




A Review on Synthesis, Characterization and Applications of Polycaprolactone as a Novel Drug Delivery System and Tissue Engineering

Amir Hosseini¹, Mohammad Shahverdi ², Zahra Sourani ³, Roya Habibi⁴, Zahra Ahangari⁴, Sadegh Shirian ^{3,5}, Fariba Zafari ^{1,*}

¹ Cellular and Molecular Research Center, Research Institute for Non-communicable Disease Prevention, Qazvin University of Medical Sciences, Qazvin, Iran

² Biochemistry Research Center, Basic Health Sciences Institute, Shahrekord University of Medical Sciences, Shahrekord, Iran

³ Department of Pathology, School of Veterinary Medicine, Shahrekord University, Shahrekord, Iran

⁴ Student Research Committee, School of Medicine, Qazvin University of Medical Sciences, Iran

⁵ Shiraz Molecular Pathology Research Center, Dr. Daneshbod Path Lab, Shiraz, Iran

*Corresponding author: Cellular and Molecular Research Center, Research Institute for Non-communicable Disease Prevention, Qazvin University of Medical Sciences, Qazvin, Iran. Email: fariba.zafari@yahoo.com

Received 2024 July 7; Accepted 2024 August 12.

Abstract

Biodegradable plastics play a critical role in reducing the build-up of solid wastes. Increasing efforts have been made to develop non-degradable and chemically degradable products to replace current plastics. Polycaprolactone (PCL), an aliphatic polyester and biocompatible thermoplastic, is the "green" material and one of the most promising and widely used materials with the most significant development potential. This biodegradable plastic is utilized in various applications including medication release monitoring, tissue processing, bone scaffolding, bag shielding, and composting. This study provides a summary of the latest PCL-based biomaterial architectures and their uses, including drug delivery, various organ tissue engineering applications, biodegradation, and PCL safety. It is also concluded that combining PCL with other materials to create nanocomposites enhances the properties of the resultant products and broadens the range of applications.

Keywords: Biodegradable, Morphology, Medical Application, Polycaprolactone

1. Context

Polycaprolactone (PCL) is an aliphatic polyester polymer with an alkaline hexanoate repeater assembly. Polycaprolactone has been closely studied with a wide variety of other polymers due to its unusual mechanical properties. Polycaprolactone's physical, thermal, and mechanical properties depend primarily on its molecular weight, degree of crystallinity, and degradation by hydrolysis of its ester bond under physiological conditions. Polycaprolactone has attracted increasing attention owing to its low melting temperature, excellent mixing compatibility, hydrophobicity, high solubility at room temperature, and simplicity of use (1). Polycaprolactone, alone and in combination with several polymers and co-polymers, has been extensively studied to prepare controlled drug delivery systems via their native biocompatibility and biodegradation. Its permeability to a wide variety of drugs has also ensured uniform delivery of drugs in the matrix, promising a lengthy release duration through a

phase of decay of up to several months. Polycaprolactone has been widely used to prepare long-term implants and scaffolds that imitate the natural extracellular matrix, thereby promoting 3D cell tissue culture (2). Polycaprolactone has already been licensed for use in a wide range of medical supplies and instruments. However, few FDA and European Community Registry Label (EC) approved scaffolds containing PCL have been marketed or widely used in clinical trials (3, 4). In recent years, PCL has been used in the manufacture of green materials/biomaterials for various tissue engineering applications. Polycaprolactone's high rheological and viscoelastic properties make it suitable for the production of aliphatic polyester equivalents in a wide variety of biodegradable devices. Additionally, the mechanical characteristics of PCL complement tissue engineering applications, such as wound dressing, contraception, and dentistry (5, 6), as well as non-medical uses, such as active packaging for food products (7, 8). Polycaprolactone has been utilized in combination with

other biopolymers, owing to the interesting biological characteristics of these materials. Customized kinetic and mechanical degradation are exhibited by these structures, which are readily fabricated and assembled, thereby facilitating the establishment of tissue-forming voids that can achieve precise and controlled delivery of therapeutic agents within the matrix (6, 9, 10). This example includes functional groups that change polymer chains to increase their hydrophilic, sticky, and biocompatible characteristics to induce a favorable cell response. Additionally, PCL formulations have been produced as micro-/nanomicellar structures after copolymerization, improving bioactive molecules and drug encapsulation (9, 11, 12). Owing to the increasing interest in the application of PCL as a biological and green material, we reviewed the available literature on PCL Drug Delivery and tissue engineering applications. We also provide insight into PCL safety, biodegradation, and recent developments in PCL-Based Biomaterials. In the first chapter, we focus on important synthetic techniques and show the chemical synthesis of key functional features, such as Biodegradation. In addition, the therapeutic alternatives of PCL have been discussed in a molten state or solution. Moreover, the essay concludes with a discussion of representative systems for biomedical and environmentally friendly chemical instruments (i.e., pore scaffolds, micro and nanocarriers, and implantable systems).

2. Synthesis

Polycaprolactone was developed using a caprolactone cyclic monomer ring-opener (Figure 1) and was investigated in the early 1930s (13). Recently, many catalysts have been tested for caprolactone ring-opening polymerization (14). Catalysts such as stannous octoates may be used to catalyze polymerization, and low-molecular-weight alcohols may also be used to regulate the molecular weight of the polymers. Various anionic, cationic, and directed mechanisms have been used to polymerize PCLs. The resulting molecular weight, molecular weight distribution, final group composition, and chemical structure are influenced by the phase (15, 16). Typically, the average molecular weight of PCL samples is between 3000 and 80000 g/mol and can be classified by molecular weight (17). Semi-crystalline polymers have a melting point between 59°C and 64°C and a transitional temperature of 60°C in glass (18). Polycaprolactone is soluble in chloroform, dichloromethane, tetrachloride carbon, benzene, toluene, cyclohexanone, and 2-nitropropane at room temperature. It is insoluble in acetone, 2-butanone,

ethyl acetate, dimethylformamide, acetonitrile as an alcoholic, oil ether, and diatomic ether (13)

Polycaprolactone is adaptable owing to its ability to be changed through copolymerization or by combining it with various polymers to exploit its electrical, chemical, and mechanical characteristics. Additionally, chemical characteristics such as crystallinity, solubility, and chain breakdown were altered, resulting in modified polymers with the expected drug transport properties (19). For tissue compositions, such as scaffolds, textiles, and films, a combination that alters the physical properties and biodegradation and strongly affects the mechanical properties is chosen. Numerous polymers have been evaluated for their ability to modify the thermal, rheological, and biophysical characteristics of PCL, according to their intended applications. Polycaprolactone has compatibility with amino polyols, hydroxyapatite, polyethylene glycol, polyethylene, polyurethane, oxazole, Polyvinyl Alcohol (PVA), and Polylactic Acid (PLA). These PCL enhancements were consistent with the biophysical characteristics anticipated for the majority of existing drug delivery formulations (17) (Figure 2).

3. Application

3.1. Drug-Delivery Systems

Polycaprolactone is used in drug delivery applications and is suitable for controlled drug release. It has superb consistency and can be excreted during resorption because of its high permeability to a range of medications. Polycaprolactone biodegradation is gradual, making it more suitable for long-term supplies of more than one year compared to other polymers. Polycaprolactone also has compatible mixtures with other polymers that can affect the degradation kinetics and rapid modifications to follow the necessary release profiles (20-22). Polycaprolactone has been modified for use in dentistry by introducing co-polymers, such as PLA or PGA, and changing the material structure with other materials, such as PEG and Ceramic (23) (Figure 3).

However, in recent drug formulations, colloidal vectors have been overlooked as effective solubilizing agents (9). Poor water solubility can limit the availability of medicinal compounds. The copolymer's micelle block can increase the number of hydrophobic molecules owing to its structural shape, which is distinguished by a sterically stable hydrophobic core. The release rate of PCL drugs depends on the formulation procedure, preparation phase, PCL quality of the medication, its size, and proportion of the drug in

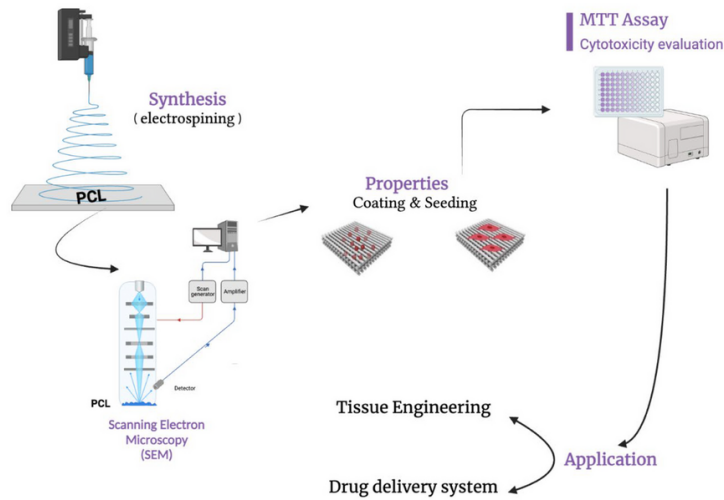


Figure 1. Synthesis, properties and applications of polycaprolactone (PCL) for tissue engineering and drug delivery system

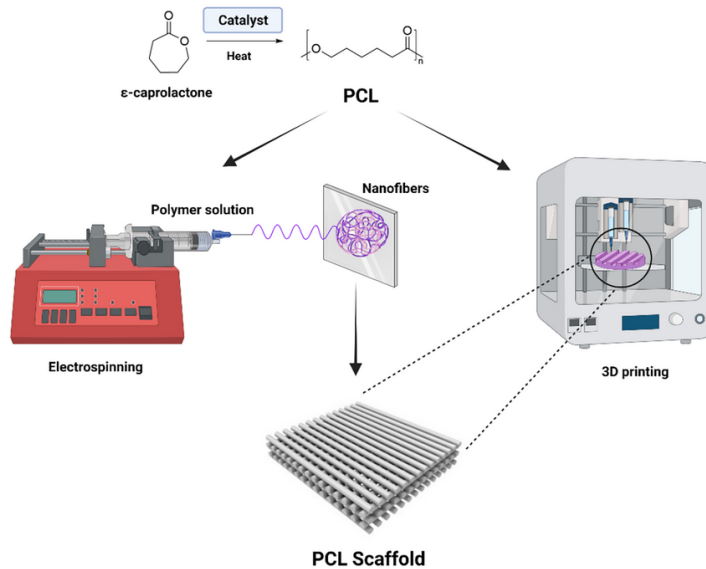


Figure 2. Fabrication of PCL scaffold: Three-dimensional (3D) fibrillates interconnects porous poly(ϵ -caprolactone) (PCL) scaffolds with desirable pore sizes and porosities prepared by blending PCL with water-soluble poly (ethylene oxide) (PEO) as a sacrificial material

the microcapsules. It is combined with additional polymers to increase stress, crack, tint, and monitor opioid release speed, resulting in improved PCL permeability. In recent decades, designing controlled

supply structures, particularly for peptides and proteins, has been a significant area of interest for PCL polymers (22). Lemmouchi et al. studied PCL-PLLA, PCL-

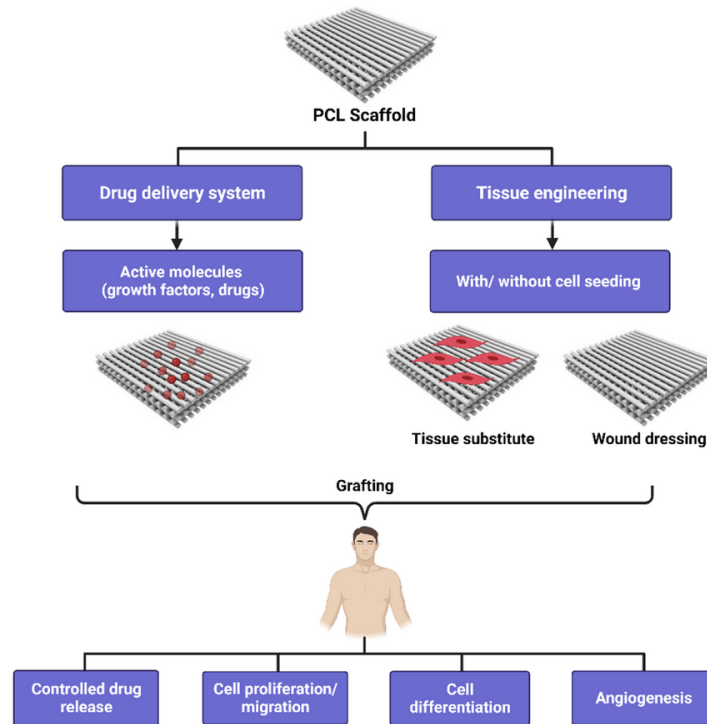


Figure 3. Application of PCL scaffolds: PCL, as a thermoplastic and linear aliphatic polyester, is one of the most widely used polymers in tissue engineering and drug delivery systems

DLLA, PCL-TMC, isometamidium chloride, and ethidium bromide both in vitro and in vivo (24).

The distribution of microspheres of degradable polymers has been studied extensively. Prescribing pharmaceutical drugs using these methods is beneficial because microspheres can be ingested or injected, tailored to the desired release profiles, and even organ-specific in certain situations (21). Microencapsulated medicine is a powerful delivery tool for medications with unique advantages including enhanced therapeutic safety and efficiency, improved biological activity, controlled medication release duration, and decreased administration frequency. In addition to biocompatibility, all of the most relevant requirements, such as medication particles, must biodegrade the product matrix for a long time in compliance with the drug release rate. Biodegradable polymers are also a subject of prescription test delivery system projects. After launching bioresorbable operating sutures in the market two decades ago, an extensive study on the distribution of biodegradable polymer materials was undertaken. Regarding their outstanding

biocompatibility, biodegradability, and "mechanical strength," thermoplastic aliphatic polymers such as PLA, PGA, and their co-polymers in particular, poly (lactide-co-glycolides) (PLGA), have attracted considerable attention among various classes of biodegradable polymers. These structures can be easily formulated to generate antibodies, peptides, proteins, and macromolecules. In particular, the FDA has approved the supply of drugs (25). The bioresorbable matrix of microparticles can be degraded into non-toxic, low-molecular-weight organisms that ultimately metabolize and eat the host. Unsurprisingly, there is considerable scientific interest in the use of biodegradable microparticles to control medication release. Polycaprolactone has many advantages over PLA and PGAs, including high dissolution permeability for small medicinal compounds and low capability to generate an acidic environment. Compared with polyesters, the degradation of PCL homopolymers is slower, making it more appropriate for longer-term supply systems lasting more than a year, and the distribution may be increased or decreased as required with correct mixing

(25). Numerous methods can be used with the PCL microspheres, which Freiberg and Zhu studied (26). In a solvent, the colloidal monomers of opposite solutions are polymerized (27). Organic monomers, such as oil in water emulsions (O/W) or water in oil emulsions (W/O), contribute to spherically distributed droplets of organic material (27). Dispersed monomers can be polymerized through various methods, including emulsions, suspensions, and dispersions. Emulsions are often used to manufacture homogeneous nanometers (10 - 100 nm). Additionally, the resultant polymer may be excessively diffractive at the nanoscale for visible light (28). The dispersion polymers have an impact of 0.5 - 10 microspheres. The reagents (monomer, initiator, and stabilizer) were dissolved in the organic medium, and polymerization occurred in the monomer droplets due to the solubility of the initiator in the monomer. Polymer pearls are insoluble in inorganic solutions and require hurrying and stabilization (29). The polymerization of supercritical CO₂ dispersions has been recently studied, which may be advantageous for medical applications due to the absence of hazardous solvents (30, 31). Suspension polymerization often results in the formation of 50 - 500 microsphere meters. Suspension polymerization distributes the monomer into a water stabilizer, whereas polymerization occurs during monomer processing. The size, quantity, and velocity of the dispersed monomer droplets define their dimensions and particle quantities (27, 32). Microspheres can be rapidly produced by evaporating organic solvents from dispersed polymers and biomolecule droplets (33). Often, biomolecules are first dissolved in water and then dispersed in a bio-solvent composed of biodegradable polymers and the original W/O emulsion (usually dichloromethane, DCM). The final O/W emulsion is produced by evaporating DCM and hardening the polymers, thereby trapping the contained medicinal product (34). Successful connectivity is a major impediment in the integration of medicines into microspheres. Numerous organizations lack the pit necessary to guarantee that microspheres will continue to develop, which is prohibitively expensive. Additionally, many studies lack a clear definition of the methodology for measuring opioid prevention effectiveness.

Polycaprolactone nanosphere 5.2 Colloidal pharmaceutical delivery structures with diameters of 10 - 1000 nm are nanospheres that function as compartments for narcotics or other active molecules. Nanospheres may include encapsulated, disseminated, or swallowed medication particles. Additionally, nanoparticles or nanocapsules can be classified

according to whether the medicine is enclosed in a shell or polymeric matrix. The adjustment of the processing settings for nanoscale outlets was similar to that for microparticulations. The spread and dispersion media have a modest ratio, but the riveting speed is significantly higher (35). The nanosphere may be utilized to specifically target the reticuloendothelial system of the liver and phagocytic cells. Unlike many other colloidal structures that conceal needles and capillaries, the dimensions of nanospheres enable intravenous injections. Nanoparticulate-injectable carriers are well-suited for the administration of basic medicines and imaging drugs. On the other hand, the reticuloendothelial system cannot be utilized routinely since it is removed after the first few injections. To overcome this limitation, amphiphilic copolymers have been synthesized using physiologically monodispersed biodegradable nanospheres. These nanospheres demonstrated enhanced blood supply and medication concentrations in a mouse model (35). Colloidal particles and their interactions with proteins and enzymes in different bodily fluids as medication carriers are inextricably linked. The combination of lysozyme, a highly condensed and positively charged enzyme, and two drug carriers, PCL-coated nanocapsules with an oily core and PCL-shaped nanoparticles, was investigated. These findings indicated that the surface loading of lysozyme on colloidal drug carriers significantly affected the mixture (34). Gref et al. (2000) investigated the capacity of PEG-coated biodegradable nanoparticles to absorb plasma proteins and particulate matter through polymorphonuclear cells (PLA, PLGA, and PCL). This study investigated the impact of PEG corona thickness and density as well as the nature of the core (36). Freezing with many cryoprotective agents maintained the conditions required to stabilize the PLGA and PCL nanoparticles. Studies have shown that sucrose, glucose, trehalose, and gelatin additives retain nanoparticles' characteristics irrespective of the freezing process (37). Polycaprolactone has also been used as an implant for targeted drug delivery. Implants are known to exhibit high biocompatibility. A new intraocular implant using porous PCL was developed by Boia et al., which can be used to deliver dexamethasone without the need for eye drops or intravitreal injections (38).

Polycaprolactone-coated chitin-lignin fibrous gel platforms appear to be good candidates for wound dressing applications based on controlled drug release (39). Manipulating the PCL-methoxypolyethylene glycol (PCL-MPEG)-based micelles ratio is an effective approach for modulating protein adsorption, phagocytosis, and biodistribution, which may be a prerequisite for drug

delivery in clinical applications (40). It has recently been shown that loading anticancer drugs on degradable PCL/magnetic nanocomposite nanoparticles effectively enhances drug function and kills cancer cells (41). Therefore, PCL is a promising polymer for pharmaceutical and biomedical applications in nanomedicine for cancer (42). Microcapsules as a controlled drug delivery carrier of Nifedipine, which is a calcium channel blocker, are widely used. Therefore, they are used to treat angina pectoris and hypertension. The core material of the microcapsules is PCL (43). Polycaprolactone nanofibers are considered carriers of oxytetracycline hydrochloride for the treatment of periodontal diseases (44).

3.2. Tissue Engineering

Usually, polymer films are formed by solvent casting, in which the polymer is molten, deposited, and evaporated from a thin film in a solvent. In Tendon, dental, skin, and Vascular tissue engineering applications, when an active surface is required to restore the patch rather than the missing tissue, biodegradable polymer films have been used (45). As they are extremely hydrophobic, films made from pure PCL do not have the requisite properties. However, the surface can be modified by shaping composites, mixtures, and copolymers to obtain the required properties. Polycaprolactone films can be used for cell binding, proliferation, and differentiation with or without external modifications. To investigate PCL's suitability for cartilage tissue engineering, the solvent effect on the film structure was compared using acetone and chloroform dissolves (46). Prabhakar et al. compared these two groups and revealed that with few cracks, PCL chloroform-cast films had a smoother coat, better mechanical properties, and were more hydrophilic than those cast from acetone. Various film chondrogenic potentials were reported; chloroform-cast PCL films showed a higher type I/II collagen ratio with a higher chondrogenic ability and type I/II collagen ratio with a higher osteogenic capacity indication (46). Polycaprolactone cast acetone solvents were also developed by Romagnoli et al. They studied PCL film attachment and their in-vitro differentiation of mesenchymal stem cells derived from human adipose (hAD-MSCs) for use in Bone tissue engineering (47). In tissue culture, cell viability, proliferation, alkaline phosphatase activity, and calcium deposition were similar to the polystyrene plate stages, indicating PCL's affinity for hAD-MSCs. These results indicate that pure PCL films can serve as tissue engineering structures and drive stem cell differentiation without chemical

changes. Chemical alteration of PCL film surfaces is one way to strengthen cell binding and enhance mechanical properties. Chemically modified PCL films for lung tissue engineering (48, 49) were prepared by Kosmala et al. Polycaprolactone was modified by aminolysis to form amine groups that connected gelatin to glutaraldehyde. Gelatin attachment can improve the viability of NCI-H292 (lung carcinoma) cells seeded on PCL. Yu et al. modified PCL films to bind a graded endothelial cell-attractive peptide on the surface. They homogeneously studied endothelial cells in one direction and concluded that target cells for a specific medical application would be assisted by this type of directional cell growth (50, 51). Composite PCL-HAp films are considered safer for Bone-Tendon Recovery. Tong et al. prepared a heat-pressing composite such as PCL and nanohydroxyapatite (nHAp) (52). The addition of nHA increased the elastic modulus of the films in parallel with the number of added HA. The films demonstrated proper cell adhesion, but higher nHA content decreased ductility and separated the PCL films. Incorporating natural polymers, biological markers, or bioceramics (calcium phosphate particles) into PCL films modifies their physical, biological, and chemical characteristics and provides ample soil for cells and tissues. Polycaprolactone films can be altered by adding metallic compounds to obtain additional properties. For example, the Fe₃O₄ antibacterial and paramagnetic properties of skin tissue engineering (53). Pai et al. used a composite PCL film with iron oxide (Fe₃O₄) (53). The spin coating created a film and examined the variation in the film properties with the rotation speed of the coating process. Bactericidal effects are shown by Iron oxide particles. Because of their capacity to withstand stress cracks, PCL composites have also been employed to reconstruct bulk materials and inorganic material surfaces. Catauro et al. coated titanium zirconium-PCL solution substrates with differing dental concentrations of PCL (54). The obtained shapes were a blend of zirconium-PCL that was transformed into a film structure to protect the titanium base utilizing a dip coating. Cell viability experiments using human mesenchymal stem cells showed no significant differences and no cytotoxic impact of pure titanium. To build engineering systems for metals with antibacterial and paramagnet properties, the soft nature of PCL can be blended. Fragments of PCL on metal surfaces may also have crack resistance characteristics, which are essential for preserving material properties and improving cell-material interactions (54). Hashiwaki et al. developed mixed films using N, N-Dimethylformamide solutions for chitin butyrate-PCL

(50: 50) (55). The film properties differed according to the butyryl substitution stage of chitin. Chitin with various degrees of butyryl substitution (deacetylation degree of 5 percent) was used, and high substitution levels were found, which improved its miscibility with PCL. For thermoplastic scaffolding, the mechanical strength of films can be enhanced by adjusting the chitin butyrate-PCL ratios. This system incorporated the functionalities of these two materials. The surface functional groups of PCL control its affinity and compatibility with cells. To achieve these characteristics, PCL can be modified from monomers. Chen et al. prepared PCL-coated co-polymer films with distinct functional groups and investigated their effects on physical, chemical, and biological properties (56). For this reason, ϵ -Caprolactone co-polymers have been prepared with modified ϵ -Caprolactone monomers with many functions (amine, methyl, carboxyl, hydroxyl, and aldehyde). Various functionalities of cell adhesion, such as aldehyde and hydroxyl groups, are classified as distinct biological responses. Increased osteogenesis was detected with amine-integrated films, chondrogenesis with methyl-incorporated films, and adipogenesis with the new PCL films. This means that the monomer, caprolactone, or the polymer, PCL, can be added to different functional groups, or the products can be tailored to achieve the correct cell-material interaction. Polycaprolactone copolymers with biocompatible chemicals may have a greater impact than mixtures and composites. Fuse et al. (2015) developed a film for cartilage tissue engineering using a PCL-PEG-PCL triblock copolymer casting of a chloroform/dimethylformamide mixture. The inclusion of PEG segments improved the hydrophilicity of adipose-derived stem cells in rats and resulted in rapid regeneration in a rat model of knee cartilage defects. Polycaprolactone has also been copolymerized with PLA to prepare films for application in bone tissue engineering (57). Collagen or fibronectin immobilization films were modified, and osteoblast MC3T3-3E1 cells were tested. Positive cell attachment has also been reported previously. It has been shown that engineered mesenchymal stem cells seeded on PCL nanofibrous alone or collagen-coated PCL scaffolds can be used for neural and skin tissue engineering (58, 59).

4. Biodegradation

Polycaprolactone is destroyed under physiological conditions (as seen in the human body) to be utilized as an implantable biomaterial. Owing to its much slower degradation rate than polylactide, the development of long-term implanted devices is an intriguing prospect.

Polycaprolactone degrades in two stages: First, by non-enzymatic hydrolysis of the ester groups and subsequent formation of a more crystalline polymer with a low molecular content (less than 3000) (60). This finding supports the hypothesis that PCL may be completely absorbed and removed through intracellular routes when its molecular weight is reduced to 3000 or less. Additionally, the degraded PCL was found to be almost identical to the *in vitro* hydrolysis at 40°C and accurate to the early stage kinetics in the first step. Polycaprolactone degradation occurs due to an unanticipated breakdown of the ester chain, resulting in a loss of molecular weight. The absolute homopolymer PCL declines during two to four years (depending on the system's weight or implant) (61, 62). Copolymerization affects the rate of hydrolysis of other lactones or glycolides/lactides. Over the past decade, over 1,000 papers on PCL-based textiles have been published in the biomass and tissue engineering literature. Polycaprolactone degradation and resorption kinetics studies were utilized (13). Improvement in the degradability of 58s glass scaffolds such as PCL by ZnO and β -TCP modification has been reported (63).

5. Current Development

The design and processing of organic plastics are gradually being studied. This was the company's exponential increase. In the timeframe up to 2020, global demand for biologically driven plastics will increase from 0.36 million to 3.45 million tons (64). In recent years, the increasing number of PCL publications has described modern biocomposites containing PCL (13, 65, 66). Polycaprolactone is a lightweight resorption polymer, particularly used in biomaterials and tissue engineering, and is popular in chemistry. Polycaprolactone can be converted into composites with better mechanical and biocompatible properties, thus diversifying its use relative to polymers. Their excellent cohesion and potential before resorbing to excretion from the PCL system are mainly used in drug delivery because of their high permeability to some drugs (67). The biodegradation of PCL is gradual and, therefore, more suitable for long-term delivery than other polymers.

It is also used in operating equipment for sutures, injured dressings, and dentistry. Tissue engineering is closely connected to tissue regeneration or repair applications in whole or part (e.g., bone, cartilage, blood vessels, and bladder). Polycaprolactone includes tissue mechanics, skeletal techniques, blood vessels, tendons, and ligaments. Over the last two decades, PCL has almost ignored the success stories of other resorbable

polymers, including polylactides and polyglycolides. Polycaprolactone and its composites deliver excellent results, which translate research into future medical applications. Recently, PCL has been widely used in Tissue Engineering. The lack of bioactivity and high degradation rate of PCL have led to the investigation of its use in bone tissue engineering (23). Recent advances in PCL-based calcium phosphate ceramics have resulted in hybrid biomaterials with good mechanical properties and improved bioactivity (68). As substitutes for bone grafts, PCL and hydroxyapatite nanoparticles can be combined to form 3D scaffolds (69). A 3D-printed PCL scaffold modified with insulin-releasing PLGA nanoparticles was successfully investigated for bone tissue repair by Wei et al. The results demonstrated that this scaffold, in addition to stimulating the proliferation of chondrocytes and differentiation of BMSCs, could enhance bone and cartilage repair *in vivo* (70). Recent studies have shown that PCL-based composites have soft (nerve, skin, and urethra) and hard (musculoskeletal and dental) tissue-repair applications. Hu et al. prepared a degradable patch with an anti-adhesive layer using PCL, polyvinyl alcohol, and soybean peptide, which has great potential for hernia repair (71). In another study, Jeong et al. illustrated that a PCL/ β -TCP scaffold could enhance bone formation in complex zygomaticomaxillary and replace traditional non-absorbable implants in the future (72). Recently, PCL-based scaffolds have attracted the attention of many researchers due to their excellent elastic properties. Several PCL-based scaffolds with excellent mechanical properties have been successfully used for wound regeneration. However, PCL applications in tissue engineering are limited by its intrinsic hydrophobic nature and slow degradation. To improve the adaptability of these scaffolds for skin tissue regeneration, PCL-based composite meshes have been developed (73). An ideal wound dressing should be biocompatible, biodegradable, antimicrobial, and removable without causing any damage to the wound (74). Polycaprolactone-based wound dressings containing Zn, Cu, and Ag exhibit enhanced antimicrobial properties and can be used for wound healing, particularly in skin infections. Muwaffak et al., who used 3D scanning to create 3D models of the ear and nose, demonstrated that PCL-based wound dressings with Cu and Ag showed excellent antimicrobial properties against *Staphylococcus aureus* (75). Polycaprolactone-based 3D-printed skin scaffolds incorporating Ag show promising applications in skin regeneration, as reported by Ninan et al. (76). A recent new area of PCL application concerns esthetic purposes in humans. Wee et al. introduced a three-dimensional

printed PCL as an easy-to-use implant with good results for aesthetic nasal lobule correction (77). In another study, PCL-based products were used to treat facial aging by stimulating collagen formation. The long-term effectiveness and safety of PCL fillers have been confirmed in clinical studies (78).

6. Polycaprolactone Safety

Polycaprolactone systems' main advantages and functionalities have led scientists and practitioners to build and use polymers (13, 51). As explained below, particularly for the latest PCL product choices, the biomedical applications of embedded PCLs endorse PCL protection as used in the new framework (79). Polycaprolactone and polyglycolide were copolymerized in Monocryl™ (Ethicon, Inc.; Somerville, New Jersey, USA) (80). This suture retains high tensile strength and allows the tissue to respond minimally to its defense. Polycaprolactone is ideal for long-term drug delivery owing to its elevated drug permeability, excellent biocompatibility, slow biological degradation, and bioresorbability. In addition, a specially formulated and scientifically produced levonorgestrel PCL biodegradable Capronor™ pill (up to 2 years) is used in the PCL drug delivery system, which offers valuable details on global PCL protection and supports the steady decline of PCL and its long-term safety. Polycaprolactone microspheres or nanospheres have been used in a range of medicines (62, 81-83). Various 3D-growing materials are used for bone, skin, or other tissues (19, 84), and autologous graft processing is used to repair and regenerate implantation tissue (83). Due to its physicochemical characteristics, mechanical nature, and effects, 3D printing scaffolding can also be used in the fabric manufacturing industry, an advanced research area, and an application field in which PCL plays a significant role. 3D printing uses computerized processes via subsequent layer deposition to create tissue and organ replacement structures. This is a promising treatment approach in PCL's life-saving (85-90). A Chinese team developed the first three-dimensional PCL airway for individuals who endured tracheomalacia, a life-threatening disease, and managed to save the lives of a newborn and a woman. The role of PCL in tracheal surgery has recently been explored. Congenital heart abnormalities, gastric wall injury (hollow organs), and periodontal repair have also been investigated (91-93). Orthopedic interest is enormous; for instance, meniscus repair affects millions of individuals worldwide (94-96).

7. Conclusions

Polycaprolactone has numerous applications, including in the pharmaceutical industry, such as medication release monitoring, tissue processing, and bone scaffolding. Combining PCL with other materials to form nanocomposites improves the properties of the resulting materials and expands the scope of applications that can benefit from the materials' superior qualities. This biomaterial may have potential applications in the years to come.

Acknowledgements

The authors would like to thank Qazvin University of Medical Sciences and Shahrekord University.

Footnotes

Authors' Contribution: Not declared by the authors.

Conflict of Interests: The authors declare no conflicts of interest.

Data Reproducibility: The dataset presented in the study is available on request from the corresponding author during submission or after publication.

Funding/Support: There is no funding or support.

References

- Nair LS, Laurencin CT. Biodegradable polymers as biomaterials. *Progress in polymer science*. 2007;**32**(8-9):762-98. <https://doi.org/10.1016/j.progpolymsci.2007.05.017>.
- Lowery JL, Datta N, Rutledge GC. Effect of fiber diameter, pore size and seeding method on growth of human dermal fibroblasts in electrospun poly(epsilon-caprolactone) fibrous mats. *Biomaterials*. 2010;**31**(3):491-504. [PubMed ID: 19822363]. <https://doi.org/10.1016/j.biomaterials.2009.09.072>.
- Kim JY, Cho D. Blended PCL/PLGA scaffold fabrication using multi-head deposition system. *Microelectronic Engineering*. 2009;**86**(4-6):1447-50. <https://doi.org/10.1016/j.mee.2008.11.026>.
- Chong C, Wang Y, Fathi A, Parungao R, Maitz PK, Li Z. Skin wound repair: Results of a pre-clinical study to evaluate electrospun collagen-elastin-PCL scaffolds as dermal substitutes. *Burns*. 2019;**45**(7):1639-48. [PubMed ID: 31076208]. <https://doi.org/10.1016/j.burns.2019.04.014>.
- Mas Estelles J, Vidaurre A, Meseguer Duenas JM, Castilla Cortazar I. Physical characterization of polycaprolactone scaffolds. *J Mater Sci Mater Med*. 2008;**19**(1):189-95. [PubMed ID: 17597379]. <https://doi.org/10.1007/s10856-006-0101-2>.
- Simoes SM, Figueiras AR, Veiga F, Concheiro A, Alvarez-Lorenzo C. Polymeric micelles for oral drug administration enabling locoregional and systemic treatments. *Expert Opin Drug Deliv*. 2015;**12**(2):297-318. [PubMed ID: 25227130]. <https://doi.org/10.1517/17425247.2015.960841>.
- Ikada Y, Tsuji H. Biodegradable polyesters for medical and ecological applications. *Macromolecular Rapid Communications*. 2000;**21**(3):17-32. [https://doi.org/10.1002/\(SICI\)1521-3927\(20000201\)21:3<17::AID-MARCI7>3.0.CO;2-X](https://doi.org/10.1002/(SICI)1521-3927(20000201)21:3<17::AID-MARCI7>3.0.CO;2-X).
- Alix S, Mahieu A, Terrie C, Soulestin J, Gerault E, Feuilloley MG, et al. Active pseudo-multilayered films from polycaprolactone and starch based matrix for food-packaging applications. *European Polymer J*. 2013;**49**(6):1234-42. <https://doi.org/10.1016/j.eurpolymj.2013.03.016>.
- Gaucher G, Dufresne MH, Sant VP, Kang N, Maysinger D, Leroux JC. Block copolymer micelles: Preparation, characterization and application in drug delivery. *J Control Release*. 2005;**109**(1-3):169-88. [PubMed ID: 16289422]. <https://doi.org/10.1016/j.jconrel.2005.09.034>.
- Annabi N, Fathi A, Mithieux SM, Weiss AS, Dehghani F. Fabrication of porous PCL/elastin composite scaffolds for tissue engineering applications. *J Supercritical Fluids*. 2011;**59**:157-67. <https://doi.org/10.1016/j.supflu.2011.06.010>.
- Gaucher G, Satturwar P, Jones MC, Furtos A, Leroux JC. Polymeric micelles for oral drug delivery. *Eur J Pharm Biopharm*. 2010;**76**(2):147-58. [PubMed ID: 20600891]. <https://doi.org/10.1016/j.ejpb.2010.06.007>.
- Gorna K, Gogolewski S. In vitro degradation of novel medical biodegradable aliphatic polyurethanes based on epsilon-caprolactone and Pluronic® with various hydrophilicities. *Polymer Degradation Stability*. 2002;**75**(1):113-22. [https://doi.org/10.1016/S0141-3910\(01\)00210-5](https://doi.org/10.1016/S0141-3910(01)00210-5).
- Woodruff MA, Hutmacher DW. The return of a forgotten polymer—Polycaprolactone in the 21st century. *Progress In Polymer Sci*. 2010;**35**(10):1217-56. <https://doi.org/10.1016/j.progpolymsci.2010.04.002>.
- Labet M, Thielemans W. Synthesis of polycaprolactone: A review. *Chem Soc Rev*. 2009;**38**(12):3484-504. [PubMed ID: 20449064]. <https://doi.org/10.1039/b820162p>.
- Storey RF, Taylor AE. Effect of stannous octoate concentration on the ethylene glycol-initiated polymerization of epsilon-caprolactone. *Abstracts of Papers of the American Chemical Society*. 1996. 114 p.
- Okada M. Chemical syntheses of biodegradable polymers. *Progress in polymer science*. 2002;**27**(1):87-133. [https://doi.org/10.1016/S0079-6700\(01\)00039-9](https://doi.org/10.1016/S0079-6700(01)00039-9).
- Hayashi T. Biodegradable polymers for biomedical uses. *Progress in polymer science*. 1994;**19**(4):663-702. [https://doi.org/10.1016/0079-6700\(94\)90030-2](https://doi.org/10.1016/0079-6700(94)90030-2).
- Pillai CK, Sharma CP. Review paper: Absorbable polymeric surgical sutures: Chemistry, production, properties, biodegradability, and performance. *J Biomater Appl*. 2010;**25**(4):291-366. [PubMed ID: 20971780]. <https://doi.org/10.1177/0885328210384890>.
- Dash TK, Konkimalla VB. Poly-small je, Ukrainian-caprolactone based formulations for drug delivery and tissue engineering: A review. *J Control Release*. 2012;**158**(1):15-33. [PubMed ID: 21963774]. <https://doi.org/10.1016/j.jconrel.2011.09.064>.
- Merkli A, Tabatabay C, Gurny R, Heller J. Biodegradable polymers for the controlled release of ocular drugs. *Progress In Polymer Sci*. 1998;**23**(3):563-80. [https://doi.org/10.1016/S0079-6700\(97\)00048-8](https://doi.org/10.1016/S0079-6700(97)00048-8).
- Freiberg S, Zhu XX. Polymer microspheres for controlled drug release. *Int J Pharm*. 2004;**282**(1-2):1-18. [PubMed ID: 15336378]. <https://doi.org/10.1016/j.ijpharm.2004.04.013>.
- Sinha VR, Bansal K, Kaushik R, Kumria R, Trehan A. Poly-epsilon-caprolactone microspheres and nanospheres: An overview. *Inter J Pharmaceutics*. 2004;**278**(1):1-23. <https://doi.org/10.1016/j.ijpharm.2004.01.044>.
- Manivasagam G, Reddy A, Sen D, Nayak S, Mathew MT, Rajamanikam A. Dentistry: Restorative and regenerative approaches. *Restorative Regenerative Approaches*. 2019. <https://doi.org/10.1016/B978-0-12-801238-3.11017-7>.
- Lemmouchi Y, Schacht E, Kageruka P, De Deken R, Diarra B, Diall O, et al. Biodegradable polyesters for controlled release of trypanocidal

- drugs: in vitro and in vivo studies. *Biomaterials*. 1998;**19**(20):1827-37. [PubMed ID: 9851583]. [https://doi.org/10.1016/S0142-9612\(98\)00074-x](https://doi.org/10.1016/S0142-9612(98)00074-x).
25. Koleske JV. Blends containing poly (ϵ -caprolactone) and related polymers. *Polymer blends*. Elsevier; 1978. p. 369-89. <https://doi.org/10.1016/B978-0-12-546802-2.50018-X>.
 26. Kiminta DMO, Braithwaite G, Luckham PF. Colloidal dispersions, nanogels. *Polymer Materials Encyclopedia*. 1996;**1**:1298-309.
 27. Mark HF, Kroschwitz JI. *Encyclopedia of polymer science and engineering*. 2. New York: Wiley; 1985.
 28. Weissman JM, Sunkara HB, Tse AS, Asher SA. Thermally switchable periodicities and diffraction from mesoscopically ordered materials. *Science*. 1996;**274**(5289):959-60. [PubMed ID: 8875932]. <https://doi.org/10.1126/science.274.5289.959>.
 29. Salamone JC. *Polymeric materials encyclopedia, Twelve volume set*. CRC press; 2020.
 30. Grignard B, Stassin F, Calberg C, Jerome R, Jerome C. Synthesis of biodegradable poly-epsilon-caprolactone microspheres by dispersion ring-opening polymerization in supercritical carbon dioxide. *Biomacromolecules*. 2008;**9**(11):3141-9. [PubMed ID: 18937397]. <https://doi.org/10.1021/bm800730m>.
 31. Kwon S, Lee K, Kim H, Lee Y, Bae W. Synthesis of biocompatible and biodegradable polymer particles in supercritical carbon dioxide. *Colloid and Polymer Science*. 2008;**286**:1181-91. <https://doi.org/10.1007/s00396-008-1888-9>.
 32. Vasir JK, Tambwekar K, Garg S. Bioadhesive microspheres as a controlled drug delivery system. *Int J Pharm*. 2003;**255**(1-2):13-32. [PubMed ID: 12672598]. [https://doi.org/10.1016/S0378-5173\(03\)00087-5](https://doi.org/10.1016/S0378-5173(03)00087-5).
 33. Bai X, Yang Y, Chung T, Ng S, Heller J. Effect of polymer compositions on the fabrication of poly (ortho-ester) microspheres for controlled release of protein. *J Applied Polymer Sci*. 2001;**80**(10):1630-42. <https://doi.org/10.1002/app.1257>.
 34. Calvo P, Sanchez A, Martinez J, Lopez MI, Calonge M, Pastor JC, et al. Polyester nanocapsules as new topical ocular delivery systems for cyclosporin A. *Pharm Res*. 1996;**13**(2):311-5. [PubMed ID: 8932455]. <https://doi.org/10.1023/a:1016015803611>.
 35. Zhang S, Uludag H. Nanoparticulate systems for growth factor delivery. *Pharm Res*. 2009;**26**(7):1561-80. [PubMed ID: 19415467]. <https://doi.org/10.1007/s11095-009-9897-z>.
 36. Gref R, Luck M, Quellec P, Marchand M, Dellacherie E, Harnisch S, et al. 'Stealth' corona-core nanoparticles surface modified by polyethylene glycol (PEG): influences of the corona (PEG chain length and surface density) and of the core composition on phagocytic uptake and plasma protein adsorption. *Colloids Surf B Biointerfaces*. 2000;**18**(3-4):301-13. [PubMed ID: 10915952]. [https://doi.org/10.1016/S0927-7765\(99\)00156-3](https://doi.org/10.1016/S0927-7765(99)00156-3).
 37. Saez A, Guzman M, Molpeceres J, Aberturas MR. Freeze-drying of polycaprolactone and poly(D,L-lactic-glycolic) nanoparticles induce minor particle size changes affecting the oral pharmacokinetics of loaded drugs. *Eur J Pharm Biopharm*. 2000;**50**(3):379-87. [PubMed ID: 11072195]. [https://doi.org/10.1016/S0939-6411\(00\)00125-9](https://doi.org/10.1016/S0939-6411(00)00125-9).
 38. Boia R, Dias PAN, Martins JM, Galindo-Romero C, Aires ID, Vidal-Sanz M, et al. Porous poly(epsilon-caprolactone) implants: A novel strategy for efficient intraocular drug delivery. *J Control Release*. 2019;**316**:331-48. [PubMed ID: 31715277]. <https://doi.org/10.1016/j.jconrel.2019.09.023>.
 39. Abdullah T, Gauthaman K, Mostafavi A, Alshahrie A, Salah N, Morganti P, et al. Sustainable drug release from polycaprolactone coated chitin-lignin gel fibrous scaffolds. *Sci Rep*. 2020;**10**(1):20428. [PubMed ID: 33235239]. [PubMed Central ID: PMC7686307]. <https://doi.org/10.1038/s41598-020-76971-w>.
 40. Hou Z, Zhou W, Guo X, Zhong R, Wang A, Li J, et al. Poly(epsilon-caprolactone)-Methoxypolyethylene Glycol (PCL-MPEG)-based micelles for drug-delivery: The effect of pcl chain length on blood components, phagocytosis, and biodistribution. *Int J Nanomedicine*. 2022;**17**:1613-32. [PubMed ID: 35411141]. [PubMed Central ID: PMC8994631]. <https://doi.org/10.2147/IJN.S349516>.
 41. Nikzamir N, Khojasteh H, Nobakht Vakili M, Azimi C, Ghanbari E. Preparation of degeredable polyprolactone polymer (PCL)/magnetic nanocomposite for drug delivery systems against anticancer compounds. *J Nanostructures*. 2021;**11**(3):456-69. <https://doi.org/10.22052/JNS.2021.03.005>.
 42. Espinoza SM, Patil HI, San Martin Martinez E, Casanas Pimentel R, Ige PP. Poly- ϵ -caprolactone (PCL), a promising polymer for pharmaceutical and biomedical applications: Focus on nanomedicine in cancer. *Inter J Polymeric Materials Polymeric Biomat*. 2020;**69**(2):85-126. <https://doi.org/10.1080/00914037.2018.1539990>.
 43. Astuti SH, Rahma WA, Budianto E. Biodegradable Microcapsules from D, L-PLA/PCL as controlled nifedipine drug delivery carrier. *Macromolecular Symposia*. Wiley Online Library; 2020. 1900132 p.
 44. Dias AM, da Silva FG, Monteiro APF, Pinzon-Garcia AD, Sinisterra RD, Cortes ME. Polycaprolactone nanofibers loaded oxytetracycline hydrochloride and zinc oxide for treatment of periodontal disease. *Mater Sci Eng C Mater Biol Appl*. 2019;**103**:109798. [PubMed ID: 31349501]. <https://doi.org/10.1016/j.msec.2019.109798>.
 45. Welman T, Michel S, Segaren N, Shanmugarajah K. Bioengineering for organ transplantation: Progress and challenges. *Bioengineered*. 2015;**6**(5):257-61. [PubMed ID: 26259720]. [PubMed Central ID: PMC4825836]. <https://doi.org/10.1080/21655979.2015.1081320>.
 46. Prabhakar A, Lynch AP, Ahearne M. Self-assembled infrapatellar fat-pad progenitor cells on a poly-epsilon-caprolactone film for cartilage regeneration. *Artif Organs*. 2016;**40**(4):376-84. [PubMed ID: 26516689]. <https://doi.org/10.1111/aor.12565>.
 47. Romagnoli C, Zonefrati R, Galli G, Puppi D, Piroso A, Chiellini F, et al. In vitro behavior of human adipose tissue-derived stem cells on poly(epsilon-caprolactone) film for bone tissue engineering applications. *Biomed Res Int*. 2015;**2015**:323571. [PubMed ID: 26558266]. [PubMed Central ID: PMC4617699]. <https://doi.org/10.1155/2015/323571>.
 48. Kosmala A, Fitzgerald M, Moore E, Stam F. Evaluation of a gelatin-modified poly (ϵ -caprolactone) film as a scaffold for lung disease. *Analytical Letters*. 2017;**50**(1):219-32. <https://doi.org/10.1080/00032719.2016.1163363>.
 49. Zafari F, Shirian S, Sadeghi M, Teimourian S, Bakhtiyari M. CD93 hematopoietic stem cells improve diabetic wound healing by VEGF activation and downregulation of DAPK-1. *J Cell Physiol*. 2020;**235**(3):2366-76. [PubMed ID: 31549396]. <https://doi.org/10.1002/jcp.29142>.
 50. Yu S, Gao Y, Mei X, Ren T, Liang S, Mao Z, et al. Preparation of an arg-glu-asp-val peptide density gradient on hyaluronic acid-coated poly(epsilon-caprolactone) film and its influence on the selective adhesion and directional migration of endothelial cells. *ACS Appl Mater Interfaces*. 2016;**8**(43):29280-8. [PubMed ID: 27723284]. <https://doi.org/10.1021/acsami.6b09375>.
 51. Malikmammadov E, Tanir TE, Kiziltay A, Hasirci N. PCL and PCL-based materials in biomedical applications. *J Biomater Sci Polym Ed*. 2018;**29**(7-9):863-93. [PubMed ID: 29053081]. <https://doi.org/10.1080/09205063.2017.1394711>.
 52. Tong SY, Wang Z, Lim PN, Wang W, Thian ES. Uniformly-dispersed nanohydroxapatite-reinforced poly(epsilon-caprolactone) composite films for tendon tissue engineering application. *Mater Sci Eng C Mater Biol Appl*. 2017;**70**(Pt 2):1149-55. [PubMed ID: 27772716]. <https://doi.org/10.1016/j.msec.2016.03.051>.
 53. Pai BG, Kulkarni AV, Jain S. Study of smart antibacterial PCL-xFe(3)O(4) thin films using mouse NIH-3T3 fibroblast cells in vitro. *J Biomed Mater Res B Appl Biomater*. 2017;**105**(4):795-804. [PubMed ID: 26762566]. <https://doi.org/10.1002/jbm.b.33615>.

54. Catauro M, Bollino F, Papale F, Mozetic P, Rainer A, Trombetta M. Biological response of human mesenchymal stromal cells to titanium grade 4 implants coated with PCL/ZrO(2) hybrid materials synthesized by sol-gel route: In vitro evaluation. *Mater Sci Eng C Mater Biol Appl.* 2014;**45**:395-401. [PubMed ID: [25491844](#)]. <https://doi.org/10.1016/j.msec.2014.09.007>.
55. Hashiwaki H, Teramoto Y, Nishio Y. Fabrication of thermoplastic ductile films of chitin butyrate/poly(varepsilon-caprolactone) blends and their cytocompatibility. *Carbohydr Polym.* 2014;**114**:330-8. [PubMed ID: [25263898](#)]. <https://doi.org/10.1016/j.carbpol.2014.08.028>.
56. Chen M, Zhang Y, Zhou Y, Zhang Y, Lang M, Ye Z, et al. Pendant small functional groups on poly(ϵ -caprolactone) substrate modulate adhesion, proliferation and differentiation of human mesenchymal stem cells. *Colloids Surf B Biointerfaces.* 2015;**134**:322-31. [PubMed ID: [26209965](#)]. <https://doi.org/10.1016/j.colsurfb.2015.07.018>.
57. Fuse M, Hayakawa T, Hashizume-Takizawa T, Takeuchi R, Kurita-Ochiai T, Fujita-Yoshigaki J, et al. MC3T3-E1 cell assay on collagen or fibronectin immobilized poly (lactic acid- ϵ -caprolactone) film. *Journal of Hard Tissue Biology.* 2015;**24**(3):249-56. <https://doi.org/10.2485/jhtb.24.249>.
58. Shirian S, Ebrahimi-Barough S, Saberi H, Norouzi-Javidan A, Mousavi SM, Derakhshan MA, et al. Comparison of capability of human bone marrow mesenchymal stem cells and endometrial stem cells to differentiate into motor neurons on electrospun poly(epsilon-caprolactone) scaffold. *Mol Neurobiol.* 2016;**53**(8):5278-87. [PubMed ID: [26420037](#)]. <https://doi.org/10.1007/s12035-015-9442-5>.
59. Sharif S, Ai J, Azami M, Verdi J, Atlasi MA, Shirian S, et al. Collagen-coated nano-electrospun PCL seeded with human endometrial stem cells for skin tissue engineering applications. *J Biomed Mater Res B Appl Biomater.* 2018;**106**(4):1578-86. [PubMed ID: [28792664](#)]. <https://doi.org/10.1002/jbm.b.33966>.
60. Woodward SC, Brewer PS, Moatamed F, Schindler A, Pitt CG. The intracellular degradation of poly(epsilon-caprolactone). *J Biomed Mater Res.* 1985;**19**(4):437-44. [PubMed ID: [4055826](#)]. <https://doi.org/10.1002/jbm.820190408>.
61. Gunatillake PA, Adhikari R. Biodegradable synthetic polymers for tissue engineering. *Eur Cell Mater.* 2003;**5**:1-16. discussion 16. [PubMed ID: [14562275](#)]. <https://doi.org/10.22203/ecm.v005a01>.
62. Middleton JC, Tipton AJ. Synthetic biodegradable polymers as orthopedic devices. *Biomaterials.* 2000;**21**(23):2335-46. [PubMed ID: [11055281](#)]. [https://doi.org/10.1016/S0142-9612\(00\)00101-0](https://doi.org/10.1016/S0142-9612(00)00101-0).
63. Shuai C, Cao Y, Dan G, Gao C, Feng P, Wu P. Improvement in degradability of 58s glass scaffolds by ZnO and beta-TCP modification. *Bioengineered.* 2016;**7**(5):342-51. [PubMed ID: [27710432](#)]. [PubMed Central ID: [PMC5060980](#)]. <https://doi.org/10.1080/21655979.2016.1197032>.
64. Faruk O, Bledzki AK, Fink H, Sain M. Biocomposites reinforced with natural fibers: 2000-2010. *Progress In Polymer Sci.* 2012;**37**(11):1552-96. <https://doi.org/10.1016/j.progpolymsci.2012.04.003>.
65. Martínez-Abad A, Sánchez G, Fuster V, Lagaron JM, Ocio MJ. Antibacterial performance of solvent cast polycaprolactone (PCL) films containing essential oils. *Food Control.* 2013;**34**(1):214-20. <https://doi.org/10.1016/j.foodcont.2013.04.025>.
66. Kelly CA, Murphy SH, Leeke GA, Howdle SM, Shakesheff KM, Jenkins MJ. Rheological studies of polycaprolactone in supercritical CO₂. *European Polymer J.* 2013;**49**(2):464-70.
67. Ruckh TT, Kumar K, Kipper MJ, Popat KC. Osteogenic differentiation of bone marrow stromal cells on poly(epsilon-caprolactone) nanofiber scaffolds. *Acta Biomater.* 2010;**6**(8):2949-59. [PubMed ID: [20144747](#)]. <https://doi.org/10.1016/j.actbio.2010.02.006>.
68. Hajiali F, Tajbakhsh S, Shojaei A. Fabrication and properties of polycaprolactone composites containing calcium phosphate-based ceramics and bioactive glasses in bone tissue engineering: A review. *Polymer Reviews.* 2018;**58**(1):164-207. <https://doi.org/10.1080/15583724.2017.1332640>.
69. Murugan S, Parcha SR. Fabrication techniques involved in developing the composite scaffolds PCL/HA nanoparticles for bone tissue engineering applications. *J Mater Sci Mater Med.* 2021;**32**(8):93. [PubMed ID: [34379204](#)]. [PubMed Central ID: [PMC8357662](#)]. <https://doi.org/10.1007/s10856-021-06564-0>.
70. Wei P, Xu Y, Zhang H, Wang L. Continued sustained insulin-releasing PLGA nanoparticles modified 3D-Printed PCL composite scaffolds for osteochondral repair. *Chem Engineering J.* 2021;**422**:130051. <https://doi.org/10.1016/j.cej.2021.130051>.
71. Hu Q, Wu J, Zhang H, Dong W, Gu Y, Liu S. Designing double-layer multimaterial composite patch scaffold with adhesion resistance for hernia repair. *Macromol Biosci.* 2022;**22**(6). e2100510. [PubMed ID: [35471592](#)]. <https://doi.org/10.1002/mabi.202100510>.
72. Jeong WS, Kim YC, Min JC, Park HJ, Lee EJ, Shim JH, et al. Clinical application of 3D-printed patient-specific polycaprolactone/beta tricalcium phosphate scaffold for complex zygomatico-maxillary defects. *Polymers (Basel).* 2022;**14**(4). [PubMed ID: [35215652](#)]. [PubMed Central ID: [PMC8875444](#)]. <https://doi.org/10.3390/polym14040740>.
73. Zhang Y, Ouyang H, Lim CT, Ramakrishna S, Huang ZM. Electrospinning of gelatin fibers and gelatin/PCL composite fibrous scaffolds. *J Biomed Mater Res B Appl Biomater.* 2005;**72**(1):156-65. [PubMed ID: [15389493](#)]. <https://doi.org/10.1002/jbm.b.30128>.
74. Dong Y, Zheng Y, Zhang K, Yao Y, Wang L, Li X, et al. Electrospun nanofibrous materials for wound healing. *Advanced Fiber Materials.* 2020;**2**:212-27. <https://doi.org/10.1007/s42765-020-00034-y>.
75. Muwaffak Z, Goyanes A, Clark V, Basit AW, Hilton ST, Gaisford S. Patient-specific 3D scanned and 3D printed antimicrobial polycaprolactone wound dressings. *Int J Pharm.* 2017;**527**(1-2):161-70. [PubMed ID: [28461267](#)]. <https://doi.org/10.1016/j.ijpharm.2017.04.077>.
76. Ninan N, Joseph B, Visalakshan RM, Bright R, Denoual C, Zilm P, et al. Plasma assisted design of biocompatible 3D printed PCL/silver nanoparticle scaffolds: In vitro and in vivo analyses. *Materials Advances.* 2021;**2**(20):6620-30. <https://doi.org/10.1039/D1MA00444A>.
77. Wee SY, Kim TH, Kang HY, Park ES. Aesthetic nasal lobule correction using a three-dimensional printed polycaprolactone implant. *J Craniofac Surg.* 2021;**32**(8):e808-12. [PubMed ID: [34292245](#)]. <https://doi.org/10.1097/SCS.00000000000007855>.
78. Christen MO, Vercesi F. Polycaprolactone: How a well-known and futuristic polymer has become an innovative collagen-stimulator in esthetics. *Clin Cosmet Investig Dermatol.* 2020;**13**:31-48. [PubMed ID: [32161484](#)]. [PubMed Central ID: [PMC7065466](#)]. <https://doi.org/10.2147/CCID.S229054>.
79. Ulery BD, Nair LS, Laurencin CT. Biomedical applications of biodegradable polymers. *J Polym Sci B Polym Phys.* 2011;**49**(12):832-64. [PubMed ID: [21769165](#)]. [PubMed Central ID: [PMC3136871](#)]. <https://doi.org/10.1002/polb.22259>.
80. Bezwada RS, Jamiolkowski DD, Lee IY, Agarwal V, Persivale J, Trenka-Benthin S, et al. Monocryl suture, a new ultra-pliable absorbable monofilament suture. *Biomaterials.* 1995;**16**(15):1141-8. [PubMed ID: [8562789](#)]. [https://doi.org/10.1016/0142-9612\(95\)93577-z](https://doi.org/10.1016/0142-9612(95)93577-z).
81. Mogosanu GD, Grumezescu AM. Natural and synthetic polymers for wounds and burns dressing. *Int J Pharm.* 2014;**463**(2):127-36. [PubMed ID: [24368109](#)]. <https://doi.org/10.1016/j.ijpharm.2013.12.015>.
82. Hernandez I, Kumar A, Joddar B. A bioactive hydrogel and 3D printed polycaprolactone system for bone tissue engineering. *Gels.* 2017;**3**(3). [PubMed ID: [29354645](#)]. [PubMed Central ID: [PMC5770986](#)]. <https://doi.org/10.3390/gels3030026>.
83. Avolio E, Caputo M, Madeddu P. Stem cell therapy and tissue engineering for correction of congenital heart disease. *Front Cell Dev Biol.* 2015;**3**:39. [PubMed ID: [26176009](#)]. [PubMed Central ID: [PMC4485350](#)]. <https://doi.org/10.3389/fcell.2015.00039>.

84. Abedalwafa M, Wang F, Wang L, Li C. Biodegradable poly-epsilon-caprolactone (PCL) for tissue engineering applications: A review. *Rev Adv Mater Sci.* 2013;**34**(2):123-40.
85. He Y, Kilsby S, Tuck C, Wildman R, Christie S, Edmonson S, et al. Processing biodegradable polycaprolactone through 3D printing. *Proceedings of the 24th International SFF Symposium-An Additive Manufacturing Conference.* Austin, TX, USA. 2013.
86. Vijayavenkataraman S, Fuh JYH, Lu WF. 3D Printing and 3D Bioprinting in Pediatrics. *Bioengineering (Basel).* 2017;**4**(3). [PubMed ID: 28952542]. [PubMed Central ID: PMC5615309]. <https://doi.org/10.3390/bioengineering4030063>.
87. Bikiaris D, Papageorgiou G, Achilias D, Pavlidou E, Stergiou A. Miscibility and enzymatic degradation studies of poly(epsilon-caprolactone)/poly(propylene succinate) blends. *European Polymer J.* 2007;**43**:2491-503. <https://doi.org/10.1016/j.eurpolymj.2007.03.051>.
88. Dawood A, Marti Marti B, Sauret-Jackson V, Darwood A. 3D printing in dentistry. *Br Dent J.* 2015;**219**(11):521-9. [PubMed ID: 26657435]. <https://doi.org/10.1038/sj.bdj.2015.914>.
89. Huang Y, Zhang XF, Gao G, Yonezawa T, Cui X. 3D bioprinting and the current applications in tissue engineering. *Biotechnol J.* 2017;**12**(8). [PubMed ID: 28675678]. <https://doi.org/10.1002/biot.201600734>.
90. Li C, Cheung TF, Fan VC, Sin KM, Wong CW, Leung GK. Applications of three-dimensional printing in surgery. *Surg Innov.* 2017;**24**(1):82-8. [PubMed ID: 27913755]. <https://doi.org/10.1177/1553350616681889>.
91. Kankala RK, Zhu K, Li J, Wang CS, Wang SB, Chen AZ. Fabrication of arbitrary 3D components in cardiac surgery: From macro-, micro- to nanoscale. *Biofabrication.* 2017;**9**(3):32002. [PubMed ID: 28770811]. <https://doi.org/10.1088/1758-5090/aa8113>.
92. Hendow EK, Guhmann P, Wright B, Sofokleous P, Parmar N, Day RM. Biomaterials for hollow organ tissue engineering. *Fibrogenesis Tissue Repair.* 2016;**9**:3. [PubMed ID: 27014369]. [PubMed Central ID: PMC4806416]. <https://doi.org/10.1186/s13069-016-0040-6>.
93. Rasperini G, Pilipchuk SP, Flanagan CL, Park CH, Pagni G, Hollister SJ, et al. 3D-printed Bioresorbable Scaffold for Periodontal Repair. *J Dent Res.* 2015;**94**(9 Suppl):153S-7S. [PubMed ID: 26124215]. <https://doi.org/10.1177/0022034515588303>.
94. Sun J, Vijayavenkataraman S, Liu H. An overview of scaffold design and fabrication technology for engineered knee meniscus. *Materials (Basel).* 2017;**10**(1). [PubMed ID: 28772388]. [PubMed Central ID: PMC5344568]. <https://doi.org/10.3390/ma10010029>.
95. Szojka A, Lalh K, Andrews SH, Jomha NM, Osswald M, Adesida AB. Biomimetic 3D printed scaffolds for meniscus tissue engineering. *Bioprinting.* 2017;**8**:1-7. <https://doi.org/10.1016/j.bprint.2017.08.001>.
96. Zhang ZZ, Wang SJ, Zhang JY, Jiang WB, Huang AB, Qi YS, et al. 3D-Printed Poly(epsilon-caprolactone) Scaffold Augmented With Mesenchymal Stem Cells for Total Meniscal Substitution: A 12- and 24-Week Animal Study in a Rabbit Model. *Am J Sports Med.* 2017;**45**(7):1497-511. [PubMed ID: 28278383]. <https://doi.org/10.1177/0363546517691513>.