

## Synthesis of Some Spiro Indeno[1,2-*b*]pyrido[2,3-*d*]Pyrimidine-5,3'-Indolines as New Urease Inhibitors

Ghodsi Mohammadi Ziarani<sup>a,b\*</sup>, Shima Asadi<sup>a</sup>, Alireza Badieci<sup>c</sup>,  
Amirhossein Sharifi<sup>d</sup> and Massoud Amanlou<sup>d\*</sup>.

<sup>a</sup>Department of Chemistry, Alzahra University, Vanak Square, Tehran, Iran. <sup>b</sup>National Laboratory of Pharmaceutical Research, Alzahra University, Tehran, Iran. <sup>c</sup>School of Chemistry, College of Science, University of Tehran, Tehran, Iran. <sup>d</sup>Drug Design and Development Research Centre and Department of Medicinal Chemistry, Faculty of Pharmacy, Tehran University of Medical Sciences, Tehran, Iran.

---

### Abstract

New series of spiro indeno[1,2-*b*]pyrido[2,3-*d*]pyrimidine-5,3'-indolines as new urease inhibitors were synthesized by the catalytic procedure in high yield and short reaction time. In this method, biacidic carbon was prepared as a novel heterogeneous acid and was subsequently used as an efficient catalyst. The inhibitory activities of synthesized compounds were tested against Jack bean urease using Berthelot colorimetric assay and docking simulation using AutoDock 4.2. The compound 4a with  $IC_{50} = 1.94 \mu M$  has the most inhibitor activity in this study. Other derivatives such as 4b, 4d, 4e and 7a were found to be more potent urease inhibitors than the standard inhibitor hydroxyurea, yielding  $IC_{50}$  values of 4.35, 5.557, 7.44, 2.81 and 14.46  $\mu M$ , respectively ( $IC_{50}$  of hydroxyurea = 100  $\mu M$ ).

**Keywords:** Urease inhibitor; Spiro indenopyridopyrimidine; Multicomponent reaction (MCR); Autodock; Biacidic nano catalyst.

---

### Introduction

Cancer is a leading cause of death worldwide, accounting for 7.6 million deaths (around 13% of all deaths) in 2008. Deaths from cancer worldwide are projected to continue rising, with an estimated 13.1 million deaths in 2030. Cancer causes 1 in 8 deaths worldwide and is rapidly becoming a global pandemic. According to the International Agency for Research on Cancer, there were 12.7 million new cancer cases in 2008. If rates don't change, the global cancer burden is expected to nearly double to 21.4 million cases and 13.5 million deaths by 2030.

Urease is an enzyme which produces numerous amounts of ammonia in the *Helicobacter pylori* by catalysis the hydrolyzing of urea into carbon dioxide and ammonia. The produced ammonia makes the growth and survival of bacteria in highly acidic media of gastric wall, by increasing the pH of environment (1). Hence, urease activity plays an important role in gastric diseases, formation of kidney stones (2) and development of urolithiasis, pyelonephritis and hepatic encephalopathy (3).

Discovering a new agent for inhibition of urease and potent anti-ulcer drugs are of special demand because *H. pylori* has become resistant to many antibiotics (4).

Pyridopyrimidine derivatives have received ample attention in pharmaceutical and

---

\* Corresponding author:

E-mail: gmziarani@hotmail.com and amanlou@tums.ac.ir

agrochemical sectors (5) due to their prominent biological applications such as antibacterial (6), antiallergic (7), CNS stimulants (8), inhibitors of adenosine kinase enzyme (9) or dihydrofolate reductase inhibitors (10). Preparation of compounds with promising drug like activity without tedious and time consuming purification is in great interest.

Recently using new catalysts opened exciting opportunities in the field of synthesizing medicinal compounds or green chemistry by minimizing waste, cost, and time (11). The catalytic systems offer the opportunity of synthesizing molecules via novel routes, which may be difficult or impossible with traditional methods and also increase the yield of the resultant compounds.

As a part of our program to develop catalytic and environmentally friendly methods in the synthesis of biologically active heterocycles (12, 13), we explored novel synthetic strategies for the synthesis of pyridopyrimidine derivatives (14) as urease inhibitor by using biacidic carbon catalyst.

### Experimental

GC-Mass analysis was performed on a GC-Mass model: 5973 network mass selective detector, GC 6890 Agilent. IR spectra were recorded from KBr disk using a FT-IR Bruker Tensor 27 instrument. Melting points were measured using the capillary tube method with an electro thermal 9200 apparatus. The <sup>1</sup>H NMR (400 MHz) was run on a Bruker DPX, 250 MHz.

#### Synthesis of the catalyst

The biacidic carbon catalyst is prepared as the following procedure (15). The mixture of glucose (10 g), citric acid (5 g), hydroxyethylsulfonic acid (3 g), and deionized water (80 mL) was placed in a 100 mL Teflon-lined stainless steel autoclave, which was heated in an oven at 180 °C for 4 h. The resulting products were filtered, washed with water and methanol, and dried in a vacuum oven at 100 °C for 4 h to give biacidic carbonaceous material (2.3 g) as a black powder.

#### General procedure for preparation of Spiro pyridopyrimidine-indolines 4a-e

A mixture of 1,3-indandione (1 mmol),

6-aminouracil (1 mmol), isatin derivatives (1 mmol) and catalytic amount of biacidic carbon (20 mg) in ethanol (5 mL) was refluxed for appropriate time (Table 2). After the completion of reaction as indicated by TLC, the catalyst was recovered by filtering and washing with acetone, and then dried in an oven at 80 °C for about 1 h followed by crystallization using ethanol and water.

#### 5>-Chloro-1H-spiro[indeno[1,2-b] pyrido [2,3-d]pyrimidine-5,3>-indoline]-2,2>,4,6 (3>H,10>H)-tetraone (4d)

Orange powder, Mp 315-319 °C, IR (KBr) ( $\nu_{\max}$  /cm<sup>-1</sup>): 3549, 2800, 1720, 1701, 1666, 1617. MS, m/z: 418 (M<sup>+</sup>), 341, 293, 271, 255, 243, 229, 195, 161, 75, 51. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta_{\text{H}}$  (ppm) 6.77-7.96 (7H, m, H-Ar), 10.55-10.58 (3H, s, 3NH), 10.99 (1H, s, NH).

#### 8>,8>-Dimethyl-8>,9>-dihydro-1>H-spiro [indoline-3,5>-pyrimido[4,5-b] quinoline]-2,2>,4>, 6>(3>H,7>H,10>H)-tetraone (7a)

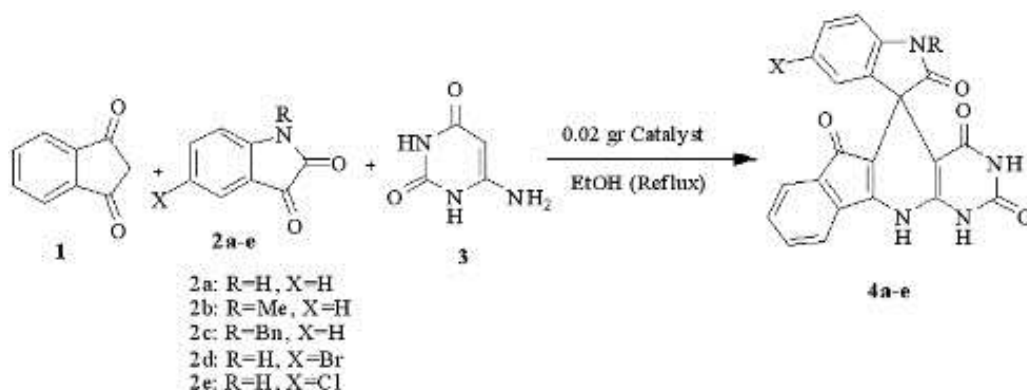
Pale orange powder, Mp 310-312 °C, IR (KBr) ( $\nu_{\max}$  /cm<sup>-1</sup>): 3402, 3126, 2955, 1720, 1673, 1613, 1400. MS, m/z: 378 (M<sup>+</sup>), 320, 308, 307, 292. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta_{\text{H}}$  (ppm) 1.06 (6H, 2CH<sub>3</sub>), 6.72-7.44 (4H, m, H-Ar), 8.88 (1H, s, NH), 10.11 (2H, s, 2NH), 10.29 (1H, s, NH).

#### 1>H-spiro[indoline-3,5>-pyrido[2,3-d:6,5-d>]dipyrimidine-2,2>,4>,6>,8>(3>H,7>H,9>H,10>H)-pentaone (7b)

Pale orange powder, Mp 333-334 °C, IR (KBr) ( $\nu_{\max}$  /cm<sup>-1</sup>): 3435, 3200, 2806, 1732, 1649, 1546. MS, m/z: 366 (M<sup>+</sup>), 352, 295, 283, 252, 43. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta_{\text{H}}$  (ppm) 6.18-7.17 (4H, dd, H-Ar), 7.95 (1H, s, NH), 9.09 (1H, s, NH), 10.08 (1H, s, NH), 10.60 (1H, s, NH).

#### 1',3'-Dimethyl-1>H-spiro[indoline-3,5>-pyrido[2,3-d:6,5-d>]dipyrimidine- 2,2>,4>,6>,8> (3>H,7>H,9>H,10>H)-pentaone (7c)

Pale orange powder, Mp 327-330 °C, IR (KBr) ( $\nu_{\max}$  /cm<sup>-1</sup>): 3489, 3184, 1772, 1702, 1664, 1501. MS, m/z: 394 (M<sup>+</sup>), 367, 352, 284. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta_{\text{H}}$  (ppm) 3.09



**Scheme 1.** Synthesis of spiro indeno[1,2-b]pyrido[2,3-d] pyrimidine-5,3'-indolines using biacidic carbon as catalyst.

(3H, s, CH<sub>3</sub>), 3.37 (3H, s, CH<sub>3</sub>), 6.82-7.28 (4H, dd, H-Ar), 9.32 (1H, s, NH), 10.46 (1H, s, NH), 11.06 (1H, s, NH), 11.95 (1H, s, NH).

#### Urease inhibition assay

In this work, the total released ammonia from urease activity has been determined by the Berthelot colorimetric method (16). In brief, solution composed of 800  $\mu$ L urea as substrate and 15  $\mu$ L enzyme and inhibitors with various concentrations were incubated in water bath at 37 °C for 30 minutes. Fresh phenol solution (solution A) was prepared by dissolving 1000 mg of phenol and 7 mg of sodium nitroprusside in 100 mL of deionized water and fresh solution B was prepared by dissolving 500 mg sodium hydroxide pellets and adding 840  $\mu$ L of sodium hypochlorite solution (11-14%) to 100 mL of water. 100  $\mu$ L sample from incubated solution was transferred to microplate well and followed by adding 500  $\mu$ L of solution A and B. The mixture was stirred and kept at 37 °C for 30 minutes. After that, the plate was read at 625 nm using the Biotech Synergy H1 micropalthe reader. All experiments were performed in triplicate.

#### Docking simulation procedure

All of the processes include of preparation of input files (receptor and ligand preparation) and docking parameters were similar to our previous study (17). Briefly, to validate observed activity, molecular docking was performed using Auto Dock 4.2 using Lamarckian genetic algorithm (18) on PDB structure 3LA4 (<http://rcsb.org>) with resolution of 2 Å using default docking parameters values. The graphical user interface AutoDockTools 1.4.5 (19) was

employed to set up the enzyme and ligand input files as reported previously (20). Ligand Scout 3.0 program was used for demonstration ligand-enzyme interaction (21). Inhibitor has sketched by Marvin sketch applet (Marvin package, Chemaxon Company).

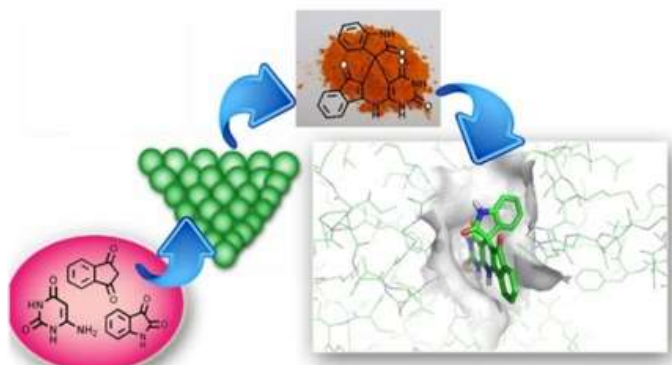
## Results and Discussion

### Chemistry

The titled compounds were synthesized in a mild, convenient and simple procedure using a three-component reaction (Scheme 1). In this method indandione 1, various isatin 2a-e and 6-aminouracil 3 were mixed and refluxed in ethanol for a short periods of time (10-30 min, Table 1) in presence of nano biacidic carbon catalyst yielding pyridopyrimidine derivatives 4a-e (Table 2) in high yields (Figure 1, Table 2).

To optimize the reaction procedure, different conditions were applied as shown in Table 1. The best results were obtained under a solvent-free condition at 130 °C (Table 1, entry 5) in EtOH as a solvent (Table 1, entry 1) and used for preparation of other derivatives.

With these optimized conditions, the scope of the reaction was inspected using various isatin as the acceptors. As can be seen from the data reported in Table 2, the use of 5-chloroisatin, resulted in improved condensation yield (95%) but the yield could be decrease by using other isatin derivatives such as *N*-benzylisatin, 5-bromoisatin and *N*-methylisatin (70-76%). It's noteworthy that in the all following cases the reaction is carried out in a very short reaction time (10-30 min).



**Figure 1.** Biacidic carbon catalyzed the synthesis of spiro pyridopyrimidine.

The mechanism of the reaction and formation of compound 4a can be explained by the condensation, addition, cyclization and dehydration reactions which illustrated in Scheme 2. First, according to acidic property of the catalyst, the carbonyl group of isatin 2a was protonated, which then condenses with enamine group of 6-aminouracil 3 to afford 5. The addition of 5 and indandione then furnished the intermediate 6. The cyclization of 6 gives product 4a after dehydration of intermediate 7.

This method also works well for the other  $\beta$ -diketone compounds 6a-c instead of indandione under the similar reaction condition (Scheme 3). The desired products 7a-c were obtained in good to excellent yields (50-90%) and at the moderate reaction times (3-4 h) as shown in Table 3.

#### *Biological activity*

All synthetic compounds were tested against urease inhibitory activity (Table 4) and shows good urease inhibitory potency in the range of  $IC_{50} = 1.94-14.46 \mu\text{M}$  in comparison to hydroxyurea as a standard inhibitor ( $\mu\text{M}$ ). To confirm obtained results and extract a preliminary structure activity relationships

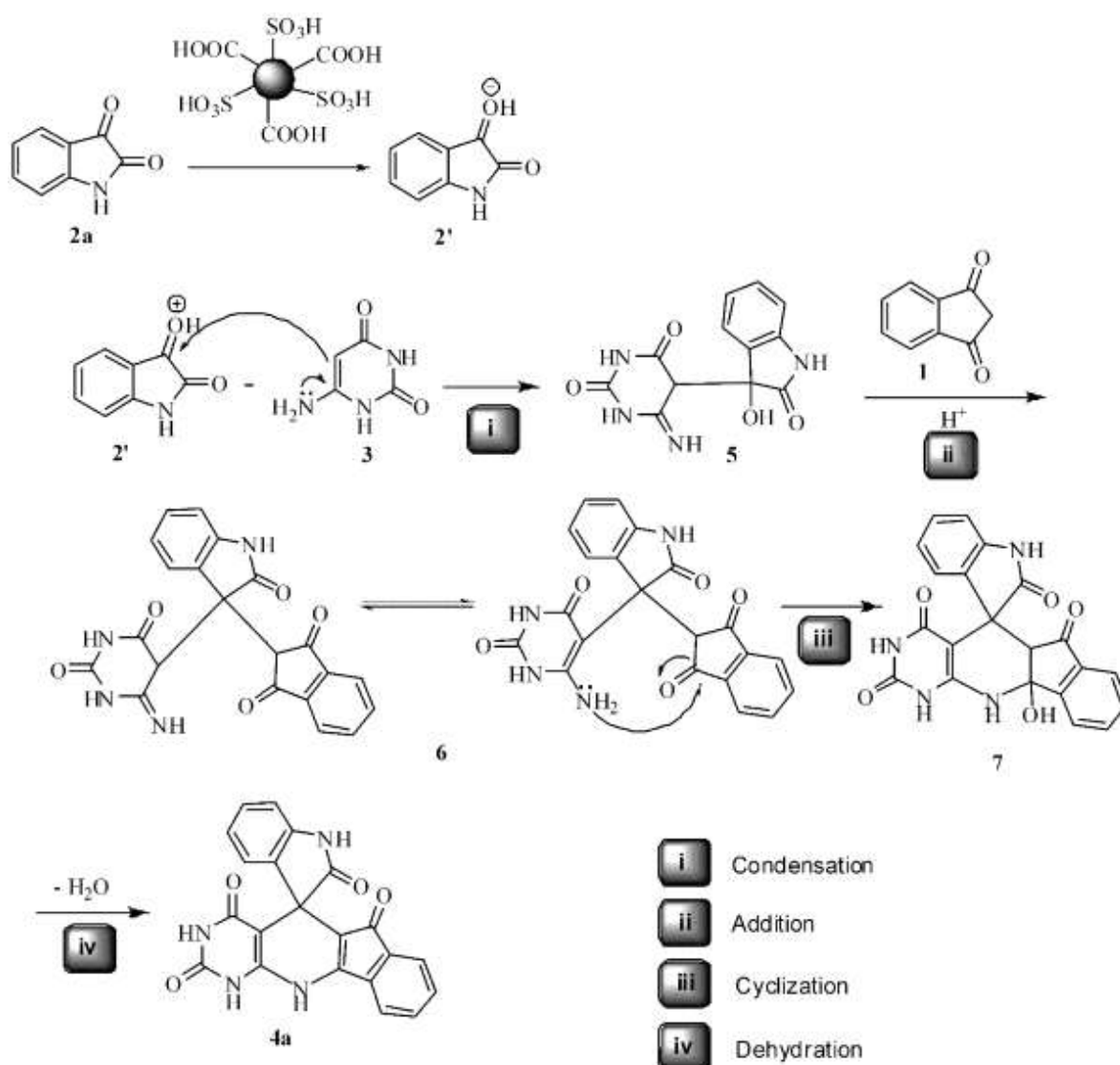
various parameters such as  $\log P$ , MR, CLog  $P$  and CMR were calculated (Table 4) using ChemOffice packages (www.cambridgesoft.com). On the other hand, the docking studies were applied for all synthesis compounds (4a-7c).

Correlation between urease inhibitory activity and physiochemical properties of tested compounds reveals that compounds with lipophilicity values in the range of  $\text{Log } P = -0.44$  to 0.61, shows good urease inhibitory potency (Table 4). Based on this deduction, compound 7b with lipophilic character was less active than 4a (Table 4). On the other hand, molar refractivity (MR) which mentioned in Table 4, has a prominent influence on the activity whereas the compound 4c with  $\text{MR} = 131.16$  hasn't any inhibition activity because it couldn't provide a good interaction with the active site.

According to our previous studies, the compounds with methyl bearing substitute on the barbituric acid ring not only haven't inhibitory activity but also activate the urease enzyme (22). This rule is shown in compound 7c which hasn't any urease inhibition activity. This could be explained by existence of two

**Table 1.** Optimization of the reaction conditions in the synthesis of 4a using biacidic carbon as catalyst.

Entry	Solvent	Temperature (°C)	Time (min)	Yield (%)
1	EtOH	Reflux	20	91
2	H <sub>2</sub> O	Reflux	60	70
3	CH <sub>3</sub> CN	100	400	71
4	neat	80	60	91
5	neat	130	30	96



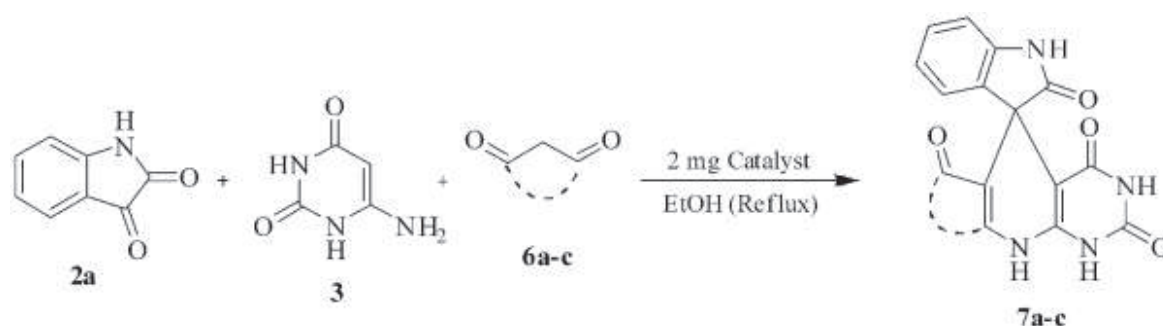
Scheme 2. The proposed reaction mechanism.

barbiturate rings in its structure, which one of barbiturate acts as an activator and the other is inhibitor thus results inactivation of compound. Meanwhile, the compound 7b which hasn't any substitution on the barbiturate ring shows the inhibitory potency with  $IC_{50} = 14.46 \mu\text{M}$ . In

compound 7a compares to 4a the indenone ring is replaced by 5,5-dimethylcyclohex-2-enone and this replacement hasn't any significant changes to the inhibitory activity ( $IC_{50}$  4a/7a is 1.94/2.81). Changing the substitution on the oxindole ring also causes considerable alteration

Table 2. Synthesis of 4 catalyzed by biacidic carbon.

Entry	R	X	Product	Time (min)	Yield (%)
1	H	H	4a	20	91
2	Me	H	4b	30	70
3	Bn	H	4c	30	76
4	H	Cl	4d	10	95
5	H	Br	4e	10	73



**Scheme 3.** Extension of this method using different β-diketones in the optimized conditions.

in the inhibitory activity since the replacement of H in 4a with halogen (5-Cl and 5-Br, 4d and 4e, respectively) decreases the inhibitory activity as a result of the steric hindrance or increase in lipophilicity due to size of halogen groups.

Our results reveal that changing the substitution on nitrogen atom of oxindole ring alters the inhibition value for instance conversion of NH to *N*-Me (4b) causes to decrease activity and conversion of NH to *N*-Bn (4c) causes the totally abolishing of the inhibition. It seems that hydrogen binding ability of nitrogen of isatin rings is necessary for higher activity and increasing the sizes of substituted groups will decrease binding of molecule to active site. These properties have been reported narcotic antagonist and agonist previously (23).

#### Docking studies

Herein, the molecular docking was used in order to predict the orientation of spiro indeno[1,2-*b*]pyrido[2,3-*d*]pyrimidine-5,3'-indolines family in the urease binding pocket. In this way, the most potent compound 4a was used as the family symbol. Figure 2A represents the binding pose of 4a in the active site. The planner structure of this ligand lets it to sit deeply inside of active site cleft. Moreover,

the suitable positions of chemical groups led to favorable interactions with the receptor. One of the carbonyl group in the barbituric ring formed a HBA with Met637. The other carbonyl moiety comprises a HBA with His545 alongside a weak ionic interaction with the Ni<sup>2+</sup> atom. The two nitrogen atoms in the barbituric ring donate their hydrogen atoms in hydrogen bonds with His492 and Asp633 residues. Thus, the barbituric cycle which is included in all synthesized compounds is responsible for four hydrogen bonds with urease protein. Besides, the hydrogen of nitrogen atom in 9 position of the ligand is involved a HBD toward Asp494. The last hydrogen bond is seen between CME592 and carbonyl group connected to the cyclopentadiene moiety. In addition, 4a can be more stabilized due to presence of π-π interaction with His593.

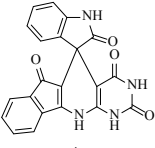
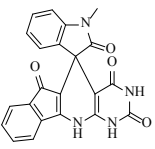
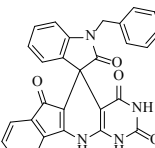
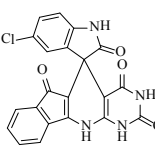
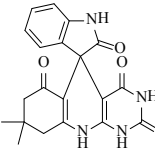
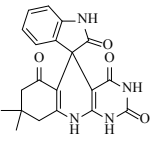
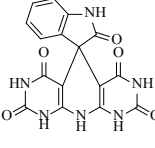
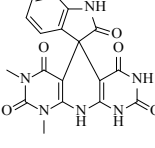

#### Conclusions

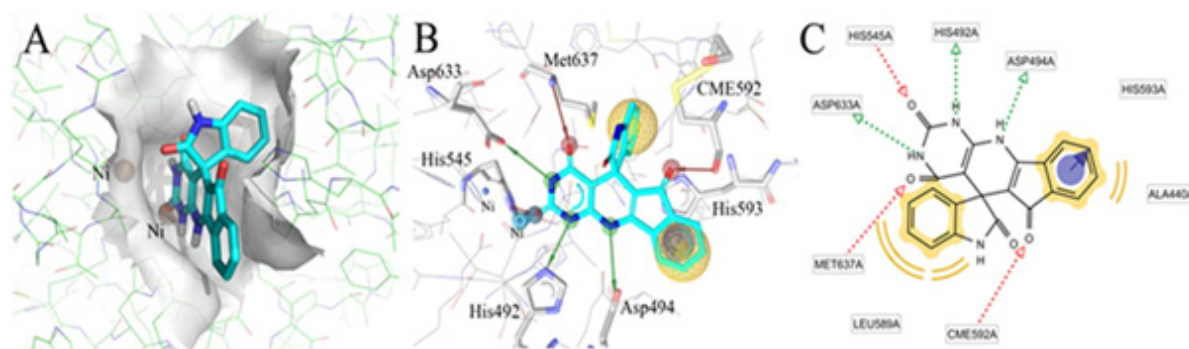
As a conclusion the spiro indenopyridopyrimidine derivatives are valuable compounds which show inhibition potency through urease and offer the opportunity of synthesizing the new molecules of this group. In the continuous of other studies by this investigation the key role of barbiturate ring in

**Table 3.** Exploration of the reaction scope in relation to the β-diketone components using biacidic carbon as catalyst.

Entry	β-diketone compounds	Product	Time (h)	Yield (%)
1		7a	3	60
2		7b	4	90
3		7c	3	50

**Table 4.** Urease inhibitory activity of spiro pyridopyrimidine compounds.

Entry	Products	IC <sub>50</sub> (μM)	Log P	MR (cm <sup>3</sup> /mol)	CLog P	CMR	Binding energies (kcal not Kcal)
1	 4a	1.94	0.38	101.67	- 0.049	10.1332	-6.40
2	 4b	4.35	0.61	106.67	0.29099	10.597	-6.71
3	 4c	000	2.35	131.16	2.265	13.1082	-8.05
4	 4d	5.557	0.94	116.27	0.9216	10.6246	-6.43
5	 7a	7.44	1.21	109.36	1.0716	10.9102	-6.52
6	 7a	2.81	-0.44	98.4	0.6825	10.169	-6.31
7	 7b	14.46	- 1.64	85.13	- 2.388	9.0649	-6.71
8	 7c	000	- 1.17	95.13	- 1.336	9.9925	-5.98
9	 Std	100					-5.29



**Figure 2.** Optimal binding model for compound 4a into active site of Jack-bean urease enzyme docked by Autodock 4.2 (A), interaction of ligand with key residues are shown in 3D and 2D models (B, C). The yellow sphere indicates hydrophobic region, the red line shows hydrogen bond acceptor, the green line point out hydrogen bond donor features and purple cycle is  $\pi$ - $\pi$  interaction.

preparation of inhibitory potent compounds was either confirmed.

### Acknowledgments

We gratefully acknowledge for financial support from the Research Councils of Alzahra University and University of Tehran and Tehran University of Medical Sciences.

### References

- (1) Biglar M, Soltani K, Nabati F, Bazl R, Mojab F and Amanlou M. A Preliminary Investigation of the Jack-bean Urease Inhibition by Randomly Selected Traditional used Herbal Medicine. *Iran. J. Pharm. Res.* (2012) 11: 831-837.
- (2) Kuwahara H, Miyamoto Y, Akaike T, Kubota T, Sawa T, Okamoto S and Maeda H. Helicobacter pylori urease suppresses bactericidal activity of peroxydinitrite via carbon dioxide production. *Infect. Immun.* (2000) 68: 4378-4383.
- (3) Rosenstein IJ, Hamilton-Miller JM and Brumfitt W. Role of urease in the formation of infection stones: comparison of ureases from different sources. *Infect. Immun.* (1981) 32: 32-37.
- (4) Musher DM, Griffith DP, Yawn D and Rossen RD. Role of urease in pyelonephritis resulting from urinary tract infection with proteus. *J. Infect. Dis.* (1975) 131: 177-181.
- (5) Kusters JG, van Vliet AH and Kuipers EJ. Pathogenesis of Helicobacter pylori infection. *Clin. Microbiol. Rev.* (2006) 19: 449-490.
- (6) Tripathi G, Singh S and Nath G. Formulation and *In-vitro* evaluation of pH-Sensitive oil entrapped polymeric blend amoxicillin beads for the eradication of Helicobacter pylori. *Iran. J. Pharm. Res.* (2012) 11: 447-455.
- (7) Megraud F. H pylori antibiotic resistance: prevalence, importance, and advances in testing. *Gut* (2004) 53: 1374-1384.
- (8) Müller CE. General synthesis and properties of 1-monosubstituted xanthenes. *Synthesis* (1993): 125-128.
- (9) Simo O, Rybár A and Alföldi J. New [c,d]-fused purinediones: 2-Substituted 9-methyl-4,5-dihydro-6H,8H-pyrimido-[1,2,3-cd]purine-8,10(9H)-diones. *Synthesis* (1995): 837-840.
- (10) Simo O, Rybár A and Alföldi J. 2-Triazolylpyrimido[1,2,3-cd]purine-8,10-diones via 1,3-dipolar cycloadditions to 2-ethynylpyrimido[1,2,3-cd]purine-8,10-dione. *J. Heterocycl. Chem.* (2000) 37: 1033-1039.
- (11) Beauglehole AR, Baker SP and Scammells PJ. Fluorosulfonyl-substituted xanthenes as selective irreversible antagonists for the A1-adenosine receptor. *J. Med. Chem.* (2000) 43: 4973-4980.
- (12) Daves GD, Robins RK and Cheng CC. Antibiotics. I. Synthesis of 1,6-dimethyl-5,7-dioxo-1,5,6,7-tetrahydropyrimido [5,4-e] as-triazine (Toxoflavin) and related compounds. *J. Am. Chem. Soc.* (1962) 84: 1724-1729.
- (13) Müller CE and Sandoval-Ramírez J. A new versatile synthesis of xanthenes with variable substituents in the 1-, 3-, 7- and 8-positions. *Synthesis* (1995): 1295-1299.
- (14) Sheldon RA, Arends IWCE, ten Brink G-J and Dijkman A. Green, Catalytic oxidations of alcohols. *Acc. Chem. Res.* (2002) 35: 774-781.
- (15) Mohammadi Ziarani G, Faramarzi S, Asadi S, Badiei A, Bazl R and Amanlou M. Three-component synthesis of pyrano [2, 3-d]-pyrimidine dione derivatives facilitated by sulfonic acid nanoporous silica (SBA-Pr-SO<sub>3</sub>H) and their docking and urease inhibitory activity. *DARU* (2013) 21: 3-16.
- (16) Mohammadi Ziarani G, Asadi S, Badiei A, Mousavi S and Gholamzadeh P. One-pot four-component synthesis of 2,5-dioxo-1,2,3,4,5,6,7,8-octahydroquinolines catalyzed by silica-based sulfonic acid. *Res Chem Intermed* (2015) 41: 637-645.
- (17) Imani Shakibaei G, Feiz A, Reza Khavasi H, Abolhasani Soorki A and Bazgir A. Simple three-component



- method for the synthesis of spiro indeno[1,2-b]pyrido[2,3-d]pyrimidine-5,3'-indolines. *ACS Comb. Sci.* (2010) 13: 96-99.
- (18) Xiao H, Guo Y, Liang X and Qi C. One-step synthesis of novel biacidic carbon via hydrothermal carbonization. *J. Solid State Chem.* (2010) 183: 1721-1725.
- (19) Weatherburn MW. Phenol-hypochlorite reaction for determination of ammonia. *Anal. Chem.* (1967) 39: 971-974.
- (20) Azizian H, Nabati F, Sharifi A, Siavoshi F, Mahdavi M and Amanlou M. Large-scale virtual screening for the identification of new *Helicobacter pylori* urease inhibitor scaffolds. *J. Mol. Model.* (2012) 18: 2917-27.
- (21) Golbabaei S, Bazl R, Golestanian S, Nabati F, Omrany ZB, Yousefi B, Hajiaghache R, Rezazadeh S and Amanlou M. Urease inhibitory activities of  $\beta$ -boswellic acid derivatives. *DARU* (2013) 21: 2-8.
- (22) Azizian H, Bahrami H, Pasalar P and Amanlou M. Molecular modeling of *Helicobacter pylori* arginase and the inhibitor coordination interactions. *J. Mol. Graph. Model.* (2010) 28: 626-635.
- (23) Morris GM, Huey R, Lindstrom W, Sanner MF, Belew RK, Goodsell DS and Olson AJ. AutoDock4 and AutoDockTools4: Automated docking with selective receptor flexibility. *J. Comput. Chem.* (2009) 30: 2785-2791.
- (24) Anon. Crystal structure of the first plant urease from Jack bean (*Canavalia ensiformis*). <http://www.rcsb.org/pdb/explore.do?structureId=3la4>. Access date: Jul 15, 2014.
- (25) Wolber G and Langer T. LigandScout: 3-D pharmacophores derived from protein-bound ligands and their use as virtual screening filters. *J. Chem. Inf. Model.* (2005) 45: 160-9.
- (26) Chemoffice, 2004, version 8.0, has been supplied by Cambridge Software Company, 100, Cambridge Park Drive, Cambridge, MA 02140-2317, USA.
- (27) Williams DA, Roche VF, Roche EB. Central Analgesics. In: Lemke TL, Williams DA, Roche VF, Zito SW, eds. *Foye's Principles of Medicinal Chemistry* (7<sup>th</sup> ed.). Wolters Kluwer, Lippincott Williams & Wilkins. Philadelphia (2012) Ch. 21:658-699.

---

This article is available online at <http://www.ijpr.ir>

---

**Back issues?**  
**Visit <http://www.ijpr.ir>**  
**or**  
**[http:// ijpr.sbm.ac.ir](http://ijpr.sbm.ac.ir)**