

Triterpenes and Steroids from *Euphorbia denticulata* Lam. with Anti-Herpes Simplex Virus Activity

Sara Shamsabadipour^a, Mustafa Ghanadian^{b*}, Hojjatollah Saeedi^c, Mohammad Reza Rahimnejad^c, Marzieh Mohammadi-Kamalabadi^d, Seyed Majid Ayatollahi^e and Loghman Salimzadeh^f

^aIsfahan Pharmaceutical Sciences Research Center, Isfahan University of Medical Sciences, Isfahan, Iran. ^bDepartment of Pharmacognosy, Isfahan Faculty of Pharmacy and Pharmaceutical Sciences, Isfahan University of Medical Sciences, Isfahan Iran. ^cDepartment of Biology, Faculty of Science, University of Isfahan, Isfahan, Iran. ^dMicrobiology, Islamic Azad University, Falavarjan Branch, Young Researchers and Elites Club, Isfahan, Iran. ^ePhytochemistry Research Center and School of Pharmacy, Shahid Beheshti University of Medical Sciences, Tehran, Iran. ^fMedical Plants Research Center, Shahrekord University of Medical Sciences, Shahrekord, Iran.

Abstract

In this research, dried acetone:chloroform extract of aerial parts of *E. denticulata* as one of the endemic plants to Iran, afforded a number of triterpenes and steroids including: betulin, 24-methylene-cycloart-3-ol, cycloart-23Z-ene-3 β ,25-diol, cycloart-23E-ene-3 β ,25-diol, ergosta-8,24-dien-3-ol (obtusifoliol) and beta-sitosterol which were reported for the first time from this plant. The structure of these compounds was elucidated by NMR and mass spectroscopic methods. The MTS assay was used to determine the toxicity and antiviral activity of betulin and (3 β ,23E)-cycloarta-23-ene-3,25-diol. Betulin showed anti-HSV-1 activity with EC₅₀ value of 84.37 \pm 0.02 μ g/mL, and toxicity on normal vero cells with CC₅₀ value of 660.718 \pm 0.072 μ g/mL. (3 β ,23E)-Cycloarta-23-ene-3,25-diol showed antiviral effect with EC₅₀ value of 86.63 \pm 0.03 μ g/mL, and toxicity with CC₅₀ value of 1089.21 \pm 0.25 μ g/mL. The results revealed that these two compounds have the antiviral activity far below the CC₅₀ value with selectivity index (CC₅₀/EC₅₀) values of 7.83, and 12.57, respectively.

Keywords: *Euphorbia denticulata*; triterpenes; cycloartanes; steroids; toxicity; anti-HSV-1 activity.

Introduction

Euphorbiaceae is one of the largest families of the phylum Anthophyta. In this family the largest genus is *Euphorbia* which comprises well over 2000 species in tropical and temperate zones of Asia and other parts of the world (1). In Iran 70 species are reported that 17 of them are

endemic. In traditional medicine *Euphorbia* was used to treat inflammations or as an antiviral or antitumor (2, 3). There are reports on *Euphorbia* species with anti HIV effects or anti herpes simplex virus (3-5). Currently coordinate with the medicinal chemists which rapidly build and develop new synthetic drugs, researchers of the natural product chemistry are also discovering secondary metabolites in plants with their subsequent biological effects. In this context, the paper in hand was also aimed to isolate and

* Corresponding author:

E-mail: ghannadian@gmail.com

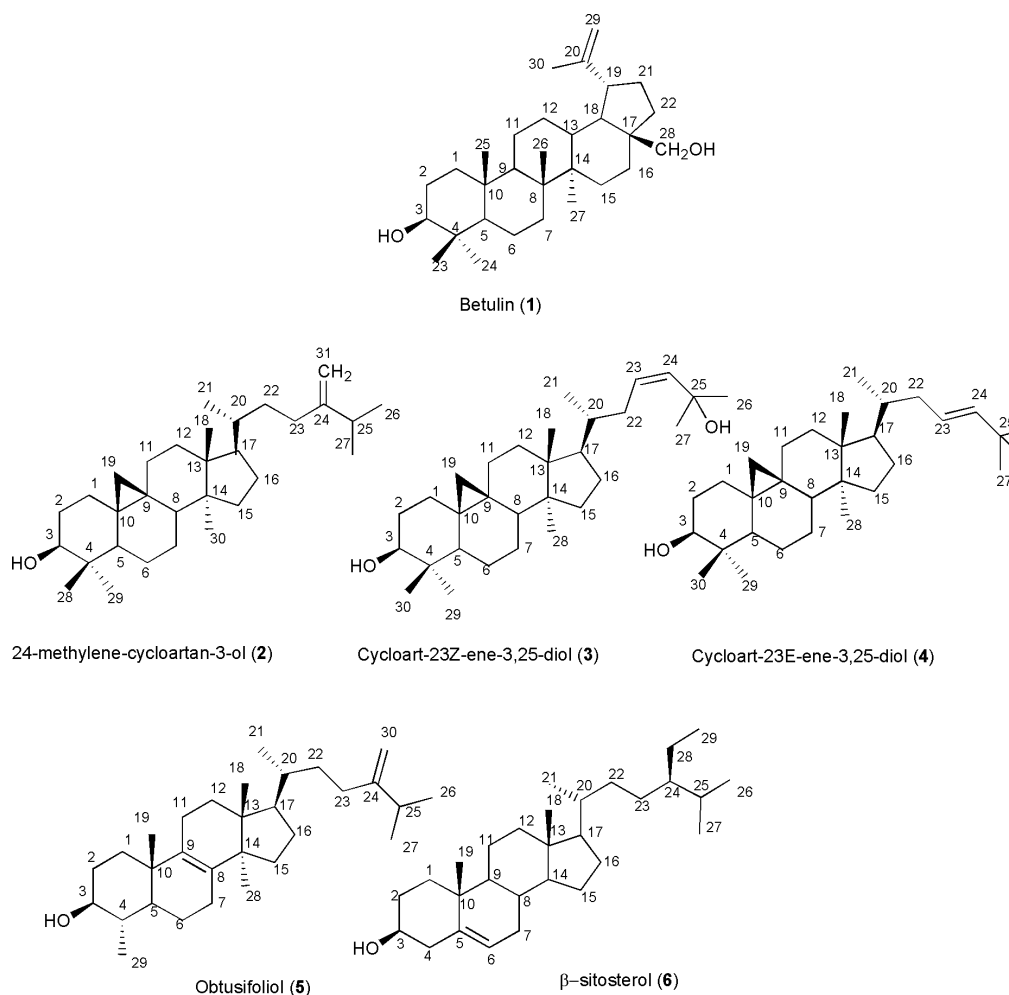


Figure 1. Triterpenes and steroids from *Euphorbia denticulate*.

detect this type of compounds from *Euphorbia denticulata* Lam., considering that *Euphorbia* genus is one of the rich and economic sources of triterpenoids specially cycloartanes that as intermediates convert to steroids in the plant metabolic pathways (6).

Experimental

General experimental procedures

The NMR spectra were recorded on a Bruker Avance AV 400, using CDCl_3 as solvent. HPLC was carried out on a waters 515 using a Pack-Sil column (250×20 mm i.d.) packed with 5 μm silica (YMC Co., Ltd., Kyoto, Japan) and hexane:EtOAc as mobile phase. Chromatographic materials were silica gel (Merck Co., Germany).

Thin layer chromatography detection was achieved by spraying the silica gel plates with cerium sulfate in 10% aq. H_2SO_4 , followed by heating.

Materials

Dulbecco's modified eagle's growth medium (DMEM), and Aciclovir purchased from Sigma-aldrich company (St Louis, MO, USA), fetal bovine serum (FBS), streptomycin, and amphotericin B from the GIBCO/Invitrogen (Karlsruhe, Germany), and MTS [3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl) 2H-tetrazolium] from Promega (Madison, WI, USA). African green monkey kidney cell line (Vero cell line, ATCC C102) was obtained from

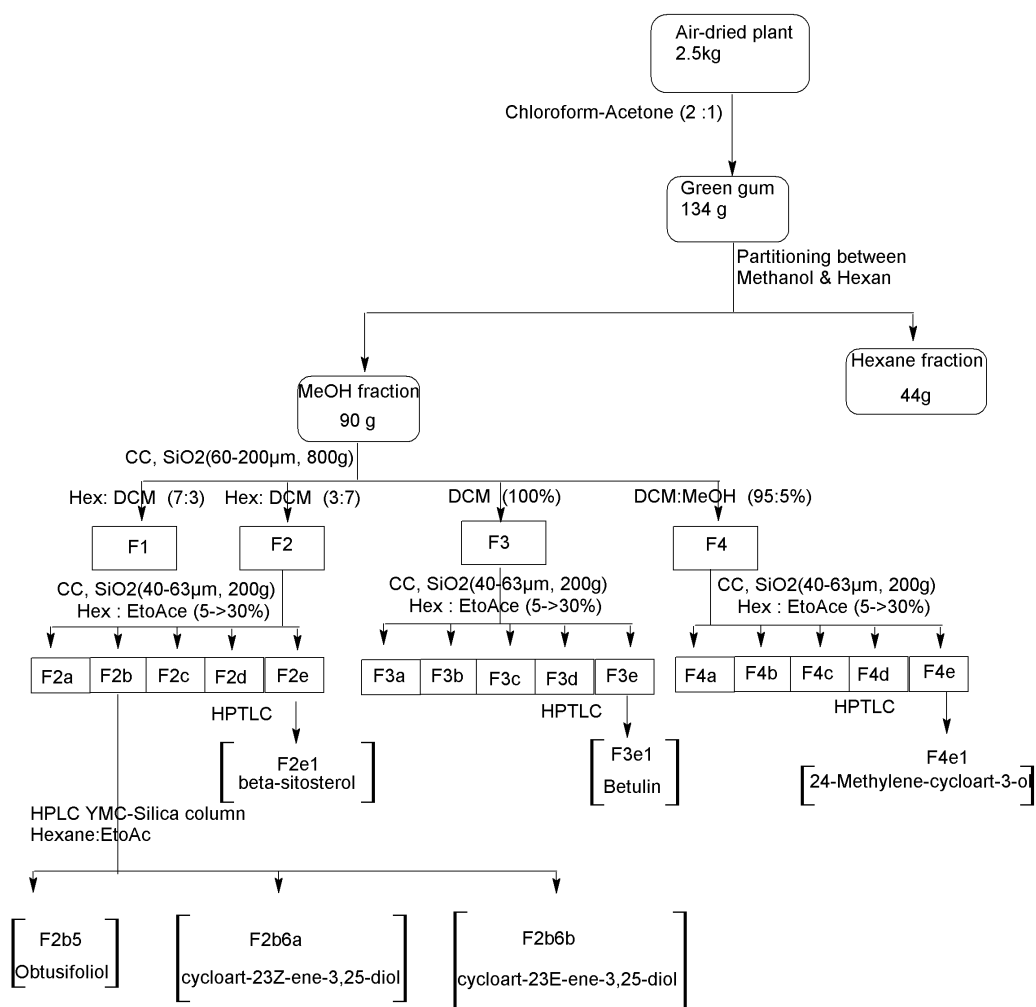


Figure 2. Flowchart representative of extraction and purification processes of triterpenes and steroids from *Euphorbia denticulata*.

cell repository of tissue culture department, Pasture Institute, Iran. Herpes simplex virus type 1 (HSV-1, strain KOS) obtained from Virology Department of Tarbiat Modares University (Tehran, Iran).

Plant material

Plant material was collected from populations growing in Sanandaj (Iran) at the West part of Iran and identified by Dr. Hojatollah Saeedi in the Department of Biology, Faculty of Science, University of Isfahan and a voucher specimen (#19001) is preserved in the herbarium of the University of Isfahan (Iran).

Extraction and isolation

The air-dried plant material (2.5 kg) was macerated with dichloromethane/acetone 2:1

(20L×3) at room temperature for 5 days. Filtration and *in-vacuo* concentration resulted in a green gum (134 g) which was partitioned between methanol and n-hexane. The defatted methanolic extract was concentrated (90 g) and subjected on silica gel CC (60-200 µm, 800 g) eluting with hexane/dichloromethane, 0→100 to give four fractions: Fr.1-Fr.4. Inferred from TLC and ¹H-NMR, Fr.1 (21.3 g) contained alkanes and fats, Fr.2 (15.2 g) containing steroids, and Fr.3 (12.6 g) as well as Fr.4 (18.1 g) triterpenes. Fr.2, Fr.3, and Fr.4 were chromatographed on flash silica gel (40-63 µm, 200 g) using hexane/ethyl acetate, 5→30. Finally steroids and triterpenes were further purified on preparative layer chromatography or high pressure liquid chromatography (HPLC) with YMC-Pak-Sil column (250 × 20 mm) and hexane:ethyl acetate (80:20) as mobile phase to

yield compounds 1-6 (Figure 2).

Betulin: lup-20(29)-ene-3 β ,28-diol (1)

White crystals; MW(g/mol): 442; yield: 0.01%; ¹H-NMR (CDCl₃, 400 MHz): δ_{H} 4.61 (1H, d, $J=2.4$ Hz, H-29a), 4.51 (1H, dd, $J=2.4, 1.6$ Hz, H-29b), 3.73 (1H, dd, $J=10.8, 1.6$ Hz, H-28a), 3.26 (1H, br d, $J=10.8$ Hz, H-28b), 3.12 (1H, dd, $J=11.6, 5.2$ Hz, H-3), 2.32 (1H, dt, $J=10.8, 6.0$ Hz, H-19), 1.61 (3H, s, H-30), 0.95 (3H, s, H-26), 0.91 (3H, s, H-23), 0.90 (3H, s, H-27), 0.75 (3H, s, H-25), 69 (3H, s, H-24); ¹³C-NMR data: see Table 1. EIMS m/z 442 [M], 411, 234, 220, 207, 203, 189, 175, 165, 135, 105, 67, 55, 41.

24-methylene-cycloart-3 β -ol (2)

White crystals; MW(g/mol): 426; yield: 0.001%; ¹H-NMR (CDCl₃, 400 MHz): δ_{H} 4.69 and 4.64 (each 1H, br s, H-31a,b), 3.25 (1H, dd, $J=11.0, 4.0$ Hz, H-3), 1.01 (3H, d, $J=3$ Hz, H-27), 0.99 (3H, d, $J=3$ Hz, H-26), 0.94 (3H, s, 6H: H-18, H-30), 0.88 (3H, s, H-28), 0.86 (3H, d, $J=6$ Hz, H-21) and 0.79 (3H, s, H-29), 0.53 and 0.31 (each 1H, d, $J=4.5$ Hz, H-19a,b); ¹³C-NMR data: see Table 1; EIMS m/z : 426, 379, 365, 353, 300, 286, 203, 175, 161, 147, 121, 107, 95, 81, 69, 55, 41.

Cycloart-23Z-ene-3 β ,25-diol (3)

White crystals; MW(g/mol): 442; yield: 0.00024%; ¹H-NMR (CDCl₃, 400 MHz): δ_{H} 5.62 (2H, br s, H-23,24), 3.30 (1H, dd, $J=4.0, 11.2$ Hz, H-3), 1.33 (6H, s, H-26, H-27), 0.99 (6H, s, H-18, H-30), 0.90 (3H, s, H-28), 0.89 (3H, d, $J=6.4$ Hz, H-21), 0.83 (3H, s, H-29), 0.57, 0.35 (each 1H, d, $J=4.4$ Hz, H-19a, b); ¹³C-NMR data: see Table 1; EIMS m/z : 442 [M]⁺, 427, 424, 381, 363, 355, 315, 313, 302, 269, 175.

Cycloart-23E-ene-3 β ,25-diol (4)

White crystals; MW(g/mol): 442; yield: 0.0002%; ¹H-NMR (CDCl₃, 400 MHz): δ_{H} 5.72 (1H, m, H-23), 5.54 (1H, d, $J=15.6$ Hz, H-24), 3.29 (1H, m, H-3), 1.59 (3H, s, H-26), 1.37 (3H, s, H-27), 1.0 (3H, s, H-29), 0.99 (3H, s, H-18), 0.90 (3H, s, H-30), 0.88 (3H, d, $J=6.4$ Hz, H-21), 0.83 (3H, s, H-28), 0.57 and 0.35 (each 1H, d, $J=4.4$ Hz, H-19a, b); ¹³C-NMR

data: see Table 1. EIMS m/z : 442, 424, 409, 407, 315, 302, 297, 255, 203, 187, 175, 145, 43.

Obtusifoliol: 4,14-dimethyl ergosta-8,24(28)-dien-3 β -ol (5)

White crystals; MW(g/mol): 426; yield: 0.00044%; ¹H-NMR (CDCl₃, 400 MHz): δ_{H} 4.56 and 4.51 (each 1H, br s, H-30a,b), 2.96 (1H, ddd, $J=4.8, 10.0, 14.8$ Hz, H-3), 0.89 (3H, d, $J=6.8$ Hz, H-27), 0.88 (3H, d, $J=6.8$ Hz, H-26), 0.85 (3H, d, $J=6.4$ Hz, H-21), 0.82 (3H, s, H-19), 0.79 (3H, d, $J=6.4$, H-29), 0.74 (3H, s, H-28), 0.55 (3H, s, H-18); ¹³C-NMR data: see Table 1; EIMS m/z : 426, 393, 327, 259, 245, 233, 173, 159, 69.

β -sitosterol: Stigmast-5-en-3 β -ol (6)

White crystals; MW(g/mol): 414; yield: 0.0013%; ¹H-NMR (CDCl₃, 400 MHz): δ_{H} 5.38 (1H, br d, $J=5.2$ Hz, H-6), 3.54 (1H, m, H-3), 1.03 (3H, s, H-19), 0.95 (3H, d, $J=6.4$ Hz, H-21), 0.88 (3H, d, $J=7.6$ Hz, H-24), 0.86 (3H, d, $J=6.8$ Hz, H-27), 0.84 (3H, d, $J=6.8$ Hz, H-26), 0.69 (3H, s, H-18); ¹³C-NMR data: see Table 1; EIMS m/z : 414, 329, 303, 273, 255, 231, 213, 161, 119, 107, 105, 95, 79, 71, 69, 67, 57, 55.

Virus and cells

For cytotoxicity and antiviral assays cells were grown in Dulbecco's Modified Eagle's growth Medium (DMEM; Sigma, USA) supplemented with 2% of fetal bovine serum (FBS; Gibco, Germany), 100 $\mu\text{g}/\text{mL}$ of streptomycin, 100 UI/mL of penicillin and 0.25 $\mu\text{g}/\text{mL}$ amphotericin B (Gibco, Germany) and 0.14% (v/v) sodium bicarbonate. All the cells were cultured at 37°C in a humidified atmosphere supplied with 5% CO₂.

Stock preparations of the herpes simplex virus type 1 (HSV-1, strain KOS), were generated by incubating in Vero cells (75 cm² culture flasks seeded with 350000 cells/mL). After 72 h infection, the cultures were frozen and thawed twice before centrifugation and the resulting supernatant aliquots stored at -70°C. Virus titers were determined by cytopathic effects in Vero cells and were expressed as 50% Tissue Culture Infective Dose (TCID₅₀) per mL (7, 8).

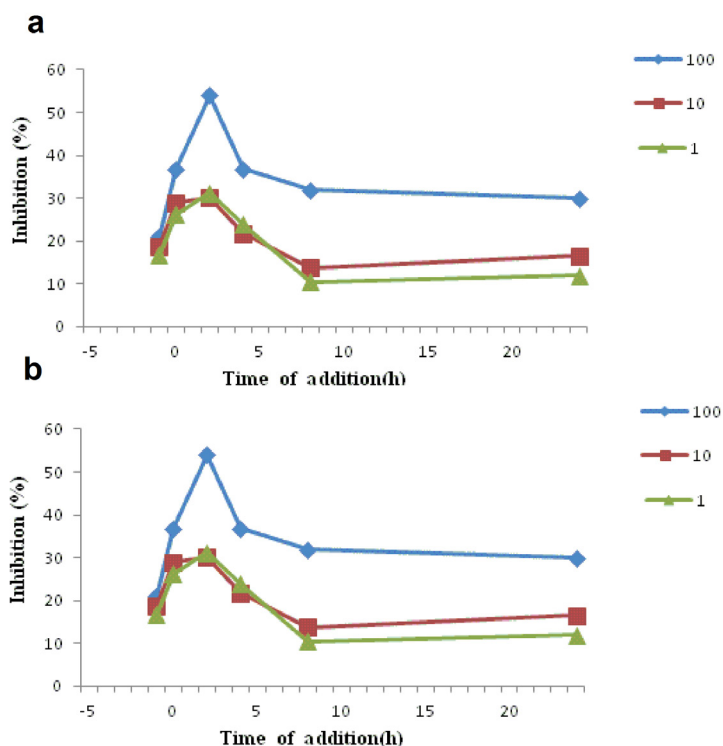


Figure 3. Inhibitory effect of test compounds at different concentrations (1, 10, 100 µg/mL) at various times pre-infection, co-infection and post-infection of herpesvirus (HSV-1) to Vero cells. a) betulin, and b) (3β,23E)-Cycloarta-23-ene-3,25-diol were added with the HSV-1 infected Vero cells at various times period like pre-infection (-1 h), co-infection (0 h) or post-infection (2-24 h). After 3 days of incubation at 37°C, inhibition was evaluated by MTS assay and expressed as the inhibition percentage. Each point represents the mean of three independent experiments.

Cytotoxicity assay

The Vero cells were seeded onto a 96-well plate at a concentration of 3.5×10^5 Vero cells per mL and a volume of 100 µL per well. Following 24 h incubation at 37 °C, a confluent cell monolayer was confirmed and cell media was removed. Test compounds were serially diluted with the culture medium supplemented with 2% serum to reach the different concentrations. Negative control dilution of DMSO at 0.1 % was also included. An aliquot of 100 µL/well of each diluted compound or DMSO was added to the plates in triplicate. After incubation at 37 °C with 5% CO₂ for 3 days, MTS (Cell Titer 96; Promega, USA) was added to each well with a volume of 20 µL. The trays were further incubated for 2 h to allow MTS production. The absorbances were determined with an ELISA reader (Stata

Fax 2100, USA) at a test wavelength of 490 nm. Data were calculated as the percentage of inhibition using the following formula: inhibition % = $[100 - (A_t/A_s) \times 100]$ %. A_t and A_s refer to the absorbances of the test substances and the solvent control, respectively. CC50 values, defined as the concentration of 50% cellular cytotoxicity (CC50) of test compounds (7, 8).

Antiviral assay using MTS method

The antiviral activity of compounds isolated from *E. denticulata* against HSV-1 were evaluated by the MTS method. Vero cells, treated with trypsin, were seeded onto 96-well plates with a concentration of 5×10^3 cells in a volume of 100 µL per well. After incubation at 37 °C with 5% CO₂ for 24 h, when the cell monolayers were confluent, the medium was removed from

Table 1. ¹³C-NMR chemical shifts of the triterpenes and steroids in *Euphorbia denticulata*.

| C | 1 | 2 | 3 | 4 | 5 | 6 | C | 1 | 2 | 3 | 4 | 5 | 6 |
|----|------|------|------|------|-------|-------|----|-------|-------|-------|-------|-------|-------|
| 1 | 38.7 | 32.0 | 32.0 | 32.0 | 35.0 | 37.3 | 16 | 29.2 | 26.5 | 26.4 | 26.4 | 31.0 | 28.2 |
| 2 | 25.2 | 30.4 | 30.4 | 30.4 | 31.1 | 31.7 | 17 | 47.8 | 52.3 | 52.1 | 52.1 | 50.4 | 56.8 |
| 3 | 79.0 | 78.8 | 78.8 | 78.8 | 76.5 | 71.8 | 18 | 47.8 | 18.1 | 18.1 | 18.1 | 15.7 | 12.2 |
| 4 | 38.9 | 40.5 | 40.1 | 40.5 | 39.2 | 42.3 | 19 | 48.7 | 29.9 | 29.9 | 29.9 | 18.7 | 19.4 |
| 5 | 55.3 | 47.1 | 47.1 | 47.1 | 47.0 | 140.8 | 20 | 150.5 | 36.4 | 36.3 | 36.3 | 36.5 | 36.1 |
| 6 | 18.3 | 21.1 | 21.1 | 21.1 | 20.7 | 121.7 | 21 | 29.7 | 18.2 | 18.4 | 18.4 | 18.2 | 18.8 |
| 7 | 34.2 | 28.0 | 28.1 | 28.1 | 28.2 | 31.9 | 22 | 34.0 | 35.6 | 39.4 | 39.4 | 35.0 | 34.0 |
| 8 | 40.9 | 48.0 | 48.0 | 48.0 | 133.6 | 31.9 | 23 | 28.0 | 31.3 | 130.9 | 130.9 | 30.8 | 26.1 |
| 9 | 50.4 | 20.0 | 20.0 | 20.0 | 134.6 | 50.2 | 24 | 15.4 | 157.0 | 134.4 | 134.4 | 156.9 | 45.9 |
| 10 | 37.2 | 25.8 | 26.1 | 26.1 | 36.4 | 36.5 | 25 | 16.0 | 33.8 | 70.9 | 70.9 | 33.8 | 29.2 |
| 11 | 20.8 | 26.0 | 26.0 | 26.0 | 21.8 | 21.1 | 26 | 16.1 | 22.0 | 24.4 | 24.4 | 21.9 | 19.8 |
| 12 | 25.2 | 32.9 | 32.8 | 32.8 | 25.5 | 39.8 | 27 | 14.8 | 19.3 | 24.3 | 24.3 | 22.0 | 19.1 |
| 13 | 37.3 | 45.3 | 45.5 | 45.3 | 44.6 | 42.3 | 28 | 60.6 | 18.3 | 19.3 | 19.3 | 15.1 | 231 |
| 14 | 42.7 | 48.8 | 47.9 | 48.8 | 49.9 | 56.1 | 29 | 109.7 | 14.0 | 14.0 | 14.0 | 105.9 | 12.1 |
| 15 | 27.0 | 35.0 | 35.6 | 35.6 | 31.3 | 24.3 | 30 | 19.1 | 25.5 | 25.4 | 25.4 | 24.4 | |
| | | | | | | | | | | | | | 105.9 |

the wells and 100 μ L of test virus was added and incubated for another 2 h. Different non-cytotoxic concentrations (\leq CC50 values) of test compounds were then added to culture wells in triplicate. The maximum concentration of DMSO (0.1%) was used as a negative control. Aciclovir; (Sigma, USA) was used as a positive control for HSV-1. After incubation at 37°C with 5% CO₂ for 3 days, the MTS test was carried out as previously described. The percentages of protection were calculated as [(A - B) \times 100/(C-B)], where A, B, and C indicate the absorbances of test compounds, virus and cell controls, respectively. Each obtained EC50 value was defined as the effective concentration that reduced the absorbance of infected cells to 50% when compared with cell and virus controls (7, 9).

Time- course anti-virus antanalysis of isolated compounds

Different non-cytotoxic concentrations (\leq CC50 values) of test compounds were added to culture cells in triplicate at different times pre-infection or post-infection. HSV-1 (10 TCID₅₀ per well) was inoculated onto confluent monolayers of Vero cells for 2 h. After 3 days, MTS test and antiviral activity were carried out

as previously described (3).

Statistical analysis

The selectivity index (SI) was determined as the ratio of CC50 to EC50. The statistically different effects of test compounds on the inhibition of HSV -1 were compared with the control group or together using the Student's t-test.

Result and Discussion

Compound 1, was identified as C₃₀H₅₀O₂ on the basis of EI-MS of *m/z* 442, and ¹³C-NMR (BB & DEPT) spectral data indicating six methyls, twelve methylenes, six methines, and six quaternary carbons which two of them were oxygenated. ¹H-NMR revealed six singlet methyls at δ_H 1.61 (H-30), 1.18 (s, H-27), 0.95 (H-26), 0.91 (H-23), 0.90 (H-27), 0.75 (H-25), and 69 (H-24) along with a pair of olefinic protons as part of an exocyclic-methylene group at δ_H 4.66 and 4.56 characteristic for lupane triterpenes (10). Two geminal oxymethylene was also detected at δ_H 3.77 and 3.31 with coupling constant of 10.8 Hz, and one doublet of doublet oxymethine proton at δ_H 3.12 indicating of 3 β -hydroxyl group. Taken together, and confirmed from the literature the

Table 2. Cytotoxicity and anti-HSV-1 activity of compounds isolated from *E.denticulata*.

| Test compounds | CC50 ^a ±SEM | HSV-1 | |
|---|------------------------|------------------------|-----------------|
| | | EC50 ^b ±SEM | SI ^c |
| Betulin | 660.72±0.072 | 84.37±0.018 | 7.83 |
| β ,23E)-Cycloarta-23-ene-3,25-diol 3) | 1±0.2501089.2 | 86.63±0.03 | 12.57 |
| Acyclovir | 128.00±0.215 | ±0.001 3.00 | 42.66 |

^aThe 50% cytotoxic concentration on vero cells in $\mu\text{g/mL}$ (n=3).

^bConcentration of compound ($\mu\text{g/mL}$) producing 50% inhibition of virus included cytopathic effects of three separate experiments.

^cSelectivity index (SI) = CC50/EC50

structure of compound 1 detected as betulin (10, 11). The resonances of compound 2 with EI-MS m/z 426, encompassed thirty-one carbons including seven methyls, twelve methylenes, six methines and six quaternary carbons. ¹H-NMR revealed four singlet methyls, a pair of doublets δ_{H} 0.53 and 0.31 in the upfield area indicative of cyclopropane ring characteristic of cycloartanes. A doublet of doublet proton at δ_{H} 3.25 (dd, J_{ax} =11.0, $J_{\text{ax,eq}}$ =4.0 Hz, H₃), was indicative of 3 β -hydroxyl group, and one pair of olefinic protons δ_{H} 4.64 and 4.69 (each 1H, br s) related to exocyclic terminal methylene. Therefore, comparing to the literature compound 2 determined as 24-methylene-cycloart-3-ol (12). ¹H-NMR of compound 3 showed two downfield singlet methyls at δ_{H} 1.33 (6H, H-26, H-27) together with four other singlet methyls at δ_{H} 0.99 (6H, H-18, H-30), 0.90 (H-28), 0.83 (H-29), and one secondary methyl δ_{H} 0.89 (d, J =6.4 Hz, H-21) along with a pair of doublets in the upfield area with constant coupling of 4.4 Hz at δ_{H} 0.57, 0.35 ppm, characteristic of cycloartane cyclopropane ring. Carbinolic proton at δ_{H} 3.30 was related to 3-hydroxyl group and two vicinal olefinic protons overlapping on each other at δ_{H} 5.62 (1H, m, H-23), and 5.62 (1H, brs, H-24), with low coupling constants indicative of the *cis* geometry. Therefore based on EI-MS m/z 442 and consistency of ¹³C- and ¹H-NMR with other reported data in the literature, compound 3 was identified as cycloart-23Z-ene-3 β ,25-diol (11). Compound 4 with EI-MS of m/z 442 was also identified as cycloart-23E-ene-3 β ,25-diol based on its similarities with compound 3 except for vicinal olefinic protons at δ_{H} 5.72 (1H, m, H-23), and 5.54 (1H, d, J =15.6 Hz, H-24), with large coupling constant which was indicative of *trans* Δ 23-geometry (12,13).

The resonances of compound 5, with EI-MS of m/z 426, encompassed thirty carbons including seven methyls, ten methylenes, seven methines, and six quaternary carbons. ¹H-NMR revealed three singlet methyls, δ_{H} 0.82 (H-19), 0.74 (H-28), and 0.55 (H-18) with a doublet of doublet of doublet proton at δ_{H} 2.96 (ddd, J = 4.8, 10.0, 14.8 Hz), indicative of the hydroxyl group at C-3 of 4 α -methyl steroids, along with one pair of olefinic protons δ_{H} 4.51 (br-s) and 4.56 (br-s) related to exocyclic terminal methylene which were in good agreement with obtusifoliol (14). Compound 6, was also determined as β -sitosterol based on EIMS m/z 414, and ¹H-NMR comprised of two singlet methyls δ_{H} 1.03 (H-19), and 0.69 (H-18), four secondary methyls at δ_{H} 0.95 (d, J = 6.4 Hz, H-21), 0.88 (d, J = 7.6 Hz, H-24), 0.86 (d, J = 6.8 Hz, H-27), and 0.84 (d, J = 6.8 Hz, H-26), with one multiplete oxymethine proton at δ_{H} 3.54 indicative of 3-hydroxy group and one broad doublet olefinic proton at δ_{H} 5.38 (br d, J = 5.2 Hz, H-6) (15).

Assessment of cytotoxicity and anti-HSV-1 activity by MTS assay on Vero cell

The MTS assay was used to determine the toxicity and antiviral activity of the tested agents (9, 16). Betulin and (3 β ,23E)-Cycloarta-23-ene-3,25-diol isolated from *E.denticulata* were investigated for anti-HSV-1 activity, were firstly tested for their cytotoxic effect alone on Vero cells. The results revealed that both compounds have antiviral activity far below the CC50 doses (Table 2). Results presented in Table 2 revealed that the CC50 of betulin and (3 β ,23E)-Cycloarta-23-ene-3,25-diol were 660.718±0.072 and 1089.205±0.250 $\mu\text{g/mL}$, respectively. The results also showed that the rate of cells death increased with increasing the concentration of

the tested compounds.

After studying the cytotoxicity effect of samples on normal Vero cells, the experiments to assess the antiviral effect of samples on HSV-1 were done.

The anti-HV-1 activity tested by MTS assay showed that betulin and (3 β ,23E)-Cycloarta-23-ene-3,25-diol had anti-HSV-1 activity at different dose levels, based on their EC₅₀ value and selectivity index (SI). In comparison with acyclovir as a standard drug with the SI value of 42.66, the SI value of (3 β ,23E)-Cycloarta-23-ene-3,25-diol was 12.57 and betulin was 7.831 (Table 2).

Time- course anti-virus analysis of isolated compounds

In order to investigate the mechanism of how each compound inhibits the infection of HSV-1 a study was conducted to investigate the time-course effect at 1 h before to 24 h after the virus infection and using different concentrations of each compound (1, 10, 100 μ g/mL). Inhibition was evaluated by MTS assay after 3 days of infection and expressed as percentage inhibition. As it is clear from Figure 3a and Figure 3b, the both compounds betulin and (3 β ,23E)-Cycloarta-23-ene-3,25-diol, exhibited the highest inhibition against HSV-1 infection within 2.0 h post-infection which were during the early period of virus replication.

Conclusion

E. denticulata as one of the endemic plants to Iran, could be a new source of 4,4 dimethyl steroids like 24-methylene-cycloart-3-ol, cycloart-23Z-ene-3 β ,25-diol, and cycloart-23E-ene-3 β ,25-diol as well as obtusifoliol as 4 α -methyl steroid and beta-sitosterol as 4-desmethyl steroid which were reported for the first time from this plant. In addition *E. denticulata* could be considered as one on the economic sources of betulin (0.01 % of dry weight of the plant).

Betulin and (3 β ,23E)-Cycloarta-23-ene-3,25-diol showed EC₅₀ value of 84.37 \pm 0.02, and 86.63 \pm 0.03 μ g/mL, and selectivity index (CC₅₀/EC₅₀) values of 7.83, and 12.57,

respectively. It shows that both compounds assert their antiviral activity with ignorable toxicity on vero cells.

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