

# The Effect of Caffeic Acid Phenethyl Ester on Development of Left Ventricular Dysfunction in Cirrhotic Rats

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#### ARTICLE INFO

*Article Type:* Research Article

Article History: Received: 18 Dec 2015 Accepted: 17 Jul 2016

*Keywords:* Caffeic Acid Phenethyl Ester Left Ventricular Function Cardiomyopathy

#### ABSTRACT

**Background:** Cardiomyopathy is one of the major complications of cirrhosis. Free radical scavenging and antioxidant substances can reduce cardiomyopathy and cardiovascular mortality of cirrhosis. Caffeic Acid Phenethyl Ester (CAPE) is one of the major compounds of polyphenols with anti-inflammatory and free-radical scavenging activities.

**Objectives:** The present study aimed to identify the association between mitochondrial mutations in tRNA<sup>Gha</sup> and Cytb genes and CHD in Iranian patients.

Materials and Methods: In this study, thirty male Sprague-Dawley rats (200 - 250 g) were divided into three groups (sham, cirrhotic, and cirrhotic treated with CAPE). CAPE was administrated (1  $\mu$ g/kg/day, ip) for 5 weeks. Bile Duct Ligation (BDL) was applied in order to induce cirrhosis. Five weeks after surgery, left ventricular function was evaluated by Langendorff system, and data of the sham group were considered as ventricular function before the BDL surgery. For histopathological evaluation also, livers of all the rats were removed and processed for light microscopy. Then, all the serum biochemical parameters were measured. After all, the data were entered into the SPSS statistical software, version 18 and were analyzed using one-way ANOVA followed by LSD test. The data were presented as mean  $\pm$  SEM and P < 0.05 was considered to be statistically significant.

**Results:** The maximum rate of increase and decrease in ventricular pressure ( $\pm$  dp.dt-1), which was reduced by cirrhosis (P < 0.001) improved significantly by CAPE (P < 0.001). **Conclusions:** The findings indicated that CAPE improved the left ventricular function in cirrhotic rats. Thus, it may be considered to be a new therapeutic approach in future cirrhotic cardiomyopathy.

► Implication for health policy/practice/research/medical education:

These results may be considered as a new therapeutic app roach in future cirrhotic cardiomyopathy.

#### 1. Background

Cirrhosis is associated with numerous cardiovascular abnormalities, which are often followed by chronotropic and inotropic dysfunction known as cirrhotic cardiomyopathy (1). Cirrhotic cardiomyopathy consists of systolic and diastolic dysfunction and electrophysiological abnormalities in the absence of any known cardiac diseases (2). This syndrome is characterized by portal hypertension and cirrhosis as well as intrinsic alterations in myocardial function. Several lines of evidence have supported some pathological etiologies for cirrhotic cardiomyopathy, such as abnormalities in the  $\beta$ -adrenergic signaling pathway, changes in cardiomyocyte membrane fluidity, and ion channels defect. Besides, many researchers have indicated that various compounds, such as Nitric Oxide (NO) and Carbon Monoxide (CO) that are

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elevated in cirrhosis, had negative inotropic effects on the myocardium (3). Moreover, many studies have demonstrated that impairment of the antioxidant defense system was involved in a number of pathological conditions, including cardiovascular diseases (4, 5).

Caffeic Acid Phenethyl Ester (CAPE) is the active component of the polyphenol purified from the hives of honeybees. The composition and properties of CAPE have been extensively investigated and reported to have many beneficial effects on human health, including reduction of cardiovascular diseases. Many researchers have also shown that CAPE compounds possessed a broad spectrum of antioxidant (6, 7), anti-inflammatory (7-9), anticarcinogenic (10, 11), antiviral (7, 12), immunomodulatory (13, 14), and free radical scavenging properties (15) that protected organs against free radicals and oxidative stress. Furthermore, CAPE had protective effects on tissues, such as brain, heart (16, 17), and spinal cord (18), and was a potent anti-arrhythmic agent (19). CAPE also had a protective effect against carbon tetrachloride-induced liver (20) and kidney injuries in rats and mice (21).

# 2. Objectives

Many studies have indicated that impairment of the antioxidant defense system was engaged in cardiovascular diseases and that CAPE had a significant effect on scavenging free radicals. Therefore, we hypothesized that CAPE could play a striking role in myocardial dysfunction. This study aims to determine the effects of CAPE on the left ventricular function in cirrhotic rats.

# 3. Materials and Methods

# 3.1. Animal Model

This study was performed on 30 male Sprague-Dawley rats weighing 200 - 250 g prepared by the research center for proliferation and maintenance of laboratory animals, Ahvaz Jundi Shapour University of Medical Sciences. The animals were kept in separate cages under standard laboratory conditions ( $25 \pm 2^{\circ}$ C and 12:12 h light-dark cycle) with free access to food and water. The study protocol was approved by the local Ethics Committee of Ahvaz Jundi Shapour University of Medical Sciences.

Chemical Agents: CAPE (C8221) was purchased from Sigma-Aldrich Company, Germany.

# 3.2. Experimental Design

Extra hepatic cholestasis was induced by double ligation. At first, all the animals were anesthetized by ketamine (50 mg/kg) and xylazine hydrochloride (10 mg/kg) (22). After a midline abdominal incision close to the sternum, the common bile duct was identified, a double ligature was made with 4 - 0 silk, and a cut was made between both ligatures (23, 24). The muscle and skin were then sutured separately with 3 - 0 silk. Sham-operated control rats also received the same laparotomy, except that the bile duct was not ligated and sectioned. This group was used for evaluation of the ventricular function before Bile Duct Ligation (BLD). It was previously shown that rats underwent typical histological biliary cirrhotic changes during the 5-week period after ligation. Accordingly, all the experiments were performed

for 5 weeks postoperatively (25, 26).

Then animals were randomly divided into three groups, namely sham-operated (sham) (n = 10), BDL, and BDL treated with CAPE. In the CAPE-treated group, CAPE was administrated daily (1  $\mu$ g/kg/day, ip) for 5 weeks (19) and other groups received a similar volume of saline (16, 27).

# 3.3. Cardiac Contractility Measurement

On the trial day, the animals were anesthetized with ketamine (50 mg/kg) and xylazine hydrochloride (10 mg/ kg). After opening the chest, the hearts were rapidly excised and placed into a petri dish containing ice-cold oxygenated modified Krebs-Henseleit Solution (KHS). After being washed with ice-cold KHS and arrested, the hearts were cannulated via the ascending aorta and immediately transferred to Langendorff system. A modified KHS of the following composition (mM): NaCl 118.4, KCl 4.7, MgSO4 H2O 1.2, KH2PO4 2H2O 1.2, NaHCO3 25, CaCl2 2.5, and glucose 11.1 in distilled water retro perfused the aorta. This solution was maintained at 37 °C, bubbled with 5% CO2 and 95% O2, and perfused the isolated hearts via peristaltic pump (Gilson-France) at a constant flow rate (4 mL/min). The maximum time between the excision of the heart and the beginning of perfusion was 2 minutes (28).

To record the Left Ventricular Pressure (LVP), a latex balloon filled with water was placed in the left ventricle and was connected to a pressure transducer (Power lab 8/30 AD Instrument, Australia). After the stabilization period (30 minutes with a diastolic pressure between 1 - 6 mm Hg), left ventricular diastolic pressure was increased by 10 mmHg every 5 minutes, reaching 10, 20, 30, and 40 mmHg.10 For each diastolic pressure, the maximum rate of pressure rise (+dP/dT max) and fall (-dP/dT max) was calculated as the first derivation of the LVP (28).

# 3.4. Determination of Serum Biochemical Parameters

After opening the chest, blood samples were collected in tubes and the sera were removed by centrifugation at 3000 rpm for 5 min. Prior to determining the biochemical parameters, all the serum samples were kept at 4 °C. Serum Aspartate Aminotransferase (AST), Alanine Aminotransferase (ALT), and Alkaline Phosphatase (ALP) were checked with an AU600 multiparameter analyzer (Olympus, Hamburg, Germany). Additionally, serum bilirubin was measured as a marker to show the occurrence of cholestasis (27, 29). Then, total and conjugated bilirubin were tested by kits (pars azmoon, Iran) through colorimetric method using an auto-analyzer (Olympus, Hamburg, Germany) (30).

# 3.5. Evaluation of Tissue Damage

Livers of all the rats were removed immediately after the blood collection process. Then, liver tissue specimens were washed twice with a cold saline solution, placed into glass bottles containing formalin 10%, and stored at 25 °C for histopathological examination (31).

# 3.6. Statistics Analysis

The data were presented as mean  $\pm$  SEM and were analyzed through one-way analysis of variance. In case

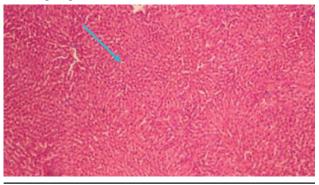
of significant difference, comparisons were made using post-hoc (LSD) test. All the analyses were performed using the SPSS statistical software, version 18) and  $P \le 0.05$  was considered to be statistically significant.

#### 4. Results

#### 4.1. Biochemical Findings

Serum levels of AST, ALT, and AlP significantly increased in the cirrhosis rats (P = 0.001) compared to the sham group. However, a significant reduction was observed in ALT (P = 0.038), AST (P = 0.0001), and ALP (P = 0.008) in the CAPE-treated group compared to the sham rats. Moreover, conjugated and total bilirubin levels were significantly higher in the cirrhotic and CAPE-treated groups than in the sham group (P = 0.001). However, no significant difference was found between the cirrhotic and CAPE-treated groups regarding bilirubin levels (P = 0.999) (Table 1).

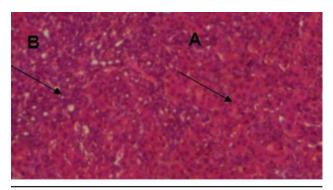
The results of conventional histopathological examination confirmed the protective effects of CAPE on cirrhosis induced by BDL, as indicated by the biochemical data. In this regard, livers of the rats in the sham group showed a normal histological appearance (Figure 1), whereas those of the rats with BDL showed classic histology of cirrhosis, coagulated necrosis, fibrosis, deposition of connective tissue, and formation of regenerative nodules (Figure 2, 3). Nevertheless, these changes decreased in the CAPEtreated group.



**Figure 1.** Section of a Control Liver Showing a Normal Histological Appearance; Hematoxylin Eosin Staining; Magnification × 100. CV, Central Vein.

# 4.2. The Maximum Rate of Pressure Rise (+dP/dt Max) and Fall (-dP/dt Max) in Different Groups

Based on the results, BDL-induced cirrhosis significantly reduced maximum and minimum rates of pressure rise ( $\pm$  dP/dT) compared to the sham group (P = 0.048). Yet,  $\pm$  dP/dt was completely improved by CAPE (P = 0.002) (Figures 4, 5).



**Figure 2.** Liver Section of a Rat after BDL Classic Cirrhotic Appearance with the Presence of Coagulative Necrosis, Mononuclear Cell Infiltration (Small Arrows), Hemorrhage (h), Microvesicular Steatosis, and Hydropic Degeneration (Large Arrow); Hematoxylin Eosin Staining; magnification × 100.

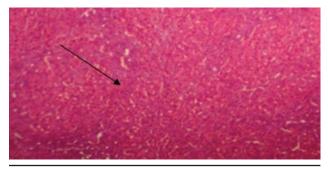


Figure 3. A Liver Section in the Cirrhotic Group Treated with CAPE ( $\mu$ g/kg) Hematoxylin Eosin Staining; Magnification × 100.

# 5. Discussion

The results of the present study showed that treatment with CAPE caused a partial correction of liver enzymes, contractility, and tissue injuries induced by cirrhosis, but had no effects on conjugated and total bilirubin levels. Previous studies indicated that closure of the bile duct through lipid peroxidation caused disorders in TNF- $\alpha$ , IL1- $\beta$ , IL- $\beta$  cytokines, and necrosis (30).

In the current study, serum aminotransferases as sensitive indicators of damage to hepatocytes were evaluated to determine the degree of inflammation (32). Coban et al. reported that oxidative stress associated with lipid peroxidation was involved in development of liver damage in cirrhotic rats by BDL (33). Additionally, other studies demonstrated that CAPE prevented the formation of Reactive Oxygen Species (ROS) and Malondialdehyde (MDA), as one of the secondary products of oxidative stress that is formed during lipid peroxidation and is one of the factors

Table 1. Comparison of the Groups Regarding Serum Biochemical Parameters			
Parameter	Sham	Cirrhotic	Cirrhotic + CAPE
AST (U/L)	$127 \pm 15.8$	$386.6 \pm 15.2^{a}$	$391.4 \pm 34.2^{a}$
ALT (U/L)	$47.2 \pm 3.6$	$103 \pm 14.3^{\mathrm{b}}$	$89.1 \pm 5.3$ <sup>b</sup>
ALP (U/L)	$621 \pm 60$	$1141 \pm 69.2^{a}$	$1108 \pm 69.7$ <sup>a</sup>
Conjugated bilirubin (mg/dL)	$0.06\pm0.008$	$4.9 \pm 1.02^{\text{ b}}$	$4.9\pm0.86^{\mathrm{b}}$
Total bilirubin (mg/dL)	$0.17\pm0.007$	$7.9 \pm 1.3^{\mathrm{b}}$	$7.9 \pm 1.2^{\text{ b}}$

Abbreviations: AST, aspartate aminotransferase; ALT, alanine aminotransferase; ALP, alkaline phosphatase

<sup>a</sup> Significantly different compared to the sham group (P < 0.01).; <sup>b</sup> Significantly different compared to the sham group (P < 0.05). Values are expressed as mean  $\pm$  SD (n = 10).

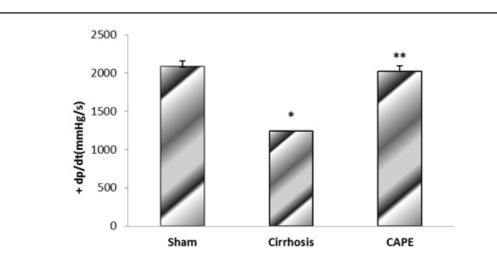


Figure 4. Comparison of the Maximum Rate of Pressure Rise (+dP/d Max) in Different Groups

Values have been expressed as mean  $\pm$  SEM. \*Significantly different compared to the sham group at P < 0.05. \*\*Significantly different compared to the sham group at P < 0.01.

Sham Cirrhosis CAPE 400 -400 -1200 -1600 -2000 -

Figure 5. Comparison of the minimum rate of pressure rise (-dP/dt) in different groups

Values have been expressed as mean  $\pm$  SEM. \*Significantly different compared to the sham group at P < 0.05. \*Significantly different compared to the sham group at P < 0.01.

of toxic ROS in rats after BDL (18, 34, 35). Thus, reduction of aminotransferases found in the CAPE-treated group in our study can be associated with the antioxidative effect of CAPE. Moreover, a previous study showed that CAPE injection (2.84 mg / kg for 4 weeks) reduced the total and conjugated bilirubin levels in liver toxicity induced by carbon tetrachloride in rats (34). In our study, however, CAPE could not change the elevation of bilirubin levels, which might be due to the difference in the administrated doses.

Cirrhotic cardiomyopathy is characterized by heart dysfunction caused by cirrhosis in the absence of any known cardiovascular diseases, such as coronary artery disease, congenital or valvular heart disease, and hypertension (2). Indeed, heart is one of the body organs with a high oxygen consumption rate and slow turnover of antioxidant enzymes. Thus, it is highly liable to oxidative damage. On the other hand, cirrhosis not only increases the production of free radicals, but also impairs the antioxidant defense system (36). NO and CO are produced in the cirrhotic heart by increased inducible NO synthase and hemoxygenase, respectively. Many researchers have asserted that stimulation of the NO pathway prompted an increase in cytokines in cirrhosis (30). For instance, increased Tumor Necrosis Factor- $\alpha$  (TNF- $\alpha$ ) and cyclic Guanosine Monophosphate (cGMP) contents have been reported in the BDL model of cirrhotic rats (3). These findings supported the role of NO and CO in mediating the reduced contractile response in cirrhosis. Both NO and CO stimulate guanylate cyclase to produce cGMP, which in turn phosphorylates protein kinase G to inhibit calcium influx into the cytosol of the cardiomyocyte (37).

Liu et al. indicated that activation of the hemeoxygenasecarbon monoxide pathway played a significant role in pathogenesis of cirrhotic cardiomyopathy in rats (29). Heme Oxygenase (HO) degrades heme to biliverdin and CO and then, CO depresses cardiac contraction through cGMP. Furthermore, proinflammatory cytokines increase their own production as well as the synthesis of small inflammatory mediators, such as Platelet-Activating Factor (PAF) and oxidative radicals. These results suggest that increased CO production caused by HO activation might play an important role in the pathogenesis of cirrhotic cardiomyopathy (29).

Ward et al. investigated the status of cellular Ca2+regulatory system in a rat model of cirrhotic cardiomyopathy. Their results revealed that L-type Ca2+ channels protein expression was reduced and the peak of inward Ca2+ current was significantly less in cirrhotic rats compared to the controls. Thus, they concluded that reduction of cardiac contractility in cirrhotic cardiomyocytes was caused by Ca2+ regulatory system dysfunction (38).

Chang et al. disclosed that CAPE reduced the occurrence of reperfusion-induced ventricular fibrillation and decreased LVP in isolated hearts. In papillary muscles, CAPE shortened the potential duration of operation and reduced both the maximum upstroke velocity and contractile force. In single ventricular myocytes, CAPE shortened this period whereby the Ca2+ transient amplitude was reduced. Patchclamp experiments revealed that CAPE produced a usedependent decrement in L-type Ca2+ current (ICa, L)  $(IC50 = 1.1 \ \mu M)$  and Na+ current (INa)  $(IC50 = 0.43 \ \mu M)$ , which caused a negative shift of the voltage-dependent inactivation and a delay in recovery from inactivation. CAPE also decreased the delayed outward K+ current (IK) slightly without affecting the inward rectifier K+ current (IK1). In the present study, CAPE as an antioxidant (39) could improve myocardial contractility determined by dp/dt, which was attributed to the above-mentioned mechanisms (40). Therefore, these findings indicated that CAPE could reduce oxidative stress in cholestatic rats through interfering with the free radical species (34).

Besides, several studies have demonstrated that this substance led to a decrease in liver damage following bile duct closure in rats. Another study reported that CAPE had anti-inflammatory properties by inhibiting the release of arachidonic acid from cell membranes. Reduction of arachidonic acid metabolism was also observed in cultured human oral epithelial cells as well as in carrageenaninduced inflammation of air pouch model in rats. The antiinflammatory properties of CAPE have been attributed to suppression of eicosanoid synthesis (8).

Since NO synthesized by inducible Nitric Oxide Synthase (iNOS) has been known to be involved in inflammatory responses, it seems that modulation of NO synthesis can be a new approach for treatment of inflammatory diseases. CAPE has been identified to show anti-inflammatory activities (25). Other studies have shown that CAPE might exert its anti-inflammatory effect by inhibiting the iNOS gene expression at the transcriptional level through suppression of NF- $\kappa$ B activation and direct inhibition of the catalytic activity of iNOS (35, 41). Hence, reduction of necrosis, edema, and degenerative changes in the liver tissues observed in our study was caused by the antiinflammatory effect of caffeic acid. Since other studies have shown that cardiovascular complications in cirrhotic patients resulted from changes in ions permeability (such as potassium and calcium), ion channels behavior (such as L-type calcium channel), cardio depressant agents, and

increased free radicals (42), it has been suggested that CAPE prevented these complications with free radical scavenging, antioxidant, and anti-inflammatory properties.

#### 5.1. Conclusion

The findings of the present study indicated that CAPE improved the left ventricular function in cirrhotic rats. Thus, it may be regarded as a new therapeutic approach in future cirrhotic cardiomyopathy.

#### Acknowledgements

This study was extracted from Samira Saadatfard's M.Sc. thesis. Hereby, the authors would like to acknowledge the scholarly and financial support of the Physiology Research Center of Ahvaz Jundishapour University of Medical Sciences (grant No. PRC-61).

#### **Authors' Contribution**

Mohammad Badavi: Conception and design of the study (50%), drafting, critical revision for important intellectual content (80%), final approval of the study, provision of the study material (30%); Najmeh Sadeghi: Collection, assembly, and possession of the raw data (doing experiments), provision of the study material (20%), critical revision of the manuscript for important intellectual content (10%); Mahin Dianat: Critical revision (10%), provision of the study material (20%), analysis and interpretation of the data (10%), statistical expertise (10%), obtaining funding for the study (10%), conception and design of the study (50%); Samira Saadatfard: Study design (60%), provision of the materials, interpretation the data (100%).

#### **Funding/Support**

This research was supported by Physiology Research Center (PRC) of Ahvaz Jundishapour University of Medical Sciences.

#### **Financial Disclosure**

The authors declare that they have no conflict of interests.

#### References

- Lee RF, Glenn TK, Lee SS. Cardiac dysfunction in cirrhosis. Best Pract Res Clin Gastroenterol. 2007;21(1):125-40.
- Moller S, Henriksen JH. Cardiovascular dysfunction in cirrhosis. Pathophysiological evidence of a cirrhotic cardiomyopathy. *Scand J Gastroenterol*. 2001;36(8):785-94.
- 3. Liu H, Ma Z, Lee SS. Contribution of nitric oxide to the pathogenesis of cirrhotic cardiomyopathy in bile duct-ligated rats. *Gastroenterology*. 2000;**118**(5):937-44.
- Moller S, Henriksen JH. Cirrhotic cardiomyopathy: a pathophysiological review of circulatory dysfunction in liver disease. *Heart*. 2002;87(1):9-15.
- 5. Moller S, Henriksen JH. Cirrhotic cardiomyopathy. *J Hepatol.* 2010;**53**(1):179-90.
- Oktar S, Aydin M, Yonden Z, Alcin E, Ilhan S, Nacar A. Effects of caffeic acid phenethyl ester on isoproterenol-induced myocardial infarction in rats. *Anadolu Kardiyol Derg.* 2010;10(4):298-302.
- Okutan H, Ozcelik N, Yilmaz HR, Uz E. Effects of caffeic acid phenethyl ester on lipid peroxidation and antioxidant enzymes in diabetic rat heart. *Clin Biochem.* 2005;38(2):191-6.
- Michaluart P, Masferrer JL, Carothers AM, Subbaramaiah K, Zweifel BS, Koboldt C, *et al.* Inhibitory effects of caffeic acid phenethyl ester on the activity and expression of cyclooxygenase-2 in human oral epithelial cells and in a rat model of inflammation. *Cancer Res.* 1999;**59**(10):2347-52.

- Orban Z, Mitsiades N, Burke TR, Jr., Tsokos M, Chrousos GP. Caffeic acid phenethyl ester induces leukocyte apoptosis, modulates nuclear factor-kappa B and suppresses acute inflammation. *Neuroimmunomodulation*. 2000;7(2):99-105.
- Chen YJ, Shiao MS, Wang SY. The antioxidant caffeic acid phenethyl ester induces apoptosis associated with selective scavenging of hydrogen peroxide in human leukemic HL-60 cells. *Anticancer* Drugs. 2001;12(2):143-9.
- Orsolic N, Terzic S, Mihaljevic Z, Sver L, Basic I. Effects of local administration of propolis and its polyphenolic compounds on tumor formation and growth. *Biol Pharm Bull*. 2005;28(10):1928-33.
- Fesen MR, Pommier Y, Leteurtre F, Hiroguchi S, Yung J, Kohn KW. Inhibition of HIV-1 integrase by flavones, caffeic acid phenethyl ester (CAPE) and related compounds. *Biochem Pharmacol*. 1994;48(3):595-608.
- Albukhari AA, Gashlan HM, El-Beshbishy HA, Nagy AA, Abdel-Naim AB. Caffeic acid phenethyl ester protects against tamoxifen-induced hepatotoxicity in rats. *Food Chem Toxicol*. 2009;47(7):1689-95.
- 14. Park EH, Kahng JH. Suppressive effects of propolis in rat adjuvant arthritis. *Arch Pharm Res.* 1999;**22**(6):554-8.
- Wu W-M, Lu L, Long Y, Wang T, Liu L, Chen Q, *et al.* Free radical scavenging and antioxidative activities of caffeic acid phenethyl ester (CAPE) and its related compounds in solution and membranes: a structure–activity insight. *Food Chemistry*. 2007;105(1):107-15.
- 16. Fadillioglu E, Oztas E, Erdogan H, Yagmurca M, Sogut S, Ucar M, *et al.* Protective effects of caffeic acid phenethyl ester on doxorubicininduced cardiotoxicity in rats. *J Appl Toxicol.* 2004;**24**(1):47-52.
- Mollaoglu H, Gokcimen A, Ozguner F, Oktem F, Koyu A, Kocak A, *et al.* Caffeic acid phenethyl ester prevents cadmium-induced cardiac impairment in rat. *Toxicology*. 2006;**227**(1-2):15-20.
- Ilhan A, Koltuksuz U, Ozen S, Uz E, Ciralik H, Akyol O. The effects of caffeic acid phenethyl ester (CAPE) on spinal cord ischemia/reperfusion injury in rabbits. *Eur J Cardiothorac Surg.* 1999;16(4):458-63.
- Huang SS, Liu SM, Lin SM, Liao PH, Lin RH, Chen YC, et al. Antiarrhythmic effect of caffeic acid phenethyl ester (CAPE) on myocardial ischemia/reperfusion injury in rats. *Clin Biochem*. 2005;**38**(10):943-7.
- Lee KJ, Choi JH, Khanal T, Hwang YP, Chung YC, Jeong HG. Protective effect of caffeic acid phenethyl ester against carbon tetrachloride-induced hepatotoxicity in mice. *Toxicology*. 2008;248(1):18-24.
- Ogeturk M, Kus I, Colakoglu N, Zararsiz I, Ilhan N, Sarsilmaz M. Caffeic acid phenethyl ester protects kidneys against carbon tetrachloride toxicity in rats. *J Ethnopharmacol*. 2005;97(2):273-80.
- 22. Mitchell GF, Jeron A, Koren G. Measurement of heart rate and Q-T interval in the conscious mouse. *Am J Physiol.* 1998;**274**(3 Pt 2):H747-51.
- Liu Y, Binz J, Numerick MJ, Dennis S, Luo G, Desai B, et al. Hepatoprotection by the farnesoid X receptor agonist GW4064 in rat models of intra- and extrahepatic cholestasis. J Clin Invest. 2003;112(11):1678-87.
- Rivera-Huizar S, Rincon-Sanchez AR, Covarrubias-Pinedo A, Islas-Carbajal MC, Gabriel-Ortiz G, Pedraza-Chaverri J, *et al.* Renal dysfunction as a consequence of acute liver damage by bile duct ligation in cirrhotic rats. *Exp Toxicol Pathol.* 2006;**58**(2-3):185-95.
- 25. Koyu A, Ozguner F, Yilmaz H, Uz E, Cesur G, Ozcelik N. The protective effect of caffeic acid phenethyl ester (CAPE) on oxidative stress in rat liver exposed to the 900 MHz electromagnetic field.

Toxicol Ind Health. 2009;25(6):429-34.

- 26. Payabvash S, Kiumehr S, Nezami BG, Zandieh A, Anvari P, Tavangar SM, *et al.* Endogenous opioids modulate hepatocyte apoptosis in a rat model of chronic cholestasis: the role of oxidative stress. *Liver Int.* 2007;**27**(4):538-47.
- 27. Ebrahimkhani MR, Moezi L, Kiani S, Merat S, Dehpour AR. Opioid receptor blockade improves mesenteric responsiveness in biliary cirrhosis. *Dig Dis Sci.* 2008;**53**(11):3007-11.
- De Angelis K, Oliveira A, Dall'Ago P, Peixoto L, Gadonski G, Lacchini S, et al. Effects of exercise training on autonomic and myocardial dysfunction in streptozotocin-diabetic rats. *Brazilian Journal of medical and biological research*. 2000;33(6):635-41.
- 29. Liu H, Song D, Lee SS. Role of heme oxygenase-carbon monoxide pathway in pathogenesis of cirrhotic cardiomyopathy in the rat. *Am J Physiol Gastrointest Liver Physiol*. 2001;**280**(1):G68-74.
- Fernández-Martínez E, Pérez-Álvarez V, Tsutsumi V, Shibayama M, Muriel P. Chronic bile duct obstruction induces changes in plasma and hepatic levels of cytokines and nitric oxide in the rat. *Experimental and Toxicologic Pathology*. 2006;**58**(1):49-58.
- Rabani V, Baharvand H, Pournasr B, Farzaneh Z. Animal Models of Liver Cirrhosis and Fibrosis. *Journal of Iranian Anatomical Sciences*.7:163-84.
- 32. Braunwald EF, Kasper DLL DL. Harrison principle of internal medicine. In: Fauci ASK DLL DL, Braunwald E, Hauser SL, Jameson J, Larry Loscalzo J, editors. *Liver and biliary tract disorders*. 17th ed. Tehran; 2008. p. 12-4.
- Coban S, Yildiz F, Terzi A, Al B, Ozgor D, Ara C, *et al.* The effect of caffeic acid phenethyl ester (CAPE) against cholestatic liver injury in rats. *J Surg Res.* 2010;**159**(2):674-9.
- Kus I, Colakoglu N, Pekmez H, Seckin D, Ogeturk M, Sarsilmaz M. Protective effects of caffeic acid phenethyl ester (CAPE) on carbon tetrachloride-induced hepatotoxicity in rats. *Acta Histochem*. 2004;**106**(4):289-97.
- Russo A, Longo R, Vanella A. Antioxidant activity of propolis: role of caffeic acid phenethyl ester and galangin. *Fitoterapia*. 2002;73 Suppl 1:S21-9.
- 36. Lu Y, Cederbaum AI. CYP2E1 and oxidative liver injury by alcohol. *Free Radic Biol Med.* 2008;**44**(5):723-38.
- Mery P, Pavoine C, Belhassen L, Pecker F, Fischmeister R. Nitric oxide regulates cardiac Ca2+ current. Involvement of cGMPinhibited and cGMP-stimulated phosphodiesterases through guanylyl cyclase activation. *Journal of Biological Chemistry*. 1993;268(35):26286-95.
- Ward CA, Liu H, Lee SS. Altered cellular calcium regulatory systems in a rat model of cirrhotic cardiomyopathy. *Gastroenterology*. 2001;**121**(5):1209-18.
- Zhou Y, Fang SH, Ye YL, Chu LS, Zhang WP, Wang ML, et al. Caffeic acid ameliorates early and delayed brain injuries after focal cerebral ischemia in rats. *Acta Pharmacol Sin*. 2006;27(9):1103-10.
- Chang GJ, Chang CJ, Chen WJ, Yeh YH, Lee HY. Electrophysiological and mechanical effects of caffeic acid phenethyl ester, a novel cardioprotective agent with antiarrhythmic activity, in guinea-pig heart. *Eur J Pharmacol*. 2013;**702**(1-3):194-207.
- 41. Natarajan K, Singh S, Burke TR, Grunberger D, Aggarwal BB. Caffeic acid phenethyl ester is a potent and specific inhibitor of activation of nuclear transcription factor NF-kappa B. *Proceedings* of the National Academy of Sciences. 1996;**93**(17):9090-5.
- Zambruni A, Trevisani F, Caraceni P, Bernardi M. Cardiac electrophysiological abnormalities in patients with cirrhosis. J Hepatol. 2006;44(5):994-1002.