Published Online: 2024 October 13 **Research Article Research Article** 



# Dendrosomal Curcumin Showed Cytotoxic Effects on Breast Cancer Cell Line by Inducing Mitochondrial Apoptosis Pathway and Cell Division Arrest

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Received 2024 July 27: Revised 2024 August 24: Accepted 2024 September 1

# Abstract

Background: Mutations in the p53 gene have been linked to the initiation and progression of breast cancer, as well as resistance to chemotherapy. Therefore, the development of novel treatment approaches is essential to combat this disease.

Objectives: This study aimed to evaluate the effects of dendrosomal curcumin (DNC) on the breast cancer cell line MDA-MB231.

Methods: MDA-MB231 cells were treated with 20 μM DNC, and the apoptosis rate and cell proliferation cycles were assessed using flow cytometry. Additionally, after RNA extraction and cDNA synthesis, the expression levels of Lnc-DANCR, EZH2, Noxa, bcl-2, bax, PUMA, p21, and p53 genes were analyzed using RT-PCR. Protein expression levels of P53, P21, Bcl-2, and Bax were evaluated through western blotting.

Results: Dendrosomal curcumin induced apoptosis in MDA-MB231 cells and caused cell cycle arrest at the SubG1 phase. Dendrosomal c[u](#page-7-0)rcumin treatment downregulated Lnc-DANCR, EZH2, bcl-2, and p53 gene expression, while upregulating bax, Noxa, PUMA, and p21 gene expression in a time-dependent manner. Bax and P21 protein levels were significantly upregulated following DNC treatment, whereas Bcl-2 and P53 protein levels were downreg[ula](#page-7-1)ted in DNC-treated breast cancer cells.

Conclusions: In summary, dendrosomal nanocurcumin demonstrated potent anti-tumor effects against breast cancer cells, suggesting its potential as a therapeutic agent in breast cancer treatment.

Keywords: Cancer,Nanocurcumin,Apoptosis,Gene,Protein

# 1. Background

Breast cancer is diagnosed in approximately a quarter of all women with cancer and causes the death of around 570,000 patients annually ([1\)](#page-7-0). Early diagnosis is crucial in reducing mortality and improving patient survival ([2](#page-7-1)). One of the primary risk factors for this malignancy is genetic mutations, particularly in the apoptosis pathway, which allows cancer cells to resist apoptosis and contribute to tumor development [\(3\)](#page-7-2). The polyproline region of the P53 gene plays a vital role in tumor suppressor activity  $(4)$  $(4)$  $(4)$ , promoting apoptosis by releasing various pro-apoptotic factors ([5\)](#page-7-4).

Consequently, mutations in this domain are linked to the development of multiple cancers [\(6](#page-7-5)). Notably, P53 gene mutations are more frequently observed in breast cancer, with over 80% of triple-negative breast cancer (TNBC) cases exhibiting such mutations ([7](#page-7-6)). The MDA-MB-231 cell line (a TNBC cell model) has a P53 gene mutation known as R280K ([8\)](#page-7-7). It is important to note that TNBC patients are often resistant to chemotherapy and have a lower survival rate  $(9)$  $(9)$  $(9)$ . Therefore, the development of new treatment strategies for this disease is essential.

Curcumin is a natural compound known for its wide range of pharmacological properties, including

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antioxidant ([10](#page-7-9)), analgesic ([11\)](#page-7-10), anti-inflammatory [\(12\)](#page-7-11), antimicrobial [\(13\)](#page-7-12), anti-mutagenic ([14](#page-8-0)), and anti-tumor effects [\(15\)](#page-8-1). It has demonstrated anticancer activity against various malignancies, including head and neck squamous cell carcinoma (HNSCC) ([16\)](#page-8-2), prostate ([17\)](#page-8-3), colorectal  $(18)$  $(18)$  $(18)$ , breast  $(19)$  $(19)$ , and brain cancers  $(20)$  $(20)$ . Mechanisms underlying curcumin's anticancer effects include downregulation of NF-κB [\(21\)](#page-8-7), induction of apoptosis through increased oxidative stress, upregulation of P53 [\(22](#page-8-8)), and inhibition of CXCL1/2 production ([23](#page-8-9)). However, its therapeutic use has been hindered by poor intestinal absorption, bioavailability, and water solubility  $(24)$  $(24)$ .

Recent advancements in nanotechnology have addressed these limitations by developing curcuminloaded nanoformulations. For example, Montazeri et al. (2016) introduced dendrosomal curcumin (DNC), which possesses amphiphilic properties and high bioavailability, and demonstrated its anticancer effects on hepatocellular carcinoma by inducing apoptosis and inhibiting cell proliferation [\(24\)](#page-8-10). Dendrosomal curcumin's effects on breast cancer have also been investigated. For instance, DNC was found to inhibit metastasis in breast cancer by modulating immune responses, upregulating STAT4 and IL-2, and reducing the expression of STAT3 and IL-10 genes ([25\)](#page-8-11). Moreover, DNC induced apoptosis in MCF7 cells ([26](#page-8-12)), and downregulation of LncHOTAIR was proposed as one of its mechanisms of action ([27](#page-8-13)). Additionally, a combination of DNC with exogenous P53 in the MDA-MB-231 cell line induced cell death via apoptosis and decreased the expression levels of ZEB1 and BMI1 genes [\(28](#page-8-14)).

Despite these promising findings, further studies are needed to fully elucidate DNC's mechanisms of action in breast cancer.

# 2. Objectives

This study aimed to investigate the effects of DNC on the MDA-MB-231 cell line and evaluate changes in gene and protein expression related to cell cycle regulation and mitochondrial apoptosis processes.

#### 3. Methods

#### 3.1. Dendrosomal Curcumin and Cell Preparations

Dendrosomal curcumin was generously provided by Babaei et al., who previously synthesized and evaluated DNC's effects on the mouse fibrosarcoma cell line WEHI-164 [\(29\)](#page-8-15). MDA-MB231 cells were cultured immediately in DMEM medium containing 1% penicillin/streptomycin and 10% FBS (GIBCO, USA) and incubated in a humidified atmosphere of 5% CO<sub>2</sub> at 37°C. The cells (80  $\times$  10<sup>6</sup> cells/mL) were seeded into 12-well plates and treated with 20  $\mu$ M DNC ([30](#page-8-16)) and free curcumin for 240 minutes. To assess curcumin uptake by the cells, fluorescence microscopy was used [\(24](#page-8-10))

# 3.2. Cell Viability

MDA-MB231 cells ( $4 \times 10^4$  cells/mL) were seeded into a 96-well plate with 200 µL of DMEM medium and incubated overnight under the conditions described above. The following day, the cells were treated with 20 µM DNC and incubated for three days. Afterward, 5 mg/mL of MTT solution was added, and the cells were incubated for 240 minutes. Finally, the upper layer was removed, and 200 µL of DMSO was added. After 15 minutes, the optical density (OD) was measured using an ELISA reader at 490 nm.

# 3.3. Cell Proliferation Cycles

The effects of DNC on MDA-MB231 cell proliferation cycles were evaluated using the propidium iodide (PI) staining method and flow cytometry. Briefly,  $3 \times 10^5$ cells/mL were exposed to 20 µM DNC for different time periods. The cells were washed with PBS and fixed with 75% ethanol for 15 minutes at 4°C, then washed again with PBS. The cells were treated with PI (50 µg/mL in PBS) for 15 minutes and analyzed using a FACSCalibur flow cytometer (Becton Dickinson, USA).

# 3.4. Apoptosis

The percentage of apoptotic cells in DNC-treated samples was measured using flow cytometry and Annexin V staining, following the manufacturer's instructions with the Annexin-V-FITC kit (Roche, Germany).

#### 3.5. Gene Expression

We used an RNA extraction kit (Invitrogen, USA) for isolating total RNA based on the instructions provided by the manufacturer. After ensuring the high quality and quantity of extracted RNA, cDNA was synthesized using a commercial kit (Takara, Japan). The primers specific for Lnc-DANCR (F-5'-CCTCAGTTCTTAGCGCAGGTTG-3, R-5'-ACTGCTCTAGCTCCTGTGGC-3'); EZH2 (F-5'- CACGGGGATAGAGAATGTGGGTT -3', R-5'-AGTTCTTCTGCTGTGCCCTTATCTG -3); bax (F-5'-GTGGATGACTGAGTACCTGAAC -3', R-5'-GCCAGGAGAAATCAAACAGAGG -3); Noxa (F-5'- GAGCTGGAAGTCGAGTGTG -3', R-5'-CTCTTTTGAAGGAGTCCCCTC -3); PUMA (F-5'- -3', R-5'- -3);



CATGAGTCCTTCCACGATACC-3') were designed using Oligo software and blasted on the NCBI website. The reaction mixture for RT-PCR included 0.5 µL of each F and R primer, 5 µL Master Mix, 0.75 ng cDNA, and 3.5 µL deionized water. The time-temperature program of the RT-PCR device (Applied Biosystems, USA) was 1 cycle of 95°C for 15 min, 40 cycles of 95°C for 5 s, 63°C for 23 s, and 72°C for 34 s, with a final cycle of 72°C for 10 min. The 2<sup>-ΔΔCT</sup> method was used for gene expression data analysis.

#### 3.6. Western Blot

Proteins from DNC-treated cells were extracted using RIPA buffer via the immunoprecipitation method and quantified by the Bradford assay. Extracted proteins were separated by SDS-PAGE and transferred to nitrocellulose membranes. The membranes were blocked with 5% skim milk for 90 minutes and incubated overnight at 4°C with primary antibodies for β-actin, Bcl-2, P21, P53, and bax proteins. The membranes were then washed with TBST and incubated with secondary goat HRP anti-mouse IgG. The chemiluminescence method and ECL kit were used to visualize the protein bands.

# 3.7. Statistical Analysis

Data were analyzed using the ANOVA procedure, and Tukey's post hoc test was used for mean comparisons at P < 0.05. The analyses were conducted using GraphPad Prism V.8 software.

# 4. Results

#### 4.1. Uptake Kinetics

The uptake of dendrosomal curcumin compared to bulk curcumin was assessed using fluorescence microscopy, leveraging curcumin's inherent fluorescent properties. In this experiment, cells were cultured in 24 well plates, and after 24 hours, they were treated with 20 μM curcumin and 20 μM DNC. Four hours posttreatment, the cells were examined under a fluorescence microscope. The results demonstrated that DNC exhibited significantly higher penetration

efficiency into the cells compared to bulk curcumin [\(Figure](#page-3-0) 1).

#### 4.2. Cell Viability, Apoptosis, and Necrosis

Dendrosomal curcumin (20 μM) reduced the cell viability of MDA-MB231 breast cancer cells in a timedependent manner. Before treatment, cell viability was measured at 99.51  $\pm$  4.11%, and 72 hours after DNC treatment, it decreased to 21.45  $\pm$  4.22% [\(Figure](#page-3-1) 2), highlighting the potent anticancer effects of this nanoformulation. The primary cause of cell death was apoptosis, with  $57.80 \pm 5.22\%$  of cells undergoing apoptosis 72 hours post-treatment. In comparison, the percentage of necrotic cells was  $20.7 \pm 3.44\%$  at the same time point [\(Figure](#page-3-1) 2), indicating that DNC predominantly induces cell death through apoptosis.

# 4.3. Cell Cycles

Dendrosomal curcumin treatment arrested MDA-MB231 cancer cells at the SubG1 phase of the cell cycle, effectively preventing cell division. As depicted in [Figure](#page-4-0) [3](#page-4-0), approximately 70% of DNC-treated cells were in the SubG1 phase 72 hours post-treatment. The inhibition of cell division began 24 hours after DNC exposure and persisted for the entire 72-hour period. These results further confirm the anti-tumor effects of DNC against MDA-MB231 cells. The flow cytometry histograms illustrating these findings are shown in [Figure](#page-4-1) 4.

# 4.4. Gene Expression Analysis

Dendrosomal curcumin treatment downregulated the expression of both Lnc-DANCR and EZH2 genes in a time-dependent manner in MDA-MB231 cells. The lowest expression levels of Lnc-DANCR were observed 48 and 72 hours after DNC treatment [\(Figure](#page-5-0) 5). Similarly, the lowest expression levels of the EZH2 gene were also recorded at these time points ([Figure](#page-5-0) 5). These findings suggest that DNC effectively suppresses the expression of key genes involved in tumor progression.

One day after DNC treatment, significant changes were observed in the expression levels of bcl-2, bax, Noxa, PUMA, p21, and p53 genes, with these changes becoming more pronounced over time (up to 72 hours). Specifically, the expressions of bax, Noxa, PUMA, and p21 genes increased significantly in a time-dependent manner following DNC treatment compared to untreated cells. In contrast, the expressions of bcl-2 and p53 genes were significantly downregulated compared to untreated cells ([Figure](#page-5-1) 6). These results highlight the role of DNC in regulating key genes involved in apoptosis and cell cycle arrest.

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<span id="page-3-1"></span>



Figure 2. The impacts of dendrosomal curcumin (DNC, 20 μM) on MDA-MB-231 cells' viability, apoptosis and necrosis [before](#page-6-0) treatment and 24, 48 and 72 hours after (n = 3).

# 4.5. Western Blot Analyses

We also measured the expressions of Bcl-2, Bax, P21, and P53 proteins in the DNC-treated cells, and the results are shown in [Figure](#page-6-0) 7. As can be seen, Bax and P21 were significantly upregulated after DNC treatment; however, both Bcl-2 and P53 were downregulated in the 20 µM DNC-exposed MDA-MB-231 cells.

# 5. Discussion

The findings of this study indicated improved penetration of curcumin when loaded into

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<span id="page-4-1"></span>Figure 3. The effects of dendrosomal curcumin (DNC, 20 μM) on the MDA-MB-231 cells proliferation cycle at SubG1, G1, S and G2/M stages (n = 3); \*P < 0.05; \*\*\*\*P < 0.001; \*\*\*\*P < 0.0001.



**Figure 4.** The histograms obtained by flow-cytometry for evaluating the impacts of dendrosomal curcumin (DNC, 20 μM) during 0, 24, 48 and 72 h on the MDA-MB-231 cells<br>proliferation cycle at SubG1, G1,S and G2/M stages (n

dendrosomes, significantly enhancing its entry into MDA-MB-231 breast cancer cells and inducing apoptosis. Additionally, DNC effectively arrested the cells in the SubG1 phase of mitotic division, preventing cell proliferation. Dendrosomal curcumin treatment also

led to decreased expression of Lnc-DANCR and EZH2 genes, while upregulating bax, Noxa, PUMA, and p21 genes and downregulating bcl-2 and p53 in the breast cancer cell line. Western blot analysis further confirmed

<span id="page-5-0"></span>

<span id="page-5-1"></span>F**igure 5.** The impacts of dendrosomal curcumin (DNC, 20 µM) on expressions of *Lnc-DANCR* and *EZH2* genes of MDA-MB-231 cells (n = 3). GAPDH gene was used as the<br>housekeeping gene.\*\*\*\*P<0.0001;\*\*\*P<0.001;\*\*P<0.05;\*P<0.05



F**igure 6.** The impacts of dendrosomal curcumin (DNC, 20 µM) on expressions of bcl-2, bax, Noxa, PUMA, p21 and p53 genes of MDA-MB-231 cells (n = 3). GAPDH gene was used as the<br>housekeeping gene. \*\*\*P < 0.0001; \*\*P < 0.01;

downregulation of Bcl-2 and P53 proteins and upregulation of Bax and P21 in DNC-treated cells.

Curcumin is a natural compound with extensive pharmacological properties, including anticancer effects ([31\)](#page-8-17). However, its low solubility and poor intestinal absorption have been significant barriers to fully utilizing its therapeutic potential ([32\)](#page-8-18). One solution to this issue has been the development of curcumin-loaded nanoformulations ([32\)](#page-8-18), which enhance curcumin's solubility through amphipathic properties. In this study, DNC uptake was evaluated in breast cancer cells, and the results demonstrated that DNC showed superior cell entry compared to free curcumin, likely due to improved solubility in its dendrosomal form [\(29\)](#page-8-15).

Furthermore, DNC significantly reduced the viability of breast cancer cells, demonstrating its strong potential to induce apoptosis. Similar findings were reported by Baghi et al. (2018), who highlighted DNC's anticancer effects in breast cancer cells through apoptosis induction, with cytotoxicity being enhanced by exogenous p53 ([28\)](#page-8-14). In another study, DNC demonstrated cytotoxic effects on T47D and MCF-7 breast cancer cells by inducing apoptosis [\(33\)](#page-8-19). Seyed Hosseini et al. (2023) also reported DNC's anticancer effects against ovarian cancer, upregulating matrix metalloproteinase-2 (MMP-2) [\(34\)](#page-8-20). These researchers further observed decreased expression of HOTAIR, bcl-2, and H19, along with overexpression of MEG3, after treating ovarian cancer cells with DNC [\(35](#page-8-21)), indicating

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**Figure 7.** The expressions of Bcl-2, Bax, P53 and P21 proteins in the MDA-MB-231 cells before and after treatment with dendrosomal curcumin (DNC, 20 μM) (n = 3). \*\*\*[\\*P](#page-9-0) < 0.0001;<br>\*\*P < 0.001; \*P < 0.05.

that curcumin exerts its anticancer effects via multiple pathways.

In summary, DNC shows great potential as an adjuvant therapy in cancer treatment by inducing apoptosis in cancerous cells.

Dendrosomal curcumin -treated cells exhibited cell division arrest at the SubG1 phase, suggesting that curcumin disrupts the mitotic process. This finding aligns with other studies reporting the arrest of cancer cells, such as Huh/HepG2, in the SubG1 and G2/M phases after curcumin treatment ([24,](#page-8-10) [36\)](#page-8-22), demonstrating curcumin's ability to inhibit cancer cell development by disrupting mitosis. Additionally, G2/M phase arrest has been observed in pancreatic ([37](#page-8-23)) and head and neck squamous carcinoma cell lines [\(38](#page-8-24)). The ATM/Chk2-P53 axis is thought to play a key role in this process  $(38)$  $(38)$ , while curcumin-induced cell cycle arrest also involves inhibition of NF-κB and histone deacetylase 4 [\(39,](#page-8-25) [40\)](#page-8-26).

Lnc-DANCR is known to be upregulated in advanced breast cancer and is associated with metastasis [\(41](#page-9-0)). The MDA-MB-231 cell line, used in this study, is highly

malignant and metastatic. Here, we observed high expression of the Lnc-DANCR gene in these cells, which significantly decreased 48 hours after DNC treatment. Previous research has shown that Lnc-DANCR knockdown leads to breast cancer cell cycle arrest in the G0/G1 phase [\(41](#page-9-0)). Thus, the arrest of DNC-treated cells in the SubG1 phase may be linked to the reduction in Lnc-DANCR expression. Lnc-DANCR knockdown prevents EZH2 from binding to the SOCS3 promoter, leading to SOCS3 upregulation, which in turn inhibits tumor growth ([41](#page-9-0), [42](#page-9-1)). Therefore, the downregulation of Lnc-DANCR and EZH2 in DNC-treated cells further explains the nanoformulation's anticancer effects.

We also evaluated the gene and protein expressions related to the intrinsic apoptotic pathway, including bax, Noxa, PUMA, p21, bcl-2, and p53. Cancer cells often evade programmed cell death through the attenuation of the apoptosis pathway, making the induction of apoptosis a key mechanism in anticancer therapies ([37](#page-8-23)). Many chemotherapy drugs exert their effects by inducing apoptosis  $(43)$  $(43)$ . In this study, DNC effectively

induced apoptosis in breast cancer cells, showcasing its potential as an antitumor agent. The gene expression analysis revealed that DNC upregulated bax, Noxa, PUMA, and p21, while downregulating bcl-2 and p53. Correspondingly, at the protein level, DNC increased Bax and P21 expressions and decreased Bcl-2 and P53 levels. P53 plays a central role in both intrinsic and extrinsic apoptosis by stimulating the production of proapoptotic proteins such as Bax, Noxa, and PUMA, which promote cytochrome C release and lead to cell death ([44\)](#page-9-3). Furthermore, the reduction of Bcl-2 expression is associated with increased apoptosis. Thus, the upregulation of pro-apoptotic genes and proteins, and the downregulation of anti-apoptotic factors, indicates the activation of the intrinsic apoptosis pathway in DNC-treated cells. Notably, the P53 protein in the MDA-MB-231 cell line is a mutant form with potential oncogenic activity ([45](#page-9-4)), and the observed reduction in both mRNA and protein levels of P53 after DNC treatment may explain the anticancer effects seen in this study.

#### 5.1. Conclusions

In conclusion, dendrosomal nanocurcumin exhibited strong anti-tumor effects against breast cancer cells. The mechanisms of action were linked to cell division arrest at the SubG1 phase and the induction of apoptosis by upregulating pro-apoptotic factors, including bax, Noxa, and PUMA, while downregulating the anti-apoptotic factor bcl-2. Further investigation of the anticancer effects of DNC in animal models is recommended to validate its potential as a therapeutic agent for cancer treatment.

# Acknowledgements

The authors would like to express their appreciation to the staff of Tehran Medical Sciences, Islamic Azad University, for their valuable assistance during the conduct of this study.

# Footnotes

Authors' Contribution: Acquisition of data: H. A. and F. H.; analysis and interpretation of data: S. T. and B. I. F.; drafting of the manuscript: S. T.; critical revision of the manuscript for important intellectual content: M. M.; statistical analysis: S. T. and B. I. F.; administrative, technical, and material support: M. M.

Conflict of Interests Statement: The authors declared that there is no conflict of interest.

Data Availability: The dataset presented in the study is available on request from the corresponding author during submission or after publication.

Funding/Support: The authors declared that they have no funding/support.

#### References

- <span id="page-7-0"></span>1. McGuire S. World Cancer Report 2014. Geneva, Switzerland: World Health Organization, International Agency for Research on Cancer, WHO Press, 2015. Adv Nutr. 2016;7(2):418-9. [PubMed ID: [26980827\]](http://www.ncbi.nlm.nih.gov/pubmed/26980827). [PubMed Central ID: [PMC4785485\]](https://www.ncbi.nlm.nih.gov/pmc/PMC4785485). [https://doi.org/10.3945/an.116.012211.](https://doi.org/10.3945/an.116.012211)
- <span id="page-7-1"></span>2. DeSantis CE, Fedewa SA, Goding Sauer A, Kramer JL, Smith RA, Jemal A. Breast cancer statistics, 2015: Convergence of incidence rates between black and white women. CA Cancer J Clin. 2016;66(1):31-42. [PubMed ID: [26513636\]](http://www.ncbi.nlm.nih.gov/pubmed/26513636). [https://doi.org/10.3322/caac.21320.](https://doi.org/10.3322/caac.21320)
- <span id="page-7-2"></span>3. Barnes DM, Camplejohn RS. P53, apoptosis, and breast cancer. J Mammary Gland Biol Neoplasia. 1996;1(2):163-75. [PubMed ID: [10887490](http://www.ncbi.nlm.nih.gov/pubmed/10887490)]. <https://doi.org/10.1007/BF02013640>.
- <span id="page-7-3"></span>4. Kaur RP, Vasudeva K, Kumar R, Munshi A. Role of p53 Gene in Breast Cancer: Focus on Mutation Spectrum and Therapeutic Strategies.<br>Curr Pharmaceutic Design. 2018;24(30):3566-75. Curr Pharmaceutic Design. 2018;24(30):3566-75. [https://doi.org/10.2174/1381612824666180926095709.](https://doi.org/10.2174/1381612824666180926095709)
- <span id="page-7-4"></span>5. Zilfou JT, Lowe SW. Tumor suppressive functions of p53. Cold Spring Harb Perspect Biol. 2009;1(5). a001883. [PubMed ID: [20066118\]](http://www.ncbi.nlm.nih.gov/pubmed/20066118). [PubMed Central ID: [PMC2773645\]](https://www.ncbi.nlm.nih.gov/pmc/PMC2773645). [https://doi.org/10.1101/cshperspect.a001883.](https://doi.org/10.1101/cshperspect.a001883)
- <span id="page-7-5"></span>6. Duffy MJ, Synnott NC, O'Grady S, Crown J. Targeting p53 for the treatment of cancer. Semin Cancer Biol. 2022;79:58-67. [PubMed ID: [32741700](http://www.ncbi.nlm.nih.gov/pubmed/32741700)]. <https://doi.org/10.1016/j.semcancer.2020.07.005>.
- <span id="page-7-6"></span>7. Koboldt DC, Fulton RS, McLellan MD, et al. Comprehensive molecular portraits of human breast tumours. Nature. 2012;490(7418):61-70. <https://doi.org/10.1038/nature11412>.
- <span id="page-7-7"></span>8. Bae YH, Shin JM, Park HJ, Jang HO, Bae MK, Bae SK. Gain-of-function mutant p53-R280K mediates survival of breast cancer cells. Genes  $\delta$ Genomics. 2013;36(2):171-8. <https://doi.org/10.1007/s13258-013-0154-9>.
- <span id="page-7-8"></span>9. Arnedos M, Bihan C, Delaloge S, Andre F. Triple-negative breast cancer: are we making headway at least? Ther Adv Med Oncol. 2012;4(4):195-210. [PubMed ID: [22754593](http://www.ncbi.nlm.nih.gov/pubmed/22754593)]. [PubMed Central ID: [PMC3384094](https://www.ncbi.nlm.nih.gov/pmc/PMC3384094)]. [https://doi.org/10.1177/1758834012444711.](https://doi.org/10.1177/1758834012444711)
- <span id="page-7-9"></span>10. Eke-Okoro UJ, Raffa RB, Pergolizzi JJ, Breve F, Taylor RJ, Nema Research Group. Curcumin in turmeric: Basic and clinical evidence for a potential role in analgesia. J Clin Pharm Ther. 2018;43(4):460-6. [PubMed ID: [29722036](http://www.ncbi.nlm.nih.gov/pubmed/29722036)]. <https://doi.org/10.1111/jcpt.12703>.
- <span id="page-7-10"></span>11. Menon VP, Sudheer AR. Antioxidant and anti-inflammatory properties of curcumin. Adv Exp Med Biol. 2007;595:105-25. [PubMed ID: [17569207\]](http://www.ncbi.nlm.nih.gov/pubmed/17569207). [https://doi.org/10.1007/978-0-387-46401-5\\_3.](https://doi.org/10.1007/978-0-387-46401-5_3)
- <span id="page-7-11"></span>12. Hussain Y, Alam W, Ullah H, Dacrema M, Daglia M, Khan H, et al. Antimicrobial Potential of Curcumin: Therapeutic Potential and Challenges to Clinical Applications. Antibiotics (Basel). 2022;11(3). [PubMed ID: [35326785](http://www.ncbi.nlm.nih.gov/pubmed/35326785)]. [PubMed Central ID: [PMC8944843\]](https://www.ncbi.nlm.nih.gov/pmc/PMC8944843). <https://doi.org/10.3390/antibiotics11030322>.
- <span id="page-7-12"></span>13. Ragunathan I, Panneerselvam N. Antimutagenic potential of curcumin on chromosomal aberrations in Allium cepa. J Zhejiang Univ Sci. 2007;8(7):470-5. [https://doi.org/10.1631/jzus.2007.B0470.](https://doi.org/10.1631/jzus.2007.B0470)
- <span id="page-8-0"></span>14. Momtazi AA, Sahebkar A. Difluorinated Curcumin: A Promising Curcumin Analogue with Improved Anti-Tumor Activity and Pharmacokinetic Profile. Curr Pharm Des. 2016;22(28):4386-97.<br>[PubMed ID: 27229723]. [27229723](http://www.ncbi.nlm.nih.gov/pubmed/27229723)]. <https://doi.org/10.2174/1381612822666160527113501>.
- <span id="page-8-1"></span>15. Borges GA, Elias ST, Amorim B, de Lima CL, Coletta RD, Castilho RM, et al. Curcumin downregulates the PI3K-AKT-mTOR pathway and inhibits growth and progression in head and neck cancer cells. Phytother Res. 2020;34(12):3311-24. [PubMed ID: [32628350](http://www.ncbi.nlm.nih.gov/pubmed/32628350)]. <https://doi.org/10.1002/ptr.6780>.
- <span id="page-8-2"></span>16. Termini D, Den Hartogh DJ, Jaglanian A, Tsiani E. Curcumin against Prostate Cancer: Current Evidence. Biomolecules. 2020;10(11). [PubMed ID: [33182828](http://www.ncbi.nlm.nih.gov/pubmed/33182828)]. [PubMed Central ID: [PMC7696488](https://www.ncbi.nlm.nih.gov/pmc/PMC7696488)]. [https://doi.org/10.3390/biom10111536.](https://doi.org/10.3390/biom10111536)
- <span id="page-8-3"></span>17. Weng W, Goel A. Curcumin and colorectal cancer: An update and current perspective on this natural medicine. Semin Cancer Biol. 2022;80:73-86. [PubMed ID: [32088363](http://www.ncbi.nlm.nih.gov/pubmed/32088363)]. [PubMed Central ID: [PMC7438305\]](https://www.ncbi.nlm.nih.gov/pmc/PMC7438305). [https://doi.org/10.1016/j.semcancer.2020.02.011.](https://doi.org/10.1016/j.semcancer.2020.02.011)
- <span id="page-8-4"></span>18. Farghadani R, Naidu R. Curcumin: Modulator of Key Molecular Signaling Pathways in Hormone-Independent Breast Cancer. Cancers. 2021;13(14). [https://doi.org/10.3390/cancers13143427.](https://doi.org/10.3390/cancers13143427)
- <span id="page-8-5"></span>19. Benameur T, Giacomucci G, Panaro MA, Ruggiero M, Trotta T, Monda V, et al. New Promising Therapeutic Avenues of Curcumin in Brain Diseases. Molecules. 2021;27(1). [PubMed ID: [35011468\]](http://www.ncbi.nlm.nih.gov/pubmed/35011468). [PubMed Central ID: [PMC8746812\]](https://www.ncbi.nlm.nih.gov/pmc/PMC8746812). [https://doi.org/10.3390/molecules27010236.](https://doi.org/10.3390/molecules27010236)
- <span id="page-8-6"></span>20. Liu Q, Loo WT, Sze SC, Tong Y. Curcumin inhibits cell proliferation of MDA-MB-231 and BT-483 breast cancer cells mediated by downregulation of NFkappaB, cyclinD and MMP-1 transcription. Phytomedicine. 2009;16(10):916-22. [PubMed ID: [19524420](http://www.ncbi.nlm.nih.gov/pubmed/19524420)]. [https://doi.org/10.1016/j.phymed.2009.04.008.](https://doi.org/10.1016/j.phymed.2009.04.008)
- <span id="page-8-7"></span>21. Moghtaderi H, Sepehri H, Attari F. Combination of arabinogalactan and curcumin induces apoptosis in breast cancer cells in vitro and inhibits tumor growth via overexpression of p53 level in vivo. Biomed<br>Pharmacother. 2017;88:582-94. [PubMed ID: 28152473]. 2017;88:582-94. [PubMed ID: [28152473](http://www.ncbi.nlm.nih.gov/pubmed/28152473)]. <https://doi.org/10.1016/j.biopha.2017.01.072>.
- <span id="page-8-8"></span>22. Bachmeier BE, Mohrenz IV, Mirisola V, Schleicher E, Romeo F, Höhneke C, et al. Curcumin downregulates the inflammatory cytokines CXCL1 and -2 in breast cancer cells via NFκB. Carcinogenesis. 2008;29(4):779-89. <https://doi.org/10.1093/carcin/bgm248>.
- <span id="page-8-9"></span>23. Liu W, Zhai Y, Heng X, Che FY, Chen W, Sun D, et al. Oral bioavailability of curcumin: problems and advancements. J Drug Target. 2016;24(8):694-702. [PubMed ID: [26942997](http://www.ncbi.nlm.nih.gov/pubmed/26942997)]. [https://doi.org/10.3109/1061186X.2016.1157883.](https://doi.org/10.3109/1061186X.2016.1157883)
- <span id="page-8-10"></span>24. Montazeri M, Sadeghizadeh M, Pilehvar-Soltanahmadi Y, Zarghami F, Khodi S, Mohaghegh M, et al. Dendrosomal curcumin nanoformulation modulate apoptosis-related genes and protein expression in hepatocarcinoma cell lines. Int J Pharm. 2016;509(1-2):244-54. [PubMed ID: [27234697](http://www.ncbi.nlm.nih.gov/pubmed/27234697)]. <https://doi.org/10.1016/j.ijpharm.2016.05.039>.
- <span id="page-8-11"></span>25. Shiri S, Alizadeh AM, Baradaran B, Farhanghi B, Shanehbandi D, Khodayari S, et al. Dendrosomal curcumin suppresses metastatic breast cancer in mice by changing m1/m2 macrophage balance in the tumor microenvironment. Asian Pac J Cancer Prev. 2015;16(9):3917- 22. **[PubMed** ID: [25987060](http://www.ncbi.nlm.nih.gov/pubmed/25987060)]. <https://doi.org/10.7314/apjcp.2015.16.9.3917>.
- <span id="page-8-12"></span>26. Alghanimi YK, Ghasemian A. Inhibitory Traits of Dendrosome Curcumin (DNC) on Breast Cancer Compared to Curcumin Single Compound. J Gastrointest Cancer. 2020;51(2):527-33. [PubMed ID: [31286422\]](http://www.ncbi.nlm.nih.gov/pubmed/31286422). [https://doi.org/10.1007/s12029-019-00273-2.](https://doi.org/10.1007/s12029-019-00273-2)
- <span id="page-8-13"></span>27. Mehrabi M, Alemohammad S, Dashtebozorgi R, Tahmasebi Birgani M, Hajjari MR, Mohammadi-Asl J. Evaluate the Effect of Dendrosomal Curcumin on Expression Level of HOTAIR Long Noncoding RNA in MCF-7 Breast Cancer Cell Line. J Shahid Sadoughi Univ Med Sci. 2023. [https://doi.org/10.18502/ssu.v31i6.13472.](https://doi.org/10.18502/ssu.v31i6.13472)
- <span id="page-8-14"></span>28. Baghi N, Bakhshinejad B, Keshavarz R, Babashah S, Sadeghizadeh M. Dendrosomal nanocurcumin and exogenous p53 can act synergistically to elicit anticancer effects on breast cancer cells. Gene.<br>2018;670:55-62. [PubMed ID: 29753810]. 2018;670:55-62. [https://doi.org/10.1016/j.gene.2018.05.025.](https://doi.org/10.1016/j.gene.2018.05.025)
- <span id="page-8-15"></span>29. Babaei E, Sadeghizadeh M, Hassan ZM, Feizi MA, Najafi F, Hashemi SM. Dendrosomal curcumin significantly suppresses cancer cell proliferation in vitro and in vivo. *Int Immunopharmacol.*<br>2012;12(1):226-34. [PubMed ID: 22155627].  $2012:12(1):226-34.$ [https://doi.org/10.1016/j.intimp.2011.11.015.](https://doi.org/10.1016/j.intimp.2011.11.015)
- <span id="page-8-16"></span>30. Moradi F, Sadeghizadeh P, Najafi F, Sadeghizadeh M. Dendrosomal Nano-Curcumin Downregulates CCAT2 Expression Levels and Promotes Cell Cycle Arrest and Apoptosis in Tamoxifen-Resistant MCF-7 Cells. Biomacromolecul J. 2021;7(3):138-48.
- <span id="page-8-17"></span>31. Fu YS, Chen TH, Weng L, Huang L, Lai D, Weng CF. Pharmacological properties and underlying mechanisms of curcumin and prospects in medicinal potential. Biomed Pharmacother. 2021;141:111888. [PubMed ID: [34237598\]](http://www.ncbi.nlm.nih.gov/pubmed/34237598). <https://doi.org/10.1016/j.biopha.2021.111888>.
- <span id="page-8-18"></span>32. Gornicka J, Mika M, Wroblewska O, Siudem P, Paradowska K. Methods to Improve the Solubility of Curcumin from Turmeric. Life (Basel). 2023;13(1). [PubMed ID: [36676157\]](http://www.ncbi.nlm.nih.gov/pubmed/36676157). [PubMed Central ID: [PMC9862957\]](https://www.ncbi.nlm.nih.gov/pmc/PMC9862957). [https://doi.org/10.3390/life13010207.](https://doi.org/10.3390/life13010207)
- <span id="page-8-19"></span>33. Sobhkhizi A, Babaei E, Azeez HJ, Katiraee F, Hussen BM, Hoseinpour Feizi MA. Dendrosomal Nano-Curcumin Modulates P-Glycoprotein Activity and Induces Apoptosis in Wild Type and P53-Mutant Breast Cancer Cell Lines. Jentashapir J Cell Mol Biol. 2020;11(4). e109143. [https://doi.org/10.5812/jjcmb.109143.](https://doi.org/10.5812/jjcmb.109143)
- <span id="page-8-20"></span>34. Seyed Hosseini E, Alizadeh Zarei M, Tarrahimofrad H, Zamani J, Haddad Kashani H, Ahmad E, et al. Synergistic effects of dendrosomal nanocurcumin and oxaliplatin on oncogenic properties of ovarian cancer cell lines by down-expression of MMPs. Biol Res. 2023;56(1):3. [PubMed ID: [36658640\]](http://www.ncbi.nlm.nih.gov/pubmed/36658640). [PubMed Central ID: [PMC9854214](https://www.ncbi.nlm.nih.gov/pmc/PMC9854214)]. [https://doi.org/10.1186/s40659-023-00412-x.](https://doi.org/10.1186/s40659-023-00412-x)
- <span id="page-8-21"></span>35. Seyed Hosseini E, Alizadeh Zarei M, Haddad Kashani H, Salimian M, Riahi Kashani N, Nikzad H. Altered Long Non-coding RNAs Expression and Cytotoxic and Anti-proliferative Activity of Dendrosomal Nano-curcumin in Ovarian Cancer Cells. India J Gynecolo Oncolo. 2021;19(2). [https://doi.org/10.1007/s40944-021-00511-](https://doi.org/10.1007/s40944-021-00511-1) [1.](https://doi.org/10.1007/s40944-021-00511-1)
- <span id="page-8-22"></span>36. Wang WZ, Cheng J, Luo J, Zhuang SM. Abrogation of G2/M arrest sensitizes curcumin-resistant hepatoma cells to apoptosis. FEBS Lett. 2008;582(18):2689-95. [PubMed ID: [18602917\]](http://www.ncbi.nlm.nih.gov/pubmed/18602917). [https://doi.org/10.1016/j.febslet.2008.06.048.](https://doi.org/10.1016/j.febslet.2008.06.048)
- <span id="page-8-23"></span>37. Zhu Y, Bu S. Curcumin Induces Autophagy, Apoptosis, and Cell Cycle Arrest in Human Pancreatic Cancer Cells. Evid Based Complement Alternat Med. 2017;2017:5787218. [PubMed ID: [29081818](http://www.ncbi.nlm.nih.gov/pubmed/29081818)]. [PubMed Central ID: [PMC5610853\]](https://www.ncbi.nlm.nih.gov/pmc/PMC5610853). <https://doi.org/10.1155/2017/5787218>.
- <span id="page-8-24"></span>38. Hu A, Huang JJ, Zhang JF, Dai WJ, Li RL, Lu ZY, et al. Curcumin induces G2/M cell cycle arrest and apoptosis of head and neck squamous cell carcinoma in vitro and in vivo through ATM/Chk2/p53-dependent pathway. Oncotarget. 2017;8(31):50747-60. [PubMed ID: [28881600\]](http://www.ncbi.nlm.nih.gov/pubmed/28881600). [PubMed Central ID: [PMC5584201\]](https://www.ncbi.nlm.nih.gov/pmc/PMC5584201). [https://doi.org/10.18632/oncotarget.17096.](https://doi.org/10.18632/oncotarget.17096)
- <span id="page-8-25"></span>39. Lee SJ, Krauthauser C, Maduskuie V, Fawcett PT, Olson JM, Rajasekaran SA. Curcumin-induced HDAC inhibition and attenuation of medulloblastoma growth in vitro and in vivo. BMC Cancer. 2011;11:144. [PubMed ID: [21501498\]](http://www.ncbi.nlm.nih.gov/pubmed/21501498). [PubMed Central ID: [PMC3090367\]](https://www.ncbi.nlm.nih.gov/pmc/PMC3090367). [https://doi.org/10.1186/1471-2407-11-144.](https://doi.org/10.1186/1471-2407-11-144)
- <span id="page-8-26"></span>40. Spiller SE, Logsdon NJ, Deckard LA, Sontheimer H. Inhibition of nuclear factor kappa-B signaling reduces growth in medulloblastoma in vivo. BMC Cancer. 2011;11:136. [PubMed ID: [21492457](http://www.ncbi.nlm.nih.gov/pubmed/21492457)]. [PubMed Central ID: [PMC3094324\]](https://www.ncbi.nlm.nih.gov/pmc/PMC3094324). [https://doi.org/10.1186/1471-2407-11-136.](https://doi.org/10.1186/1471-2407-11-136)
- <span id="page-9-0"></span>41. Zhang KJ, Tan XL, Guo L. The long non-coding RNA DANCR regulates the inflammatory phenotype of breast cancer cells and promotes breast cancer progression via EZH2-dependent suppression of SOCS3 transcription. Mol Oncol. 2020;14(2):309-28. [PubMed ID: [31860165](http://www.ncbi.nlm.nih.gov/pubmed/31860165)]. [PubMed Central ID: [PMC6998389](https://www.ncbi.nlm.nih.gov/pmc/PMC6998389)]. [https://doi.org/10.1002/1878-](https://doi.org/10.1002/1878-0261.12622) [0261.12622](https://doi.org/10.1002/1878-0261.12622).
- <span id="page-9-1"></span>42. Barclay JL, Anderson ST, Waters MJ, Curlewis JD. SOCS3 as a tumor suppressor in breast cancer cells, and its regulation by PRL. Int J<br>Cancer. 2009;124(8):1756-66. [PubMed ID: 19115200]. 2009;124(8):1756-66. [https://doi.org/10.1002/ijc.24172.](https://doi.org/10.1002/ijc.24172)
- <span id="page-9-2"></span>43. Kaufmann SH, Earnshaw WC. Induction of apoptosis by cancer chemotherapy. Exp Cell Res. 2000;256(1):42-9. [PubMed ID: [10739650\]](http://www.ncbi.nlm.nih.gov/pubmed/10739650). <https://doi.org/10.1006/excr.2000.4838>.
- <span id="page-9-3"></span>44. Marvalim C, Datta A, Lee SC. Role of p53 in breast cancer progression: An insight into p53 targeted therapy. Theranostics. 2023;13(4):1421-42. [PubMed ID: [36923534](http://www.ncbi.nlm.nih.gov/pubmed/36923534)]. [PubMed Central ID: [PMC10008729\]](https://www.ncbi.nlm.nih.gov/pmc/PMC10008729). <https://doi.org/10.7150/thno.81847>.
- <span id="page-9-4"></span>45. Goh AM, Coffill CR, Lane DP. The role of mutant p53 in human cancer.<br>  $\int$  Pathol. 2011;223(2):116-26. [PubMed ID: 21125670]. Pathol. 2011;223(2):116-26. <https://doi.org/10.1002/path.2784>.