Original Article

Procerenone: a Fatty Acid Triterpenoid from the Fruit Pericarp of *Omphalocarpum procerum* (Sapotaceae)

Rosine Fotsing Ngamgwe^{*a*}, Raoul Yankam^{*b*}, Jean Rodolphe Chouna^{*c*}, Christian Lanz^{*b*}, Julien Furrer^{*d*}, Stefan Schürch^{*d*}, Marcel Kaiser^{*ef*}, Bruno Ndjakou Lenta^{*g**}, Silvère Ngouela^{*b*}, Etienne Tsamo^{*a*} and Rudolf Brenneisen^{*b*}

^aDepartment of Organic Chemistry, Faculty of Science, TWAS Research Unit, University of Yaoundé 1, P. O. Box 812, Yaoundé, Cameroon. ^bDepartment of Clinical Research, University of Bern, Murtenstrasse 35, CH-3010 Bern, Switzerland. ^cDepartment of Chemistry, Faculty of Science, University of Dschang, P. O. Box 67, Dschang, Cameroon. ^dDepartment of Chemistry and Biochemistry, University of Bern, Freiestrasse 3, CH-3012 Berne, Switzerland. ^eSwiss Tropical and Public Health Institute, Socinstrasse 57, CH-4002 Basel, Switzerland. ^fUniversity of Basel, Petersplatz 1, CH-4003 Basel, Switzerland. ^gDepartment of Chemistry, Higher Teacher Training College, University of Yaoundé 1, P. O. Box 47, Yaoundé, Cameroon.

Abstract

Phytochemical investigation of a dichloromethane-methanol (1:1) extract of the fruit pericarp of *Omphalocarpum procerum* which exhibited antiplasmodial activity during preliminary screening led to the isolation of the new fatty ester triterpenoid 3β -hexadecanoyloxy-28hydroxyolean-12-en-11-one (1), together with five known compounds 2-6. The structure of the new compound as well as those of the known compounds was established by means of spectroscopic methods and by comparison with previously reported data. Compounds 1- 4 were evaluated *in-vitro* for their cytotoxicity against L6 cell lines and antiprotozoal activities against *Plasmodium falciparum*, *Leishmania donovani*, *Trypanosoma brucei rhodesiense* and *Trypanosoma cruzi* (species responsible for human malaria, visceral leishmaniasis, African trypanosomiasis and Chagas disease, respectively). The tested compounds showed weak to moderate antiprotozoal activity and, no significant effect was detected regarding their cytotoxic potency.

Keywords: Sapotaceae; *Omphalocarpum procerum*; Triterpenoid; Procerenone; Antiparasitic activity.

Introduction

Omphalocarpum procerum (Sapotaceae) is a tree confined to humid tropical Africa including Cameroon. It can grow up to 30 m in height and 40 cm in diameter. Its fruits are hard and often consumed by elephants (1, 2).

In Africa, plants of the genus *Omphalocarpum* are prepared for various purposes such as decoctions, powders, macerations, and are used for years in traditional medicine to treat headaches; wounds skin diseases, constipation, elephantiasis, fever, cough, and rheumatism (3-6). Previous phytochemical investigations of plants of the Sapotaceae family have revealed the presence of alkaloids, phenolic compounds, glycerol, fatty acids, flavonoids, saponins and

^{*} Corresponding author:

E-mail: lentabruno@yahoo.fr

triterpenes (7-12). To the best of our knowledge, no phytochemical or pharmacological study has been reported on the species *O. procerum* so far. In a continuing search for bioactive compounds from Cameroonian medicinal plants, we have investigated the CH_2Cl_2 -MeOH (1:1) extract of the pericarps of the fruits of *O. procerum*. Herein, we report on the isolation and structure elucidation of a new triterpenoid, procerenone (1), together with the antiparasitic activity of some isolated compounds.

Experimental

General procedure

Melting points were determined on an M-540 melting-point unit (Buchi, Flawil, Switzerland). Optical rotations were measured, in chloroform solution, on a DIP-3600 digital polarimeter (JASCO, Tokyo, Japan). Infrared (IR) spectra were determined on a Fourier transform IR spectrometer (JascoP-2000). The mass spectra were acquired on a LTQ Orbitrap XL mass spectrometer (Thermo Fisher Scientific, Bremen, Germany) equipped with a nanoelectrospray ion source. Spectra were recorded in the positive ion mode with the resolution set to 100 000. Calibration of the instrument was performed with ProteoMass LTQ/FT-Hybrid ESI positive mode calibration mix solution (Supelco Analytical, Bellefonte, PA, USA). For MS/MS experiments, the precursor ion was isolated within a window of $\pm 1.5 \text{ m/z-units}$. Collision-induced dissociation (CID) was performed using Helium as the collision gas and relative collision energy of 35%. The Xcalibur software package V. 2.0.7 was used for data processing. ¹H and ¹³C NMR spectra were recorded on a Bruker Avance II 400 MHz spectrometer equipped with a 5-mm broadband probe head (BBI), operating at 400 (1H) and 100 MHz (¹³C), respectively. All chemical shifts are reported as relative differences to the internal standard tetramethylsilane (TMS). Silica gels of 230- to 400-mesh and 70- to 230-mesh (Merck, Darmstadt, Germany) were used for column chromatography (CC), respectively, while aluminum sheets precoated with silica gel 60 F_{254} (Merck) were used for thin-layer chromatography (TLC), with various mixtures of petroleum ether, *n*-hexane, EtOAc, and acetone as mobile phases.

Plant material

The Shell seeds of *O. procerum* were collected at Ambam in the Southern province of Cameroon. The plants were identified at the National Herbarium of Cameroon, where a voucher specimen (N°11955SFR) was deposited.

Extraction and isolation

The air-dried and powdered pericarps of the fruits (1.5 Kg) of O. procerum were extracted in a CH₂Cl₂-MeOH (1:1) mixture (5.0 L) at room temperature within 2 days. The solvents were evaporated under reduced pressure to afford 60 g of crude extract. The resulting mixture was successively extracted with *n*-hexane and EtOAc at room temperature to yield 22 g and 13 g of *n*-hexane and EtOAc extracts, respectively. The *n*-hexane soluble fraction was subjected to CC over silica gel (230 - 400 mesh) and eluted with mixtures of *n*-hexane-EtOAc and *n*-hexane-CH₂Cl₂ of increasing polarities. A total of 121 fractions of 300 mL each were collected and combined on the basis of similar TLC profiles to yield 3 main fractions (F_1-F_3) . Fraction F_1 (8.2 g) was a complex oily mixture that was not further studied. Fraction F_2 (6.4 g) was subjected to CC on silica gel (Merck, 70 - 230 mesh, Merck) and eluted with *n*-hexane- EtOAc (1:0 to 1:1) to afford procerenone (1) (19 mg), stigmasterol (5) (49 mg), and betulin (2) (27 mg). Fraction F_3 (5.8 g) was also subjected to CC on silica gel (Merck, 70 - 230 mesh) and eluted with n-hexane-CH₂Cl₂ (1:4 to 0:1) to afford β-amyrin (3) (50 mg), β-sitosterol (6) (62 mg), and lupeol acetate (4) (5 mg).

Procerenone (1): 3β -hexadecanoyloxy-28hydroxyolean-12-en-11-one

White powder. $- [\alpha]_D^{20} 41.5 (c = 0.1, \text{CHCl}_3)$. $- \text{IR} (\text{KBr, cm}^{-1})$. $- v_{\text{max}} = 3447.3 (\text{OH}), 1651.0 (\text{C} = \text{O}) \text{ and } 1698.2 (\text{C} = \text{O}) \text{ cm}^{-1}$. $- {}^{1}\text{H} \text{ NMR} (\text{CDCl}_3, 400 \text{ MHz}) \text{ and } {}^{13}\text{C} \text{ NMR} (\text{CDCl}_3, 100 \text{ MHz}) \text{ spectroscopic data, see Table 1.} - \text{HRESIMS: } [\text{M}+\text{H}]^+ \text{ at } m/z = 695.5955 (\text{calcd.} m/z = 695.5973 \text{ for } \text{C}_4\text{H}_{20}\text{O}_4)$.

Bioassays

The *in-vitro* cytotoxicity and antiprotozoal activities against the parasites *T. b. rhodesiense*,

T. cruzi, L. donovani, and *P. falciparum* were determined as earlier reported (20). The tests were carried out with the following strains,

were carried out with the following strains, parasite forms and positive controls: *T. b. rhodesiense* STIB900, trypomastigote forms, melarsoprol, IC₅₀ of 3 ng/mL; *T. cruzi*, Tulahuen C2C4, amastigote forms in L6 rat myoblasts, benznidazole, IC₅₀ of 0.531 µg/mL; *L donovani*, MHOM/ET/67/L82, axenic amastigote forms, miltefosine, IC₅₀ of 0.145 µg/mL; *P. falciparum*, NF54, erythrocytic stages, chloroquine, IC₅₀ of 6 ng/mL and L6 cells, rat skeletal myoblasts, podophyllotoxin IC₅₀ of 6 ng/mL.

Results and Discussion

The air-dried and ground fruit pericarp of *O*. procerum was extracted at room temperature with a mixture of CH_2Cl_2 –MeOH (1:1, v/v). The extract was concentrated to dryness under vacuum and the residue subjected to repeated column chromatographic separation to yield procerenone (1) along with betulin (2) (13), β -amyrin (3) (13), lupeol acetate (4) (14), stigmasterol 5 (15), and β -sistosterol (6) (16,17).

Compound (1) was obtained as a white powder. It gave a positive reaction to Liebermann-Burchard test, as usual for a triterpenoid. Its molecular formula $C_{46}H_{78}O_4$, with eight double bond equivalents, was deduced from the HRESIMS spectrum which showed the pseudo-molecular ion peak $[M+H]^+$ at m/z =695.5955 (calcd. m/z = 695.5973 for $C_{46}H_{70}O_{4}$). The IR spectrum showed characteristic absorption bands at 3447 (OH), 1651 and 1698 $(\alpha,\beta$ -unsaturated ketone), and 1613 (C = C) cm⁻ ¹. The ¹H NMR spectrum (Table 1) revealed the presence of seven singlet resonances due to seven angular triterpenoids methyl protons (δ_{μ} = 0.86, 0.90, 1.10, 1.14, 1.29, 1.39 and 1.59), a singlet of one methine proton ($\delta_{\rm H} = 2.37$) and one singlet of an olefinic proton ($\delta_{\rm H} = 5.52$). This spectrum also exhibited two doublets typical of an AB system ($\delta_{\rm H}$ = 3.45 and 3.20, *J* = 10.8 Hz) and one doublet of doublets ($\delta_{\rm H} = 4.47, J = 4.8$, 12 Hz). Furthermore, the ¹H NMR spectrum exhibited series of resonances at $\delta_{\rm H} = 0.88$ (3H, t, J = 6.0 Hz), 1.26 (brs) and 2.27 (2H, t, J = 7.5Hz) which could be assigned to protons of a long

alkyl chain. The ¹³C NMR (Table 1) and DEPT spectra of compound 1 displayed resonances characteristic of a single double bond (δ_c 128.2 and 169.6), one conjugated ketone carbonyl $(\delta_{c} = 199.7)$, one oxymethine $(\delta_{c} = 80.2)$, and one oxymethylene ($\delta_c = 69.5$). The ¹³C NMR also confirmed the presence of the long chain acyl ester with the carbon resonances at δ_c = 173.5 (-COOR), 29.8 $[(CH_2)_n]$ and 14.0 (CH_3) . The ¹³C NMR spectrum also displayed eight resonances typical for eight methyl groups, from which seven could be assigned to the triterpene pattern ($\delta_{c} = 16.3, 16.6, 18.6, 22.8,$ 23.2, 29.3 and 32.7) and one to a terminal methyl of the long acyl chain ($\delta_c = 14.0$). On the basis of these NMR data, compound 1 was assumed to be a fatty acid ester of an olean-12ene-type triterpenoid with one hydroxyl group and one α,β -unsaturated ketone group (12,18). The location of the ketone carbonyl at C-11 was deduced from the correlations observed in the HMBC spectrum between the olefinic proton H-12 ($\delta_{\rm H}$ 5.52) and the carbons C-9 ($\delta_{\rm C}$ 61.7), C-11 (δ_{c} 199.7) on one hand and between H-9 $(\delta_{\rm H} 2.37)$ and the carbonyl carbon C-11 $(\delta_{\rm C}$ 199.7) (Figure 1) on the other hand. According to these findings, the triterpenoid moiety of 1 was identified as being 11-oxoerythrodiol (2) (19). The ester function at the C-3 position was deduced from the correlation observed in the HMBC spectrum between H-3 ($\delta_{\rm H}$ 4.47) and the ester carbonyl carbon (δ_{c} 173.5). The length of the acyl chain ester was deduced from the MS/ MSⁿ spectrum which exhibits the characteristic ion peaks at m/z 677.58 [M-H₂O]⁺, m/z = 665.59 $[M-CH_2O]^+$, and $m/z = 439.36 [M-CH_2O]^+$ $C_{16}H_{31}O_{2}^{+}$ (Figure 2). The axial orientation of the H-3 proton was deduced from the coupling constants with protons H-2 (δ_{H3} = 4.47, dd, J = 4.4 and 12.0 Hz) (17). Thus, the structure of compound 1 was unambiguously assigned to 3β-hexadecanoyloxy-28-hydroxyolean-12-en-11-one, named procerenone.

Compounds 1-4 (Figure 3) were tested for cytotoxicity against L6 cell lines and for their antiprotozoal activity against *Plasmodium falciparum*, *Leishmania donovani*, *Trypanosoma brucei rhodesiense* and *Trypanosoma cruzi*. The tested compounds showed weak to moderate antiprotozoal activity against the tested parasites



Figure 1. Selected HMBC correlations observed in compound 1.



Figure 2. Fragmentation patterns of compound 1.



Figure 3. Structure of compounds 1-4.

with $IC_{50}s$ in range of 9 to 80 µg/mL. No significant effect was detected regarding their cytotoxic potency.

Conclusion

The phytochemical study of the fruit pericarp of *O. procerum* (Sapotaceae) led to the isolation and characterization of six compounds including one new fatty acid triterpenoids, procerenone. This class of secondary metabolite has been isolated from other genera of the Sapotaceae family like *Gambeya* and could be considered as one chemotaxonomic marker (12). The antiprotozoal activities of the tested compounds were nearly equal to that of the extract on each tested strain of parasite. Despite the moderate antiprotozoal potency of the extract it exhibited weak cytotoxicity to L-6 cell lines. The present results partially validate the use of *O. procerum* in folk medicine.

Acknowledgements

The authors acknowledge the Swiss National Science Foundation for awarding a fellowship to B. N. Lenta, grant number: IZKOZ2-13857O/I, and the Third World Academy of Science (TWAS) for the research grant Nr. 07-141 LDC/ CHE/AF/AC-UNESCO FR: 3240171776 to our TWAS Research Unit.

References

- Normand D and Paquis J. Manuel d'identification des bois commerciaux, Afrique guinéo-congolaise Tome 2. Centre Technique Forestier Tropical France (1976) 335.
- (2) Nchanji A and Plumptre A. Seed germination and early seedling establishment of some elephant-dispersed species in Banyang-Mbo wildlife sanctuary, southwestern Cameroon. J. Trop. Ecol. (2003) 19: 229-237.
- (3) Irvine F. Woody Plants of Ghana with Special Reference to their Uses. London Oxford University press (1961) 594-595.
- (4) Bouquet A and Debray M. Plantes Médicinales de la Côte d'Ivoire, Travaux et Documents Paris (1974) 231.
- (5) Aubréville A. Flore du Cameroun: Sapotaceae, Volume 2. ed. Museum national d'histoire naturelle, Paris (1964) 143.
- (6) Betti J. An ethnobotanical study of medicinal plants among the Baka pygmies in the Dja biosphere reserve, Cameroon. *Afr. Study Monogr.* (2004) 25: 1-27.
- (7) Baliga MS, Pai RJ, Bhat PL, Palatty HP and Boloor R. Chemistry and medicinal properties of the Bakul (*Mimusops elengi* Linn): A review. *Food Res. Int.* (2011) 44: 1823-1829.
- (8) Charrouf Z and Guillaume D. Phenols and polyphenols from *Argania spinosa*. *Am. J. Food Technol*. (2007) 2: 679-680.
- (9) Charrouf Z and Guillaume D. Secondary metabolites from Argania spinosa (L.) Skeels. Phytochem. Rev. (2002) 1: 345-354.
- (10) Akihisa T, Kojima N, Kikuchi T, Yasukawa K, Tokuda H, Fukatsu ET, Masters ET, Manosroi M and Manosroi J. Anti-Inflammatory and Chemopreventive effects of Triterpene cinnamates and acetates from Shea Fat. J. Oleo. Sci. (2010) 59: 273-280.

- (11) Akihisa T, Kojima N, Katoh N, Kikuchi T, Fukatsu M, Shimizu N and Masters ET. Triacylglycerol and triterpene ester composition of Shea nuts from seven African countries. J. Oleo. Sci. (2011) 60: 385-386.
- (12) Wandji J, Tillequin F, Mulholland AD, Wansi JD, Fomum TZ, FeundjiepV, Libot F and Tsabang N. Fatty esters of triterpenoids and steroids glycosides from *Gambeya africana*. *Planta Med*.(2002) 68: 822-826.
- (13) Mahato SB and Kundu AP. ¹³C-NMR spectra of pentacyclic triterpenoids-a compilation and some saliente features. *Phytochem.* (1994) 37: 1517-1575.
- (14) Mitra CR and Misra G. Mimusops hexandra-I: Constituents of fruit and seed. Phytochem. (1965) 4: 345-348.
- (15) Susidarti RA, Rahmani M, Ali AM, Sukari MA, Ismail HBM, Kulip J and Waterman P G. 8-Methoxycapnolactone and stigmasterol from *Micromelum minutum. Indonesian J. Pharm.* (2007) 18: 105-109.
- (16) Moghaddam FM, Farimani MM, Salahvarzi S and

Amin G. Chemical constituents of dichloromethane extract of cultivated *Satureja khuzistanica. e-CAM* (2007) 4: 95-98.

- (17) Wang XL, Hay AE, Matheeussen A, Gupta MP and Hostettman K. Structure elucidation and NMR assignments of two new triterpenoids from the stems of *Paragonia pyramidata* (Bignoniaceae). *Magn. Reson. Chem.* (2011) 49: 184-189.
- (18) Barreiros LM, David JM, Guedes MLS and David PJ. Fatty acid esters of triterpenes from *Erythroxylum* passerinum. J. Braz. Chem. Soc. (2002) 13: 669-673.
- (19) Kawaga M, Minami H, Nakahara M, Takahashi H, Takoaka S and Fukuyama Y. Oleanane - type triterpenes from *Viburnum awabuki*. *Phytochem*. (1998) 47: 1101-1105.
- (20) Orhan I, Şener B, Kaiser M, Brun R and Tasdemir D. Inhibitory activity of marine sponge-derived natural products against parasitic protozoa. *Mar. Drugs* (2010) 8: 47-58.
- This article is available online at http://www.ijpr.ir