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**Research Article** 

# Genotypic and Phenotypic Features of Both *NPHS1* and *NPHS2* Genes in Infantile Nephrotic Syndrome and Prognostic Effect of *E117K* Polymorphism in *NPHS1* Gene

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#### Abstract

**Background:** Infantile nephrotic syndrome (INS) refers to disease that is present after the first three months of life up to one year of age. There is genetic heterogeneity and genotype-phenotype correlation is not clear.

**Objectives:** The focus of the present study was to analyze genotypic and phenotypic features of both *NPHS1* and *NPHS2* genes in INS. **Methods:** Clinical data, mutational analysis, histology, treatments, and outcomes of 48 children with NS are evaluated. A direct sequencing of *NPHS1* gene and *NPHS2* gene was performed. Patients were classified into 3 groups; group 1: cases having only *NPHS1* mutation; group 2: cases with only *NPHS2* mutation; group 3: cases without any mutation.

**Results:** The mean age at onset of the disease was  $8.7 \pm 2.3$  months, and mean follow-up time was 8.3 years. Seven familial and 41 sporadic cases of INS were found. Kidney biopsy was performed in 45 out of 48 patients and pathological investigations revealed focal segmental glomerulosclerosis in 29 (65%), IgM nephropathy in 6 (13%), and minimal change disease in 10 patients (22%). There were 5 (10.4%) cases in group 1 (patients having only mutations of *NPHS1*) and 13 cases (27%) in group 2 (patients having only mutations of *NPHS2*). Thirty cases (62.5%) had neither *NPHS1* nor *NPHS2* mutation (group 3).

**Conclusions:** The genotypic and phenotypic features of INS were demonstrated. We found that INS with podocin mutation has poor prognosis according to exonal distribution. *NPHS1* mutations caused a severe disease but with a more favorable prognosis.

Keywords: Infantile Nephrotic Syndrome, Infants, Genetics

# 1. Background

Infantile nephrotic syndrome (INS) is defined as nephrotic syndrome (NS) that refers to disease being present after the first three months of life up to one year of age (1). About 90% of patients are steroid responsive and approximately one third of the remaining 10% do not respond to corticosteroids; the so called steroid resistant nephrotic syndrome (SRNS). It progresses to end-stage renal disease (ESRD) after about 10 years (2-6). It is well known that inherited structural defects of the glomerular filtration barrier are responsible for a large proportion of SRNS cases. Glomerular filtration barrier structurally consists of glomerular basement membrane (GBM), endothelial cells and podocytes. There are slit diaphragms between foot processes of podocytes. Main structural elements of the slit diaphragm are nephrin, podocin and CD2AP encoded by NPHS1, NPHS2 and CD2AP genes, respectively (7). Indeed, mutations in genes highly expressed in podocytes have been found in two thirds of patients presenting with SRNS in the first year of life (8-10).

## 2. Objectives

In this study, we aimed to analyze genotypic and phenotypic features of both *NPHS1* and *NPHS2* genes in INS and prognostic effect of E117K polymorphism in *NPHS1* gene as E117K change was accepted as polymorphism of *NPHS1* gene.

## 3. Methods

3.1. Study Design, Inclusion and Exclusion Criteria and Data Collection

The study was enrolled retrospectively, with forty eight infantile NS cases diagnosed and followed in our clinic. An

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informed consent was obtained after a complete explanation for the patients' parents. The institutional committee has approved the study. Children with syndromic NS, secondary NS and having irregular follow-up or missing data on their records were excluded from the study. Demographic features, clinical data, genetic analysis and outcomes were extracted from their charts.

The diagnosis of infantile NS was done by the presence of severe proteinuria (> 40 mg/m<sup>2</sup>/hour or urine protein/urine creatinine ratio of > 2 mg/mg), edema, hypoalbuminemia (< 2.5 mg/dL), and hyperlipidemia. A complete response was defined as both clinical healing and disappearance of proteinuria over three consecutive days. Presence of proteinuria below nephrotic range without edema and hypoalbuminemia was defined as partial response. Failure of remission after 4 weeks of prednisone treatment indicates steroid resistance (11). The glomerular filtration rate (GFR) was measured according to guidelines published by K/DOQI (12).

Patients were classified into three groups; group 1: cases having only *NPHS1* mutation; group 2: cases with only *NPHS2* mutation; group 3: cases with any mutation. Patients having E117K polymorphism in *NPHS1* gene were separately compared with each of the 3 groups.

#### 3.2. Mutational Analysis of NPHS1 and NPHS2

Peripheral blood leukocytes were used for genomic DNA extractions from patients and healthy controls by using Purelink Genomic DNA Mini Isolation Kits (Invitrogen, Carlsbad, CA). Thermo Scientific Nanodrop spectrophotometer (Wilmington, USA) was used for quantification extracted DNA purity at 260/280 nm. For quality assessment, 2% agarose gel electrophoresis was used. The direct sequencing of all 29 exons of NPHS1 gene and 8 exons of the NPHS2 gene were made. The direct DNA sequencing reactions were performed from controlled DNA of NPHS1 and NPHS2 genes. Nucleotide comparison of the NPHS1 cDNA sequence (NCBI reference sequence: NM\_004646) and NPHS2 cDNA sequence (NCBI reference sequence: NM\_-014625) were performed using GeneMapper SeaScape Software v3.0. We carried out amino-acid comparisons with NCBI reference sequence: NP\_004637 protein database for NPHS1 gene and NCBI reference sequence: NP\_055440 for NPHS2 gene protein database.

## 3.3. Statistical Analysis

Results were shown as mean  $\pm$  SD. For the comparisons between two continuous variables *t*-test was used. Genetic association between *NPHS1* variants and NS were analyzed by chi-square test or Fisher's exact test. The difference was considered to be significant if P < 0.05. Patients having *NPHS1* mutation were compared with other patients. Statistical analyses were performed using SNPStats software.

## 4. Results

# 4.1. Baseline Characteristics

Forty-eight children with infantile NS (18 girls) were included in the study. The mean age of onset was 8.7  $\pm$  2.3 months and the follow-up time was 8.3 years. Consanguinity was seen in 12.5%. Seven familial and 41 sporadic cases were included in the study. Kidney biopsy was carried out in 45 patients; there were focal segmental glomerulosclerosis in 29 (65%), IgM nephropathy (Ig MN) in 6 (13%), and minimal change disease (MCD) in 10 (22%) (Table 1).

Mutation analysis of *NPHS1* and *NPHS2* was performed in all cases, with a mutation rate of 58.3% (28 out of 48). Different DNA sequence variant analyses were performed. Missense, nonsense, insertions and deletions and splice site mutations resulting in nucleotide changes were expressed either as homozygous or compound heterozygous sequence changes.

Initially, all patients had normal renal function except one (patient 11). Patients were given corticosteroids, cyclosporine, cyclophosphamide, and rituximab. At the end of the follow-up period (8.3 years), there were 32 children with partial (n = 20) and complete response (n = 12) and 16 without any response. Among the 16 non-responsive cases, 6 developed chronic kidney disease (CKD) stages of 2 to 4, and 10 progressed to ESRD (Table 2).

Among the ESRD cases (n = 10) there was only one case with *NPHS1* (V709G) mutation. Half of the ESRD cases had no mutation, though 40% revealed *NPHS2* mutation (P20L, R168H, 467/7 insertion T). The median time for progression to ESRD was  $36.1 \pm 51.9$  months. All 10 ESRD cases were transplanted at a median age of  $68.9 \pm 43.6$  months, with only one rejection due to recurrence of the disease (Table 2). The relationship between histological diagnosis, age of onset, familial or sporadic form, clinical course, and mutational analysis are shown in Table 1.

#### 4.2. Genotype-Phenotype Correlations of Groups

## 4.2.1. Group 1: Cases with Only NPHS1 Mutation (Patients 1 - 5)

There were 5 patients, three of them male, and three familial, with the mean disease onset of 7.19  $\pm$  2.73 months. Patients 1 - 3 were heterozygotes for *V*709*G*, *R*408*Q*, *R*800*C*, and patients 4 - 5 were both heterozygotes for *N*1077S.

Kidney biopsy showed focal segmental glomerulosclerosis in four and immunoglobulin M nephropathy in one patient. There were two cases with chronic kidney disease

Patient	Candan	E/C	Discourse (Mantha)	Tour a form any f 10 - or a man	<b>B</b> i	CVD/Whan2 (Mantha)	Tu(4 (35 4 )	NDUC	NBUCa
Patient	Gender	F/S	Disease Onset (Months)	ireatment/kesponse	вюрѕу	CKD/when? (Months)	IX/Age (Months)	NPHS1	NPH52
Pt. 1. Y.G.	F	F	4	ARB,ACEI/-	FSGS	ESRD/II	+/28	V709G het.	
Pt. 2 Y.E.T.	М	S	11.8	CS/+	FSGS			R408Q het.	•
Pt. 3 A.S.	F	S	8	CS/+	FSGS	•		R800C het.	•
Pt. 4 R.Ç.	М	S	11	CS+	FSGS			N1077S het.	
Pt. 5 K.B.	М	S	11	CS,CyC,CsA,ritux/-,-,-,	IgM	•		N1077S het.	•
Pt. 6 S.A.	F	S	11	CS,CyC,CsA/-,-,+	FSGS	•		•	V180M homo.
Pt. 7 K.Ç.	М	S	7	CS,CyC,CsA/-,+,+	FSGS	Stage 2/180		•	P89T het.
Pt. 8 B.N.	F	F	11.7	CS,CyC,CsA/-,+,+	FSGS	•	•	•	P89T het.
Pt. 9 D.Ç.	М	S	8	CS,CyC,CsA/-,-,-	MCN	ESRD/ 90	+/121	•	467 ins. 7 T homo.
Pt. 10 A.Ö.	F	F	8		FSGS	ESRD/22	+/60	•	P20L/R168H comp het.
Pt. 11 K.Ö.	E	F	6			ESRD 5/11	+/96	•	P20L/R168H comp het.
Pt. 12 A.K.	E	S	5.6		FSGS				P118L homo.
Pt. 13 E.Ü.	E	S	3		MCN				P20L homo.
Pt. 14 A.A.	F	S	12	CS,CsA/-,-	FSGS				P20L/R168H comp het.
Pt.15 Ö.I.	F	S	10.9	CS/CyC/-	FSGS	ESRD/ 25	+/76/reject		P20 L homo.
Pt. 16 C.Ç.	F	S	6	CS,CyC/-,-	FSGS	Stage 4/9		E117K het.	P118L het.
Pt. 17 A.A.	М	S	11	CS,CyC,CsA /-,-,+	MCN			E117K het.	V64E/K289X het.
Pt. 18 İ.G.	М	S	9	CS,CyC,CsA/-,+,+	FSGS			E117K het.	R229Q het.
Pt. 19 D.P.	F	S	6	CS,CyC,CsA,CQ10/-,-,+,+	FSGS	Stage 2/13			
Pt. 20 S.Ç.	М	S	12	CS,CyC,CsA,CQ10/-,-,+,+	FSGS				
Pt. 21 M.B.	М	S	9	CS,CyC/-,-	FSGS	ESRD 5/12	+/20		
Pt. 22 S.K.	М	S	п	CS,CsA,CQ10/-,+,+	FSGS	Stage 2/17			
Pt. 23 S.A.	М	S			FSGS				
Pt. 24 Y.D.	М	F	6	ARB/ACEI	FSGS	ESRD/18	+/26	•	
Pt. 25 S.G.	F	S	12	CS,CyC/+,+	IgM				
Pt. 26 A.A.	М	S	10.8	CS,CyC,CsA/-,+,+	IgM				
Pt. 27 H.Ç.	М	S	12	CS/+	MCN				
Pt. 28 F.A.	F	S	11	CS/+					
Pt. 29 B.Ç.	М	S	10.5	CS,CyA/-,+	FSGS				
Pt. 30 E.A.	М	S	11	CS,CyC,CsA/-,+,+	FSGS				
Pt. 31 H.Ç.	F	S	12	CS/+	MCN				
Pt. 32 B.N.	F	F	4	CS,CyC,CsA/-,+,+	FSGS	ESRD/20	+/60		
Pt. 33 C.D.	F	S	11		FSGS				
Pt. 34 M.B.	М	S	11.3	CS+	MCN				
Pt. 35 M.E.	М	F	11	CS/+	FSGS				
Pt. 36 A.K.	F	S	12	CS,CyC,CsA/-,-,+		ESRD/21	+/100	•	
Pt. 37 M.P.	М	S	11		FSGS	ESRD/35	+/122		
Pt. 38 O.D.	М	S	10		IgM				
Pt. 39 M.A.	М	S	9	CS,CyC,CsA,ritux/-,-,?,-,	FSGS	Stage 3/144		E117K het.	•
Pt. 40 A.H.	М	S	7	CS/+	MCN			E117K het.	•
Pt. 41 R.K.	М	S	6.1	CS/+	FSGS	•		E117K het.	•
Pt. 42 Ö.Y.	М	S	7.2	CS,CyC/+-,+	MCN			E117K het.	
Pt. 43 G.Ç.	F	S	5.7	CS,CsA/-,+	IgM		-	E117K het.	
Pt. 44 D.A.	F	S	3	CS,CsA/-,+	MCN			E117K het.	
Pt. 45 E.Y.	М	s	5.1	CS,CyC/-,+	MCN		-	E117K het.	
Pt. 46 A.B.	М	S	4	ARB/ACEI	FSGS	Stage 4/10		E117K het.	•
Pt. 47 A.Y.	М	s	9	CS,CyC/+,+	IgM	•		E117K het.	•
Pt. 48 K.A.	F	S	6	CS,CyC/+,+	FSGS			E117K het.	

Table 1. The Relationship Between Histological Diagnosis, Age of Onset, Familial or Sporadic Forms, Clinical Course, and Results of Mutational Analysis of the Individual Patients

Abbreviations: ARB, angiotensin receptor blocker; CKD, chronic kidney disease; CS, corticosteroid; CyA, cyclosporine A; CyC, cyclophosphamide; ESRD, end stage renal disease; FSGS, focal segmental glomerulosclerosis; GFR, glomerular filtration rate; het, heterozygous; homo, homozygous; IgMN, IgM nephropathy; MCD, minimal change disease; NS, nephrotic syndrome; PCR, polymerase chain reaction; Tx, transplantation.

(CKD); one progressed to stage 4 CKD in 10 months and had both E117K and N1077S. The second one, who progressed to ESRD within 11 months of disease onset and was transplanted with a well-functioning graft from her mother for four years, had V709G mutation on the 16th exon (patient 1) (Table 1). Thus, ESRD ratio was 20% (n = 1) in the group.

4.2.2. Group 2: Cases with Only NPHS2 Mutation (Patients 6 - 18)

There were 13 patients, eight of them male, and three familial cases with the mean disease onset of 8.3  $\pm$  3.0

Variables	Patients with NPHS1	Patients with NPHS2 M	utation (Group 2)	Patients Without Any Mutation (Group 3)				
variabits	Mutation (Group 1)	No E117K	With E117K	With	No E117K			
Patient number	5	10	3	6	4	20		
Gender, male/female	3/2	5/5	2/1	4/2	2/2	13/7		
Age of disease onset, month	$7.9\pm3.9$	$8.32\pm3.01$	$8.6\pm2.5$	$6.2\pm1.8$	$6.3\pm0.8$	$9.9\pm2.4$		
Number of relapses	4/patient	6.2/patient	4.3/patient	6.1/patient	3.8/patient	7.1/patient		
Response to treatment, complete/partial/no	3 - 2	-/3/7	-/2/1	2/3/1	-/3/1	7/9/4		
Proteinuria, mg/m²/h/ Cr. Cle. mL/min/m²	$51.7 \pm 14.3   67.1 \pm 29.0$	$67.2 \pm 21/32.3 \pm 16$	$37.1 \pm 9.7 / 35.8 \\\pm 11.2$	$\begin{array}{c} 12.3 \pm 6.1 /  86 \pm \\ 9.2 \end{array}$	$43.2 \pm 11.9 / 87.1 \\ \pm 12.1$	$\begin{array}{c} 128.3 \pm 22.9 \\ 22.3 \pm 27.1 \end{array}$		
CKD	-	1	1	1	1	2		
ESRD	1	4				5		

## Table 2. Demographic, Clinical, and Genetic Features of Groups

Abbreviations: CKD, chronic kidney disease; ESRD, end stage renal disease.

months in this group. Five cases (pt. 6, 9, 12, 13, 15) were homozygous, two heterozygous (pt. 7, 8), and three compound heterozygous (pt. 10, 11, 14). The most common mutation was P20L found in five cases; 3 of 5 had compound mutation with *R168H* (pt. 10, 11, 14). Pt. number 16, carrying both *E117K* and *P118L*, went to stage 4 CKD within 9 months, whereas the others were partially responsive to treatment (Table 1).

As patient number 11 was admitted in ESRD status, no biopsy was taken. Among the remaining nine cases, nine samples had FSGS. Four of them progressed to ESRD at a mean age of 37  $\pm$  35.8 months. Patient number 10 and patient number 11 were siblings, having P20L and R168H compound heterozygous mutations, progressed to ESRD. All three patients had *E117K* single nucleotide polymorphism of NPHS1, accompanied with various NPHS2 mutations. Two other patients with ESRD were patients 9 and 15, with mutations of 467/7 insertion homozygous and P20L homozygous, respectively. All of the ESRD cases were transplanted and followed with a well-functioning graft, except for one rejection at the 76th month due to recurrence of the disease. There were only three patients that had partial response, having mutations of V180M homozygous, P89T heterozygous, and TT/AA 951T > C. A case with *P118L* homozygous, another case with P20L, and a third case from the compound mutated cases (P20L and R168H) were also not responsive to treatment (Table 1). Thus, ESRD ratio was 30.8% (n = 4) in the group.

#### 4.2.3. Group 3: Cases Without any Mutation (19 - 48)

There were 30 patients, with a male: female ratio of 19:11, with three familial cases, and the mean onset of dis-

ease was 9.9  $\pm$  2.4 months.

There were cases having renal biopsy, FSGS, IgM nephropathy, and MCD. Two cases progressed to stage 2 CKD within 13 and 17 months, whereas five patients progressed to ESRD and were transplanted with well-functioning grafts.

Pathogenic mutations in *NPHS1* were found in 5/48 (10.4%) of cases. Also, the ratio of *NPHS2*-detected patients was 13/48. There were four *NPHS2* mutated cases that progressed to ESRD (4/10; 40%). Whereas only one out of five *NPHS1* mutated patients progressed to ESRD, this equals to 20% of cases. All 10 cases with *E117K* were sporadic, resistant to treatment, and had earlier onset (Tables 1 and 2).Thus, ESRD ratio was 16.7% (n = 5) in the group.

#### 5. Discussion

NPHS1 gene mutations are the cause of the Finnish type nephrotic syndrome. In Finland, two mutations, Fin major and Fin minor were seen in 78% and 16% of the cases respectively (13). NPHS1 gene mutations account for 39% - 55% cases of childhood NS in European, North American, and Turkish societies and nearly 40% of CNS cases (14, 15). Nephrin encoding of mutations of NPHS1 are responsible for the most of congenital NS and result in infantile and childhood steroid-resistant nephrotic syndrome (SRNS) (16-19). Also, there are some infantile NS cases having both nephrin and podocin mutations, causing triallelic abnormality (homozygous mutations in one gene and heterozygous mutations in the other) (20). Because of INS genetic heterogeneity there is no clear genotypephenotype correlations. A few patients with typical INS cases were found to lack NPHS1 mutations but were found to have recessive NPHS2 mutations (20). Philippe et al. reported NPHS1 mutations in 7% - 14% of the patients with SRNS at least 3 months after birth (age at onset of NS in mutated patients 0.5 - 8 years (mean 3 years)) (19). Santin et al. also supported the former with same mutation ratio with the age of onset of 0.7 - 27 years (mean 8 years) (21). Lahdenkari et al. pointed to the immunogenic stimuli caused by hypomorphic mutation in cases with NPHS1 variants (22). Also, infections trigger the attacks, Kitamura et al. reported two siblings bearing nephrin mutations with spontaneous partial remissions, but repeated relapses concurrently with respiratory infections. One of them presented NS at birth and the other at 10 months of age. Both were compound heterozygous for the p.C265R and p.V822M mutations. The p.C625R mutant protein was predominantly trapped within the ER, while the p.V822M protein reached the plasma membrane, explaining the milder phenotype (23). Another factor is environmental influences that play a role in phenotype variability. Santin et al. followed an adult case without any renal impairment during 2 years, diagnosed as FSGS at 27 years of age, with two NPHS1 mutations (p.R827X and p.R976S) (21). On the other hand, Philippe et al. identified one patient with infantile-onset NS with the same two mutations (19).

NPHS2 gene mutations account for an autosomal recessive steroid resistant nephrotic syndrome (SRNS) with early disease onset and focal segmental glomerulosclerosis (FSGS) (24). Infantile nephrotic syndrome cases can be derived from either isolated nephrin or podocin mutations, or both (25). There are other genes that cause INS as LAMB2 or WT. In this study, we evaluated both NPHS1 and NPHS2 mutations in the infantile group to determine nephrin and podocin mutational profile and outcome. A recent study reported 57% rate of mutations in INS, with 14% NPHS1, 29% NPHS2, and 14% WT1 (26). Our total mutation rate was 37.5%. Mbarek et al. reported ten different pathological mutations including NPHS1 and NPHS2 in 24 Tunisian children, and only two infantile cases without any NPHS1 or NPHS2 mutation (27). In our cohort, cases without mutations made up 62.5% (30/48) of our population, which might be explained so that we have only analyzed NPHS1, NPHS2 genes of INS.

Santin et al. analyzed podocyte genes in SRNS, with patient ages ranging from congenital to adult onset (21) Infantile group had various podocyte mutations with a rate of 57%. Also, *NPHS1* mutation rate was found to be 14% and *NPHS2* rate was 29% only in the infantile onsets. The range of *NPHS1* gene mutations prevalence was 39 to 55% and the *NPHS2* gene mutations was 10 to 28% in European and American populations (11, 12, 25, 28). In contrast to these, Abid et al. reported that the *NPHS1* gene mutations were approximately 20% and *NPHS2* gene mutations 5.5% of the patients with early onset NS (29). The prevalence of *NPHS1* and *NPHS2* were low in studies from Japan and China (30, 31). Our study group consisted of only infantile cases. We found the mutation rate to be 32%, nearly equal to the study reported by Santin et al. (21). Also, our *NPHS2* rate was similar (20.8%). Our cohort was more relevant to European and American than Asian populations (11, 12, 19, 25, 30, 31). It shows geographic and ethnic genetic diversity of NS in the world.

Nowadays, papers have reported general nonresponsiveness to intensive immunosuppressive therapy regimens, and many studies observed the low recurrence rate of NS after transplantation (22, 29, 32, 33). Similarly, our cohort showed a 16% rate of intensive immunosuppressive drug response and only one recurrence of disease after transplantation.

Tryggvason et al. reported the faster progression to ESRD of *NPHS1* mutations compared to *NPHS2* (34). We found a worse prognosis in the *NPHS2* positive group; the difference is due to the population group, as the majority of *NPHS1* positive cases in their study were congenital nephrotic syndrome cases, not infantile ones (34). *NPHS1* gene mutations progressed rapidly to ESRD within one to three years of age in children in some studies (1, 33, 35). In addition, Abid et al. reported that *NPHS1* gene mutations carriage in children result in preserving renal function up to 2.5 years of age (29). We had only one patient who progressed to ESRD within 11 months of disease onset.

Koziell et al. reported digenic inheritance of *NPHS1* and *NPHS2* genes (31). In the study of a cohort from Pakistan, they observed a patient with both heterozygous R408Q *NPHS1* gene mutation and a heterozygous P321S mutation in the *NPHS2* gene together (30). We had no digenic cases, but three of our cases had heterozygous *E117K* polymorphism in *NPHS1* and P118L in *NPHS2* in one, K289X in another, and R229Q in the third case.

Today, there are more than 173 various mutations of *NPHS1* reported in the Human Gene Mutation Database. The clinical course associated with *NPHS1* mutations is not restricted to classical CNS. *NPHS1* mutations were causing a mild disease in adulthood onset NS with the FSGS histology (28). We can explain this by underlying mutations; predominately missense mutations result in minor protein modifications.

Abid et al. screened mutations of 145 patients, including 36 early-onset NS cases (CNS cases included). Mutations in the *NPHS1* gene accounted for approximately 20% of cases with early-onset NS. They showed a heterozygous mutation, R408Q, in three patients with childhood onset (29). Lenkkeri et al. reported this mutation as a compound heterozygous condition in CNS cases (15). In our cohort, we found one patient with R408Q with disease onset of 11.8 months. Other *NPHS1* mutations of our cohort were N1077S, V709G, and R800C.

Here, we also would like to discuss the phenotypic features of patients with the E117K genotype, which has been accepted as a polymorphism since Lenkkeri et al. reported it as a single nucleotide polymorphism (15). As our cohort did not deal with CNS cases, we questioned whether it is also a polymorphism in the infantile group or if it affects the protein as a hot mutation. E117K was found in 6 homozygous and 21 heterozygous conditions in the study by Abid et al. (29). However, this was a common variant, as it was found in normal individuals (1). In our cohort, E117K cases showed a clinical course that had no statistical differences compared with other nephrin mutations, whereas, E117K had statistically significant differences compared with non-mutated cases (Table 3). Also, we had previously reported patient number 26 in a case report, when she was in stage 2 CKD, and emphasized that, if E117K change in nephrin diverges the podocyte signaling pathways and causes P118L mutation of NPHS2, it behaves different and suggested that it might be called a genetic modifier in future (36). Pettersson-Fernholm et al. examined diabetic patients with nephrin polymorphism (37) and found that the onset of diabetes in patients with E117K polymorphism K genotype occurred later compared to patients with the wild genotype. Downregulation of the nephrin gene was seen in experimental nephrosis of rats. Animal models showed that the expression levels of nephrin-specific mRNA were associated with early changes of diabetic nephropathy (35). In the literature, it was reported that any changes in amino acid sequence might affect the nephrin protein confirmation. Even if the polymorphisms have unknown functional implications, they may have a role in proteinuria via influence on the slit diaphragm permeability (1, 36).

Nephrin is a transmembrane adhesion protein (10) and directly participates in slit diaphragm structure by its ability to homo- or hetero-dimerization (1). *NPHS1* missense mutations result in abnormal endoplasmic reticulum nephrin retention, and failure of trafficking out to the cell surface (20). Therefore, nephrin dysfunction may explain severe and early onset phenotypes resulted from mostly truncated and missense *NPHS1* mutations. Beside these, extracellular Ig domains 2, 4 and 7 have clusters of mutations. More than 50% of missense mutations are ex-

tracellular, and 66% of them are in Ig domains and this leads to the hotspot mutations. The structure of nephrin is highly flexible, most mutations can affect it. E117K polymorphism is a missense mutation of nephrin and placed in immunoglobulin motif and the transmembrane domain of the polypeptide chain (22). Koziell et al. showed that missense *NPHS1* mutation decreases nephrin expression in podocyte cell cultures (31).

Pettersson-Fernholm et al. reported that all the polymorphisms of E117K, N1077S, and R408Q were changing the amino acid (37). Any change in amino acid sequence such as G > A substitution in E117K is called a mutation, whether it leads to the protein malfunctioning (hot/disease-causing mutation) or not (polymorphism). In our patients having E117K polymorphism, there were statistically significant differences of disease onset when compared with other mutations. E117K had a similar phenotype to other known mutations of nephrin and podocin and differed from the non-mutated group (38).

## 5.1. Conclusions

In the current study, the genotypic and phenotypic features of infantile NS were displayed. *NPHS1* mutations cause severe and early disease type but with better prognosis. Additionally,*E117K* polymorphism of *NPHS1* showed a similar course as other *NPHS1* and *NPHS2* mutations, with the only difference being that *E117K* polymorphism manifested relatively earlier onset. Also, among NPHS1 mutations, *E117K* had been reported as a polymorphism, but we showed our contrary findings and ask: Is it still a polymorphism?

#### Footnotes

Authors' Contribution: Study concept and design: Ebru Yılmaz, Nida Dinçel, İpek Kaplan Bulut, Afig Berdeli and Sevgi Mir; analysis and interpretation of data: Ebru Yılmaz and Nida Dinçel; drafting of the manuscript: Ebru Yılmaz and Nida Dinçel; critical revision of the manuscript for important intellectual content: Ebru Yılmaz; statistical analysis: Nida Dinçel.

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able 3. Comparison of Patients with E117K Polymorphism with 3 Groups											
	Patients with E117K (N:10)										
Group	Disease Onset (Months)			Familial / Sporadic		Histology (FSGS/Others)		ESRD		CKD	
	$\mathbf{SD}^2$	t	Р	$\chi^2$	Р	$\chi^2$	Р	$\chi^2$	Р	$\chi^2$	Р
Group 1, n = 5	5.72	0.91	> 0.1	3.65	> 0.05	3.36	> 0.05	3.64	> 0.05	0.75	> 0.2
Group 2, n = 13	6.3	1.87	< 0.05	3.52	> 0.05	1.82	> 0.1	5	< 0.05	0.39	> 0.2
Group 3, n = 30	6.41	3.47	< 0.05	3.34	> 0.1	1.68	> 0.1	3.34	>0.05	0.68	> 0.2

Abbreviations: CKD, chronic kidney disease; ESRD, end stage renal disease; FSGS, focal segmental glomerulosclerosis.

# References

- Hinkes B, Vlangos C, Heeringa S, Mucha B, Gbadegesin R, Liu J, et al. Specific podocin mutations correlate with age of onset in steroidresistant nephrotic syndrome. *J Am Soc Nephrol.* 2008;19(2):365-71. doi: 10.1681/ASN.2007040452. [PubMed: 18216321]. [PubMed Central: PMC2396749].
- Sahay M, Gowrishankar S, Narayen GA. Nephrotic syndrome in the first year of life. J Acad Med Sci. 2012;2(1):22. doi: 10.4103/2249-4855.104011.
- 3. [No Authors Listed]. The primary nephrotic syndrome in children. Identification of patients with minimal change nephrotic syndrome from initial response to prednisone. A report of the International Study of Kidney Disease in Children. *J Pediatr.* 1981;**98**(4):561-4. [PubMed: 7205481].
- Cattran DC, Rao P. Long-term outcome in children and adults with classic focal segmental glomerulosclerosis. *Am J Kidney Dis.* 1998;**32**(1):72-9. [PubMed: 9669427].
- Mendoza SA, Reznik VM, Griswold WR, Krensky AM, Yorgin PD, Tune BM. Treatment of steroid-resistant focal segmental glomerulosclerosis with pulse methylprednisolone and alkylating agents. *Pediatr Nephrol.* 1990;4(4):303–7. [PubMed: 2206894].
- Mekahli D, Liutkus A, Ranchin B, Yu A, Bessenay L, Girardin E, et al. Long-term outcome of idiopathic steroid-resistant nephrotic syndrome: A multicenter study. *Pediatr Nephrol.* 2009;24(8):1525–32. doi: 10.1007/s00467-009-1138-5. [PubMed: 19280229].
- Machuca E, Benoit G, Antignac C. Genetics of nephrotic syndrome: Connecting molecular genetics to podocyte physiology. *Hum Mol Genet.* 2009;18(R2):R185–94. doi: 10.1093/hmg/ddp328. [PubMed: 19808795].
- Hinkes BG, Mucha B, Vlangos CN, Gbadegesin R, Liu J, Hasselbacher K, et al. Nephrotic syndrome in the first year of life: Two thirds of cases are caused by mutations in 4 genes (NPHS1, NPHS2, WT1, and LAMB2). *Pediatrics*. 2007;119(4):e907-19. doi: 10.1542/peds.2006-2164. [PubMed: 17371932].
- Gbadegesin RA, Winn MP, Smoyer WE. Genetic testing in nephrotic syndrome-challenges and opportunities. *Nat Rev Nephrol.* 2013;9(3):179–84. doi: 10.1038/nrneph.2012.286. [PubMed: 23321566]. [PubMed Central: PMC3702380].
- Niaudet P. Genetic forms of nephrotic syndrome. *Pediatr Nephrol.* 2004;**19**(12):1313-8. doi: 10.1007/s00467-004-1676-9. [PubMed: 15503167].
- [No Authors Listed]. Short versus standard prednisone therapy for initial treatment of idiopathic nephrotic syndrome in children. Arbeitsgemeinschaft fur Padiatrische Nephrologie. *Lancet.* 1988;1(8582):380– 3. [PubMed: 2893190].
- National Kidney Foundation. K/DOQI clinical practice guidelines for chronic kidney disease: Evaluation, classification, and stratification. *Am J Kidney Dis.* 2002;**39**(2 Suppl 1):SI-266. [PubMed: 11904577].
- 13. Kestila M, Lenkkeri U, Mannikko M, Lamerdin J, McCready P, Putaala

H, et al. Positionally cloned gene for a novel glomerular proteinnephrin-is mutated in congenital nephrotic syndrome. *Mol Cell*. 1998;1(4):575-82. [PubMed: 9660941].

- Heeringa SF, Vlangos CN, Chernin G, Hinkes B, Gbadegesin R, Liu J, et al. Thirteen novel NPHS1 mutations in a large cohort of children with congenital nephrotic syndrome. *Nephrol Dial Transplant*. 2008;**23**(11):3527–33. doi: 10.1093/ndt/gfn271. [PubMed: 18503012]. [PubMed Central: PMC2720813].
- Lenkkeri U, Mannikko M, McCready P, Lamerdin J, Gribouval O, Niaudet PM, et al. Structure of the gene for congenital nephrotic syndrome of the Finnish type (NPHS1) and characterization of mutations. *Am J Hum Genet*. 1999;**64**(1):51–61. [PubMed: 9915943]. [PubMed Central: PMC1377702].
- Benoit G, Machuca E, Heidet L, Antignac C. Hereditary kidney diseases: Highlighting the importance of classical Mendelian phenotypes. *Ann N Y Acad Sci.* 2010;**1214**:83–98. doi: 10.1111/j.1749-6632.2010.05817.x. [PubMed: 20969579].
- [No authors listed]. Nephrotic syndrome in children: Prediction of histopathology from clinical and laboratory characteristics at time of diagnosis. A report of the International Study of Kidney Disease in Children. *Kidney Int*. 1978;13(2):159–65. [PubMed: 713276].
- Savin VJ, Sharma R, Sharma M, McCarthy ET, Swan SK, Ellis E, et al. Circulating factor associated with increased glomerular permeability to albumin in recurrent focal segmental glomerulosclerosis. *N Engl J Med.* 1996;**334**(14):878–83. doi: 10.1056/NEJM199604043341402. [PubMed: 8596570].
- Philippe A, Nevo F, Esquivel EL, Reklaityte D, Gribouval O, Tete MJ, et al. Nephrin mutations can cause childhood-onset steroidresistant nephrotic syndrome. J Am Soc Nephrol. 2008;19(10):1871–8. doi: 10.1681/ASN.2008010059. [PubMed: 18614772]. [PubMed Central: PMC2551572].
- McKinney PA, Feltbower RG, Brocklebank JT, Fitzpatrick MM. Time trends and ethnic patterns of childhood nephrotic syndrome in Yorkshire, UK. *Pediatr Nephrol.* 2001;16(12):1040-4. doi: 10.1007/s004670100021. [PubMed: 11793096].
- Santin S, Garcia-Maset R, Ruiz P, Gimenez I, Zamora I, Pena A, et al. Nephrin mutations cause childhood- and adult-onset focal segmental glomerulosclerosis. *Kidney Int.* 2009;**76**(12):1268–76. doi: 10.1038/ki.2009.381. [PubMed: 19812541].
- Lahdenkari AT, Kestila M, Holmberg C, Koskimies O, Jalanko H. Nephrin gene (NPHS1) in patients with minimal change nephrotic syndrome (MCNS). *Kidney Int.* 2004;65(5):1856–63. doi: 10.1111/j.1523-1755.2004.00583.x. [PubMed: 15086927].
- Kitamura A, Tsukaguchi H, Hiramoto R, Shono A, Doi T, Kagami S, et al. A familial childhood-onset relapsing nephrotic syndrome. *Kidney Int.* 2007;**71**(9):946–51. doi: 10.1038/sj.ki.5002110. [PubMed: 17290294].
- Boute N, Gribouval O, Roselli S, Benessy F, Lee H, Fuchshuber A, et al. NPHS2, encoding the glomerular protein podocin, is mutated in autosomal recessive steroid-resistant nephrotic syndrome. *Nat Genet*. 2000;**24**(4):349–54. doi: 10.1038/74166. [PubMed: 10742096].

- Wartiovaara J, Ofverstedt LG, Khoshnoodi J, Zhang J, Makela E, Sandin S, et al. Nephrin strands contribute to a porous slit diaphragm scaffold as revealed by electron tomography. *J Clin Invest.* 2004;**114**(10):1475-83. doi: 10.1172/JCl22562. [PubMed: 15545998]. [PubMed Central: PMC525744].
- Tsukaguchi H, Sudhakar A, Le TC, Nguyen T, Yao J, Schwimmer JA, et al. NPHS2 mutations in late-onset focal segmental glomerulosclerosis: R229Q is a common disease-associated allele. *J Clin Invest.* 2002;**110**(11):1659–66. doi: 10.1172/JCl16242. [PubMed: 12464671]. [PubMed Central: PMC151634].
- Mbarek IB, Abroug S, Omezzine A, Pawtowski A, Gubler MC, Bouslama A, et al. Novel mutations in steroid-resistant nephrotic syndrome diagnosed in Tunisian children. *Pediatr Nephrol.* 2011;26(2):241–9. doi: 10.1007/s00467-010-1694-8. [PubMed: 21125408].
- Brown EJ, Schlondorff JS, Becker DJ, Tsukaguchi H, Tonna SJ, Uscinski AL, et al. Mutations in the formin gene INF2 cause focal segmental glomerulosclerosis. *Nat Genet*. 2010;**42**(1):72–6. doi: 10.1038/ng.505. [PubMed: 20023659]. [PubMed Central: PMC2980844].
- Abid A, Khaliq S, Shahid S, Lanewala A, Mubarak M, Hashmi S, et al. A spectrum of novel NPHS1 and NPHS2 gene mutations in pediatric nephrotic syndrome patients from Pakistan. *Gene.* 2012;**502**(2):133–7. doi:10.1016/j.gene.2012.04.063. [PubMed: 22565185].
- Liu G, Kaw B, Kurfis J, Rahmanuddin S, Kanwar YS, Chugh SS. Neph1 and nephrin interaction in the slit diaphragm is an important determinant of glomerular permeability. *J Clin Invest*. 2003;**112**(2):209– 21. doi: 10.1172/JCI18242. [PubMed: 12865409]. [PubMed Central: PMC164293].
- Koziell A, Grech V, Hussain S, Lee G, Lenkkeri U, Tryggvason K, et al. Genotype/phenotype correlations of NPHS1 and NPHS2 mutations in nephrotic syndrome advocate a functional inter-relationship in

glomerular filtration. *Hum Mol Genet*. 2002;**11**(4):379–88. [PubMed: 11854170].

- Schoeb DS, Chernin G, Heeringa SF, Matejas V, Held S, Vega-Warner V, et al. Nineteen novel NPHSI mutations in a worldwide cohort of patients with congenital nephrotic syndrome (CNS). *Nephrol Dial Transplant*. 2010;25(9):2970–6. doi: 10.1093/ndt/gfq088. [PubMed: 20172850]. [PubMed Central: PMC2948833].
- 33. Mao J, Zhang Y, Du L, Dai Y, Gu W, Liu A, et al. NPHS1 and NPHS2 gene mutations in Chinese children with sporadic nephrotic syndrome. *Pediatr Res.* 2007;61(1):117-22. doi: 10.1203/01.pdr.0000250041.19306.3d. [PubMed: 17211152].
- Tryggvason K, Patrakka J, Wartiovaara J. Hereditary proteinuria syndromes and mechanisms of proteinuria. N Engl J Med. 2006;354(13):1387-401. doi: 10.1056/NEJMra052131. [PubMed: 16571882].
- Sako M, Nakanishi K, Obana M, Yata N, Hoshii S, Takahashi S, et al. Analysis of NPHS1, NPHS2, ACTN4, and WT1 in Japanese patients with congenital nephrotic syndrome. *Kidney Int.* 2005;67(4):1248–55. doi: 10.1111/j.1523-1755.2005.00202.x. [PubMed: 15780077].
- Dincel N, Mir S, Berdeli A, Bulut IK, Sozeri B. Does NPHS1 polymorphism modulate P118l mutation in NPHS2? *Saudi J Kidney Dis Transpl.* 2013;24(6):1210-3. doi: 10.4103/1319-2442.121300. [PubMed: 24231487].
- Pettersson-Fernholm K, Forsblom C, Perola M, Groop PH, FinnDiane Study G. Polymorphisms in the nephrin gene and diabetic nephropathy in type 1 diabetic patients. *Kidney Int.* 2003;63(4):1205–10. doi: 10.1046/j.1523-1755.2003.00855.x. [PubMed: 12631336].
- Berdeli A, Mir S, Yavascan O, Serdaroglu E, Bak M, Aksu N, et al. PNPHS2 (podicin) mutations in Turkish children with idiopathic nephrotic syndrome. *Pediatr Nephrol*. 2007;22(12):2031-40. doi: 10.1007/s00467-007-0595-y. [PubMed: 17899208].