



# The Association of Apolipoprotein A1 Gene Polymorphisms with Acute Lymphocytic Leukemia in Tehran Province

Mostafa Bahrebar<sup>1\*</sup> and Tahmineh Chalak<sup>2</sup>

<sup>1</sup>Department of Biology, Faculty of Science, Dehdasht Branch, Islamic Azad University, Dehdasht, Iran

<sup>2</sup>Department of Biology, Faculty of Sciences, Gachsaran Branch, Islamic Azad University, Gachsaran, Iran

\*Corresponding author: Assistant Professor, Department of Biology, Faculty of Science, Dehdasht Branch, Islamic Azad University, Dehdasht, Iran. Tel: +98-9171484517, Email: m.bahrebar448@gmail.com

Received 2020 June 17; Revised 2020 August 19; Accepted 2020 August 20.

## Abstract

**Background:** Acute lymphocytic leukemia (ALL) is a cancer of the lymphoid line of blood cells. It is characterized by the development of large numbers of immature lymphocytes. It is one of the most common cancers among children. Molecular markers can play an important role in determining the nature of leukemia-related tumors and can be used as a diagnostic adjuvant along with precise pathological methods. Some research has shown that apolipoprotein A1 gene polymorphisms were associated with the formation of various types of diseases.

**Objectives:** This study aimed to evaluate the association of ApoA1 gene insertion/deletion polymorphisms with ALL disease.

**Methods:** The salting-out method was used for DNA extraction from the blood samples. Questionnaires were prepared with the approval of a demographic data expert. The genotype distribution and allele frequencies for the ApoA1 gene insertion/deletion polymorphisms were determined by Gap-polymerase chain reaction (Gap-PCR) to compare patient's cases with ALL disease and healthy subjects. This case-control study was performed by odds ratio (OR, with a confidence interval of 0.95) to reveal the association of these polymorphisms with ALL disease.

**Results:** In this study, two ApoA1 gene polymorphisms, including II and DD genotypes, were observed, which II genotype had the highest frequency in both groups of healthy subjects and patients. However, no significant differences were found between the two groups ( $P > 0.05$ ). ALL and both genotypes had a higher frequency in males than females. Despite the high frequency of both genotypes in males, there were no significant differences between healthy subjects and patients in terms of age and sex ( $P > 0.05$ ).

**Conclusions:** Based on the results of this study, it can be concluded that ApoA1 gene polymorphisms were not associated with ALL disease. It was not effective in the formation or exacerbation of it in this population. Also, it could not be identified as a prognostic marker. It should be noted that these findings are the first report of the degree of association of ApoA1 polymorphisms with ALL.

**Keywords:** Apolipoprotein, Lymphocytic, Leukemia, Polymorphism, ApoA1 Gene

## 1. Background

Cancer is the third leading cause of death in Iran after cardiovascular disease and accidents and is also the second leading cause of death worldwide (1, 2). Acute lymphocytic leukemia (ALL) that is known as acute lymphoblastic leukemia is the most common type of leukemia, which is more common among people under the age of 15 years in Iran (3). In Iran, as a developing country, cancer is on the rise. Leukemia is about 8% of all human cancers (4). Leukemia, based on the rate of progression of disease to acute and chronic and also based on the involved cells to lymphoid and myeloid, is divided into two types (5). ALL is a cancer of the lymphoid line of blood cells. It is characterized by the development of large numbers of immature lymphocytes. The accumulation of these cells in the

lymphatic tissues causes these organs to enlarge. Also, a significant increase in lymphocytes leads to a decrease in the number of other blood cells such as red blood cells and platelets, and this imbalance in blood cells leads to anemia, bleeding, and non-coagulation. This type of leukemia progresses rapidly and is typically fatal within weeks or months if left untreated (6).

Various studies have shown that the time between the appearance of the ALL disease and its spreading is very short, and the most common type of leukemia is in children, which is often seen in children between the ages of 2 and 6. ALL can also occur in adults, though the chance of a cure is greatly reduced, and people over the age of 75 are more vulnerable to the disease (7).

Some genes or proteins as tumor markers are some-

how involved in the progression of cancers. Molecular markers can play an important role in determining the nature of leukemia-related tumors and can be used as a diagnostic adjuvant along with precise pathological methods. These markers must be simple and fast in terms of technique and interpretation of results and have high sensitivity (8).

Apolipoprotein A1 (ApoA1) is a protein and the major component of HDL particles that, in humans, is encoded by the ApoA1 gene. This gene is located on the 11th chromosome with its specific location, 11q23-q24. It contains four exons. Some studies elucidated that genetic changes in the ApoA1 gene led to triglycerides increasing and HDL decreasing (9-12). Numerous studies have shown that lower HDL and higher triglycerides are associated with cancers (13-17). Naik demonstrated that serum cholesterol, HDL, and LDL are significantly decreased and associated inversely with the incidence of cancer, whereas triglycerides were significantly elevated in cancer patients compared to the normal population (13), which is in agreement with the findings of Musolino et al. (14) and Fiorenza and colleagues (15). The relationship between hypocholesterolemia and the degree of maturation of leukemic blast cells in acute myeloid leukemia was demonstrated in the other studies (16). Also, Peterson et al. demonstrated that hypocholesterolemia in cancer patients may be caused by elevated LDL receptor activates in malignant cells. The inverse association between cancer and serum cholesterol may reflect a physiological response to early undiagnosed stages of cancer (17).

Two siblings with HDL deficiency and no plasma apolipoprotein A-I (Apo A-I) were found to be homozygous for a cytosine deletion in exon 3 of Apo A-I gene (c.85 del C, Q5FsX11). This mutation causes a frameshift leading to a premature stop codon and abolishes the synthesis of ApoA1. In two other unrelated subjects, HDL deficiency was due to heterozygosity for a nucleotide substitution in exon 4 of Apo A-I gene (c.494 T > G, L141R) (18). A family with hereditary non-neuropathic amyloidosis has reported that hereditary hepatic and systemic amyloidosis caused by a new deletion/insertion mutation in the ApoA1 gene in which 12 residues have been deleted and two residues inserted in the amyloidogenic NH<sub>2</sub>-terminal fragment. The 5' end of exon four of the ApoA1 gene reported as abnormal, thereby producing a PCR product of 361bp fragment as the deletion and 396bp as the insertion mutation (19).

## 2. Objectives

In the present study, the role of different modes of ApoA1 gene polymorphisms in patients with ALL, and the frequency of different alleles of insertion/deletion mutation of the ApoA1 gene was investigated.

## 3. Methods

The present study was a case-control (retrospective) study consisting of a case study and a control population. The 50 blood samples from patients with ALL and healthy subjects were taken. The patients were diagnosed by a physician specialized in diseases, and they were under treatment (chemotherapy of maintenance phase in Ali Asghar Hospital affiliated to Iran University of Medical Sciences and Imam Khomeini Hospital in Tehran). Among the healthy people who referred to the health centers in the west of Tehran affiliated to Iran University of Medical Sciences, the healthy subjects were selected through simple randomization. The study population was examined from March 2017 to January 2017, and written informed consent was obtained from all patients before the sampling. The required sample size was obtained to study 25 patients with ALL and 25 healthy subjects based on the sampling size of similar studies and limited access to patient samples (9, 20). The alpha level was set at 0.05, and the study strength was 80%.

DNA was extracted from blood samples by salting out method. A rapid, safe, and inexpensive method was developed to simplify the deproteinization procedure. This method involves salting out of the cellular proteins by dehydration and precipitation with a saturated NaCl solution according to the following procedure: A total of blood samples were collected and kept at 4°C. Then 1.5 mL of cell lysis buffer was added to 500  $\mu$ L of each blood sample. The cell lysates were digested overnight at 37°C with 0.2 mL of 10% SDS and 0.5 mL of a protease K solution. After digestion was complete, 1 mL of saturated NaCl (approximately 6M) was added to each tube and shaken vigorously for 15 seconds, followed by centrifugation at 12,000 rpm for 15 minutes. The precipitated protein pellet was left at the bottom of the tube, and the supernatant containing the DNA was transferred to another tube. Exactly, 2 volumes of absolute ethanol were added, and the tubes were inverted several times until DNA precipitated. The precipitated DNA strands were removed and transferred to a 1.5 mL microcentrifuge tube containing 100 - 200  $\mu$ L TE buffer. The DNA was allowed to dissolve for 2 hours at 37°C before quantitating. Qualitative analysis of the DNA extracted was determined with the help of Agarose gel 1.5% of electrophoresis, and its concentration was measured using Nanodrop (21, 22).

Genotype distribution and allele's frequencies were determined by the Gap-PCR method. Using a pair of primers presented in Table 1, which was designed and optimized with the help of Gene Runner software and the BLAST method of the National Website of the Biotechnology Information Center (<http://www.ncbi.nlm.nih.gov>). The Gap-PCR method is a technique used to detect large mutations.

The oligonucleotide primers used in this technique are designed to cover both sides of the removed area. If deleted, the two primers come close together, and DNA replicates. This method is considered a simple, fast, and inexpensive alternative compared to Southern blotting and fluorescent in situ hybridization (FISH) methods (23, 24).

The promoter area to the intron 2 of the 856 bp ApoA1 gene were amplified through Gap-PCR under the following conditions: 100 nanograms of genomic DNA were added to the reactive mixture containing Taq buffer (10 mM tris-chloride, 50 mM KCL, 0.1 triton X100, 25  $\mu$ L) (2x Master mix Red, Taq DNA Pol) and then 5 pM/ $\mu$ L was added from each of the primers and the final volume was increased to 50  $\mu$ L (20). PCR was performed in 35 cycles with optimal conditions according to Table 2.

Finally, the Gap-PCR product was detected based on electrophoresis on agarose gel and ethidium bromide staining against ultraviolet light.

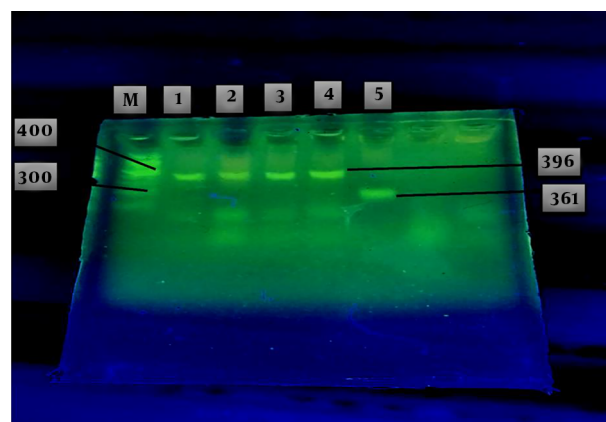
Moreover, SPSS software version 16 was used to record data and statistical analysis. A chi-square test was used to check the frequency of the genotype and its differences in the healthy subjects and patients. The confidence level in all experiments was 95%, and the level of significance was less than 0.05.

#### 4. Results

The results of agarose gel electrophoresis of the GAP-PCR products related to the insertion and deletion mutations of the ApoA1 gene in patients are shown in Figure 1. Two fragments were achieved, one of which was 361 bp related to the deletion mutation of the ApoA1 gene, called allele D, as a genotype of DD, and the other fragment, with 396 bp, was related to the insertion mutation of the ApoA1 gene, named I allele, as a genotype of II (Figure 1). Heterozygote state was not observed in these cases like other subjects.

The results of the genotype distribution among the patients showed that there were 21 homozygotes with genotype II and 4 homozygotes with genotype DD and nobody with heterozygote genotype DI. Among the healthy subjects, 22 homozygotes with genotype II and 3 homozygotes with genotype DD, also nobody with heterozygote genotype DI was found. Based on the relevant statistical analysis, despite the high frequencies of both genotypes in males than in females, there was no significant difference between genotype distribution of healthy subjects and patients ( $P = 0.516$ ) (Table 3).

The results of sex-based genotype distribution showed that ALL disease had a higher relative frequency percentage in males than females. However, there were no significant differences between healthy subjects and patients ( $P = 0.508$ ) (Table 4).



**Figure 1.** Agarose gel electrophoresis of the GAP-PCR products related to Insertion and Deletion mutation of the ApoA1 gene in the patients with ALL (lanes 1-5). A fragment with 396bp yielded in lanes 1-4 related to the insertion mutation as a genotype of II. The other fragment with 361 bp (lane 5), related to deletion mutation as a genotype of DD. Markers of known length were run in lane M.

The results of frequency subjects of the sample population based on age are presented in Table 5. The percentage of absolute frequency of 6-8 years was the lowest, and 8-10 years was the highest rate compared to the other subjects. The results of genotype frequency and its relationship with ALL based on age showed that there was no significant difference between healthy subjects and patients ( $P = 0.662$ ) (Table 5).

#### 5. Discussion

Studies on different modes of a genetic predisposition suggest that factors such as genetic polymorphisms may indicate individual differences in susceptibility, premature onset and progression of cancer, and even drug resistance. Acute lymphocytic leukemia (ALL) is more than forty cases per million children per year (5). Eighty percent of ALL cases occur in children and 20 percent in adults, and the disease is more common in males than in females that is in agreement with the findings of this study (4). The disease does not have a good prognosis in adults; despite significant advances in treatment, its mortality is more common in adults. In various studies, the rate of mortality has been stated to be 25% to 40% (7). According to research conducted in Iran, this type of leukemia accounts for the largest percentage of all cancers in children under 15 years of age (25). Previous studies have reported that ApoA1 is a potential marker for many types of cancer, including breast and pancreatic cancer (26, 27). Another study reported an association between HDL-C/ApoA1 plasma levels and the risk of developing a wide range of cancers (28). Sheikhha et al. (29) reported no significant relationship be-

**Table 1.** Sequence of the Primers Used in Gap-PCR

	Sequence (5' → 3')	Length	Tm	GC	Name
<b>Forward primer</b>	AGGACCTGCTGGGGACTAAAG	21	61.1	57.1	ApoA1
<b>Reverse primer</b>	TGAGAACTGCTGCCTCTGC	21	62.9	57.1	

**Table 2.** Protocol for Performing Gap-PCR

Number of Cycles/Steps	Steps of Gap-PCR	Temperature (°C)	Time
1/1	Primary denaturation	95	5 min
35/2	Denaturation	95	0.5 min
	Annealing	62	0.5 min
	Extension	72	1.5 min
1/3	Final extension	72	5 min

**Table 3.** Genotype Distribution of ApoA1 Gene Polymorphisms in the Healthy Subjects and Patients Based on Sex

Sex/Group	II Genotype	II Percent	DD Genotype	DD Percent	Total	Percent	P
<b>Male</b>							
Patients	17	34	3	6	20	40	0.516
Healthy	15	30	3	6	18	36	
Total	32	64	6	12	38	76	
<b>Female</b>							
Patients	4	8	1	2	5	10	0.516
Healthy	7	14	0	0	7	14	
Total	11	22	1	2	12	24	
<b>Total of male and female</b>	43	86	7	14	50	100	

**Table 4.** Frequency of Sex-Based ALL Distribution in the Healthy Subjects and Patients

Gender	Cumulative Frequency	Relative Frequency Percentage	Relative Frequency	Absolute Frequency	P
<b>Male</b>					
Patient	20	40	0.4	20	0.508
Healthy	38	36	0.36	18	
<b>Female</b>					
Patient	43	10	0.1	5	0.508
Healthy	50	14	0.14	7	
<b>Total</b>				50	

tween the polymorphism of the ApoA1 gene and the risk of diabetes that is consistent with the results of the present study.

Owing to the fact that in a few studies conducted in this regard (26-28), no significant differences were observed, and therefore, the results of this study are consistent with the results of similar studies.

While many studies have indicated that polymor-

phism of ApoA1 does not have a significant relationship with ALL disease in different communities, the opposite is also true in one study (30) that this observed inconsistency is due to differences in the related allele frequency connected to the genetic background of different populations.

Owing to the fact that the risk of ALL disease is influenced by a combination of genetic and environmental fac-

**Table 5.** Age-Based Genotypic Distribution of ApoA1 Gene Polymorphisms in the Healthy Subjects and ALL Patients

Age (Year)	Samples Mode	Absolute Frequency	Genotype II	Percent II	Genotype DD	Percent DD	P
6-8	7	Patient	3	6	0	0	0.662
		Healthy	4	8	0	0	
8-10	15	Patient	4	8	1	2	
		Healthy	9	18	1	2	
10-12	14	Patient	7	14	1	2	
		Healthy	4	8	2	4	
12-14	14	Patient	7	14	2	4	
		Healthy	5	10	0	0	
Total	25	Patient	21	42	4	8	
	25	Healthy	22	44	3	6	

tors, for this reason, different results have been obtained among the populations in some studies.

However, in order to check the serum level of ApoA1 protein, a specific kit was needed, and the preparation of this kit was not possible. Therefore, it was not possible for us to measure serum levels of it. Nevertheless, owing to the non-significant difference of this gene in the healthy subjects and patients, it does not seem necessary to measure the relevant protein.

The results of this study and other similar studies (26-28) showed that ApoA1 polymorphism does not play a role in ALL incidence in the community. Although the ApoA1 gene plays an important role in fat metabolism and lymphocyte metabolism, it does not play a significant role in the processes involved in the reproduction, hematopoiesis, and natural immune system (leukemia). One important reason for this could be the small sample size of the study population. Finally, it is suggested that in order to resolve the existing ambiguities, in future studies, sequencing should be performed in all areas of the ApoA1 gene, and the interventions of ApoA1 gene polymorphisms in a larger Iranian population should be investigated.

In this study, there was no significant difference between the healthy subjects and patients in terms of the frequency of alleles and genotypes, which was consistent with some studies (27, 29). One of the reasons that can be stated is that the genetic association is population-dependent (31). In this regard, the subjects studied are important reasons. Thus the subjects were randomly selected in this study, while matching has been done in many other studies so that individuals are matched in terms of various factors such as Body Mass Index (BMI), fat index, gender, and age characteristics.

### 5.1. Conclusion

Based on the result of this study, it can be concluded that ApoA1 gene polymorphisms were not associated with ALL and were not effective in the formation or exacerbation of it in this population. Also, it could not be identified as a prognostic marker.

It should be noted that these findings are the first report of the degree of association of ApoA1 gene insertion/deletion polymorphisms with ALL, and it is recommended that this study should be performed on more individuals.

### Acknowledgments

This study and its findings come from a master's thesis in Cellular Molecular Biology at the IAU Gachsaran branch. We extend our gratitude to all those who participated in this study.

### Footnotes

**Authors' Contribution:** Study concept and design: MB and TCh. Analysis and interpretation of data: MB and TCh. Drafting of the manuscript: MB and TCh. Critical revision of the manuscript for important intellectual content: MB and TCh.

**Conflict of Interests:** The authors declared no conflict of interest.

**Ethical Approval:** This article comes from a master's thesis of Cellular Molecular Biology. It has been registered with code 14930554962004 at the Research Committee of the Islamic Azad University Gachsaran Branch. All of the subjects in this study voluntarily filled out a written consent form to participate in this study.

**Funding/Support:** This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

**Informed Consent:** All the subjects in this study gave voluntary informed consent to participate in this research.

## References

- Diaz Mdel P, Osella AR, Aballay LR, Munoz SE, Lantieri MJ, Butinof M, et al. Cancer incidence pattern in Cordoba, Argentina. *Eur J Cancer Prev.* 2009;**18**(4):259–66. doi: [10.1097/CEJ.0b013e3283152030](https://doi.org/10.1097/CEJ.0b013e3283152030). [PubMed: [19404198](https://pubmed.ncbi.nlm.nih.gov/19404198/)].
- Mousavi SM, Gouya MM, Ramazani R, Davanlou M, Hajsadeghi N, Seddighi Z. Cancer incidence and mortality in Iran. *Ann Oncol.* 2009;**20**(3):556–63. doi: [10.1093/annonc/mdn642](https://doi.org/10.1093/annonc/mdn642). [PubMed: [19073863](https://pubmed.ncbi.nlm.nih.gov/19073863/)].
- Mousavi SM, Pourfeizi A, Dastgiri S. Childhood cancer in Iran. *J Pediatr Hematol Oncol.* 2010;**32**(5):376–82. doi: [10.1097/MPH.0b013e318e003f7](https://doi.org/10.1097/MPH.0b013e318e003f7). [PubMed: [20588194](https://pubmed.ncbi.nlm.nih.gov/20588194/)].
- Babaei M, Mousavi S, Malek M, Tosi G, Zolfaghari M, Danaei N, et al. [Cancer occurrence in Semnan Province, Iran: results of a population-based cancer registry]. *Asian Pacific Journal of Cancer Prevention.* 2005;159–64. Persian.
- Davis AS, Viera AJ, Mead MD. Leukemia: An overview for primary care. *American family physician.* 2014;**89**(9):731–8.
- Hoffbrand A. Non-Hodgkin's lymphoma. In: Hoffbrand AV, Moss PAH, Pettit JE, editors. *Essential Hematology.* Blackwell, Publisher, Massachusetts, USA; 2006.
- Moricke A, Zimmermann M, Reiter A, Henze G, Schrauder A, Gadner H, et al. Long-term results of five consecutive trials in childhood acute lymphoblastic leukemia performed by the ALL-BFM study group from 1981 to 2000. *Leukemia.* 2010;**24**(2):265–84. doi: [10.1038/leu.2009.257](https://doi.org/10.1038/leu.2009.257). [PubMed: [20010625](https://pubmed.ncbi.nlm.nih.gov/20010625/)].
- Rostami E, Mohamadi M, Bagheri S. [Investigation of MDM2 gene promoter polymorphism (SNP309) in patients with acute myeloid leukemia in Khuzestan province]. *Jundishapur Sci Med J.* 2018;**17**(2):187–94. Persian.
- Firoozrai M, Ehsani Zanoz AV, Hesabi B, Kheradmand Kia S. [Polymorphism of CIII and AI apolipoprotein genes in Iranian patient with coronary artery disease]. *Razi Journal of Medical Sciences.* 2003;**10**(34):247–55.
- Jeenah M, Kessling A, Miller N, Humphries S. G to A substitution in the promoter region of the apolipoprotein AI gene is associated with elevated serum apolipoprotein AI and high density lipoprotein cholesterol concentrations. *Mol Biol Med.* 1990;**7**(3):233–41. [PubMed: [1977072](https://pubmed.ncbi.nlm.nih.gov/1977072/)].
- Halley P, Kadakkuzha BM, Faghihi MA, Magistri M, Zeier Z, Khorkova O, et al. Regulation of the apolipoprotein gene cluster by a long non-coding RNA. *Cell Rep.* 2014;**6**(1):222–30. doi: [10.1016/j.celrep.2013.12.015](https://doi.org/10.1016/j.celrep.2013.12.015). [PubMed: [24388749](https://pubmed.ncbi.nlm.nih.gov/24388749/)]. [PubMed Central: [PMC3924898](https://pubmed.ncbi.nlm.nih.gov/PMC3924898/)].
- van der Vorst EPC. High-Density Lipoproteins and Apolipoprotein AI. *Subcell Biochem.* 2020;**94**:399–420. doi: [10.3389/fphar.2016.00323](https://doi.org/10.3389/fphar.2016.00323).
- Naik PP, Ghadge MS, Raste AS. Lipid profile in leukemia and Hodgkin's disease. *Indian J Clin Biochem.* 2006;**21**(2):100–2. doi: [10.1007/BF02912921](https://doi.org/10.1007/BF02912921). [PubMed: [23105623](https://pubmed.ncbi.nlm.nih.gov/23105623/)]. [PubMed Central: [PMC3453994](https://pubmed.ncbi.nlm.nih.gov/PMC3453994/)].
- Musulino C, Calabro L, Bellomo G, Cincotta M, Di VG, Pezzano C, et al. Lipid profile in hematologic neoplasms. *Recenti Prog Med.* 2002;**93**(5):298–301. doi: [10.1007/BF02912921](https://doi.org/10.1007/BF02912921).
- Fiorenza AM, Branchi A, Sommariva D. Serum lipoprotein profile in patients with cancer. A comparison with non-cancer subjects. *Int J Clin Lab Res.* 2000;**30**(3):141–5. doi: [10.1007/s005990070013](https://doi.org/10.1007/s005990070013). [PubMed: [11196072](https://pubmed.ncbi.nlm.nih.gov/11196072/)].
- Zyada LE, Hassan HT, Rees JK, Ragab MH. The relation between hypcholesterolemia and degree of maturation in acute myeloid leukemia. *Hematol Oncol.* 1990;**8**(1):65–9. doi: [10.1002/hon.2900080109](https://doi.org/10.1002/hon.2900080109). [PubMed: [2298420](https://pubmed.ncbi.nlm.nih.gov/2298420/)].
- Peterson C, Vitols S, Rudling M, Blomgren H, Edsmyr F, Skoog L. Hypocholesterolemia in cancer patients may be caused by elevated LDL receptor activities in malignant cells. *Medical oncology and tumor pharmacotherapy.* 1985;**2**(3):143.
- Pisciotta L, Miccoli R, Cantafora A, Calabresi L, Tarugi P, Alessandrini P, et al. Recurrent mutations of the apolipoprotein A-I gene in three kindreds with severe HDL deficiency. *Atherosclerosis.* 2003;**167**(2):335–45. doi: [10.1016/s0021-9150\(03\)00020-0](https://doi.org/10.1016/s0021-9150(03)00020-0).
- Booth DR, Tan SY, Booth SE, Tennent GA, Hutchinson WL, Hsuan JJ, et al. Hereditary hepatic and systemic amyloidosis caused by a new deletion/insertion mutation in the apolipoprotein AI gene. *J Clin Invest.* 1996;**97**(12):2714–21. doi: [10.1172/JCI118725](https://doi.org/10.1172/JCI118725). [PubMed: [8675681](https://pubmed.ncbi.nlm.nih.gov/8675681/)]. [PubMed Central: [PMC507363](https://pubmed.ncbi.nlm.nih.gov/PMC507363/)].
- Hamidi L, Khatamsaz S, Mokhtari MJ, Babaei Beigi MA. The Association of Apolipoprotein AI Polymorphisms with Diabetes and Hypertension in Patients with Coronary Artery Disease in Fars Province. *Journal of Arak University of Medical Sciences.* 2017;**20**(8):20–8.
- Miller SA, Dykes DD, Polesky HF. A simple salting out procedure for extracting DNA from human nucleated cells. *Nucleic Acids Res.* 1988;**16**(3):1215. doi: [10.1093/nar/16.3.1215](https://doi.org/10.1093/nar/16.3.1215). [PubMed: [3344216](https://pubmed.ncbi.nlm.nih.gov/3344216/)]. [PubMed Central: [PMC334765](https://pubmed.ncbi.nlm.nih.gov/PMC334765/)].
- Gaib JN. Simple salting-out method for genomic DNA extraction from whole blood. *Tikrit J Pure Sci.* 2011;**16**(2):9–11.
- Hartevelde CL, Voskamp A, Phylipsen M, Akkermans N, den Dunnen JT, White SJ, et al. Nine unknown rearrangements in 16p13.3 and 11p15.4 causing alpha- and beta-thalassaemia characterised by high resolution multiplex ligation-dependent probe amplification. *J Med Genet.* 2005;**42**(12):922–31. doi: [10.1136/jmg.2005.033597](https://doi.org/10.1136/jmg.2005.033597). [PubMed: [15894596](https://pubmed.ncbi.nlm.nih.gov/15894596/)]. [PubMed Central: [PMC1735959](https://pubmed.ncbi.nlm.nih.gov/PMC1735959/)].
- Craig JE, Barnetson RA, Prior J, Raven JL, Thein SL. Rapid detection of deletions causing delta beta thalassemia and hereditary persistence of fetal hemoglobin by enzymatic amplification. *Blood.* 1994;**83**(6):1673–82. [PubMed: [7510147](https://pubmed.ncbi.nlm.nih.gov/7510147/)].
- Pui CH, Relling MV, Downing JR. Acute lymphoblastic leukemia. *N Engl J Med.* 2004;**350**(15):1535–48. doi: [10.1056/NEJMra023001](https://doi.org/10.1056/NEJMra023001). [PubMed: [15071128](https://pubmed.ncbi.nlm.nih.gov/15071128/)].
- Hamrita B, Ben Nasr H, Gabbouj S, Bouaouina N, Chouchane L, Chahed K. Apolipoprotein AI -75 G/A and +83 C/T polymorphisms: susceptibility and prognostic implications in breast cancer. *Mol Biol Rep.* 2011;**38**(3):1637–43. doi: [10.1007/s11033-010-0274-0](https://doi.org/10.1007/s11033-010-0274-0). [PubMed: [20842444](https://pubmed.ncbi.nlm.nih.gov/20842444/)].
- Moore LE, Fung ET, McGuire M, Rabkin CC, Molinaro A, Wang Z, et al. Evaluation of apolipoprotein AI and posttranslationally modified forms of transthyretin as biomarkers for ovarian cancer detection in an independent study population. *Cancer Epidemiol Biomarkers Prev.* 2006;**15**(9):1641–6. doi: [10.1158/1055-9965.EPI-05-0980](https://doi.org/10.1158/1055-9965.EPI-05-0980). [PubMed: [16985025](https://pubmed.ncbi.nlm.nih.gov/16985025/)].
- Ordovas JM, Corella D, Cupples LA, Demissie S, Kelleher A, Coltell O, et al. Polyunsaturated fatty acids modulate the effects of the APOAI G-A polymorphism on HDL-cholesterol concentrations in a sex-specific manner: the Framingham Study. *Am J Clin Nutr.* 2002;**75**(1):38–46. doi: [10.1093/ajcn/75.1.38](https://doi.org/10.1093/ajcn/75.1.38). [PubMed: [11756058](https://pubmed.ncbi.nlm.nih.gov/11756058/)].
- Sheikhha MH, Afkhami-Ardekani M, Mirjalili SMR, Dehghani SMR, Ghadimi HR. Investigating the frequency of MSPI polymorphism of APOAI gene in type II diabetic patients and comparing it with this frequency in nondiabetics. *Genetics in the 3rd Millennium.* 2013;**11**(2):3078–83.
- Cine N, Baykal AT, Sunnetci D, Canturk Z, Serhatli M, Savli H. Identification of ApoAI, HPX and POTE genes by omic analysis in breast cancer. *Oncol Rep.* 2014;**32**(3):1078–86. doi: [10.3892/or.2014.3277](https://doi.org/10.3892/or.2014.3277). [PubMed: [24969553](https://pubmed.ncbi.nlm.nih.gov/24969553/)].
- Deschler B, Lübbert M. Acute Myeloid Leukemia: Epidemiology and Etiology. *Acute Leukemias.* Springer; 2008. p. 47–56. doi: [10.1007/978-3-540-72304-2\\_3](https://doi.org/10.1007/978-3-540-72304-2_3).