M, Cynober L, Allaouchiche B, Coeffier M, Hecksweiler B, et al. L-alanyl-L-glutamine dipeptide-supplemented total parenteral nutrition reduces infectious complications and glucose intolerance in critically ill patients: the French controlled, randomized, double-blind, multicenter study. *Crit Care Med.* 2006;**34**(3):598-604. doi: [10.1097/01.CCM.0000201004.30750.D1](https://doi.org/10.1097/01.CCM.0000201004.30750.D1). [PubMed: [16505644](https://pubmed.ncbi.nlm.nih.gov/16505644/)].
- Calder PC, Newsholme P. Glutamine and the immune system. *Nutrition and immune function*. CABI; 2002. p.109-32.
- Sifa D, Bai X, Zhang D, Hu H, Wu X, Wen A, et al. Dietary glutamine improves meat quality, skeletal muscle antioxidant capacity and glutamine metabolism in broilers under acute heat stress. *J Appl Anim Res.* 2018;**46**(1):1412-7. doi: [10.1080/09712119.2018.1520113](https://doi.org/10.1080/09712119.2018.1520113).
- Wischmeyer PE, Kahana M, Wolfson R, Ren H, Musch MM, Chang EB. Glutamine reduces cytokine release, organ damage, and mortality in a rat model of endotoxemia. *Shock.* 2001;**16**(5):398-402. doi: [10.1097/00024382-200116050-00014](https://doi.org/10.1097/00024382-200116050-00014). [PubMed: [11699081](https://pubmed.ncbi.nlm.nih.gov/11699081/)].

12. Alipanah-Moghadam R, Molazadeh L, Jafari-Suha Z, Naghizadeh-Baghi A, Mohajeri M, Nemati A. Glutamine supplementation can reduce some atherosclerosis markers after exhaustive exercise in young healthy males. *Nutrition*. 2022;**94**:111506. doi: [10.1016/j.nut.2021.111506](https://doi.org/10.1016/j.nut.2021.111506). [PubMed: [34844156](https://pubmed.ncbi.nlm.nih.gov/34844156/)].
13. Ren W, Xia Y, Chen S, Wu G, Bazer FW, Zhou B, et al. Glutamine Metabolism in Macrophages: A Novel Target for Obesity/Type 2 Diabetes. *Adv Nutr*. 2019;**10**(2):321-30. doi: [10.1093/advances/nmy084](https://doi.org/10.1093/advances/nmy084). [PubMed: [30753258](https://pubmed.ncbi.nlm.nih.gov/30753258/)]. [PubMed Central: [PMC6416106](https://pubmed.ncbi.nlm.nih.gov/PMC6416106/)].
14. Raizel R, Leite JS, Hypolito TM, Coqueiro AY, Newsholme P, Cruzat VF, et al. Determination of the anti-inflammatory and cytoprotective effects of L-glutamine and L-alanine, or dipeptide, supplementation in rats submitted to resistance exercise. *Br J Nutr*. 2016;**116**(3):470-9. doi: [10.1017/S0007114516001999](https://doi.org/10.1017/S0007114516001999). [PubMed: [27215379](https://pubmed.ncbi.nlm.nih.gov/27215379/)].
15. Wernerman J. Role of glutamine supplementation in critically ill patients. *Curr Opin Anaesthesiol*. 2008;**21**(2):155-9. doi: [10.1097/ACO.0b013e3282f54fd6](https://doi.org/10.1097/ACO.0b013e3282f54fd6). [PubMed: [18443481](https://pubmed.ncbi.nlm.nih.gov/18443481/)].
16. Engel JM, Pitz S, Muhling J, Menges T, Martens F, Kwapisz M, et al. Role of glutamine administration on T-cell derived inflammatory response after cardiopulmonary bypass. *Clin Nutr*. 2009;**28**(1):15-20. doi: [10.1016/j.clnu.2008.08.007](https://doi.org/10.1016/j.clnu.2008.08.007). [PubMed: [18835506](https://pubmed.ncbi.nlm.nih.gov/18835506/)].
17. Cruzat V, Macedo Rogerio M, Noel Keane K, Curi R, Newsholme P. Glutamine: Metabolism and Immune Function, Supplementation and Clinical Translation. *Nutrients*. 2018;**10**(11). doi: [10.3390/nu10111564](https://doi.org/10.3390/nu10111564). [PubMed: [30360490](https://pubmed.ncbi.nlm.nih.gov/30360490/)]. [PubMed Central: [PMC6266414](https://pubmed.ncbi.nlm.nih.gov/PMC6266414/)].
18. Chen AC, Burr L, McGuckin MA. Oxidative and endoplasmic reticulum stress in respiratory disease. *Clin Transl Immunology*. 2018;**7**(6). e1019. doi: [10.1002/cti2.1019](https://doi.org/10.1002/cti2.1019). [PubMed: [29928501](https://pubmed.ncbi.nlm.nih.gov/29928501/)]. [PubMed Central: [PMC5992202](https://pubmed.ncbi.nlm.nih.gov/PMC5992202/)].
19. Ozsurekci Y, Aykac K. Oxidative Stress Related Diseases in Newborns. *Oxid Med Cell Longev*. 2016;**2016**:2768365. doi: [10.1155/2016/2768365](https://doi.org/10.1155/2016/2768365). [PubMed: [27403229](https://pubmed.ncbi.nlm.nih.gov/27403229/)]. [PubMed Central: [PMC4926016](https://pubmed.ncbi.nlm.nih.gov/PMC4926016/)].
20. Bwititi P, Chinkwo K. Oxidative stress markers in infectious respiratory diseases: current clinical practice. *Int J Res Med Sci*. 2016;**6**(4):1802-13. doi: [10.18203/2320-6012.ijrms20161727](https://doi.org/10.18203/2320-6012.ijrms20161727).
21. Gan WQ, Man SF, Senthilselvan A, Sin DD. Association between chronic obstructive pulmonary disease and systemic inflammation: a systematic review and a meta-analysis. *Thorax*. 2004;**59**(7):574-80. doi: [10.1136/thx.2003.019588](https://doi.org/10.1136/thx.2003.019588). [PubMed: [15223864](https://pubmed.ncbi.nlm.nih.gov/15223864/)]. [PubMed Central: [PMC1747070](https://pubmed.ncbi.nlm.nih.gov/PMC1747070/)].
22. Chung KF. Cytokines as targets in chronic obstructive pulmonary disease. *Curr Drug Targets*. 2006;**7**(6):675-81. doi: [10.2174/13894500677435263](https://doi.org/10.2174/13894500677435263). [PubMed: [16787167](https://pubmed.ncbi.nlm.nih.gov/16787167/)].
23. Nemati A, Alipanah-Moghadam R, Molazadeh L, Naghizadeh Baghi A. The Effect of Glutamine Supplementation on Oxidative Stress and Matrix Metalloproteinase 2 and 9 After Exhaustive Exercise. *Drug Des Devel Ther*. 2019;**13**:4215-23. doi: [10.2147/DDDT.S218606](https://doi.org/10.2147/DDDT.S218606). [PubMed: [31849453](https://pubmed.ncbi.nlm.nih.gov/31849453/)]. [PubMed Central: [PMC6912001](https://pubmed.ncbi.nlm.nih.gov/PMC6912001/)].
24. Kumar S, Kumar R, Sharma SB, Jain BK. Effect of oral glutamine administration on oxidative stress, morbidity and mortality in critically ill surgical patients. *Indian J Gastroenterol*. 2007;**26**(2):70-3. [PubMed: [17558069](https://pubmed.ncbi.nlm.nih.gov/17558069/)].
25. Javanamani R, Nakhostin-Roohi B. [The Effect of One-week Glutamine Supplementation on Oxidative Stress Indices in Healthy Young Men]. *J Ardabil Univ Med Sci*. 2015;**15**(1):83-9. Persian.
26. Jordan I, Balaguer M, Esteban ME, Cambra FJ, Felipe A, Hernandez L, et al. Glutamine effects on heat shock protein 70 and interleukines 6 and 10: Randomized trial of glutamine supplementation versus standard parenteral nutrition in critically ill children. *Clin Nutr*. 2016;**35**(1):34-40. doi: [10.1016/j.clnu.2015.01.019](https://doi.org/10.1016/j.clnu.2015.01.019). [PubMed: [25701159](https://pubmed.ncbi.nlm.nih.gov/25701159/)].
27. Aghamohammadi khiavi V, Pourghassem Gargari B, Aliasgharzadeh A. [Effect of Folic Acid Supplementation on Indices of Glycemic Control, Insulin Resistance and Lipid Profile in Patients With Type 2 Diabetes Mellitus]. *Iran J Endocrinol Metab*. 2011;**13**(4):354-60. Persian.
28. Haidari F, Aghamohammadi V, Mohammadshahi M, Ahmadi-Angali K. Effect of whey protein supplementation on levels of endocannabinoids and some of metabolic risk factors in obese women on a weight-loss diet: a study protocol for a randomized controlled trial. *Nutr J*. 2017;**16**(1):70. doi: [10.1186/s12937-017-0294-x](https://doi.org/10.1186/s12937-017-0294-x). [PubMed: [29061179](https://pubmed.ncbi.nlm.nih.gov/29061179/)]. [PubMed Central: [PMC5654050](https://pubmed.ncbi.nlm.nih.gov/PMC5654050/)].
29. Wilson MM, Thomas DR, Rubenstein LZ, Chibnall JT, Anderson S, Baxi A, et al. Appetite assessment: simple appetite questionnaire predicts weight loss in community-dwelling adults and nursing home residents. *Am J Clin Nutr*. 2005;**82**(5):1074-81. doi: [10.1093/ajcn/82.5.1074](https://doi.org/10.1093/ajcn/82.5.1074). [PubMed: [16280441](https://pubmed.ncbi.nlm.nih.gov/16280441/)].
30. Shafiei Sabet M. [Relationship Between Nutritional Status And Appetite Among Hiv Patients On Haart]. *Nutr Food Sci Res*. 2014;**1**(Suppl. (1)):236. Persian.
31. Tamaddon A, Nasser E, Mohammadi E, Quej D, Zayeri F, Zand H, et al. A Double-blind Randomized Controlled Trial of Curcumin for Improvement in Glycemic Status, Lipid Profile and Systemic Inflammation in β -Thalassemia Major. *J Herb Med*. 2020;**21**. doi: [10.1016/j.hermed.2019.100324](https://doi.org/10.1016/j.hermed.2019.100324).
32. Soferman R, Glatstein M, Sivan Y, Weisman Y. HsCRP levels: measurement of airway inflammation in asthmatic children. *Pediatr Int*. 2008;**50**(1):12-6. doi: [10.1111/j.1442-200X.2007.02517.x](https://doi.org/10.1111/j.1442-200X.2007.02517.x). [PubMed: [18279198](https://pubmed.ncbi.nlm.nih.gov/18279198/)].
33. Das CS. Acute Respiratory Ailments in Pediatric Age Group and Role of CRP in Diagnosis and Management. In: Ansar W, Ghosh S, editors. *Clinical Significance of C-reactive Protein*. Singapore: Springer; 2020. p. 213-48.
34. Li Y, Deng W, Xiong H, Li H, Chen Z, Nie Y, et al. Immune-related factors associated with pneumonia in 127 children with coronavirus disease 2019 in Wuhan. *Pediatr Pulmonol*. 2020;**55**(9):2354-60. doi: [10.1002/ppul.24907](https://doi.org/10.1002/ppul.24907). [PubMed: [32543756](https://pubmed.ncbi.nlm.nih.gov/32543756/)]. [PubMed Central: [PMC7323435](https://pubmed.ncbi.nlm.nih.gov/PMC7323435/)].
35. Abdollahi A, Shoar S, Nayyeri F, Shariat M. Diagnostic Value of Simultaneous Measurement of Procalcitonin, Interleukin-6 and hs-CRP in Prediction of Early-Onset Neonatal Sepsis. *Mediterr J Hematol Infect Dis*. 2012;**4**(1). e2012028. doi: [10.4084/MJHID.2012.028](https://doi.org/10.4084/MJHID.2012.028). [PubMed: [22708043](https://pubmed.ncbi.nlm.nih.gov/22708043/)]. [PubMed Central: [PMC3375671](https://pubmed.ncbi.nlm.nih.gov/PMC3375671/)].
36. Xia R, Guo F, Li Z, Wu R, Fang Q. Application of c-reactive protein test in children with acute respiratory infection [J]. *Journal of Southeast University*; 2004.
37. Lu W, Li L, Chen Y, Ye J. *Combined Detection of hs-CRP and WBC in Pediatric Infectious Diseases*. Guide of China Medicine; 2013.
38. Cai GL, Yan J, Yu YH, Zhang ZC, Gong SJ, Dai HW, et al. [Influence of glutamine and growth hormone intensified nutrition support on immunomodulation in critically ill elderly patients]. *Zhongguo Wei Zhong Bing Ji Jiu Yi Xue*. 2006;**18**(10):595-8. Chinese. [PubMed: [17038243](https://pubmed.ncbi.nlm.nih.gov/17038243/)].
39. Fuentes-Orozco C, Cervantes-Guevara G, Mucino-Hernandez I, Lopez-Ortega A, Ambriz-Gonzalez G, Gutierrez-de-la-Rosa JL, et al. L-alanyl-L-glutamine-supplemented parenteral nutrition decreases infectious morbidity rate in patients with severe acute pancreatitis. *JPEN J Parenter Enteral Nutr*. 2008;**32**(4):403-11. doi: [10.1177/0148607108319797](https://doi.org/10.1177/0148607108319797). [PubMed: [18596311](https://pubmed.ncbi.nlm.nih.gov/18596311/)].
40. Zhao G, Zhang JG, Wu HS, Tao J, Qin Q, Deng SC, et al. Effects of different resuscitation fluid on severe acute pancreatitis. *World J Gastroenterol*. 2013;**19**(13):2044-52. doi: [10.3748/wjg.v19.i13.2044](https://doi.org/10.3748/wjg.v19.i13.2044). [PubMed: [23599623](https://pubmed.ncbi.nlm.nih.gov/23599623/)]. [PubMed Central: [PMC3623981](https://pubmed.ncbi.nlm.nih.gov/PMC3623981/)].
41. Wischmeyer PE, Lynch J, Liedel J, Wolfson R, Riehm J, Gottlieb L, et al. Glutamine administration reduces Gram-negative bacteremia in severely burned patients: a prospective, randomized, double-blind trial versus isonitrogenous control. *Crit Care Med*. 2001;**29**(11):2075-80. doi: [10.1097/00003246-200111000-00006](https://doi.org/10.1097/00003246-200111000-00006). [PubMed: [11700398](https://pubmed.ncbi.nlm.nih.gov/11700398/)].
42. Ameho CK, Adjei AA, Harrison EK, Takeshita K, Morioka T, Arakaki Y, et al. Prophylactic effect of dietary glutamine supplementation on interleukin 8 and tumour necrosis factor alpha production in trinitrobenzene sulphonic acid induced colitis. *Gut*. 1997;**41**(4):487-93. doi: [10.1136/gut.41.4.487](https://doi.org/10.1136/gut.41.4.487). [PubMed: [9391247](https://pubmed.ncbi.nlm.nih.gov/9391247/)]. [PubMed Central:

- [PMCI891521](#)].
43. Zhao G, Wang CY, Wang F, Xiong JX. Clinical study on nutrition support in patients with severe acute pancreatitis. *World J Gastroenterol*. 2003;**9**(9):2105–8. doi: [10.3748/wjg.v9.i9.2105](#). [PubMed: [12970916](#)]. [PubMed Central: [PMC4656684](#)].
 44. Sahin H, Mercanligil SM, Inanc N, Ok E. Effects of glutamine-enriched total parenteral nutrition on acute pancreatitis. *Eur J Clin Nutr*. 2007;**61**(12):1429–34. doi: [10.1038/sj.ejcn.1602664](#). [PubMed: [17311061](#)].
 45. Pearce CB, Sadek SA, Walters AM, Goggin PM, Somers SS, Toh SK, et al. A double-blind, randomised, controlled trial to study the effects of an enteral feed supplemented with glutamine, arginine, and omega-3 fatty acid in predicted acute severe pancreatitis. *JOP*. 2006;**7**(4):361–71. [PubMed: [16832133](#)].
 46. Nakhostin-Roohi B, Javanamani R. The Effect of Glutamine Supplementation on Exercise-Induced Oxidative Stress. *J Adv Agric Technol*. 2015;**2**(1). doi: [10.12720/joaat.2.1.8-12](#).