



In vitro Effects of Propofol and Bupivacaine on Pregnant Women's Plasma Cholinesterase Activity and Malondialdehyde Level

Rozheen Shukri Karam ¹ and Fouad Kasim Mohammad ^{2,*}

¹Department of Pharmacology, College of Pharmacy, University of Duhok, KRG, Iraq

²Department of Physiology, Biochemistry and Pharmacology, College of Veterinary Medicine, University of Mosul, Mosul, Iraq

*Corresponding author: Department of Physiology, Biochemistry and Pharmacology, College of Veterinary Medicine, University of Mosul, Mosul, Iraq. Email: fouadmohammad@yahoo.com

Received 2024 January 22; Accepted 2024 February 03.

Abstract

Background: Propofol and bupivacaine are commonly used anesthetics for cesarean section (CS), and they might modulate plasma cholinesterase (ChE) activity and oxidative stress during the last stage of pregnancy. This study aimed to assess the *in vitro* effects of propofol and bupivacaine on plasma ChE activity and malondialdehyde (MDA) levels in pregnant women before undergoing elective CS.

Methods: The plasma samples of 20 women set for elective CS were pooled for the *in vitro* determination of the effects of propofol and bupivacaine separately on plasma ChE activity (10 minutes of incubation with different concentrations at 37°C) and the MDA level after the *in vitro* exposure of plasma samples containing different anesthetic concentrations to H₂O₂ (100 μM, incubated for 1 hour at 37°C).

Results: Bupivacaine at 1.1 and 2.2 μM significantly inhibited plasma ChE *in vitro* in a concentration-dependent manner by 13% and 20%, respectively. Propofol at 25 and 50 μM did not affect plasma ChE. A unique finding in this study was that both propofol and bupivacaine revealed an antioxidant effect, as both propofol at concentrations of 25, 50, and 100 μM and bupivacaine at 1.1, 2.2, and 4.4 μM reduced the MDA level in a concentration-dependent manner *in vitro* after the incubation of plasma samples with H₂O₂ as a source of oxidant.

Conclusions: The *in vitro* findings suggest that bupivacaine exerts anti-ChE activity that should be taken into consideration in CS anesthesia, and both propofol and bupivacaine possess antioxidant properties that need additional clinical studies.

Keywords: Anesthesia, Cesarean Section, Cholinesterase Inhibition, Pregnancy, Oxidative Stress

1. Background

Normal pregnancy is associated with various physiologic and hemodynamic changes, especially during the last month very close to the delivery (1, 2), which might affect the response to drugs or anesthetics (3, 4). Decreased plasma cholinesterase (ChE) activity and increased oxidative stress (OS) biomarkers in the plasma malondialdehyde (MDA) have been reported in the last trimester of pregnancy (5-7). The decrease in plasma ChE activity might predispose pregnant women to the possibility of adverse drug reactions when neuromuscular blocking agents or anesthetics are used, especially in cesarean section (CS) delivery (8-10). A low level of plasma ChE is considered a risk factor in pregnant women (11), and the condition can be complicated when preeclampsia (pregnancy hypertension) coexists (12).

On the other hand, OS biomarkers were reported to increase in pregnancy and were observed to be involved in the pathogenesis of preeclampsia complications (13, 14). The general anesthetic propofol and the spinal one bupivacaine are widely used in CS delivery (15, 16) and might affect plasma ChE activity and oxidative status (7, 17-19). These anesthetics even affect the quality of life after the CS delivery (20, 21).

A recent report implicates the possibility of the existence of health risks from reduced plasma ChE activity during propofol anesthesia and increased OS in women undergoing elective CS delivery (7). However, despite some *in vivo* findings on plasma ChE and MDA levels (5-7), limited information is available on the *in vitro* assessment of anesthetics on pregnant women's plasma. The *in vitro* assessment of plasma ChE activity

and OS biomarkers have been used to minimize invasive activities in patients or experimental animals (22-26). Furthermore, the *in vitro* experimental paradigms avoid the possibility of interference via maternal conditions involving anesthetics and surgical manipulations for CS delivery and the expected post-delivery biochemical changes (2, 6, 14).

2. Objectives

The purpose of the present study was to further explore and ascertain the effects of propofol and bupivacaine on plasma ChE activity and MDA level under *in vitro* conditions without *in vivo* complications.

3. Methods

3.1. Ethical Approval

Female subjects undergoing elective CS deliveries were recruited for the present study (age range: 20 - 45 years), as detailed in a previous study (7). The study subjects were informed about the purpose of the study, and written consent was obtained from each one. The Committee of Postgraduate Studies, College of Pharmacy, University of Duhok, KRG, Iraq, approved the present study (No. 470, October 6, 2021), and the Research Ethics Committee, Duhok Directorate General of Health, Duhok, KRG, Iraq, also confirmed its approval (No. 10112021-11-17, November 10, 2021). Furthermore, this study complied with the ethical standards of the Helsinki Declaration of Ethical Principles for Medical Research Involving Human Subjects, as revised in 2013.

3.2. Blood Sampling

Heparinized 5 mL venous blood samples were obtained only once from each of the 20 women undergoing elective CS just before the induction of anesthesia. The blood samples were centrifuged at 1037 g for 15 minutes to obtain plasma aliquots, which were stored frozen at -20°C for the *in vitro* ChE and MDA assays.

3.3. Used Anesthetics

Propofol 1% (Polifarma, Istanbul, Turkey) and bupivacaine 0.5% (Aguettant Corporate, Lyon, France) were used for the experiments. In this study, none of the participants received anesthetics. Blood samples were taken from them once just before undergoing anesthesia for elective CS delivery. Then, all of the anesthetic *in vitro* experiments were conducted on pooled plasma samples of 20 pregnant women.

3.4. Measurements of Plasma ChE Activity and MDA Level

A modified electrometric method was used to measure plasma ChE activity ($\Delta\text{pH}/20$ min at 37°C) as described earlier using 0.2 mL plasma sample and 0.1 mL of the substrate acetylcholine iodide (7.1%) with an incubation time of 20 minutes (27, 28). The plasma MDA level was measured spectrophotometrically at 535 nm, as described earlier (29).

3.5. *In vitro* Inhibition of Plasma ChE Activity with Propofol or Bupivacaine

On the experimental day, 20 plasma samples were pooled for propofol or bupivacaine in plain glass containers and mixed before taking out plasma aliquots for the ChE assay. We used the method of 10-minute incubation of the inhibitor with the ChE source (plasma) at 37°C (30, 31). The baseline and residual plasma ChE activity was measured electrometrically (27, 28). The inhibitor-ChE combination ($n = 5/\text{concentration}$) included propofol (0-baseline, 25 and $50 \mu\text{M}$) (22, 32) and bupivacaine (0-baseline, 1.1 and $2.2 \mu\text{M}$) (33) in the final reaction mixture (6.3 mL). The choice of these concentrations was based on preliminary experiments and the literature as mentioned above.

3.6. *In vitro* Antioxidant Property of Anesthetic Drugs by Measuring Plasma MDA Level

This method of using plasma samples for the evaluation of OS in *in vitro* tests was, in principle, according to the literature (23, 24). Pooled plasma samples were used with different concentrations of propofol or bupivacaine ($n = 4/\text{concentration}$). The OS *in vitro* test mixtures contained the plasma sample (0.25 mL), propofol at 0, 25, 50, and $100 \mu\text{M}$, bupivacaine at 0, 1.1, 2.2, and $4.4 \mu\text{M}$, or distilled water (-ve and +ve controls). The tubes containing the plasma and the anesthetic were incubated in a water bath at 37°C for one hour. Thereafter, 0.1 mL of H_2O_2 ($100 \mu\text{M}$) as a source of OS was added (32, 34). The sample-mixture contents were then subjected to another one-hour incubation in a water bath at 37°C . Thereafter, the level of MDA in the plasma was determined spectrophotometrically as described above.

3.7. Statistics

The data were statistically analyzed using the analysis of variance followed by Tukey's test, using the software program PAST4.12 (<https://www.nhm.uio.no/english/research/resources/past/>). The level of accepted statistical significance was at $P < 0.05$.

4. Results

4.1. *In vitro* Plasma ChE Activity

After incubating pooled plasma samples of pregnant women set for CS delivery with the anesthetics at 37°C for 10 minutes, propofol at 25 and 50 μM did not significantly ($P = 0.97$ and 0.474 , respectively) affect plasma ChE activity *in vitro*, when compared to the baseline-control value (Table 1). However, the *in vitro* incubation of the samples with bupivacaine at 1.1 and 2.2 μM , in a concentration-dependent manner, was significantly different ($P = 0.02$ and 0.001 , respectively) and inhibited plasma ChE activity by 13% and 20%, respectively, in comparison to the corresponding control value (Table 1).

Table 1. *In vitro* Effects of Propofol and Bupivacaine on Plasma cholinesterase (ChE) Activity of Pregnant Women^a

Treatment	Plasma ChE (Δ pH/20 min)	% Inhibition	P-Value
Propofol (μM)			
0 (Control)	0.69 ± 0.039	-	
25	0.70 ± 0.035	0	0.970
50	0.61 ± 0.065	12	0.474
Bupivacaine (μM)			
0 (Control)	0.82 ± 0.033	-	
1.1	0.71 ± 0.016^b	13	0.02
2.2	$0.66 \pm 0.018^{b,c}$	20	0.001

Abbreviation: ChE, cholinesterase.

^a Values are expressed as mean \pm SE of five measurements/concentrations. Before cesarean section anesthesia, plasma samples from 20 pregnant women were pooled prior to measuring the ChE. All samples were incubated with the anesthetic at 37°C for 10 minutes to facilitate effect on the ChE activity.

^b Significantly different from the corresponding control value, $P < 0.05$.

^c Significantly different from 1.1 μM concentration of bupivacaine, $P < 0.05$.

4.2. *In vitro* Antioxidant Effects of Propofol and Bupivacaine on Plasma MDA Level

We used the *in vitro* method of incubation of plasma samples of pregnant women set for CS delivery by exposing samples containing different concentrations of anesthetics to 100 μM H_2O_2 for 1 hour at 37°C. By using this *in vitro* experimental paradigm, the source of OS H_2O_2 elevated MDA level in the plasma, and prior treatments with both propofol and bupivacaine in a concentration-dependent manner reduced the MDA level when compared to respective control values (Table 2). Propofol at 25, 50, and 100 μM reduced MDA levels in the plasma samples by 6% ($P = 0.933$), 33% ($P = 0.02$), and 49% ($P = 0.001$), respectively (Table 2). Similarly, bupivacaine at 1.1, 2.2, and 4.4 μM reduced MDA levels in the plasma samples by 7% ($P = 0.823$), 17% ($P = 0.202$), and 45% ($P < 0.001$), respectively (Table 2).

5. Discussion

The *in vitro* experiments of the present study supported the fact that plasma ChE activity could be inhibited by the use of bupivacaine but not by propofol. Local anesthetics, specifically bupivacaine, inhibit the butyryl ChE, which is a form of pseudo-ChE, also known as the plasma ChE (17, 35). A recent finding reported reduced plasma ChE activity in women who underwent elective CS with propofol and bupivacaine (7). These *in vivo* findings, however, do not differentiate the drug's effect on blood ChE activity from those of the physiological changes observed in the last month of pregnancy (5, 36).

In light of the importance of plasma ChE, which is associated with the metabolism of anesthetic and neuromuscular blocking agents (36, 37), it seems reasonable to practice caution in dealing with CS cases due to the risk of reduced plasma ChE activity that occurs during pregnancy and from additional intrinsic input from the anesthetic itself, in this instant, bupivacaine. Furthermore, from the clinical standpoint, reduced plasma ChE in pregnancy (38) is a risk factor for women who might develop a condition of preeclampsia (11, 14), exerting additional burden on the body in dealing with neuromuscular blocking drugs (10, 37), anesthetics (8, 9), and even when exposed to pesticides (39).

As the increase in OS is an important biomarker of advanced pregnancy and its complications (12-14, 38), the unique finding of the present study was that both propofol and bupivacaine revealed an antioxidant effect by reducing MDA level after the incubation of the plasma sample with H_2O_2 as a source of oxidant. H_2O_2 has been used to induce OS both *in vivo* and *in vitro* using different cellular systems (32, 34). This study suitably used the plasma of the pregnant women just before the CS, a biological source for oxidation/antioxidation mechanisms, by monitoring the MDA level *in vitro* without the effects of the whole-body biochemical mechanisms on this OS biomarker (6).

The *in vitro* antioxidant effect of propofol correlates with its antioxidant effect reported after *in vivo* administration (7, 40-42). However, such an antioxidant effect has not been reported with bupivacaine, as reported in the present study; therefore, additional *in vitro* and *in vivo* studies are warranted with the use of different types of local anesthetics, especially those with anti-ChE activity.

The clinical relevance of the present study lies in the fact that it draws attention to the versatility of the *in vitro* experimental conditions in demonstrating reduced plasma ChE activity by the local anesthetic and showing the anesthetics' antioxidant effects that could be important in avoiding possible drug-ChE interaction

Table 2. *In vitro* Effect of Propofol and Bupivacaine on Plasma malondialdehyde (MDA) Level of Pregnant Women ^{a, b}

Treatment	MDA ($\mu\text{mol/L}$)	P-Value
Propofol (μM)		
0 (Control)	2.36 \pm 0.05	
25	2.23 \pm 0.129	0.933
50	1.58 \pm 0.21 ^c	0.02
100	1.20 \pm 0.194 ^{c, d}	0.001
Bupivacaine (μM)		
0 (Control)	2.36 \pm 0.05	
1.1	2.19 \pm 0.158	0.823
2.2	1.95 \pm 0.144	0.202
4.4	1.29 \pm 0.158 ^{c, e, f}	< 0.001

Abbreviation: MDA, malondialdehyde

^a Values are expressed as mean \pm SE of four measurements/concentrations.

^b Plasma samples were incubated *in vitro* with the anesthetics for 1 hour at 37°C and then with 100 μM H₂O₂ for another hour at 37°C.

^c Significantly different from the corresponding control value, P < 0.05.

^d Significantly different from 25 μM concentration of propofol, P < 0.05.

^e Significantly different from 1.1 μM concentration of bupivacaine, P < 0.05.

^f Significantly different from 2.2 μM concentration of bupivacaine, P < 0.05.

and for the follow-up of maternal antioxidant status in response to drugs/anesthetics, especially after the CS delivery. Further studies are also needed to determine the potential drug interactions of anti-ChE anesthetics *in vitro* using plasma samples from pregnant women.

5.1. Limitations

An important limitation of the present study is the lack of follow-up of the participants by conducting the same experimental protocol on blood samples after the CS delivery. Two common anesthetics, the general anesthetic propofol and the local anesthetic bupivacaine, were used in the present study. Further studies are needed on different types of anesthetics.

5.2. Conclusions

The *in vitro* experiments of the present study were versatile tools to assess the anti-ChE and anti-OS activity of the anesthetics, and the findings in the present study suggest that bupivacaine exerts anti-ChE activity that should be taken into consideration in CS anesthesia. Moreover, both propofol and bupivacaine possess antioxidant properties that need further assessment in clinical studies.

Acknowledgments

This report represents a portion of a thesis submitted by the first author to the University of Duhok, Iraq, as the partial fulfillment of the requirements for an MSc

degree in clinical pharmacology. The authors would like to express their gratitude to the College of Pharmacy, University of Duhok, for providing facilities and supplies to conduct this study.

Footnotes

Authors' Contribution: RSK dealt with the participants, obtained blood samples, executed laboratory assays, conducted the literature search, performed statistical analyses, and contributed to drafting the manuscript. FKM conceptualized and supervised the study and contributed to the literature search, statistical analyses, and drafting of the manuscript. Both authors have read the manuscript and approved it for publication.

Conflict of Interests: The authors declare that none of them have any competing interests.

Ethical Approval Code: The Committee of Postgraduate Studies, College of Pharmacy, University of Duhok, KRG, Iraq, approved the present study (No. 470, October 6, 2021), and the Research Ethics Committee, Duhok Directorate General of Health, Duhok, KRG, Iraq, also confirmed its approval (No. 10112021-11-17, November 10, 2021).

Funding/Support: The study was supported by the College of Pharmacy, University of Duhok, Duhok, Kurdistan Region, Iraq.

Informed Consent: Written informed consent was obtained from all participants.

References

- Troiano NH. Physiologic and Hemodynamic Changes During Pregnancy. *AACN Adv Crit Care*. 2018;**29**(3):273–83. [PubMed ID: 30185494]. <https://doi.org/10.4037/aacnacc2018911>.
- Pascual ZN, Langaker MD. *Physiology, pregnancy*. StatPearls Publishing; 2023.
- Pinheiro EA, Stika CS. Drugs in pregnancy: Pharmacologic and physiologic changes that affect clinical care. *Semin Perinatol*. 2020;**44**(3):151221. [PubMed ID: 32115202]. [PubMed Central ID: PMC8195457]. <https://doi.org/10.1016/j.semperi.2020.151221>.
- Fiat F, Merghes PE, Scurtu AD, Almajan Guta B, Dehelean CA, Varan N, et al. The Main Changes in Pregnancy-Therapeutic Approach to Musculoskeletal Pain. *Medicina (Kaunas)*. 2022;**58**(8). [PubMed ID: 36013582]. [PubMed Central ID: PMC9414568]. <https://doi.org/10.3390/medicina58081115>.
- Elton RJ. Pregnancy-induced cholinesterase deficiency. *Anaesthesia*. 1999;**54**(4):398. [PubMed ID: 10455851]. <https://doi.org/10.1046/j.1365-2044.1999.00867.x>.
- de Lucca L, Jantsch LB, Vendrame SA, de Paula HL, Dos Santos Stein C, Gallarreta FMP, et al. Variation of the Oxidative Profile in Pregnant Women With and Without Gestational Complications. *Matern Child Health J*. 2022;**26**(10):2155–68. [PubMed ID: 35969329]. <https://doi.org/10.1007/s10995-022-03475-6>.
- Karam RS, Mohammad FK. Changes in blood oxidative stress biomarker and cholinesterase activity after general vs spinal anesthesia for elective cesarean sections. *Anaesth Pain Intensive Care*. 2023;**27**(3):396–404. <https://doi.org/10.35975/apic.v27i3.2244>.
- Andersson ML, Moller AM, Wildgaard K. Butyrylcholinesterase deficiency and its clinical importance in anaesthesia: a systematic review. *Anaesthesia*. 2019;**74**(4):518–28. [PubMed ID: 30600548]. <https://doi.org/10.1111/janae.14545>.
- Inangil G, Deniz S, Kurt YG, Keskin U, Bakal O, Sen H, et al. How important is the butyrylcholinesterase level for cesarean section? *Pregnancy Hypertens*. 2016;**6**(1):26–9. [PubMed ID: 26955768]. <https://doi.org/10.1016/j.preghy.2016.01.004>.
- Zhang C, Cao H, Wan ZG, Wang J. Prolonged neuromuscular block associated with cholinesterase deficiency. *Medicine (Baltimore)*. 2018;**97**(52). e13714. [PubMed ID: 30593143]. [PubMed Central ID: PMC6314751]. <https://doi.org/10.1097/MD.00000000000013714>.
- Kurdoglu Z, Ozkol H, Kurdoglu M, Kamaci M. Evaluation of the relationship between adenosine deaminase, myeloperoxidase, cholinesterase, preeclampsia severity, and neonatal outcomes. *Clin Exp Hypertens*. 2012;**34**(7):493–7. [PubMed ID: 22681531]. <https://doi.org/10.3109/10641963.2012.666605>.
- Chiarello DI, Abad C, Rojas D, Toledo F, Vazquez CM, Mate A, et al. Oxidative stress: Normal pregnancy versus preeclampsia. *Biochim Biophys Acta Mol Basis Dis*. 2020;**1866**(2):165354. [PubMed ID: 30590104]. <https://doi.org/10.1016/j.bbadis.2018.12.005>.
- Afrose D, Chen H, Ranashinghe A, Liu CC, Henessy A, Hansbro PM, et al. The diagnostic potential of oxidative stress biomarkers for preeclampsia: systematic review and meta-analysis. *Biol Sex Differ*. 2022;**13**(1):26. [PubMed ID: 35658944]. [PubMed Central ID: PMC9167545]. <https://doi.org/10.1186/s13293-022-00436-0>.
- Rahimi Z, Ahmadi R, Vaisi-Raygani A, Rahimi Z, Bahrehmand F, Parsian A. Butyrylcholinesterase (BChE) activity is associated with the risk of preeclampsia: influence on lipid and lipoprotein metabolism and oxidative stress. *J Matern Fetal Neonatal Med*. 2013;**26**(16):1590–4. [PubMed ID: 23650977]. <https://doi.org/10.3109/14767058.2013.795534>.
- Iddrisu M, Khan ZH. Anesthesia for cesarean delivery: general or regional anesthesia—a systematic review. *Ain Shams J Anesthesiol*. 2021;**13**(1). <https://doi.org/10.1186/s42077-020-00121-7>.
- Karam RS, Mohammad FK. The use of anesthetics for cesarean section delivery in women in Duhok, Kurdistan region, Iraq. *J Ideas Health*. 2022;**5**(4). <https://doi.org/10.47108/jidhealth.Vol5.Iss4.257>.
- Kluge WH, Kluge HH, Konig U, Venbrocks RA, Bauer HI, Lange M. Effect of bupivacaine application on cholinesterase activities, total protein- and albumin concentration in serum and cerebrospinal fluid. *Scand J Clin Lab Invest*. 2002;**62**(7):495–502. [PubMed ID: 12512739]. <https://doi.org/10.1080/003655102321004503>.
- Senoner T, Velik-Salchner C, Luckner G, Tauber H. Anesthesia-Induced Oxidative Stress: Are There Differences between Intravenous and Inhaled Anesthetics? *Oxid Med Cell Longev*. 2021;**2021**:8782387. [PubMed ID: 34873432]. [PubMed Central ID: PMC8643269]. <https://doi.org/10.1155/2021/8782387>.
- Kundovic SA, Rasic D, Popovic L, Peraica M, Crnjak K. Oxidative stress under general intravenous and inhalation anaesthesia. *Arh Hig Rada Toksikol*. 2020;**71**(3):169–77. [PubMed ID: 33074169]. [PubMed Central ID: PMC7968496]. <https://doi.org/10.2478/aiht-2020-71-3437>.
- Ilska M, Kolodziej-Zaleska A, Banas-Fiebrich E, Brandt-Salmeri A, Janowska-Tyc E, Lyszczarz A, et al. Health-Related Quality-of-Life among Pregnant Women after First, Second, and Multiple Cesarean Sections in the Perinatal Period: A Short-Term Longitudinal Study. *Int J Environ Res Public Health*. 2022;**19**(24). [PubMed ID: 36554628]. [PubMed Central ID: PMC9779739]. <https://doi.org/10.3390/ijerph192416747>.
- Ghaffari S, Dehghanpisheh L, Tavakkoli F, Mahmoudi H. The Effect of Spinal versus General Anesthesia on Quality of Life in Women Undergoing Cesarean Delivery on Maternal Request. *Cureus*. 2018;**10**(12). e3715. [PubMed ID: 30788204]. [PubMed Central ID: PMC6373886]. <https://doi.org/10.7759/cureus.3715>.
- Garmavy HMS. *General anesthesia and oxidative stress status: interaction with cholinesterase inhibitors [dissertation]*. Duhok, Iraq: University of Duhok; 2009.
- Gawlik-Kotelnicka O, Mielicki W, Rabe-Jablonska J, Strzelecki D. Impact of lithium alone or in combination with haloperidol on selected oxidative stress parameters in human plasma in vitro. *Redox Rep*. 2016;**21**(1):45–9. [PubMed ID: 26193071]. [PubMed Central ID: PMC6837648]. <https://doi.org/10.1179/1351000215Y.0000000030>.
- Gawlik-Kotelnicka O, Mielicki W, Rabe-Jablonska J, Lazarek J, Strzelecki D. Impact of lithium alone or in combination with haloperidol on oxidative stress parameters and cell viability in SH-SY5Y cell culture. *Acta Neuropsychiatr*. 2016;**28**(1):38–44. [PubMed ID: 26286703]. <https://doi.org/10.1017/neu.2015.47>.
- Mohammed AA, Mohammad FK. Recognition and Assessment of Antidotal Effects of Diphenhydramine against Acute Carbaryl Insecticide Poisoning in a Chick Model. *Toxicol Int*. 2022:339–52. <https://doi.org/10.18311/ti/2022/v29i3/29732>.
- Zhang Z, Yan B, Li Y, Yang S, Li J. Propofol inhibits oxidative stress injury through the glycogen synthase kinase 3 beta/nuclear factor erythroid 2-related factor 2/heme oxygenase-1 signaling pathway. *Bioengineered*. 2022;**13**(1):1612–25. [PubMed ID: 35030972]. [PubMed Central ID: PMC8805835]. <https://doi.org/10.1080/21655979.2021.2021062>.
- Mohammad FK. Clarifying an electrometric method for determining blood cholinesterase activity: a scientific letter. *Asia Pac J Med Toxicol*. 2022;**11**(1):30–2.
- Garmavy HMS, Mohammed AA, Rashid HM, Mohammad FK. A meta-analysis of normal human blood cholinesterase activities determined by a modified electrometric method. *J Med Life*. 2023;**16**(1):22–34. [PubMed ID: 36873131]. [PubMed Central ID: PMC9979180]. <https://doi.org/10.25122/jml-2022-0215>.
- Odisho SK, Mohammad FK. Blood cholinesterase activities and oxidative stress status among farmworkers using pesticides in Duhok, KRG, Iraq. *J Ideas Health*. 2022;**5**(4):786–93. <https://doi.org/10.47108/jidhealth.Vol5.Iss4.264>.
- Mohammad FK, Al-Baggou BK, Naser AS, Fadel MA. In vitro inhibition of plasma and brain cholinesterases of growing chicks by chlorpyrifos and dichlorvos. *J Appl Anim Res*. 2014;**42**(4):423–8. <https://doi.org/10.1080/09712119.2013.875912>.

31. Mohammed AA, Mohammad FK. Monitoring Blood Cholinesterase Activity of Farmworkers: In Vitro Inhibition by Diphenhydramine and Carbaryl. *Malays Appl Biol.* 2022;**51**(2):23–32. <https://doi.org/10.55230/mabjournal.v51i2.2204>.
32. Naser AS, Mohammad FK. [The antioxidant activity of propofol in chicks]. *Iraqi J Vet Sci.* 2015;**29**(1):29–34. Arabic. <https://doi.org/10.33899/ijvs.2015.116853>.
33. Reynolds F, Hargrove RI, Wyman JB. Maternal and foetal plasma concentrations of bupivacaine after epidural block. *Br J Anaesth.* 1973;**45**(10):1049–53. [PubMed ID: 4772641]. <https://doi.org/10.1093/bja/45.10.1049>.
34. Nakamura J, Purvis ER, Swenberg JA. Micromolar concentrations of hydrogen peroxide induce oxidative DNA lesions more efficiently than millimolar concentrations in mammalian cells. *Nucleic Acids Res.* 2003;**31**(6):1790–5. [PubMed ID: 12626721]. [PubMed Central ID: PMC152865]. <https://doi.org/10.1093/nar/gkg263>.
35. Galenko-Yaroshevskii AP, Derlugov LP, Ponomarev VV, Dukhanin AS. Pharmacokinetics and pharmacodynamics of a new local anesthetic agent. *Bull Exp Biol Med.* 2003;**136**(2):170–3. [PubMed ID: 14631501]. <https://doi.org/10.1023/a:1026323124831>.
36. Evans RT, Wroe JM. Plasma cholinesterase changes during pregnancy. Their interpretation as a cause of suxamethonium-induced apnoea. *Anaesthesia.* 1980;**35**(7):651–4. [PubMed ID: 7435932]. <https://doi.org/10.1111/j.1365-2044.1980.tb03878.x>.
37. Davies P, Landy M. Suxamethonium and mivacurium sensitivity from pregnancy-induced plasma cholinesterase deficiency. *Anaesthesia.* 1998;**53**(11):1109–11. [PubMed ID: 10023281]. <https://doi.org/10.1046/j.1365-2044.1998.00581.x>.
38. Kharb S, Nanda S. Patterns of Biomarkers in Cord Blood During Pregnancy and Preeclampsia. *Curr Hypertens Rev.* 2017;**13**(1):57–64. [PubMed ID: 28128050]. <https://doi.org/10.2174/1573402113666170126101914>.
39. Kumar SN, Vaibhav K, Bastia B, Singh V, Ahluwalia M, Agrawal U, et al. Occupational exposure to pesticides in female tea garden workers and adverse birth outcomes. *J Biochem Mol Toxicol.* 2021;**35**(3). e22677. [PubMed ID: 33350548]. <https://doi.org/10.1002/jbt.22677>.
40. Brasil LJ, San-Miguel B, Kretzmann NA, Amaral JL, Zettler CG, Marroni N, et al. Halothane induces oxidative stress and NF-kappaB activation in rat liver: protective effect of propofol. *Toxicology.* 2006;**227**(1-2):53–61. [PubMed ID: 16965849]. <https://doi.org/10.1016/j.tox.2006.07.013>.
41. Li Volti G, Basile F, Murabito P, Galvano F, Di Giacomo C, Gazzolo D, et al. Antioxidant properties of anesthetics: the biochemist, the surgeon and the anesthetist. *Clin Ter.* 2008;**159**(6):463–9. [PubMed ID: 19169610].
42. Braz MG, Braz LG, Freire CMM, Lucio LMC, Braz JRC, Tang G, et al. Isoflurane and Propofol Contribute to Increasing the Antioxidant Status of Patients During Minor Elective Surgery: A Randomized Clinical Study. *Medicine (Baltimore).* 2015;**94**(31). e1266. [PubMed ID: 26252290]. [PubMed Central ID: PMC4616612]. <https://doi.org/10.1097/MD.0000000000001266>.