



# Molecular Analysis of Pathogenic Genes (lasB and exoA) in *Pseudomonas aeruginosa* Strains Isolated from Animal and Human Samples and Determination of Their Resistance Pattern

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## Abstract

*Pseudomonas aeruginosa* has a wide range of virulence factors. These factors have the potential to increase bacterial pathogenicity and serious infection. The purpose of this study was to evaluate the virulence profiles and antibiotic susceptibility of isolates of *P. aeruginosa* originated from animal and human samples. The samples were cultured on selective media before being extracted for DNA and subjected to a PCR technique to detect virulence genes. There was a significant difference in the isolation of *P. aeruginosa* isolated from human and animal sources. Where, in humans, the percentage of *P. aeruginosa* was 52 (68.42%) while in animals the percentage of *P. aeruginosa* was 24 (31.57%). In humans, the percentage of *P. aeruginosa* in blood was 26.92% (14 isolates), in urine it was 25% (13 isolates), in wound it was 40.38% (21 isolates), and in sputum it was 7.69% (4 isolates). We used a PCR technique that produced highly specific and accurate results for detecting virulence factor genes in *P. aeruginosa* isolates that cause disease in humans and animals. The percentage of *exoA* genes was (83.33%) and (81.66%) in the animal and human, and that of *lasB* was (58.33%) and (92.30%) in animal and human samples respectively. Furthermore, both the *exoA* and *lasB* genes are found in 26.31% of animal strains and 17.10% of human strains. The disc diffusion method was used to determine antimicrobial susceptibility. In both animal and human isolates, *P. aeruginosa* showed the highest resistance to amikacin and the lowest resistance to ciprofloxacin. These findings could aid in the understanding of pathogenicity processes, treatment direction, and the development of strategies to control the spread of epidemic *P. aeruginosa* strains.

**Keywords:** *Pseudomonas aeruginosa*, Infection, Antibiotic, Pathogenicity, Ciprofloxacin

## 1. Background

*Pseudomonas aeruginosa* is a nosocomial, aerobic, gram-negative bacterium that causes pneumonia, urinary tract infections, surgical site infections, ulcerative keratitis, and soft tissue infections, among other things (1). The species are often isolated from soil and water, or they colonize a variety of anatomical sites, including plants, insects, animals, and humans (2). Virulent factors are molecules produced by pathogenic *P. aeruginosa* that allow it to play a significant role in pathogenesis infection. This bacterium is normally not pathogenic, but it can cause opportunistic infections in patient with weak immune systems, such as ICU patients (3). *Pseudomonas aeruginosa* is caused by the production of several cellular and extracellular virulence factors. *Pseudomonas aeruginosa* has a higher proportion of predicted regulatory genes due to its large genome of 6.3 million base pairs which suggests that *P. aeruginosa* can adapt to a variety

of environmental stresses (4). Some of the genes that normally encode and participate in virulence factors that may contribute to its pathogenicity are *toxA*, *exoS*, *exoY*, *exoU*, *oprL*, *oprI*, *lasA*, *lasB*, *oprD*, *plcH*, *plcN*, and *nan1*. The major outer membrane lipoproteins responsible for identifying *P. aeruginosa* infections are *oprL*, *oprI*, and *oprD*. Similarly, the *toxA* gene, which encodes exotoxin A and inhibits protein biosynthesis, is the most toxic virulence gene. Elastase B (*lasB*) is an important metalloenzyme that plays a key role in pathogenesis during host infection by promoting tissue adhesion, colonization, and invasion, resulting in chronic pulmonary inflammation (5). It causes elastin lysis, protein compound destruction, complement inactivation, and clotting factor degradation (6). Exotoxin A (*exoA*) is responsible for tissue necrosis (7). *ExoA* is the major member of the type II secretion system (T2SS) which inhibits protein synthesis through ADP-ribosylation of eukaryotic elongation factor 2 (8).

Rapid detection of such virulence genes which cause hospital infections is important for subsequent treatment decisions of patients. Previously, common methods of bacterial typing such as antibiotic resistance patterns, phage typing, and serotyping were used to study the genetic linkages of bacteria, particularly in nosocomial infections. These are now being replaced by molecular methods such as ribotyping, pulse-field gel electrophoresis (PFGE), and PCR-based methods. Molecular methods are superior over phenotypic methods for identifying *P. aeruginosa* by developing a multiplex Polymerase chain reaction (PCR) (9). PCR is used to determine the contribution of several virulence genes in resistant mechanisms, which aids in the selection of appropriate antibiotics for the treatment of *P. aeruginosa*-caused infectious disease. PCR can be rapidly identify the microbial species by the amplification of sequences unique to a specific organism. The enterobacterial repetitive intergenic consensus (ERIC)-PCR technique uses genetic marker i.e., the position and number of different ERIC sequences in bacteria, to detect the bacterial diversity (10).

Antimicrobial resistance (AMR) is a public health issue that affects both human and veterinary medicine. A diverse group of *P. aeruginosa* strains have the inherent ability to form biofilms. And these biofilms boosts resistance to various classes of antibiotics or antibacterial agents, making infection control difficult (11). AMR in *P. aeruginosa* is usually the result of a combination of different imported (mobile genetic elements) and chromosomally encoded resistance mechanisms (12). *Pseudomonas aeruginosa*'s major antibiotic-resistance mechanisms can be divided into three categories: intrinsic, acquired, and adaptive resistance. The unreasonable administration of antibiotics in both human medicine and animal production for growth-promoting purposes, metaphylaxis, and prophylaxis have nourished the proliferation, spread of antibiotic-resistant bacteria, and resistance genes. Such an uncontrolled and random use of antibiotics increased public health and environmental risks (13). And also it is difficult to cure the patients whose infections are resistant to conventional antibiotics. Hence there is an urgent need to develop the advance antibiotics and effective alternative therapeutic strategies for the treatment of *P. aeruginosa* infections (14).

## 2. Objectives

The objective of this study was to look into the virulence factors and antibiotic susceptibility isolated from animal and human samples in order to determine the pathogenicity of *P. aeruginosa* using the PCR technique.

## 3. Methods

### 3.1. Sample Collection and Bacterial Isolation

The study was performed on a total of 120 specimens of *P. aeruginosa* strains. Among of, 60 animal specimens were collected from veterinary microbiology laboratories and the remaining 60 human specimens were collected from hospital laboratories. After that, the samples were then cultured on MacConkey media, eosin methylene blue EMB agar, and incubated overnight at 33°C. Following incubation, the bacterial isolates were subjected to biochemical tests to determine bacterial growth and colony formation. Finally, 76 isolates were identified by gram staining and biochemical tests of oxidase, catalase, motion, indole sulfide (SIM), methyl (TSI) triple sugar agar-agar, (VP) Vegs Prosquier, (MR) red, (OD) ornithine decarboxylase, (SiMc) Simon citrate agar, (AD) arginine dehydrogenase, (LD) lysine decarboxylation, oxidation-fermentation test (Merck, Germany). During the period of study, 76 isolates of *Pseudomonas* species were identified, out of which 24 isolates of *P. aeruginosa* were recovered from animal samples and 52 strains were recovered from human samples. These 52 strains recovered from human clinical infection samples includes isolates of wound, urine, blood, and sputum. Detected *P. aeruginosa* bacterial isolates were used for subsequent tests.

### 3.2. Antibiotic Susceptibility Testing

Antibiotic susceptibility testing was determined by disk diffusion method in Müller-Hinton agar medium examined according to Clinical and Laboratory Standard Institute (CLSI) guidelines (15). The following antibiotics disks (Kirby-Bauer) were used for antibiotic susceptibility testing: (1) ceftazidim, (2) gentamycin, (3) piperacillin, (4) amikacin, (5) ciprofloxacin, (6) imipenem, (7) cefotaxim, (8) erythromycin, (9) ofloxacin, (10) tobramycin, (11) clindamycin, and (12) tetracyclin. The isolates were divided into three groups: (1) sensitive (S), (2) intermediate (I), and (3) resistant (R).

### 3.3. DNA Extraction and Molecular Detection of Virulence-Related Genes Using PCR Technique

Bacterial genomic DNAs were extracted from *P. aeruginosa* isolates using a commercial DNA extraction kit (CinnaGen Co., Tehran, Iran). Table 1 shows two sets of primers used to amplify the genes [exoA (396bp) and lasB (300bp)] using a specific polymerase chain reaction (PCR). PCR amplification was carried out using a thermal cycler (Eppendorf, Germany) with specific forward and reverse primers with specific length coding for exoA (exoenzyme A) and

lasB (elastase) virulence genes. Each PCR mixture was prepared in a final volume of 25  $\mu$ L containing 2.5  $\mu$ L of PCR-10x buffer, 1  $\mu$ L MgCl<sub>2</sub> (2.5 mM), 1  $\mu$ L of each primer (10 pmol), 1.5  $\mu$ L dNTPS (2.5 mM), 0.3  $\mu$ L of Tq polymerase enzyme (5U), and 2  $\mu$ L of DNA template. PCR amplifications were carried out on a thermocycler under the following conditions: 95°C for 15 minutes; followed by 35 cycles at 95°C for 1 minute, 60°C for 45 seconds, 72°C for 1 minute and a final extension step at 72°C for 7 minutes. Multiple negative (water) amplification was used as a negative control (16).

## 4. Results

### 4.1. Bacterial Isolates and Distribution of Virulence-Related Genes

A total of 76 isolates were screened for the presence of different virulence genes of *P. aeruginosa* using conventional methods such as growth characteristics, colony morphology, and biochemical tests. The current study included 24 (31.57%) *P. aeruginosa* isolates from animal samples and 52 (68.42%) isolates from human samples. Table 2 shows the different percentages of virulence genes in *P. aeruginosa* strains of animal and human samples.

Figure 1 shows the percentage of *P. aeruginosa* strains isolated from a variety of clinical sites of human. Where, in human, the percentage of *P. aeruginosa* in blood was 26.92% (14 isolates), in the urine was 25% (13 isolates), in wound 40.38% (21 isolates), in sputum was 7.69% (4 isolates). Phenotypic analysis shows that highest percentage of *P. aeruginosa* strains were isolated from blood isolates.

The genes of *P. aeruginosa* are considered as important virulence factor attributed to the bacterium pathogenicity which are detected by PCR. The PCR results of virulence factor occurrence showed that *exoA* and *lasB* genes were present in both the animal samples and the human samples. Figure 2 shows the percentage of virulence genes in *P. aeruginosa* strains present in animal and human samples.

### 4.2 Antibiotic Resistance Patterns

The antibiotic susceptibility testing results of *P. aeruginosa* strains revealed varying levels of resistance, as shown in Figure 3. According to the findings of this study, Ciprofloxacin and Tubramycin were the least resistant antibiotics against both animal and human isolates, with resistance rates of 75 and 75.53%, respectively. In contrast, all the *P. aeruginosa* isolates were most resistive against Amikacin (98.33%) and Tetracycline (97.50%).

## 5. Discussion

In this study, the prevalence of *P. aeruginosa* isolates was 76, with the distribution in animal samples 24 (31.57%) was lower than in human samples 52 (68.42%). Possible explanations include different types of studied populations, different geographical locations, and different types of hospitals and veterinary laboratories. *Pseudomonas aeruginosa* is an opportunistic human pathogen competent for a wide array of infections including respiratory tract, blood, urinary tract, and skin infections. This competence for infections has made the organism to be recognized as a threat to public health (18). Phenotypic analysis shows that of a number of *P. aeruginosa* strains obtained from a variety of clinical sources, including wounds, urine, sputum, and blood. It is observe from the present study that wound patients are more liable to get infections in comparison with other patients. This was attributed to the organisms' virulence and invasive capability, the physiological state of the tissue in the wound, and the host's immunological integrity (19), lack of general hygienic measures, larger production of low quality antiseptic and medicinal solutions for treatment, difficulties in properly defining responsibilities among hospital staff (20). Wound infections are difficult to heal, and this bacterium has caused serious economic problems. *Pseudomonas aeruginosa* is a major cause of wound morbidity and mortality worldwide (21).

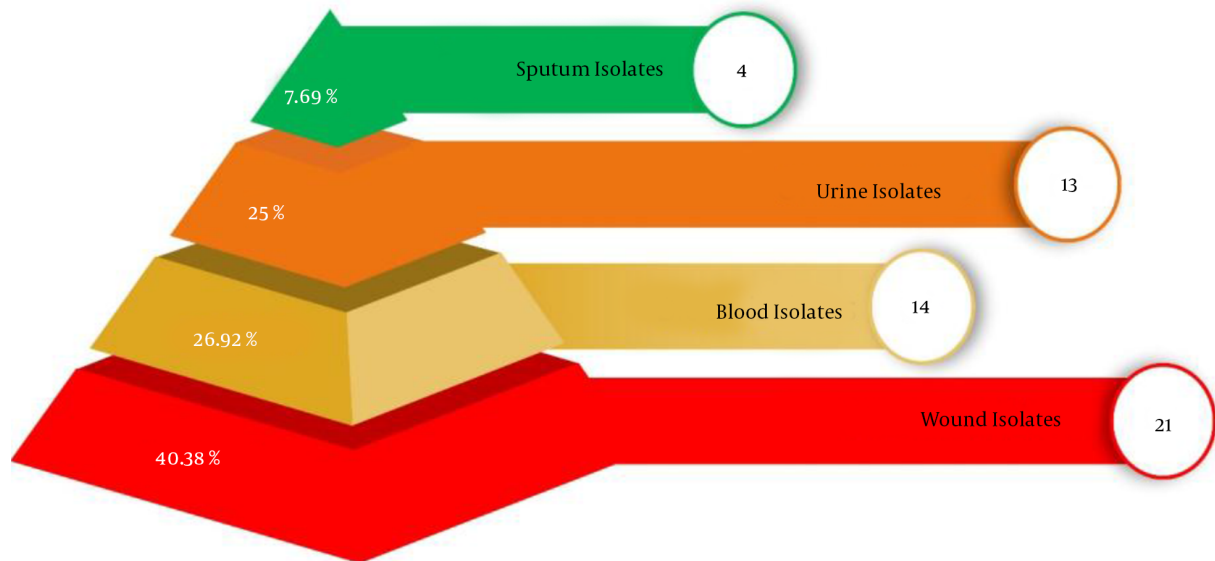
*Pseudomonas aeruginosa* is a pathogen having different virulence factors such as exotoxin A, alginate, exoenzyme S, elastase, and phospholipase. These virulence factors are regulated by particular signaling systems and can cause nosocomial problems to the world health system (22). Pathogenicity of *P. aeruginosa* is multifactorial. *ExoA* is responsible for the toxigenic trait of *P. aeruginosa* and plays a distinct role in the hindrance of wound contraction and healing. *lasB*, one of the most important proteases of *P. aeruginosa*, is responsible for invasiveness. *lasB* can facilitate bacterial attachment and immune system disruption (23). *lasB* causes elastin lysis, protein compound destruction, complement inactivation, and clotting factor degradation (6). Because of the importance of *P. aeruginosa*, genes encoding virulence factors including *exoA*, *lasB* genes as well as antibiotic resistance patterns were investigated. Herein, all examined isolates have *exoA* and *lasB* genes. The frequency of *exoA* genes in isolated strains *P. aeruginosa* strains was 83.33% (animal sample) and 81.66% (human sample). However, the frequency of *lasB* genes was found to be 58.33 and 92.30% in animal and human samples respectively. Furthermore, 26.31% of animal strains and 17.10% of human strains contain both the *exoA* and *lasB* genes (Figure 2). Divergences in the distribution of virulence factor genes across populations may be due to

**Table 1.** Nucleotide Sequences of Primers Used for Amplification of *Pseudomonas aeruginosa* Virulence

Primer	Primer Sequence	Product Size (bp)	Ref.
exoA	F: GACAACGCCCTCAGCATCACCAGC	396	(16)
	R: CGCTGGCCATTGCTCCAGCGCT		
lasB	F: GGAATGAACGAAGCGTTCTC	300	(17)
	R: GGTCCAGTAGTAGCGGTGG I		

**Table 2.** Percentage of Virulence Genes in *Pseudomonas aeruginosa* Strains

Virulence Genes	Isolated Strains of <i>P. aeruginosa</i> Total Percentage (%)	
	(Animal Sample Isolates) 24	(Human Sample Isolates) 52
exoA	83.33	81.66
lasB	58.33	92.30
exoA + lasB	26.31	17.10

**Figure 1.** Percentage *Pseudomonas aeruginosa* strains isolated from a variety of clinical sites of human.

that some *P. aeruginosa* strains are better adapted to the specific conditions found in infectious sites. The prevalence of *P. aeruginosa* and its virulence genes is affected by a variety of factors, including the nature of the environment, the degree of contamination and type, and the individual's immune status (24). There are various reports on the frequency of *exoA* and *lasB* genes in different studies. Amirmozafar et al. detected the *exoA* strains in 81% of clinical isolates of *P. aeruginosa* (25). Neamah discovered 45.7% of *exoA* gene in otitis media and 75% in cow milk isolates of *P. aeruginosa* (26). Dekhil recorded a 54.5% prevalence of *exoA* gene in *P. aeruginosa* isolates (27) which were less than our results. According to the findings of our

and other studies, *exoA* is a more common virulence factor. Sabharwal et al. reported the prevalence of *lasB* in 75% clinical isolates of *P. aeruginosa* (28). Comoé detected a high prevalence of elastase (89.2%) encoding the *lasB* genes of *P. aeruginosa*. This suggests that this protease, which cleaves elastin and collagen, may be important in the pathogenesis of *P. aeruginosa* isolated from animal products (2). Faraji et al. discovered the higher prevalence of *lasB* in *P. aeruginosa* isolated from cystic fibrosis (CF) 62 (95.4%) and burn wounds 47 (82%) than any other genes detected (22).

As *P. aeruginosa* is an opportunistic pathogen involved in clinical infections, it is critical to understand the full extent of the variation in antimicrobial resistance gene

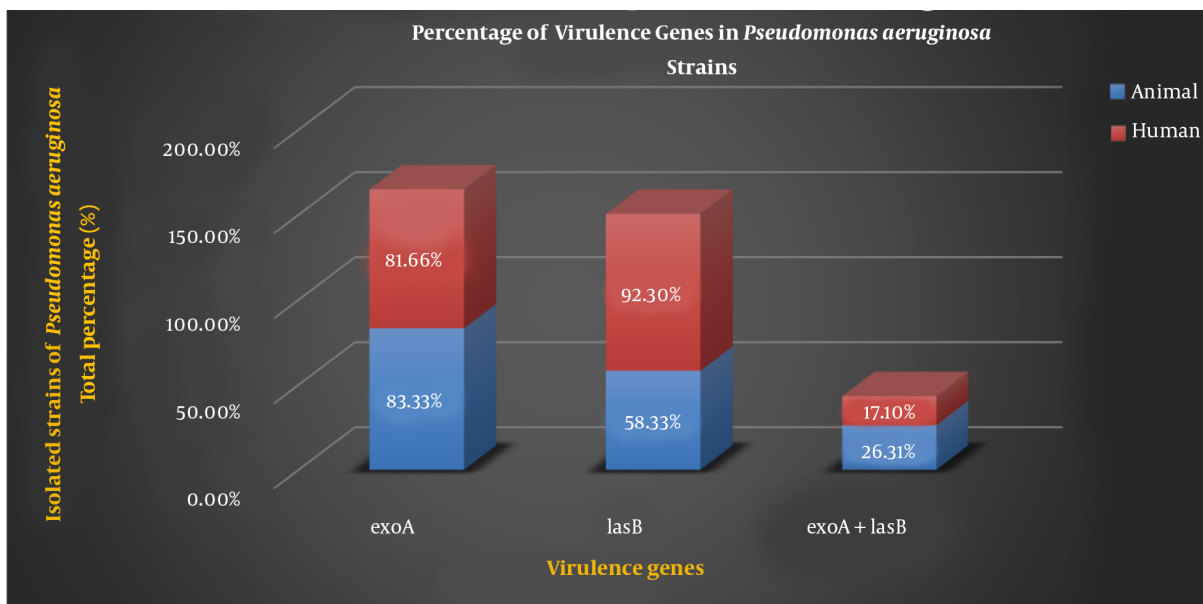


Figure 2. Percentage of virulence genes in *Pseudomonas aeruginosa* strains present in animal and human samples.

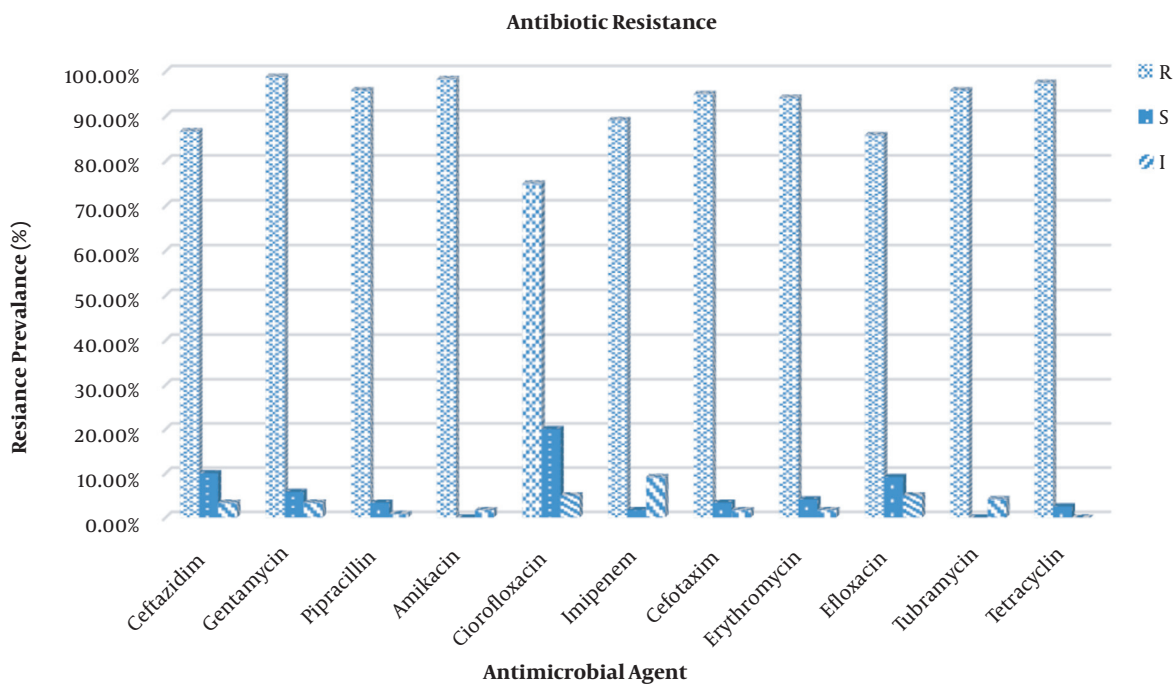


Figure 3. The antibiotic resistance (%) of isolated *Pseudomonas aeruginosa* strains.

content (29). In the current study, 76 isolates of *P. aeruginosa* were tested using disc diffusion against 11 different antibiotics in order to gain a better understanding of the

antibiotic resistance rates (Figure 1). *Pseudomonas aeruginosa* isolates showed high-level resistance to many antimicrobial agents. In the present study, the in vitro antibi-

otic resistivity test of isolated *P. aeruginosa* strains revealed the highest resistance in human and animal samples was to amikacin and there was no significant difference between them. On the other side, the lowest resistance was to ciprofloxacin and there was a significant difference between human and animal strains. Antibiotic resistance in *P. aeruginosa* could be caused by indiscriminate antibiotic use, the production of various enzymes such as carbapenamase and AmpC-lactamases, quorum sensing modification of different target sides, and so on (30, 31). A cause for concern is that the high resistance displayed to the ciprofloxacin which is one of the best options available for the treating the infections caused by *P. aeruginosa*, particularly in the treatment of urinary tract infections. Such drawbacks will result in limited treatment options (32). In 2016, Sharma et al. (33) discovered that amikacin (18.2%) and ciprofloxacin (31.7%) were the most effective drugs for routine use among the *P. aeruginosa* strains studied (34). Similarly, Hosu et al. reported 16.7 and 22.2% resistance for amikacin and ciprofloxacin respectively (13). In accordance with our findings, Mobaraki in Iran reported increased resistance for both amikacin and ciprofloxacin from the year 2007 to 2014 (35). Furthermore, Poonsuk et al. (36) showed high-level resistance of *P. aeruginosa* isolates to amikacin (92.1%), and ciprofloxacin (95%) (37). Resistance to such antibiotics increased in comparison to previous studies, which could be attributed to variations in antibiotic usage (3).

### 5.1. Conclusions

This discovery revealed differences in the virulence profiles and antibiotic resistance of *P. aeruginosa* strains isolated from animal and human samples, demonstrating the adaptive ability of this bacterial species. According to the current study, wound patients are more likely to get infections than other patients. In conclusion, isolated *P. aeruginosa* isolates exhibit several number of *exoA* and *lasB* virulence genes which may play an important role in infections caused by *P. aeruginosa*. It was observed that high percentages of resistance to amikacin and low percentages of resistance to ciprofloxacin among all the examined antibiotics which is cause for concern. As a result, antibiotic consumption and management are critical for reducing resistance and effectively treating infections caused by the *P. aeruginosa* bacterium.

### Footnotes

**Authors' Contribution:** Ciamak Ghazaei is the only author of the article and the study was solely carried out by the author.

**Conflict of Interests:** There is no conflict of interest.

**Ethical Approval:** IR.ARUMS.REC.1399.464

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