



Aptamers: Rapid and Emerging Tool for Diagnosis of *Pseudomonas aeruginosa* Keratitis

Sara Rahimi^{1,2}, Mehdi Bakht^{1,2}, Hamid Sadeghi^{1,2}, Saeideh Gholamzadeh Khoei ^{1,*}, Fatemeh Karimi Dermani ^{3,**}

¹ Medical Microbiology Research Center, Qazvin University of Medical Sciences, Qazvin, Iran

² Student Research Committee, Qazvin University of Medical Sciences, Qazvin, Iran

³ Hellen Diller Family Cancer Research, University of California San Francisco, USA

* **Corresponding Author:** Medical Microbiology Research Center, Qazvin University of Medical Sciences, Qazvin, Iran. Email: s.gholamzade@yahoo.com

** **Corresponding Author:** Hellen Diller Family Cancer Research, University of California San Francisco, USA. Email: fatemekarimi2007@yahoo.com

Received: 28 August, 2024; Accepted: 3 March, 2024

Keywords: Keratitis, *Pseudomonas aeruginosa*, Aptamers

Dear Editor,

Bacterial keratitis is a corneal disease commonly associated with the extended use of soft contact lenses (1). This bacterial infection of the cornea is a serious medical condition that requires urgent treatment due to the risk of reduced vision or even vision loss in the affected eye (2). Bacteria isolated from patients with keratitis include a variety of organisms, such as *Pseudomonas* spp., *Staphylococcus* spp., *Serratia* spp., and *Streptococcus* spp. (2). Among these, *Pseudomonas aeruginosa*, a Gram-negative pathogen, is known as an opportunistic bacterium, particularly in immunocompromised patients (3). It is responsible for a wide range of infections and is the most common causative agent of bacterial keratitis related to contact-lens use (4). *Pseudomonas aeruginosa* infections are difficult to treat due to the bacterium's ability to rapidly acquire antibiotic resistance (3). Bacterial keratitis is notorious for causing rapid, fulminant disease and vision loss, even in treated patients. Early detection of keratitis is crucial for treatment and for preventing vision loss. As such, new strategies are needed for the rapid detection of microorganisms responsible for bacterial keratitis.

Aptamers, single-stranded nucleic acids that bind to specific targets, function similarly to antibodies. Aptamers have recently gained attention due to their desirable properties, including ease of chemical modification, cost-effective production, long-term stability, and binding affinity comparable to proteins (5). Several methods exist for detecting *P. aeruginosa*, including bacterial culture, ELISA, polymerase chain reaction (PCR)-based methods, MALDI-TOF MS assays, and electrochemical biosensors (6). While these

methods are sensitive, they are also costly, time-consuming, and require specialized facilities. Therefore, it is necessary to explore new, low-cost, simple, and rapid methods for detecting *P. aeruginosa* in bacterial keratitis cases (7). This study proposes aptamers as a promising tool for the urgent detection of *P. aeruginosa*.

In a study conducted in 2018, a colorimetric aptamer was designed to detect *P. aeruginosa* in water and chicken samples, with a linear detection range of 10^2 to 10^6 CFU/mL and a detection time of 2 hours (8). Another study detected *P. aeruginosa* in water samples within just 10 minutes (9), while a separate study reported detection in milk, juice, and popsicle samples within 1.5 hours (10).

Although immunological methods offer faster results than bacterial culture, the quality of antibodies and the complexity of operations can affect the accuracy of the results. Molecular methods, such as PCR, are highly sensitive and specific. However, various factors may interfere with the results, leading to false positives or negatives (11). Additionally, PCR-based methods cannot distinguish between live and dead bacteria, which is a potential limitation for their future development (12). Therefore, it is crucial to develop a fast, accurate, sensitive, affordable, and portable method for detecting pathogenic bacteria, which is essential for both public health and clinical diagnosis.

Footnotes

Authors' Contribution: Not declared by the authors.

Conflict of Interests Statement: The authors declare no conflict of interest.

Funding/Support: This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

References

1. Fleiszig SM, Evans DJ. The pathogenesis of bacterial keratitis: studies with *Pseudomonas aeruginosa*. *Clin Exp Optom*. 2002;**85**(5):271-8. [PubMed ID: 12366347]. <https://doi.org/10.1111/j.1444-0938.2002.tb03082.x>.
2. Wang AG, Wu CC, Liu JH. Bacterial corneal ulcer: a multivariate study. *Ophthalmologica*. 1998;**212**(2):126-32. [PubMed ID: 9486553]. <https://doi.org/10.1159/000027291>.
3. Bakht M, Alizadeh SA, Rahimi S, Kazemzadeh Anari R, Rostamani M, Javadi A, et al. Phenotype and genetic determination of resistance to common disinfectants among biofilm-producing and non-producing *Pseudomonas aeruginosa* strains from clinical specimens in Iran. *BMC Microbiol*. 2022;**22**(1):124. [PubMed ID: 35525944]. [PubMed Central ID: PMC9078005]. <https://doi.org/10.1186/s12866-022-02524-y>.
4. Hilliam Y, Kaye S, Winstanley C. *Pseudomonas aeruginosa* and microbial keratitis. *J Med Microbiol*. 2020;**69**(1):3-13. [PubMed ID: 31750813]. <https://doi.org/10.1099/jmm.0.001110>.
5. Yang LF, Ling M, Kacherovsky N, Pun SH. Aptamers 101: aptamer discovery and in vitro applications in biosensors and separations. *Chem Sci*. 2023;**14**(19):4961-78. [PubMed ID: 37206388]. [PubMed Central ID: PMC10189874]. <https://doi.org/10.1039/d3sc00439b>.
6. Webster TA, Sismaet HJ, Conte JL, Chan IP, Goluch ED. Electrochemical detection of *Pseudomonas aeruginosa* in human fluid samples via pyocyanin. *Biosens Bioelectron*. 2014;**60**:265-70. [PubMed ID: 24813917]. <https://doi.org/10.1016/j.bios.2014.04.028>.
7. Labib M, Zamay AS, Kolovskaya OS, Reshetneva IT, Zamay GS, Kibbee RJ, et al. Aptamer-based impedimetric sensor for bacterial typing. *Anal Chem*. 2012;**84**(19):8114-7. [PubMed ID: 22971146]. <https://doi.org/10.1021/ac302217u>.
8. Wu Z, He D, Cui B, Jin Z. A bimodal (SERS and colorimetric) aptasensor for the detection of *Pseudomonas aeruginosa*. *Mikrochim Acta*. 2018;**185**(11):528. [PubMed ID: 30382404]. <https://doi.org/10.1007/s00604-018-3073-2>.
9. Castro ER, Manz A. Present state of microchip electrophoresis: state of the art and routine applications. *J Chromatogr A*. 2015;**1382**:66-85. [PubMed ID: 25529267]. <https://doi.org/10.1016/j.chroma.2014.11.034>.
10. Zhong Z, Gao R, Chen Q, Jia L. Dual-aptamers labeled polydopamine-polyethyleneimine copolymer dots assisted engineering a fluorescence biosensor for sensitive detection of *Pseudomonas aeruginosa* in food samples. *Spectrochim Acta A Mol Biomol Spectrosc*. 2020;**224**:117417. [PubMed ID: 31362188]. <https://doi.org/10.1016/j.saa.2019.117417>.
11. Levi K, Smedley J, Towner KJ. Evaluation of a real-time PCR hybridization assay for rapid detection of *Legionella pneumophila* in hospital and environmental water samples. *Clin Microbiol Infect*. 2003;**9**(7):754-8. [PubMed ID: 12925125]. <https://doi.org/10.1046/j.1469-0691.2003.00666.x>.
12. Wu W, Yu C, Wang Q, Zhao F, He H, Liu C, et al. Research advances of DNA aptasensors for foodborne pathogen detection. *Crit Rev Food Sci Nutr*. 2020;**60**(14):2353-68. [PubMed ID: 31298036]. <https://doi.org/10.1080/10408398.2019.1636763>.