



Design, Synthesis, and Investigation of Cytotoxic Effects of 5-Hydroxyindole-3-Carboxylic Acid and Ester Derivatives as Potential Anti-breast Cancer Agents

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Abstract

Breast cancer is a deadly disease with a high prevalence rate among females. Despite several treatments, scientists are still engaged in finding less invasive treatments for this disease. The cellular proliferation rate and cell viability survey are critical to assess the drug's effect on both normal and malignant cell populations. Indole derivatives are promising candidates for their cytotoxic effect causing on breast cancer cells; however, they are less toxic on normal cells. This study synthesized 23 novel 5-hydroxyindole-3-carboxylic acids and related esters featuring various linear, cyclic, and primary aromatic amines. The MTT assay indicated the cytotoxicity of all acid and ester derivatives against the MCF-7 cells with no significant cytotoxicity on normal human dermal fibroblasts cells. Compound 5d, an ester derivative possessing a 4-methoxy group, was the most potent compound, with a half-maximal effective concentration of 4.7 μ M. Compounds 5a, 5d, and 5l bearing ester group in their structure demonstrated cytotoxicity values < 10 μ M against the MCF-7 cell line and were safe for advanced screening.

Keywords: 5-Hydroxy Indole, Survivin, Human Breast Cancer Cell Line (MCF-7), MTT Assay

1. Background

Breast cancer is one of the most well-known cancers in females. Due to its effect on the population, this disease presents a critical health problem that requires further molecular investigation to specify its prognosis and treatment (1). Indole derivatives' diverse abilities and unique physicochemical properties make them an ideal scaffold in drug design; accordingly, their footprint can be found in many natural and synthetic therapeutic agents. Tryptophan (i.e., an essential amino acid), serotonin (i.e., a polyfunctional signaling molecule) (2-5), indole-3-acetic acid (i.e., a plant hormone), indomethacin (i.e., a nonsteroidal anti-inflammatory drug), arbidol (i.e., an antiviral) are only a few famous examples. Indoles exhibit diverse pharmacological activities, primarily highlighting their anti-cancer effects, followed by antimicrobial, antimalarial, and antituberculosis effects (Figure 1) (6-10).

In the present study, novel indole derivatives were synthesized and screened for cytotoxicity against MCF-7 (Michigan Cancer Foundation-7) cells using the MTT assay and compared to cisplatin as a commonly used chemotherapeutic agent in treating many tumors (11). Recent investigations have proven that indolic compounds do not adversely affect the kidney and liver (12). Additionally, in one study, indomethacin, containing an indole scaffold, was observed as an effective agent, with an unknown mechanism, to inhibit breast cancer (13).

Another study claimed that indomethacin is a survivin inhibitor (14). Survivin was detected as the smallest member of the inhibitors of apoptosis (IAPs) protein family. Survivin in aggressive tumors by binding to caspase-3 and 7 inhibits apoptosis (15, 16). UC-112 is a small molecule that strongly inhibits cancer cell proliferation and selectively degrades survivin among

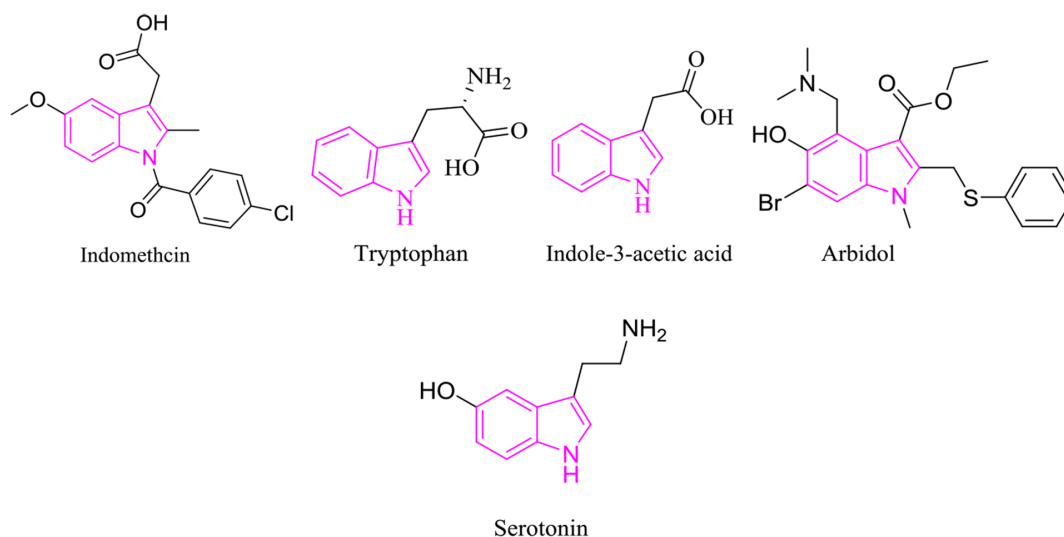


Figure 1. Natural and medicinal compounds bearing indole scaffold

other IAPs. Indole analogs of UC-112 as selective and effective survivin inhibitors indicate that indole is an essential pharmacophore in survivin inhibitory agents (17, 18). On the other hand, protocatechuic acid (PCA), a plant phenolic acid, inhibits proliferation and induces apoptotic effects on breast cancer. Through the survivin inhibition mechanism, apoptosis removes malignant cells without causing damage to normal cells or tissues (19-21). The present study applied molecular hybridization techniques to design new compounds based on indomethacin, UC-112, and PCA structures. In the design of the target molecules (6a-6k), the $-\text{CH}_2\text{-COOH}$ group in indomethacin was replaced by $-\text{COOH}$ or $-\text{COOEt}$ group, and different *N*-substituted indole cores bearing a 5-OH group to resemble PCA were synthesized (Figure 2).

2. Methods

2.1. General

Reagents and chemicals used were purchased from Merck Corporation (USA). Melting points were measured by an Electrothermal 9100 device. Cary 630 FTIR spectrometer recorded infrared spectra in potassium bromide (KBr) in ν_{max} (cm^{-1}). Bruker 400 Avance apparatus, in dimethyl sulfoxide (DMSO)- d_6 at 400.1 and 100 MHz, measured $^1\text{H-NMR}$ and $^{13}\text{C-NMR}$. HPLC 6410 Agilent device measured liquid chromatography (LC) mass spectra and the Costech (Italy) elemental analyzer measured elemental analysis. ChemDraw Professional software (version 16) was used to measure acids and

esters' log P. Discovery studio 4.5 visualizer, a feature-rich molecular modeling application, was used for the final docking image of a 5d small molecule with survivin protein.

2.2. General Experimental Approach

The process consists of three steps, namely synthesis of enamine, 5-hydroxyindoles carboxylic ester synthesis, and basic ester cleavage of 5-hydroxyindoles carboxylic ester (Figure 3). The primary enamine was obtained under ultrasonic irradiation with the catalytic amounts of acetic acid in a round-bottom flask in which ethyl acetoacetate 1 and primary amine 2 were added. Then, enamine 3 was added dropwise to benzoquinone or naphthoquinone 4 in the presence of CaI_2 as a catalyst and reacted for one hour at reflux temperature, and the product (i.e., 5-hydroxyindoles carboxylic ester 5) was obtained, which undergoes subsequent hydrolysis to form 5-hydroxyindoles carboxylic acid 6.

1-Butyl-5-Hydroxy-2-Methyl-1H-Indole-3-Carboxylic Acid (6a)

Cream powder; yield: (85%); mp: 129 - 130°C. IR (ν_{max} , cm^{-1}): 3283, 2959, 1647, 1245, 868. $^1\text{H-NMR}$ (DMSO- d_6 -400MHz) δ_{H} = 8.82 (s, 1H, OH), 7.24 (d, J = 8.4 Hz, 1H, Ar), 7.12 (d, J = 2 Hz, 1H, Ar), 6.64 (dd, J = 8.4, 2 Hz, 1H, Ar), 4.08 (t, J = 7.2 Hz, 2H, CH_2N), 2.54 (s, 3H, Me), 1.63 (m, 2H, CH_2), 1.37 - 1.24 (m, 2H, CH_2), 0.89 (t, J = 7.6 Hz, 3H, Me). $^{13}\text{C-NMR}$ (DMSO- d_6 -100MHz) δ_{C} = 166.83 (C=O), 152.10 (C), 139.57 (C=N), 130.27 (C=N), 126.76 (C), 111.30 (C), 110.50 (CH), 106.59 (CH), 104.75 (CH), 42.71 (CH_2N), 32.15 (CH_2),

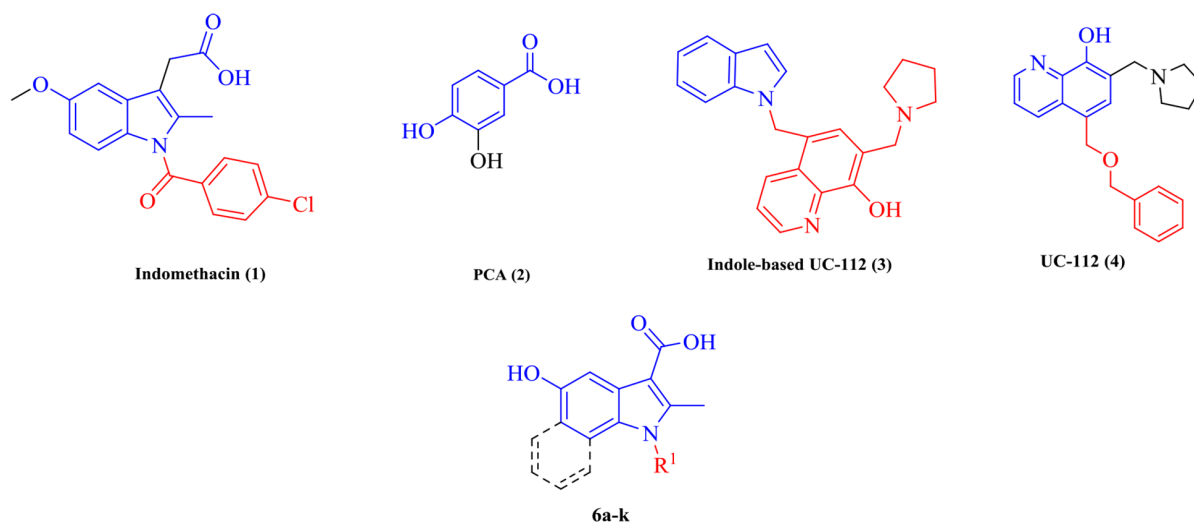


Figure 2. Structure of lead compounds (1-4) and the present study's designed structures (6a-k)

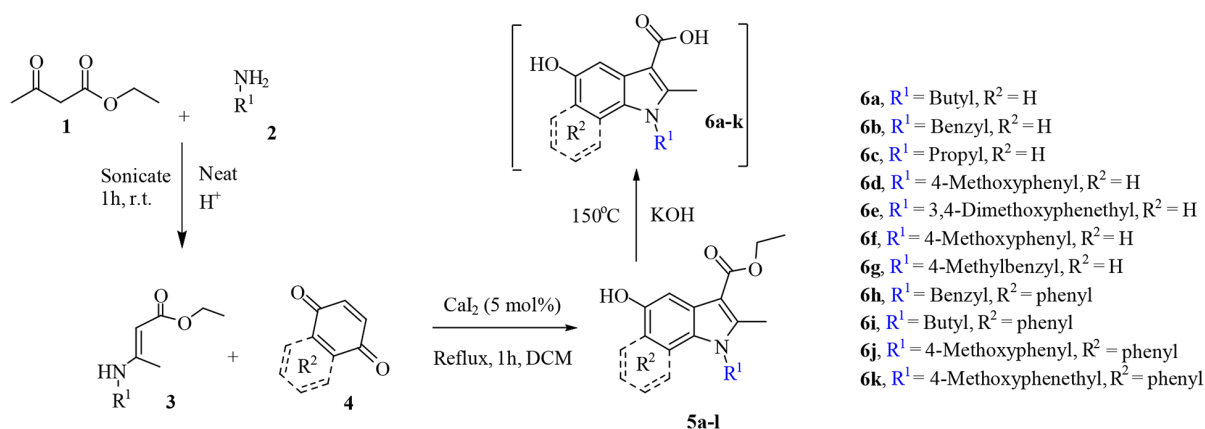


Figure 3. Synthesis of 5-hydroxyindole-carboxylic acid derivatives

20.02 (CH₂), 14.18 (CH₃), 11.88 (CH₃) LC-MS (ESI) m/z 270.12 (M+Na⁺).

72.58; H, 5.37; N, 4.98. Found: C, 72.54; H, 5.39; N, 5.00.

1-Benzyl-5-Hydroxy-2-Methyl-1H-Indole-3-Carboxylic Acid (6b)

White powder; yield: (86%); mp: 143 - 145°C. IR (KBr) (ν_{\max} , cm⁻¹): 3302, 3063, 1666, 1297, 838. ¹H-NMR δ_{H} = 8.99 (s, 1H, OH), 7.43 (s, 1H, Ar), 7.28 (d, *J* = 11.6 Hz, 1H, Ar), 7.12 - 6.9 (m, 5H, Ar), 6.67 (d, *J* = 11.6 Hz, 1H, Ar), 5.38 (s, 2H, CH₂N), 2.66 (s, 3H, Me). ¹³C-NMR δ_{C} = 165.60 (C=O), 153.25 (C), 138.10 (C), 137.60 (C=N), 132.93 (C=N), 131.70 (C=C), 130.81 (C), 129.71 (C), 127.69 (C=C), 126.62 (C=C), 125.60 (C=C), 112.05 (C), 111.26 (C=C), 106.02 (C=C), 103.10 (C=C), 59.23 (CH₂N), 12.28 (CH₃). LC-MS (ESI) m/z 282 (M+H⁺). Anal. Calc. for C₁₇H₁₅NO₃: C,

5-Hydroxy-2-Methyl-1-Propyl-1H-Indole-3-Carboxylic Acid (6c)

Light blue powder; yield: (80%); mp: 120 - 123°C. IR (ν_{\max} , cm⁻¹): 3302, 1602, 1233, 820. ¹H-NMR: δ_{H} = 8.99 (s, 1H, OH), 7.25 (d, *J* = 8.4 Hz, 1H, Ar), 6.63 (dd, *J* = 8.4, 1.6 Hz, 1H, Ar), 4.28 (t, *J* = 7.2 Hz, 2H, CH₂), 2.54 (s, 3H, Me), 1.67 (q, *J* = 7.2 Hz, 2H, CH₂), 0.88 (t, *J* = 7.2 Hz, 3H, Me). ¹³C-NMR δ_{C} = 166.83 (C=O), 152.08 (C), 139.63 (C=N), 130.34 (C=N), 126.75 (C), 111.29 (C), 110.56 (CH), 106.58 (CH), 104.73 (CH), 44.38 (CH₂N), 23.52 (CH₂), 12.94 (CH₃), 11.58 (CH₃). LC-MS (ESI) m/z 235 (M+H⁺).

5-Hydroxy-1-(4-Methoxyphenyl)-2-Methyl-1H-Indole-3-Carboxylic Acid (6d)

Cream powder; yield: (83%); mp: 189 - 191°C. IR (KBr) (ν_{\max} , cm^{-1}): 3313, 2985, 1710, 1646, 1252, 864. $^1\text{H-NMR}$ δ_{H} = 7.47 (d, J = 4 Hz, 1H, Ar), 7.36 - 7.34 (d, J = 8 Hz, 2H, Ar), 7.15 - 7.13 (d, J = 8 Hz, 2H, Ar), 6.74 (d, J = 8 Hz, 1H, Ar), 6.60 (dd, J = 8, 4 Hz, 1H, Ar), 3.85 (s, 3H, OMe), 2.45 (s, 3H, Me). $^{13}\text{C-NMR}$ δ_{C} = 167.30 (C=O), 159.58 (C), 153.28 (C), 145.26 (C), 132.20 (C=N), 129.65 (2C), 129.17 (C=N), 127.94 (C), 115.38 (2CH), 112.13 (C), 111 (CH), 105.97 (CH), 104.52 (CH), 55.93 (CH₃O), 13.24 (CH₃). MS: m/z (%) = 298 (M+H⁺). Anal. Calc. for C₁₇H₁₅NO₄: C, 68.68; H, 5.09; N, 4.71. Found: C, 68.66; H, 5.07; N, 4.75.

1-(3,4-Dimethoxyphenethyl)-5-Hydroxy-2-Methyl-1H-Indole-3-Carboxylic Acid (6e)

Cream powder; yield: (75%); mp: 140 - 142°C. IR (KBr) (ν_{\max} , cm^{-1}): 3280, 2985, 1647, 1244, 831. $^1\text{H-NMR}$ δ_{H} = 7.41 (d, J = 4 Hz, 1H, Ar), 7.33 (d, J = 8.0 Hz, 1H, Ar), 6.83 (d, J = 8.0 Hz, 1H, Ar), 6.66 - 6.63 (m, 2H, Ar), 6.54 (d, 1H, Ar), 4.30 (t, 2H, CH₂), 3.69 (s, 3H, OMe), 3.60 (s, 3H, OMe), 2.89 (t, J = 8.0 Hz, 2H, CH₂), 2.39 (s, 3H, Me). $^{13}\text{C-NMR}$ δ_{C} = 167.28 (C=O), 152.90 (C), 148.90 (C), 147.97 (C), 145.23 (C), 131.14 (C=N), 130.09 (C=N), 128.24 (C), 121.33 (C), 113.18 (CH), 112.25 (CH), 111.58 (CH), 110.93 (CH), 106.07 (CH), 103.15 (CH), 55.97 (CH₃O), 55.70 (CH₃O), 44.90 (CH₂N), 35.06 (CH₂), 11.75 (CH₃). MS: m/z (%) = 356 (M+H⁺). Anal. Calc. for C₂₀H₂₁NO₅: C, 67.59; H, 5.96; N, 3.94. Found: C, 67.60; H, 5.97; N, 3.92.

5-Hydroxy-2-Methyl-1-(4-Methylbenzyl)-1H-Indole-3-Carboxylic Acid (6f)

White powder; yield: (90%); mp: 176 - 178°C. IR (KBr) (ν_{\max} , cm^{-1}): 3298, 2981, 1729, 1658, 1174, 868. $^1\text{H-NMR}$ δ_{H} = 11.90 (s, 1H, COOH), 8.90 (s, 1H, OH), 7.42 (d, J = 2.4 Hz, 1H, Ar), 7.24 (d, J = 8.8, 1H, Ar), 7.11 (d, J = 8 Hz, 2H, Ar), 6.91 - 6.89 (d, J = 8 Hz, 2H, Ar), 6.61 (m, 1H, Ar), 5.36 (s, 2H, CH₂N), 2.63 (s, 3H, Me), 2.24 (s, 3H, Me). $^{13}\text{C-NMR}$ δ_{C} = 167.19 (C=O), 153.06 (C), 145.16 (C), 136.88 (C), 134.84 (C=N), 130.78 (C=N), 129.70 (2 CH), 128.11 (C), 126.64 (2 CH), 111.81 (C), 111.11 (CH), 106.00 (CH), 103.60 (CH), 46.07 (CH₂N), 21.08 (CH₃), 12.23 (CH₃). MS: m/z (%) = 318 (M+23). Anal. Calc. for C₁₈H₁₇NO₃: C, 73.20; H, 5.80; N, 4.74. Found: C, 73.17; H, 5.80; N, 4.77.

1-Cyclohexyl-5-Hydroxy-2-Methyl-1H-Indole-3-Carboxylic Acid (6g)

White powder; yield: (93%); mp: ND. IR (KBr) (ν_{\max} , cm^{-1}): 3268, 2996, 1647, 1244, 864. $^1\text{H-NMR}$ δ_{H} = 11.82 (s, 1H, COOH), 8.85 (s, 1H, OH), 7.48 (d, J = 8.8 Hz, 1H, Ar), 7.43 (d, J = 2.4 Hz, 1H, Ar), 6.61 (dd, J = 8.8, 2.4 Hz, 1H, Ar), 4.34 (m, 1H, NCH, Cy), 2.72 (s, 3H, Me), 1.87 - 1.68 (m, 6H, Cy), 1.49 - 1.37 (m, 4H, Cy, Me). $^{13}\text{C-NMR}$ δ_{C} = 165.73 (C=O), 152.57 (C), 145.03 (C=N), 129.11 (C=N), 128.59 (C), 113.34 (C), 111.47 (CH), 106.14 (CH), 102.57 (CH), 59.11 (CHN), 30.82 (2 CH₂), 26.13 (2 CH₂),

25.19 (CH₂), 12.57 (CH₃). MS: m/z (%) = 273 (M+H⁺). Anal. Calc. for C₁₆H₁₉NO₃: C, 70.31; H, 7.01; N, 5.12. Found: C, 70.31; H, 7.03; N, 5.10.

1-Benzyl-5-Hydroxy-2-Methyl-1H-Benzo[g]Indole-3-Carboxylic Acid (6h)

White powder; yield: (74%); mp: 179 - 181°C. IR (KBr) (ν_{\max} , cm^{-1}): 3261, 2978, 1662, 1233, 864. $^1\text{H-NMR}$ δ_{H} = 8.25-8.22 (m, 1H, Ar), 8.07-8.03 (dd, J = 12.0, 8 Hz, 2H, Ar), 7.33 - 7.21 (m, 6H, Ar), 7.03-7.01 (m, 1H, Ar), 5.83 (s, 2H, CH₂N), 2.78 (s, 3H, Me). $^{13}\text{C-NMR}$ δ_{C} = 173.96 (C=O), 169.10 (C), 148.45 (C=N), 141.60 (C=N), 138.22 (C), 129.33 (CH), 127.57 (C), 126.01 (CH), 125.99 (CH), 125.82 (2CH), 123.81 (CH), 123.49 (CH), 123.40 (CH), 122.45 (CH), 122.22 (C), 120.60 (C), 109.12 (C), 103.22 (C), 48.95 (CH₂N), 11.83 (CH₃). MS: m/z (%) = 332 (M+H⁺). Anal. Calc. for C₂₁H₁₇NO₃: C, 76.12; H, 5.17; N, 4.23. Found: C, 76.14; H, 5.15; N, 4.14.

1-Butyl-5-Hydroxy-2-Methyl-1H-Benzo[g]Indole-3-Carboxylic Acid (6i)

Blue powder; yield: (89%); mp: 200 - 202°C. IR (KBr) (ν_{\max} , cm^{-1}): 3298, 1647, 1248, 864. $^1\text{H-NMR}$ δ_{H} = 12.07 (s, 1H, COOH), 9.71 (s, 1H, OH), 8.28-8.25 (dd, J = 8.0, 4.0 Hz, 2H, Ar), 7.76 (s, 1H, Ar), 7.61 - 7.51 (dd, J = 15.6, 7.2 Hz, 1H, Ar), 7.44 - 7.42 (dd, J = 15.2, 8.0 Hz, 1H, Ar), 4.52 (t, J = 7.6 Hz, 2H, CH₂N), 2.77 (s, 3H, Me), 1.82-1.77 (m, 2H, CH₂), 1.48 - 1.38 (m, 2H, CH₂), 0.96 (t, J = 7.2 Hz, 3H, CH₃). $^{13}\text{C-NMR}$ δ_{C} = 167.36 (C=O), 148.48 (C), 143.09 (C=N), 126.71 (C=N), 125.28 (C), 123.87 (C), 123.38 (C), 123.23 (C), 122.84 (CH), 122.62 (CH), 120.63 (CH), 104.65 (CH), 102.24 (CH), 45.44 (CH₂N), 31.92 (CH₂), 19.74 (CH₂), 14.15 (CH₃), 12.12 (CH₃). MS: m/z (%) = 298 (M+H⁺). Anal. Calc. for C₁₈H₁₉NO₃: C, 72.71; H, 6.44; N, 4.71. Found: C, 72.70; H, 6.43; N, 4.73.

5-Hydroxy-1-(4-Methoxyphenyl)-2-Methyl-1H-Benzo[g]Indole-3-Carboxylic Acid (6j)

Pale purple powder; yield: (91%); mp: 210 - 212°C. IR (KBr) (ν_{\max} , cm^{-1}): 3268, 2963, 1647, 1185, 872. $^1\text{H-NMR}$ δ_{H} = 9.03 (d, J = 4 Hz, 1H, Ar), 8.6 (s, 1H, Ar), 8.25 (d, J = 8 Hz, 2H, Ar), 8.12-8.04 (m, 3H, Ar), 8.0-7.95 (m, 1H, Ar), 7.77-7.75 (m, 1H, Ar), 4.74 (s, 3H, OMe), 3.24 (s, 3H, Me). $^{13}\text{C-NMR}$ δ_{C} = 167.60 (C=O), 160.12 (C), 148.70 (C), 143.52 (C=N), 132.09 (C=N), 130.36 (2 CH), 125.85 (CH), 125.02 (C), 124.71 (C), 123.69 (C), 123.37 (C), 122.86 (C), 122.52 (CH), 120.00 (CH), 120.03 (CH), 115.82 (2 CH), 102.15 (CH), 56.02 (CH₃O), 13.03 (CH₃). MS: m/z (%) = 348 (M+H⁺). Anal. Calc. for C₂₁H₁₇NO₄: C, 72.61; H, 4.93; N, 4.03. Found: C, 72.59; H, 4.92; N, 4.06.

5-Hydroxy-1-(4-Methoxyphenethyl)-2-Methyl-1H-Benzo[g]Indole-3-Carboxylic Acid (6k)

Gold powder; yield: (79%); mp: 141 - 143°C. IR (KBr) (ν_{\max} , cm^{-1}): 3239, 1666, 1237, 861. $^1\text{H-NMR}$ δ_{H} = 8.43 (d, J = 8.4 Hz,

¹H, Ar), 8.31 (d, *J* = 8.4 Hz, 1H, Ar), 7.83 (s, 1H, Ar), 7.65 (m, 1H, Ar), 7.46 (m, 1H, Ar), 7.09 (d, *J* = 8.8 Hz, 2H, Ar), 6.89 (d, *J* = 8.8 Hz, 2H, Ar), 4.73 (t, *J* = 7.2 Hz, 2H, CH₂N), 3.72 (s, 3H, OMe), 3.09 (t, *J* = 7.2 Hz, 2H, CH₂N), 2.53 (s, 3H, Me). ¹³C-NMR δ_C = 167.59 (C=O), 158.55 (C), 148.45 (C), 143.12 (C=N), 130.32 (C=N), 130.22 (2 CH), 126.69 (C), 125.56 (C), 123.97 (C), 123.41 (C), 122.96 (CH), 122.77 (CH), 122.64 (CH), 120.65 (C), 114.46 (2 CH), 102.54 (CH), 55.51 (CH₃O), 47.33 (CH₂N), 34.91 (CH₂), 14.97 (CH₃), 11.81 (CH₃). MS: *m/z* (%) = 376 (M+H⁺). Anal. Calc. for C₂₃H₂₁NO₄: C, 73.58; H, 5.64; N, 3.73. Found: C, 73.60; H, 5.62; N, 3.76.

2.3. Cytotoxicity Evaluation

2.3.1. Cell Culture

The cytotoxicity of 5-hydroxyindole-carboxylic acids and esters' derivatives were determined against breast adenocarcinoma, MCF-7, and the results were compared to fibroblast normal cell line, human dermal fibroblasts (HDF). The cell lines were obtained from the Pasture Institute of Iran. The cells were placed on the cell culture flasks with RPMI, filled out with 10% fetal bovine serum and 1% antibiotic solution (10 mg of streptomycin and 10,000 units of penicillin in 0.9% NaCl) in a wet atmosphere of 5% CO₂ at room temperature in 37°C.

2.3.2. Cell Proliferation Assay

MTT (3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyl tetrazolium bromide) approach is a standard assay to evaluate the inhibitory effect of the compounds (22). The base of the colorimetric assay is the decrease of the yellow color MTT to purple crystals by metabolically smart cells. The live cells have NAD(P)H-dependent oxidoreductase enzymes, decreasing the MTT to purple formazan crystal.

For the MTT assay, the cells (10000 cells/well) were cultured in 96-well plates in media and incubated for 24 hours. Then, the solution of each synthesized compound was poured into the wells in three repetitions and six concentrations. The plates were incubated for 24 hours, the culture medium was drained from the wells, and 100 μL of MTT solution in phosphate-buffered saline (0.5 mg/mL) was added to each well and incubated for 4 hours. Then, 100 μL of formazan solvent containing 10% sodium dodecyl sulfate instead of DMSO (23) was added to each well and incubated for 2 hours. Finally, relative cell viability was calculated based on a colorimetric method using a Cell Imaging Multi-Mode Microplate Reader (Biotek, Cytation 3, USA) and recording the absorbance at 570 nm (Figure 4) (24).

2.4. Molecular Modeling Study

AutoDock software (version 4) was used to perform docking simulations to describe the interactions between

synthesized compounds and survivin protein. A stable conformer of the synthesized compounds was docked into the binding pocket of the biological target, survivin (PDB code: 3UIH). The crystal structure was optimized by separating all water molecules from the protein. Kollman charges and polar hydrogens were also added to the protein structure. The output file was converted to PDBQT format using AutoDock 4. In the next step, the energy of the synthesized ligands was minimized through the MM⁺ method using HyperChem 8.0 software. The ligands were converted to PDBQT using AutoDock 4. Then, a grid box of 20-20-20 Å around the protein's active site was constructed (for more efficacy, the residues with atoms bigger than 7.5 Å were removed from the grid box), and the Lamarckian genetic search algorithm with total runs of 50 was applied. Finally, the most energetically stable conformation was selected. To explain the good potency of 5d that was observed, a molecular modeling study was used. Figure 5 illustrates the complex of human survival (PDB entry: 3UIH) and the results.

3. Results and Discussion

3.1. Chemistry

3.1.1. Experimental Procedure to Achieve Compounds 5

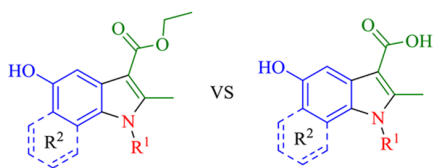
The general procedure of accessing 5-hydroxyindole 5d: Enamine 3d that achieved with the reaction between the ethyl acetoacetate 1 (3.0 mmol) and 4-methoxy phenylamine 2 (3.0 mmol) under catalytic acid, added dropwise during 15 minutes to a solution of benzoquinone (3 mmol) in CH₂Cl₂ (10 mL, reflux), with a CaI₂ (5 mol%) as a catalyst. The reaction was stirred for an additional hour, and then the mixture was cooled for 3-4 hours. The solid mixture was filtered off and then washed with CH₂Cl₂ (10 mL) (25).

3.1.2. General Procedure for Preparation of Compounds 6

To obtain 5-hydroxyindole carboxylic acid 6d, a mixture of indole 5d (0.01 mmol) was melted at 150°C with potassium hydroxide (0.1 mmol) for about an hour and then cooled. Afterward, the least amount of water was added to dissolve the mixture. Glacial acetic acid was added dropwise to the solution to precipitate the almost pure product. Analytical samples were prepared by crystallization from dilute ethanol.

3.2. Biological Evaluations

With the MTT assay, the cytotoxic effects of compounds, which were selected via docking studies, were evaluated against the MCF-7 breast cancer cell line. Furthermore, the cytotoxicity was assessed on



Structure	Compound	MCF-7	HDF
R¹	R²	Acid vs. Ester	EC ₅₀ (μM) SI EC ₅₀ (μM)
nBu	-	Ester (5a)	7.8 ± 0.078 ≥ 3.2 ≥ 25
	-	Acid (6a)	≥ 100 - ≥ 100
Bn	-	Ester (5b)	≥ 25 - ≥ 25
	-	Acid (6b)	≥ 100 - ≥ 100
Pr	-	Ester (5c)	18.8 ± 0.9 ≥ 1.32 ≥ 25
	-	Acid (6c)	≥ 100 - ≥ 100
<i>p</i>-MeOPh	-	Ester (5d)	4.7 ± 0.7 ≥ 5.3 ≥ 25
	-	Acid (6d)	≥ 100 - ≥ 100
3,4-diMeOPhEt	-	Ester (5e)	33 ± 0.2 ≥ 3 ≥ 100
	-	Acid (6e)	19 ± 0.11 ≥ 5.2 ≥ 100
<i>p</i>-MeBn	-	Ester (5f)	≥ 25 - ≥ 25
	-	Acid (6f)	≥ 100 - ≥ 100
Cyclohexyl	-	Ester (5g)	≥ 25 - ≥ 25
	-	Acid (6g)	10.25 ± 1.6 ≥ 9.7 ≥ 100
Bn	Ph	Ester (5h)	≥ 25 - ≥ 25
	Ph	Acid (6h)	11.5 ± 0.6 ≥ 8.6 ≥ 100
nBu	Ph	Ester (5i)	14.7 ± 1.06 ≥ 1.7 ≥ 25
	Ph	Acid (6i)	≥ 100 - ≥ 100
<i>p</i>-MeOPh	Ph	Ester (5j)	≥ 25 - ≥ 25
	Ph	Acid (6j)	6.9 ± 0.9 ≥ 14.49 ≥ 100
4-MeOPhEt	Ph	Ester (5k)	≥ 25 - ≥ 25
	Ph	Acid (6k)	30 ± 0.9 ≥ 3.3 ≥ 100
3,4-diMeOPhEt	Ph	Ester (5l)	5.6 ± 0.06 ≥ 4.4 ≥ 25
	Ph	-	- - -
Cisplatin		67 ± 1.3	1 67.2 ± .85

Figure 4. Half-maximal effective concentration in MCF-7 and human dermal fibroblasts for target indole esters (5) and acids (6). HDF, human dermal fibroblasts; EC₅₀, half-maximal effective concentration; SI, selectivity index.

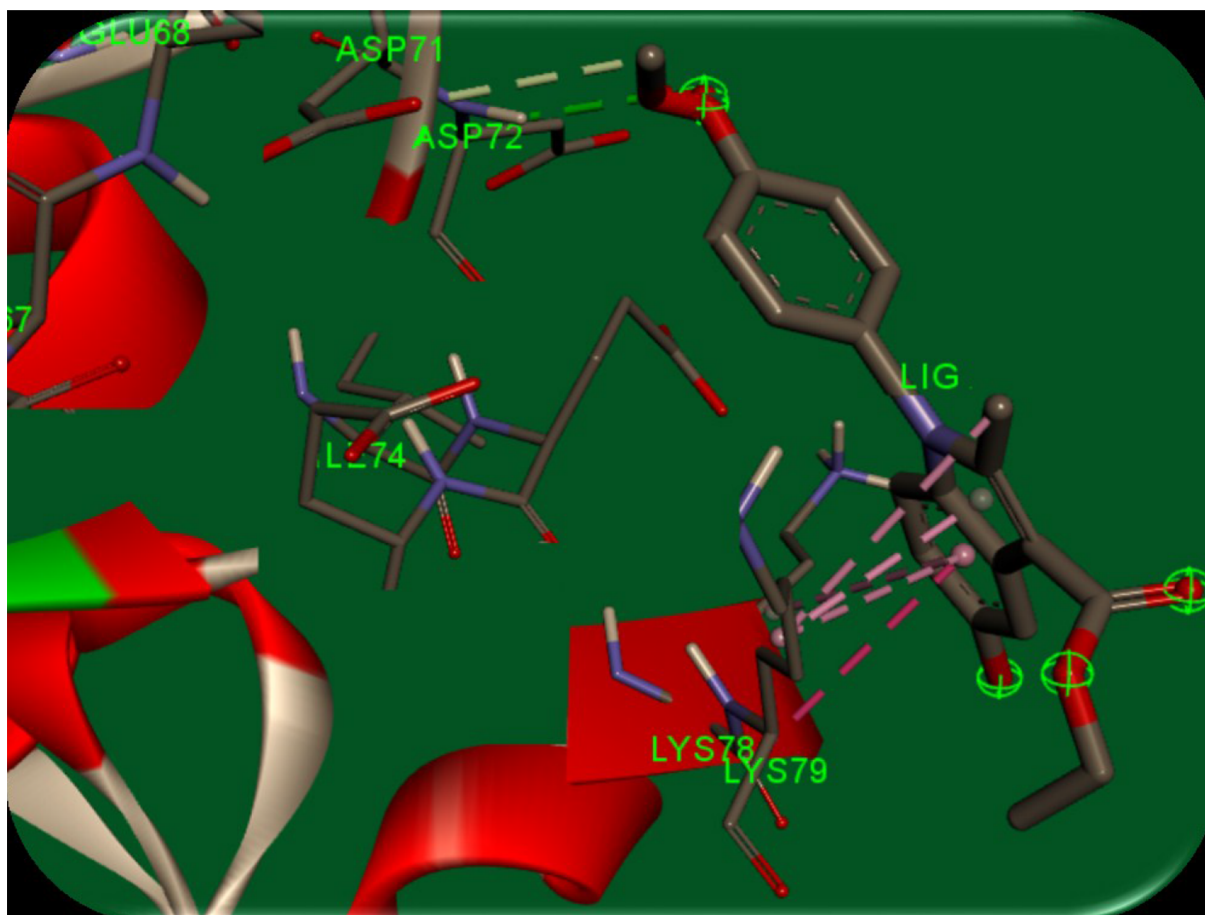


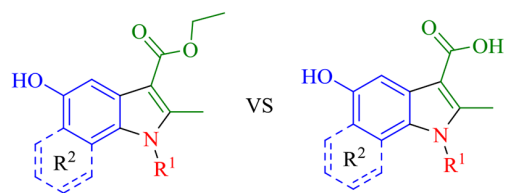
Figure 5. Binding modes of ethyl 5-hydroxy-1-(4-methoxyphenyl)-2-methyl-1H-indole-3-carboxylate (5d) in the active site of survivin protein (PDB entry: 3UIH)

the normal HDF line. The antiproliferative activities of selected compounds were compared to cisplatin at six concentrations within 31.25 - 1000 μM on MCF-7 and normal fibroblast cells. As depicted in Figure 4, all the selected compounds showed cytotoxic effects on the MCF-7 cell line, compared to cisplatin. 5a, 5d, 5l, and 6j derivatives from esters and acids showed the highest potency against the MCF-7 cell line among the other compounds. According to the structural similarity, combinations, including the *p*-methoxy phenyl group, substituted in R² generally had better potency than the benzyl or *p*-methyl benzyl group. Moreover, the results revealed that electron-donating groups in the *para* part of the indole ring had better cytotoxic activity than compounds without them. Because, on the one hand, the solubility of new synthetic compounds was very important in the cancer biological test mediums, and on the other hand, the amount of hydrophilicity or hydrophobicity of compounds must be evaluated, log P

was measured with the ChemDraw software to compare acid and ester products and then indicated that not only the solubility improved but also the amount of log P was within the allowed range of Lipinski's rule of five (log P < 5) (Figure 6).

3.3. Docking Study

It is speculated that the *para* methoxy group in the 5d ligand has hydrogen bondings between Asp71 and Asp72 residues. Furthermore, the 5-hydroxy indole group and the carboxylate group form π - π stacking interaction between Lys79 residue in the active site. Green bonds indicate hydrogen bonds between Asp71 (with 3.50 Å distance) and Asp72 (with 2.11 Å distance) amino acids with *p*-methoxy phenyl in 5d ligand, and pink bond is a π - π stacking interaction between the 5d indole and residue Lys79 with 3.58 Å distance. The results indicated the minimal cytotoxic effect of the compounds at a concentration of 100 μM against normal HDF cells. A molecular modeling



Ester product				Acid product		
Entry	(R ¹)	(R ²)	Log P	Entry	Log P	Yield ^b (%)
5a	Bu	-	3.16	6a	2.56	85
5b	Bn	-	3.65	6b	3.05	86
5c	Pr	-	2.74	6c	2.14	80
5d	<i>P</i> -MeOPh	-	3.45	6d	2.85	83
5e	3,4-diMeOPhEt	-	3.68	6e	3.07	75
5f	<i>P</i> -MeBn	-	4.14	6f	3.53	90
5g	Cy	-	3.46	6g	2.86	93
5h	Bn	<i>Ph</i>	4.65	6h	4.04	74
5i	Bu	<i>Ph</i>	4.15	6i	3.55	89
5j	<i>P</i> -MeOPh	<i>Ph</i>	4.45	6j	3.85	91
5k	4-MeOPhEt	<i>Ph</i>	4.8	6k	4.2	79
5l	3,4-MeOPhEt	<i>Ph</i>	4.67	-	-	-

Figure 6. Comparison of log P for ester and acid derivatives of 5-hydroxyindole derivatives

study on compound 5d showed proper orientation with the survivin protein active site (Figure 5).

4. Conclusions

In summary, various *N*-substituted 5-hydroxyindole-3-carboxylic acids and esters were designed and synthesized based on targeted structural modification of the UC-112 and PCA lead compounds. The structure-activity relationships were investigated by changing the amine groups inserted in the indole ring, changing the ester bond to the acid bond. Ultimately, their log P was checked to ensure that the hydrophobic properties did not decrease much. These small molecules indicated a significant cytotoxic effect on the MCF-7 cell line and low toxicity on normal fibroblast cells. The aforementioned results showed a good correlation with the docking study. The most potent investigated compound, 5d, was shown to exhibit well drug-like properties. These novel small-molecule inhibitors of survivin are currently being developed to produce more selective and effective survivin inhibitors. However, further studies are needed to find the exact mechanism and possible off-targets.

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Footnotes

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References

- Comsa S, Cimpean AM, Raica M. The Story of MCF-7 Breast Cancer Cell Line: 40 years of Experience in Research. *Anticancer Res.* 2015;**35**(6):3147-54. [PubMed ID: 26026074].
- Hornedo-Ortega R, Da Costa G, Cerezo AB, Troncoso AM, Richard T, Garcia-Parrilla MC. In Vitro Effects of Serotonin, Melatonin, and Other Related Indole Compounds on Amyloid-beta Kinetics and Neuroprotection. *Mol Nutr Food Res.* 2018;**62**(3):1700383. [PubMed ID: 29131485]. <https://doi.org/10.1002/mnfr.201700383>.
- Palmieri A, Petrini M. Tryptophol and derivatives: natural occurrence and applications to the synthesis of bioactive compounds. *Nat Prod Rep.* 2019;**36**(3):490-530. [PubMed ID: 30230504]. <https://doi.org/10.1039/c8np000032h>.
- Gershon MD. 5-Hydroxytryptamine (serotonin) in the gastrointestinal tract. *Curr Opin Endocrinol Diabetes Obes.* 2013;**20**(1):14-21. [PubMed ID: 23222853]. [PubMed Central ID: PMC3708472]. <https://doi.org/10.1097/MED.0b013e32835bc703>.
- Koopman N, Katsavelis D, Hove AST, Brul S, Jonge WJ, Seppen J. The Multifaceted Role of Serotonin in Intestinal Homeostasis. *Int J Mol Sci.* 2021;**22**(17):9487. [PubMed ID: 34502396]. [PubMed Central ID: PMC8431144]. <https://doi.org/10.3390/ijms22179487>.
- Dadashpour S, Emami S. Indole in the target-based design of anticancer agents: A versatile scaffold with diverse mechanisms. *Eur J Med Chem.* 2018;**150**:9-29. [PubMed ID: 29505935]. <https://doi.org/10.1016/j.ejmech.2018.02.065>.
- Sharma SK, Kumar P, Narasimhan B, Ramasamy K, Mani V, Mishra RK, et al. Synthesis, antimicrobial, anticancer evaluation and QSAR studies of 6-methyl-4-[1-(2-substituted-phenylamino-acetyl)-1H-indol-3-yl]-2-oxo/thioxo-1,2,3,4-tetrahydropyrimidine-5-carboxylic acid ethyl esters. *Eur J Med Chem.* 2012;**48**:16-25. [PubMed ID: 22154835]. <https://doi.org/10.1016/j.ejmech.2011.11.028>.
- Sravanthi TV, Manju SL. Indoles - A promising scaffold for drug development. *Eur J Pharm Sci.* 2016;**91**:1-10. [PubMed ID: 27237590]. <https://doi.org/10.1016/j.ejps.2016.05.025>.
- Mehndiratta S, Hsieh YL, Liu YM, Wang AW, Lee HY, Liang LY, et al. Indole-3-ethylsulfamoylphenylacrylamides: potent histone deacetylase inhibitors with anti-inflammatory activity. *Eur J Med Chem.* 2014;**85**:468-79. [PubMed ID: 25113875]. <https://doi.org/10.1016/j.ejmech.2014.08.020>.
- Liew LPP, Fleming JM, Longeon A, Mouray E, Florent I, Bourguet-Kondracki ML, et al. Synthesis of 1-indolyl substituted β -carboline natural products and discovery of antimalarial and cytotoxic activities. *Tetrahedron.* 2014;**70**(33):4910-20. <https://doi.org/10.1016/j.tet.2014.05.068>.
- Shona S, Essawy AW, Zaki SM, El-Galil TIA. Effect of cisplatin on the liver of the adult albino rat and the possible protective role of vitamin E (Histological and ultrastructural study). *Anatomy & Physiology.* 2012;**2**(3):102.
- Razak S, Afsar T, Bibi N, Abulmeaty M, Qamar W, Almajwal A, et al. Molecular docking, pharmacokinetic studies, and in vivo pharmacological study of indole derivative 2-(5-methoxy-2-methyl-1H-indole-3-yl)-N'-(E)-(3-nitrophenyl)methylidene] acetohydrazide as a promising chemoprotective agent against cisplatin induced organ damage. *Sci Rep.* 2021;**11**(1):6245. [PubMed ID: 33737575]. [PubMed Central ID: PMC7973782]. <https://doi.org/10.1038/s41598-021-84748-y>.

13. Planchon P, Veber N, Magnien V, Prevost G, Starzec AB, Israel L. Evidence for separate mechanisms of antiproliferative action of indomethacin and prostaglandin on MCF-7 breast cancer cells. *Life Sci*. 1995;**57**(12):1233–40. [PubMed ID: 7674812]. [https://doi.org/10.1016/0024-3205\(95\)02069-u](https://doi.org/10.1016/0024-3205(95)02069-u).
14. Chiou SK, Tanigawa T, Akahoshi T, Abdelkarim B, Jones MK, Tarnawski AS. Survivin: a novel target for indomethacin-induced gastric injury. *Gastroenterology*. 2005;**128**(1):63–73. [PubMed ID: 1563124]. <https://doi.org/10.1053/j.gastro.2004.10.008>.
15. Shin S, Sung BJ, Cho YS, Kim HJ, Ha NC, Hwang JI, et al. An anti-apoptotic protein human survivin is a direct inhibitor of caspase-3 and -7. *Biochemistry*. 2001;**40**(4):1117–23. [PubMed ID: 11170436]. <https://doi.org/10.1021/bi001603q>.
16. O'Connor DS, Grossman D, Plescia J, Li F, Zhang H, Villa A, et al. Regulation of apoptosis at cell division by p34cdc2 phosphorylation of survivin. *Proc Natl Acad Sci U S A*. 2000;**97**(24):13103–7. [PubMed ID: 11069302]. [PubMed Central ID: PMC27185]. <https://doi.org/10.1073/pnas.240390697>.
17. Xiao M, Wang J, Lin Z, Lu Y, Li Z, White SW, et al. Design, Synthesis and Structure-Activity Relationship Studies of Novel Survivin Inhibitors with Potent Anti-Proliferative Properties. *PLoS One*. 2015;**10**(6):e0129807. [PubMed ID: 26070194]. [PubMed Central ID: PMC4466525]. <https://doi.org/10.1371/journal.pone.0129807>.
18. Wang Q, Arnst KE, Xue Y, Lei ZN, Ma D, Chen ZS, et al. Synthesis and biological evaluation of indole-based UC-112 analogs as potent and selective survivin inhibitors. *Eur J Med Chem*. 2018;**149**:211–24. [PubMed ID: 29501942]. [PubMed Central ID: PMC5849576]. <https://doi.org/10.1016/j.ejmech.2018.02.045>.
19. Abotaleb M, Liskova A, Kubatka P, Busselberg D. Therapeutic Potential of Plant Phenolic Acids in the Treatment of Cancer. *Biomolecules*. 2020;**10**(2):221. [PubMed ID: 32028623]. [PubMed Central ID: PMC7072661]. <https://doi.org/10.3390/biom10020221>.
20. Özmen A, Değirmenci EH. In vitro anti-cancer and apoptotic activity of edible mushroom *Lepista nuda* (Bull.) Cooke on leukemia and breast cancer compared with protocatechuic acid, paclitaxel and doxorubicin. *Indian J Exp Biol*. 2021;**59**(3):147–52.
21. Allam RM, El-Halawany AM, Al-Abd AM. Chemo-sensitizing agents from natural origin for colorectal cancer: Pharmacodynamic and cellular pharmacokinetics approaches. In: Cho CH, Hu T, editors. *Drug Resistance in Colorectal Cancer: Molecular Mechanisms and Therapeutic Strategies*. London: Academic Press; 2020. p. 93–116. <https://doi.org/10.1016/B978-0-12-819937-4.00006-6>.
22. Mosmann T. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *J Immunol Methods*. 1983;**65**(1-2):55–63. [PubMed ID: 6606682]. [https://doi.org/10.1016/0022-1759\(83\)90303-4](https://doi.org/10.1016/0022-1759(83)90303-4).
23. Septisetyani EP, Ningrum RA, Romadhani Y, Wisnuwardhani PH, Santoso A. Optimization of Sodium Dodecyl Sulphate as a Formazan Solvent and Comparison of 3-(4,5-Dimethylthiazo-2-yl)-2,5-Diphenyltetrazolium Bromide (Mtt) Assay with Wst-1 Assay in MCF-7 Cells. *Indonesian J Pharm*. 2014;**25**(4):245. <https://doi.org/10.14499/indonesianjpharm25iss4pp245>.
24. Mokhtari S, Mosaddegh M, Hamzeloo Moghadam M, Soleymani Z, Ghafari S, Kobarfard F. Synthesis and cytotoxic evaluation of novel 3-substituted derivatives of 2-indolinone. *Iran J Pharm Res*. 2012;**11**(2):411–21. [PubMed ID: 24250465]. [PubMed Central ID: PMC3832156].
25. Teymori A, Sedaghat A, Kobarfard F. Ca-mediated Nenitzescu synthesis of 5-hydroxyindoles. *Chem Pap*. 2022;**77**(4):1791–5. <https://doi.org/10.1007/s11696-022-02463-y>.