



Optimization of the Alpha-Amylase Production from Microbial Source: A Systematic Review of Experimental Studies

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Abstract

Context: The growth and production of enzymes by microorganisms depend highly on the compounds and factors entering the system.

Objectives: The present systematic review aimed to assess the optimized medium content to enhance alpha-amylase production from the microorganisms.

Methods: The PubMed, Embase Web of Science, Scopus, and Google Scholar databases were searched from inception to April 2022, restricted to the English language. The reference list of the included studies was cross-checked, and a partial gray literature search was undertaken. Alpha-amylase production from the microorganisms and optimized medium content by statistical design were the key evaluated outcomes.

Results: Among 995 initially gathered litterateurs, 12 studies were included in this review, which involved bacteria (seven studies), yeasts (two studies), fungi (two studies), and protists (one study). The results indicated that the optimized medium composition by statistical design might increase alpha-amylase production.

Conclusions: Growth conditions, including pH, temperature, and starch concentration, are essential to be optimized to improve alpha-amylase yield.

Keywords: Alpha-Amylase, Microorganism, Production, Medium Culture

1. Context

Enzymes are biological catalysts that are an essential component of biological reactions (1). Most bacteria, yeasts, and fungi are capable of producing amylase. *Aspergillus Oryzae* and *Rhizopus* sp. are important fungi that produce amyolytic enzymes (2). Enzymes are now used in various parts of the industry (3). They are used in detergents, paper industries, textile industries, food industries, and many other industrial applications (4). Alpha-amylase also has important applications in the medical industry. According to studies, it is used to diagnose and treat acute pancreatitis and to differentiate pancreatitis from other causes of acute abdominal diseases. As many as 80% of patients with acute pancreatitis have high levels of amylase. This test is required for the identification of symptoms of pancreatic diseases, such as severe abdominal pain, fever, loss of appetite, and vomiting (5). Serum amylase levels are increased slightly in benign inflammation of the pancreas

and decrease after two days, and this pattern can be used to diagnose this type of disease. This enzyme is sometimes used to treat pancreatic cancer and gallstones. Alpha-amylases have been used to reduce starch, produce beverages, such as beer, treat digestive disorders, and produce cheese from milk (6). Among the most widely used enzymes, alpha-amylase hydrolyzes the carbohydrate chain of starch at random locations to produce short oligosaccharides, maltose, and glucose (7).

The demand for amylase is increasing due to its important role in starch hydrolysis and applications of this hydrolysis process. Compared to chemical methods that require harsh conditions, such as high pressure and high temperatures, the use of microorganisms in the production of enzymes will be much more economical (8). On the other hand, the demand for high-quality products leads to new methods for improving industrial products, such as protease and amylase, which are often used in industry

and medical sciences (9). Most alpha-amylases are secreted extracellularly, but some alpha-amylase are secreted intracellularly (10). Due to the prominent application of alpha-amylase, there is an urgent need to develop cost-effective techniques for the production of stable and efficient alpha-amylase (11). The immersion fermentation method is the traditional method in the production of alpha-amylase.

In general, for the industrial production of more important enzymes, the immersion fermentation method is used because indicators, such as acidity, temperature, aeration, oxygen transfer, and humidity are better controlled (12). Because of a high volume of enzyme production, it easily meets the needs of different industries. The growth and production of enzymes by microorganisms are highly dependent on the compounds and factors entering the system (13). Optimization is the use of one factor at a time so that one of the factors (response variable) changes and the other variables are kept constant (13). The aim of the present systematic review was to assess the optimized content medium to enhance alpha-amylase production from the microbial source.

2. Methods

2.1. Search Strategy

A comprehensive search of PubMed, Embase, ISI Web of Science, Scopus, and Google Scholar was done from inception to April 2022 by two researchers (B. E. and R. H.) independently. The literature search was restricted to the English language. Combinations of the following terms were used in the search strategy: 'Alpha-amylase' OR 'α-amylase' and 'optimization' OR 'improvement' AND 'production' AND 'purification'. The eligible articles were used, and their reference lists were also checked for potential additional articles.

2.2. Eligibility Criteria

2.2.1. Inclusion Criteria

Studies focused on alpha-amylase production, optimization using statistical design, and other important characterizations were worthy of inclusion. Most articles had evaluated the optimization of alpha-amylase production using statistical experimental designs.

2.2.2. Exclusion Criteria

Exclusion criteria were as follows: (1) papers that only reported alpha-amylase activity; (2) articles that only focused on enzyme screening studies or did not assay alpha-amylase activity; (3) studies that lack parts about the screening of significant factors by Plackett-Burman design; (4) conference abstracts, reviews, letters, personal opinions, and book chapters.

2.3. Data Extraction

Two researchers (B. E. and R. H.), according to the above-mentioned inclusion and exclusion criteria, independently extracted and recorded the following information from eligible papers: First author's name, publication year, microbial species, alpha-amylase activity, growth conditions, and screening and optimization of the selected factors. This systematic review was performed based on the preferred reporting items for systematic reviews and meta-analysis (PRISMA) checklist (14). The present study is a systematic review of in vitro studies because its protocol was not registered.

3. Results

3.1. Summary of the Included Studies and Optimization of Enzyme Production

The flow chart of this systematic review for the article selection process is presented in Figure 1. A total of 995 articles were identified in the reference lists of related articles and also our initial search. After 43 duplicate articles were removed, 952 articles were screened based on the title and abstract. Twelve studies were ultimately included in this systematic review, including the optimization of alpha-amylase production in bacteria (seven studies), yeasts (two studies), fungi (two studies), and protists (one study). Box-Behnken design (BBD), central composite design (CCD), and Taguchi methods were the most used statistical designs to calculate process optimization (15-26).

3.1.1. Culture Medium Condition

Several microorganisms, such as bacteria, fungi, yeast, and protist, can produce alpha-amylase; on the other hand, the condition for types of microorganisms is different from optimizing enzyme production. Growth conditions are different for several bacteria types; for instance, the best pH condition for thermophilic *Bacillus subtilis* is 8.4 (27); however, for *Bacillus amyloliquefaciens* 04BBA15 is 5.5 (28). Most research on alpha-amylase-producing microorganisms showed that the best temperature for the production of alpha-amylase is around 35°C (17, 18, 27, 29).

Studies on the optimization of alpha-amylase production by fungi indicated that the best condition to optimize alpha-amylase production is the temperature of 25°C, incubation time of 120 hours, and pH of 7.2 (21, 23, 30). The highest alpha-amylase production in yeast was observed in the pH range from 5.5 to 6 and during incubation of 72 hours (24, 31, 32). A summary of the descriptive culture medium condition of included studies is provided in Table 1.

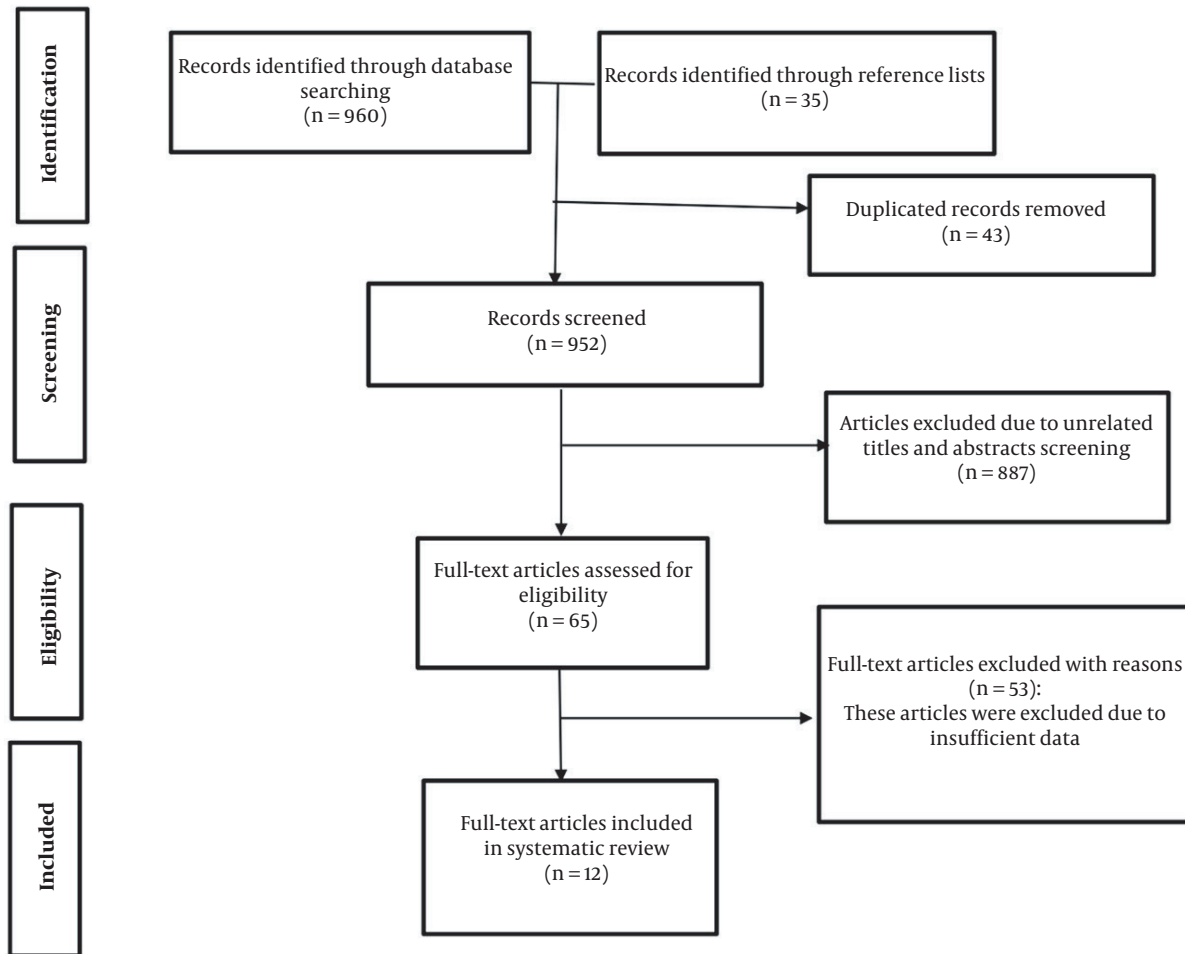


Figure 1. The flow of the search strategy

4. Discussion

According to our knowledge, this is the first systematic review to assess the alpha-amylase production and purification from the microbial source by statistical design. Ten different microorganisms had been reported in the selected studies, including *B. amyloliquefaciens* (15, 16), *Lactobacillus plantarum* (17), *Bacillus acidicola* (18), *Geobacillus thermoleovorans* (19), *Bacillus megaterium* (33), *B. subtilis* (20), *A. oryzae* (21-23), *Aspergillus flavus* (23), *Streptomyces erumpens* (24, 25), and *Ulkenia* sp. (26). Bioprocess factors, including physical conditions and medium composition, are applied for optimized alpha-amylase production. Plackett-Burman design is employed to find the most effective factors for alpha-amylase production. Six studies had used the Plackett-Burman design for screening the significant factors. Significant factors for alpha-amylase produc-

tion are substrate concentration, incubation period, CaCl_2 for the bacteria, temperature, agitation and inoculum size, urea concentration, C/N ratio for fungi, and soluble starch, peptone, NaCl for yeasts.

Several statistical experimental designs, including BBD, CCD, and Taguchi experimental design, have been used in bioprocess optimization of alpha-amylase production from the microorganisms. Seven out of thirty studies had used the central composite design (15, 17-19, 21, 24, 26), Box-Behnken was employed in five studies (16, 22, 23, 25, 33), but only one study optimized alpha-amylase production using Taguchi experimental design (20).

Starch concentration (15, 16, 18, 33) and pH (15, 17, 33) are the most repeated significant factors in the research to optimize the alpha-amylase-concentration of the bacteria. Starch is known as a carbon source, and due to improving the population of alpha-amylase-producing bacte-

Table 1. Summary of Descriptive Culture Medium Condition of Included Studies (N = 12)

Author	Microorganism	Statistical Design	Alpha-Amylase Activity	Optimized Growth Condition
Du et al. (15)	<i>Bacillus amyloliquefaciens</i> BH1	Central composite design	198.26 U/mL	Soluble starch: 12.51 g/L; Na ₂ HPO ₄ : 60.82 g/L; initial pH: 6.81
Gangadharan et al. (16)	<i>Bacillus amyloliquefaciens</i>	Box-Behnken design	965.9 U/mL	Substrate concentration: 12.5%; incubation period: 42 h; CaCl ₂ : 0.0275 M
Panda et al. (17)	<i>Lactobacillus plantarum</i> MTCC 1407	Central composite design	4022 U/min/mL	Incubation period: 36 h; pH: 7.0; temperature: 35°C
Sharma and Satyanarayana (18)	<i>Bacillus acidicola</i>	Central composite design	7440 ± 200 IU l ⁻¹	Starch: 2.75%; K ₂ HPO ₄ : 0.01%; inoculum size: 2% (v/v); temperature: 33°C
Uma Maheswar Rao and Satyanarayana (19)	<i>Geobacillus thermoleovorans</i>	Central composite design	29690 U/L	Glucose: 3%; riboflavin: 200 μg; inoculum density: 3%
Uma Maheswar Rao and Satyanarayana (19)	<i>Bacillus subtilis</i> RSKK96	Taguchi experimental design	503.26 U/mg	Casein: 1%; corn meal: 1%; glutamic acid: 0.01%; incubation time: 72 h; inoculum density: 1%
Gigras et al. (21)	<i>Aspergillus oryzae</i>	Central composite design	161 U/mL	pH: 7.3; incubation time: 120 h; starch: 6.92 g/L, yeast extract: 4.9 g/L KH ₂ PO ₄
Kammoun et al. (22)	<i>Aspergillus oryzae</i> CBS 819.72	Box-Behnken design	Not reported	Temperature, agitation, and inoculum size
Naili et al. (23)	<i>Aspergillus oryzae</i> S2	Box-Behnken design	750 U/mL	Temperature: 24°C; urea: 1 g/L; C/N ratio: 2
Kar and Ray (24)	<i>Streptomyces erumpens</i> MTCC 7317	Central composite design	4143 U	Incubation period: 36 h; pH: 6; temperature: 50°C
Manivasagan et al. (26)	<i>Streptomyces</i> sp. MBRC-82	Box-Behnken design	163.98 U/mL	Soluble starch: 5.8484 g; peptone: 3.5191 g; NaCl: 0.3829
Shirodkar and Muraleedharan (26)	<i>Ulkenia</i> sp.	Central composite design	71.93 U/mL	Glucose: 0.21%; substrate: 0.21%; yeast extract: 0.33%

ria, this factor can increase enzyme production (34). The optimum pH improves the production and secretion of alpha-amylase. Also, bacteria require a neutral pH (around seven) (35), and the best pH for alpha-amylase production is around seven. In the eukaryotic cell, the most repeated significant factor is temperature. The temperature is the most important factor in the production of alpha-amylase by eukaryotic cells (22-24). Also, the temperature is an important factor that directly affects microorganisms' growth and subsequently increases alpha-amylase production (36). The best temperature for alpha-amylase production is directly related to microorganism type; however, the best temperature for enzyme production for fungi is 24°C.

4.1. Conclusions

This systematic review discussed alpha-amylase production by several types of microorganisms, including *B. amyloliquefaciens*, *L. plantarum*, *B. acidicola*, *G. thermoleovorans*, *B. megaterium*, *B. subtilis*, *A. oryzae*, *A. flavus*, and *S. erumpens*. The most significant factors for alpha-amylase production by the bacteria were pH and starch concentration, and for the eukaryotic cell (fungi, yeasts, and *Ulkenia* sp.) was temperature. The starch concentration for

alpha-amylase production is variable by several microorganisms, but the best pH for enzyme production is around seven. Also, the best temperature for fungi to produce alpha-amylase is 24°C. The results showed that the growth conditions (such as pH, temperature, and starch concentration) need to be optimized to increase alpha-amylase yield.

Footnotes

Authors' Contribution: Babak Elyasi Far conceived and designed the evaluation and drafted the manuscript. All authors contributed to the collection and writing of the manuscript.

Conflict of Interests: The authors are four faculty members of the Dezful University of Medical Sciences and one employee of the Mashhad University of Medical Sciences.

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References

- Ye R, Mao X, Sun X, Chen P. Analogy between Enzyme and Nanoparticle Catalysis: A Single-Molecule Perspective. *ACS Catalysis*. 2019;9(3):1985-92. doi:10.1021/acscatal.8b04926.

2. Singh RS, Singh T, Pandey A. Microbial Enzymes—An Overview. In: Singh RS, Singhania RR, Larroche C, editors. *Advances in Enzyme Technology*. Amsterdam, the Netherlands: Elsevier; 2019. p. 1–40. doi: [10.1016/b978-0-444-64114-4.00001-7](https://doi.org/10.1016/b978-0-444-64114-4.00001-7).
3. Meshram A, Singhal G, Bhagyawant SS, Srivastava N. Plant-Derived Enzymes: A Treasure for Food Biotechnology. In: Kuddus M, editor. *Enzymes in Food Biotechnology*. Amsterdam, the Netherlands: Elsevier; 2019. p. 483–502. doi: [10.1016/b978-0-12-813280-7.00028-1](https://doi.org/10.1016/b978-0-12-813280-7.00028-1).
4. Kumari U, Singh R, Ray T, Rana S, Saha P, Malhotra K, et al. Validation of leaf enzymes in the detergent and textile industries: launching of a new platform technology. *Plant Biotechnol J*. 2019;17(6):167–82. doi: [10.1111/pbi.13122](https://doi.org/10.1111/pbi.13122). [PubMed: [30963679](https://pubmed.ncbi.nlm.nih.gov/30963679/)]. [PubMed Central: [PMC6523609](https://pubmed.ncbi.nlm.nih.gov/PMC6523609/)].
5. Elyasi Far B, Ahmadi Y, Yari Khosroshahi A, Dilmaghani A. Microbial Alpha-Amylase Production: Progress, Challenges and Perspectives. *Adv Pharm Bull*. 2020;10(3):350–8. doi: [10.34172/apb.2020.043](https://doi.org/10.34172/apb.2020.043). [PubMed: [32665893](https://pubmed.ncbi.nlm.nih.gov/32665893/)]. [PubMed Central: [PMC7335993](https://pubmed.ncbi.nlm.nih.gov/PMC7335993/)].
6. Arora R; Sonali. Industrial Potential of Microbial Enzymes. In: Sharma S, Sharma NR, Sharma M, editors. *Microbial Diversity, Interventions and Scope*. Singapore, Singapore: Springer; 2020. p. 301–18. doi: [10.1007/978-981-15-4099-8_17](https://doi.org/10.1007/978-981-15-4099-8_17).
7. de Souza Moretti MM, Yu W, Zou W, Franco CML, Albertin LL, Schenk PM, et al. Relationship between the molecular structure of duckweed starch and its in vitro enzymatic degradation kinetics. *Int J Biol Macromol*. 2019;139:244–51. doi: [10.1016/j.ijbiomac.2019.07.206](https://doi.org/10.1016/j.ijbiomac.2019.07.206). [PubMed: [31374280](https://pubmed.ncbi.nlm.nih.gov/31374280/)].
8. Jin M, Gai Y, Guo X, Hou Y, Zeng R. Properties and Applications of Extremozymes from Deep-Sea Extremophilic Microorganisms: A Mini Review. *Mar Drugs*. 2019;17(12). doi: [10.3390/md17120656](https://doi.org/10.3390/md17120656). [PubMed: [31766541](https://pubmed.ncbi.nlm.nih.gov/31766541/)]. [PubMed Central: [PMC6950199](https://pubmed.ncbi.nlm.nih.gov/PMC6950199/)].
9. Bilal M, Iqbal HMN. Sustainable bioconversion of food waste into high-value products by immobilized enzymes to meet bio-economy challenges and opportunities - A review. *Food Res Int*. 2019;123:226–40. doi: [10.1016/j.foodres.2019.04.066](https://doi.org/10.1016/j.foodres.2019.04.066). [PubMed: [31284972](https://pubmed.ncbi.nlm.nih.gov/31284972/)].
10. Soler MT. Comparative study of bacterial and fungal alpha-amylase industrial producers. 2019.
11. Xu T, Jing Peng JP, Zhu Y, Su Li SL, Zhou K, Cheng H, et al. Yield Enhancement of Recombinant α -Amylases in *Bacillus amyloliquefaciens* by ARTP Mutagenesis-Screening and Medium Optimization. *Sains Malays*. 2019;48(5):965–74. doi: [10.17576/jism-2019-4805-04](https://doi.org/10.17576/jism-2019-4805-04).
12. Carmona Cabello M. *Biorefinery design based on the valorization of food industry wastes*. Córdoba, Spain: University of Córdoba; 2020.
13. Al-Maqtari QA, Waleed AA, Mahdi AA. Microbial enzymes produced by fermentation and their applications in the food industry-A review. *Int J Agric Innov Res*. 2019;8(1):2319–1473.
14. Moher D, Liberati A, Tetzlaff J, Altman DG, Prisma Group. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *Ann Intern Med*. 2009;151(4):264–9. W64. doi: [10.7326/0003-4819-151-4-200908180-00135](https://doi.org/10.7326/0003-4819-151-4-200908180-00135). [PubMed: [19622511](https://pubmed.ncbi.nlm.nih.gov/19622511/)].
15. Du R, Zhao F, Qiao X, Song Q, Ye G, Wang Y, et al. Optimization and partial characterization of ca-independent alpha-amylase from *Bacillus amyloliquefaciens* BH1. *Prep Biochem Biotechnol*. 2018;48(8):768–74. doi: [10.1080/10826068.2018.1504221](https://doi.org/10.1080/10826068.2018.1504221). [PubMed: [30303444](https://pubmed.ncbi.nlm.nih.gov/30303444/)].
16. Gangadharan D, Sivaramkrishnan S, Nampoothiri KM, Sukumaran RK, Pandey A. Response surface methodology for the optimization of alpha amylase production by *Bacillus amyloliquefaciens*. *Bioresour Technol*. 2008;99(11):4597–602. doi: [10.1016/j.biortech.2007.07.028](https://doi.org/10.1016/j.biortech.2007.07.028). [PubMed: [17761415](https://pubmed.ncbi.nlm.nih.gov/17761415/)].
17. Panda SH, Swain MR, Kar S, Ray RC, Montet D. Statistical optimization of alpha-amylase production by probiotic *Lactobacillus plantarum* MTCC 1407 in submerged fermentation. *Pol J Microbiol*. 2008;57(2):149–55. [PubMed: [18646403](https://pubmed.ncbi.nlm.nih.gov/18646403/)].
18. Sharma A, Satyanarayana T. Optimization of medium components and cultural variables for enhanced production of acidic high maltose-forming and Ca²⁺-independent alpha-amylase by *Bacillus acidicola*. *J Biosci Bioeng*. 2011;111(5):550–3. doi: [10.1016/j.jbiosc.2011.01.004](https://doi.org/10.1016/j.jbiosc.2011.01.004). [PubMed: [21292551](https://pubmed.ncbi.nlm.nih.gov/21292551/)].
19. Uma Maheswar Rao JL, Satyanarayana T. Statistical optimization of a high maltose-forming, hyperthermostable and Ca²⁺-independent alpha-amylase production by an extreme thermophile *Geobacillus thermoleovorans* using response surface methodology. *J Appl Microbiol*. 2003;95(4):712–8. doi: [10.1046/j.1365-2672.2003.02036.x](https://doi.org/10.1046/j.1365-2672.2003.02036.x). [PubMed: [12969284](https://pubmed.ncbi.nlm.nih.gov/12969284/)].
20. Uysal E, Akcan N, Baysal Z, Uyar F. Optimization of alpha-amylase production by *Bacillus subtilis* RSKK96: using the Taguchi experimental design approach. *Prep Biochem Biotechnol*. 2011;41(1):84–93. doi: [10.1080/10826068.2010.534333](https://doi.org/10.1080/10826068.2010.534333). [PubMed: [21229466](https://pubmed.ncbi.nlm.nih.gov/21229466/)].
21. Gigras P, Sahai V, Gupta R. Statistical media optimization and production of ITS alpha-amylase from *Aspergillus oryzae* in a bioreactor. *Curr Microbiol*. 2002;45(3):203–8. doi: [10.1007/s00284-001-0107-4](https://doi.org/10.1007/s00284-001-0107-4). [PubMed: [12177743](https://pubmed.ncbi.nlm.nih.gov/12177743/)].
22. Kammoun R, Naili B, Bejar S. Application of a statistical design to the optimization of parameters and culture medium for alpha-amylase production by *Aspergillus oryzae* CBS 819.72 grown on gruel (wheat grinding by-product). *Bioresour Technol*. 2008;99(13):5602–9. doi: [10.1016/j.biortech.2007.10.045](https://doi.org/10.1016/j.biortech.2007.10.045). [PubMed: [18180155](https://pubmed.ncbi.nlm.nih.gov/18180155/)].
23. Naili B, Sahnoun M, Bejar S, Kammoun R. Optimization of submerged *Aspergillus oryzae* S2 alpha-amylase production. *Food Sci Biotechnol*. 2016;25(1):185–92. doi: [10.1007/s10068-016-0028-4](https://doi.org/10.1007/s10068-016-0028-4). [PubMed: [30263256](https://pubmed.ncbi.nlm.nih.gov/30263256/)]. [PubMed Central: [PMC6049366](https://pubmed.ncbi.nlm.nih.gov/PMC6049366/)].
24. Kar S, Ray RC. Statistical optimization of alpha-amylase production by *Streptomyces erumpens* MTCC 7317 cells in calcium alginate beads using response surface methodology. *Pol J Microbiol*. 2008;57(1):49–57. [PubMed: [18610656](https://pubmed.ncbi.nlm.nih.gov/18610656/)].
25. Manivasagan P, Venkatesan J, Kang KH, Sivakumar K, Park SJ, Kim SK. Production of alpha-amylase for the biosynthesis of gold nanoparticles using *Streptomyces* sp. MBRC-82. *Int J Biol Macromol*. 2015;72:71–8. doi: [10.1016/j.ijbiomac.2014.07.045](https://doi.org/10.1016/j.ijbiomac.2014.07.045). [PubMed: [25128097](https://pubmed.ncbi.nlm.nih.gov/25128097/)].
26. Shirodkar PV, Muraleedharan UD. Enhanced alpha-amylase production by a marine protist, *Ulkenia* sp. using response surface methodology and genetic algorithm. *Prep Biochem Biotechnol*. 2017;47(10):1043–9. doi: [10.1080/10826068.2017.1373293](https://doi.org/10.1080/10826068.2017.1373293). [PubMed: [29020512](https://pubmed.ncbi.nlm.nih.gov/29020512/)].
27. Al-Johani NB, Al-Seeni MN, Ahmed YM. Optimization Of Alkaline Alpha-Amylase Production By Thermophilic *Bacillus subtilis*. *Afr J Tradit Complement Altern Med*. 2017;14(1):288–301. doi: [10.21010/ajtcam.v14i1.31](https://doi.org/10.21010/ajtcam.v14i1.31). [PubMed: [28480407](https://pubmed.ncbi.nlm.nih.gov/28480407/)]. [PubMed Central: [PMC5411881](https://pubmed.ncbi.nlm.nih.gov/PMC5411881/)].
28. Fossi BT, Tavea F, Fontem LA, Ndjouenkeu R, Wanji S. Microbial interactions for enhancement of alpha-amylase production by *Bacillus amyloliquefaciens* O4BBA15 and *Lactobacillus fermentum* O4BBA19. *Biotechnol Rep (Amst)*. 2014;4:99–106. doi: [10.1016/j.btre.2014.09.004](https://doi.org/10.1016/j.btre.2014.09.004). [PubMed: [28626668](https://pubmed.ncbi.nlm.nih.gov/28626668/)]. [PubMed Central: [PMC5466130](https://pubmed.ncbi.nlm.nih.gov/PMC5466130/)].
29. Yang H, Liu L, Li J, Du G, Chen J. Heterologous expression, biochemical characterization, and overproduction of alkaline alpha-amylase from *Bacillus alcalophilus* in *Bacillus subtilis*. *Microb Cell Fact*. 2011;10:77. doi: [10.1186/1475-2859-10-77](https://doi.org/10.1186/1475-2859-10-77). [PubMed: [21978209](https://pubmed.ncbi.nlm.nih.gov/21978209/)]. [PubMed Central: [PMC3204233](https://pubmed.ncbi.nlm.nih.gov/PMC3204233/)].
30. Zaferanloo B, Bhattacharjee S, Ghorbani MM, Mahon PJ, Palombo EA. Amylase production by *Preussia minima*, a fungus of endophytic origin: optimization of fermentation conditions and analysis of fungal secretome by LC-MS. *BMC Microbiol*. 2014;14:55. doi: [10.1186/1471-2180-14-55](https://doi.org/10.1186/1471-2180-14-55). [PubMed: [24602289](https://pubmed.ncbi.nlm.nih.gov/24602289/)]. [PubMed Central: [PMC3995912](https://pubmed.ncbi.nlm.nih.gov/PMC3995912/)].
31. Chadha BS, Singh S, Vohra G, Saini HS. Shake culture studies for the production of amylases by *Thermomyces lanuginosus*. *Acta Microbiol Immunol Hung*. 1997;44(2):181–5. [PubMed: [9330667](https://pubmed.ncbi.nlm.nih.gov/9330667/)].
32. Acourene S, Ammouche A. Optimization of ethanol, citric acid, and alpha-amylase production from date wastes by strains of *Saccharomyces cerevisiae*, *Aspergillus niger*, and *Candida guilliermondii*. *J Ind Microbiol Biotechnol*. 2012;39(5):759–66. doi: [10.1007/s10295-011-1070-0](https://doi.org/10.1007/s10295-011-1070-0). [PubMed: [22193823](https://pubmed.ncbi.nlm.nih.gov/22193823/)].
33. Elyasi Far B, Dilmaghani A, Yari Khosroshahi A. In Silico Study and

- Optimization of *Bacillus megaterium* alpha-Amylases Production Obtained from Honey Sources. *Curr Microbiol.* 2020;77(10):2593-601. doi: [10.1007/s00284-020-02019-x](https://doi.org/10.1007/s00284-020-02019-x). [PubMed: [32424606](https://pubmed.ncbi.nlm.nih.gov/32424606/)].
34. Ahmed SA, Mostafa FA, Helmy WA, Abdel-Naby MA. Improvement of bacterial α -amylase production and application using two steps statistical factorial design. *Biocatal Agric Biotechnol.* 2017;10:224-33. doi: [10.1016/j.bcab.2017.03.004](https://doi.org/10.1016/j.bcab.2017.03.004).
35. Sivaramakrishnan S, Gangadharan D, Nampoothiri KM, Soccol CR, Pandey A. α -Amylases from microbial sources—an overview on recent developments. *Food Technol Biotechnol.* 2006;44(2):173-84.
36. Woods RG, Burger M, Beven CA, Beacham IR. The aprX-lipA operon of *Pseudomonas fluorescens* B52: a molecular analysis of metallo-protease and lipase production. *Microbiology (Reading)*. 2001;147(Pt 2):345-54. doi: [10.1099/00221287-147-2-345](https://doi.org/10.1099/00221287-147-2-345). [PubMed: [11158351](https://pubmed.ncbi.nlm.nih.gov/11158351/)].